PREPARATION OF ALLYL2,3,4-TRI-O-ALLYL-D-GLUCOPYRANURONATE AND OF 1-O-ACYL-D-GLUCOPYRANURONIC ACIDS: ATTEMPTED SYNTHESIS OF 1-O-BILIRUBIN-D-GLUCURONIC ACIDS

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ABSTRACT

Allyl 2,3,4-tri-O-allyl-D-glucopyranuronate (7) has been prepared (overall yield, 37%) from methyl 1,2,3,4-tetra-O-acetyl- α - or - β -D-glucopyranuronate via a six-step reaction procedure. 1-O-Acetyl-, 1-O-methacryloyl-, and 1-O-phenylacetyl- $\alpha\beta$ -D-glucopyranuronic acid are produced by esterification of 7 and removal of the allyl groups with palladium-on-carbon in an acidic medium. The reaction of 7 with the di-imidazole derivative of bilirubin yields the allyl-protected mono- and di-D-glucopyranosyluronic acid derivatives of bilirubin. Neither the bilirubin nor the corresponding biliverdin derivatives resisted the conditions required for de-allylation.

INTRODUCTION

Bilirubin, a tetrapyrrolic pigment produced by biological breakdown of haem, is excreted into bile as mono- and di-D-glucosyluronic acid derivatives having one or two propionic acid side-chains esterified with β -D-glucopyranuronic acid¹⁻³.

Esterification of the di-imidazole derivative of bilirubin with D-glucuronic acid yielded a mixture of 2-, 3-, and 4-O-acyl-D-glucosyluronic acid derivatives⁴. Clearly, HO-2,3,4 and the carboxyl group of D-glucuronic acid need to be protected if 1-esters are to be obtained. However, such protecting groups must be removable under very mild conditions, since migration of the bilirubin 1-O-acyl group occurs¹ at pH 7–9. Moreover, the asymmetric bilirubin-IX α tetrapyrrole readily undergoes⁵ dipyrrole exchange, yielding a 2:1:1 mixture of bilirubin-IX α and the symmetric bilirubin-III α and -XIII α . This reaction is catalysed by mineral acid for bilirubin and even occurs in neutral medium for the 1-O-bilirubin-D-glucuronic acids. Chain reactions are probably involved, with the formation of dipyrrylmethyl cations or radicals as the initiating species.

Although several 1-O-acyl-D-glucopyranuronic acids were prepared⁶ from benzyl 2,3,4-tri-O-benzyl-D-glucopyranuronate^{7,8}, this method is not suitable for preparation of 1-O-bilirubin-D-glucuronic acids, since hydrogenolysis of benzyl groups also results in reduction of vinyl substituents. A possible solution of these

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problems was indicated by the preparation of benzyl 2,3,4-tri-O-(chloroacetyl)- β -D-glucopyranosiduronic acid⁹.

The preparation is now reported of allyl 2,3,4-tri-O-allyl-D-glucopyranuronate (7), a compound which appears to be useful for the synthesis of 1-O-acyl-D-glucopyranuronic acids.

RESULTS AND DISCUSSION

Methyl 1,2,3,4-tetra-O-acetyl- α - or - β -D-glucopyranuronate (1), which is readily prepared from D-glucurono-6,3-lactone, was converted¹⁰ into the α -glycosyl bromide 2. Hydrolysis of 2 in the presence of silver carbonate gave the 1-hydroxy derivative 3 which, with ethyl vinyl ether, gave the acetal 4. Saponification of the ester groups in 4, followed by allylation of the hydroxyl and carboxyl groups, and liberation of HO-1 afforded 7, in an overall yield of 37%. Benzylation of 5 should give benzyl 2,3,4-tri-O-benzyl-D-glucopyranuronate, a compound that was recently prepared⁸ by an eight-step procedure starting from methyl β -D-glucopyranoside (overall yield, ~4.3%).



Esterification of 7 variously with acetic anhydride, methacryloyl chloride, and 1-phenylacetylimidazole gave $\alpha\beta$ -mixtures which were characterised by g.l.c.-m.s.

Allyl ethers are cleaved¹¹ by Rh(I)-catalysed isomerisation to 1-propenyl ethers and subsequent acid hydrolysis at pH 2. Apparently, in the first step, an equilibrium mixture is formed containing ~95% of the 1-propenyl ether^{11,12}. Cleavage of the four allyl groups in 1-O-acyl esters of 7 was effected in the presence of dilute hydrogen chloride by using a palladium-on-carbon catalyst¹³. The 1-O-acyl-D-glucopyranuronic acids were shown to be the major products by t.l.c. and m.s.

Esterification of 7 with the di-imidazole derivative of bilirubin⁴ yielded the mono- and mainly the di-D-glucosyluronic acid derivatives. Subsequent oxidation with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone gave the corresponding biliverdin derivatives. The allyl-protected 1-O-bilirubin-D-glucuronic acids were characterised further as the dipyrrolic azopigments, prepared by using a diazonium reagent derived from ethyl anthranilate¹⁴. This reagent cleaves the asymmetrical bilirubin tetrapyrrole into dipyrrolic azopigments having either an *endo*- or an *exo*-vinyl substituent¹⁵⁻¹⁸. Four azopigment isomers, corresponding to the anomeric pairs of the *exo*- and

endo-vinyl compounds, were isolated by t.l.c. The endo-vinyl isomers exhibited intense molecular ions (M⁺ at m/z 798, 20% relative abundance), whereas the less-stable, exo-vinyl isomers gave low-intensity molecular ions (M⁺ at m/z 798, 0.2%). In the n.m.r. spectra of the endo-vinyl isomers, the signals for the pyrrolinone and pyrrole methyl groups coincided (δ 2.12), whereas they occurred at δ 2.19 and 2.11, respectively, for the corresponding methyl groups of the exo-vinyl isomers¹⁶⁻¹⁸. The α and β anomers could not be assigned, because of overlap of the signals for the anomeric and various vinylic protons.

Neither the bilirubin nor the biliverdin allyl-protected D-glucosyluronic acids resisted the conditions required for removal of the allyl groups. The solutions were decolorised gradually before all protecting groups had been removed. Degradation of the chromophoric systems and/or adsorption of the pigments to carbon could have occurred.

EXPERIMENTAL

General. — Column chromatography was performed on Merck silica gel (0.06-0.2 mm). T.l.c. was conducted on pre-coated silica gel plates (Merck aluminum sheets), and detection was effected by spraying with a 3:1 mixture of 0.2% naphthalene-1,3-diol in ethanol and 20% sulphuric acid, followed by heating at 120°. G.l.c. was performed with 3% of OV-101 (glass column, 2 m × 6.3 mm) on Gaschrom Q, with helium as the carrier gas (50 ml/min) and a 1:1 outlet split to the flame-ionisation detector and mass spectrometer. Mass spectra were recorded by using either the direct insertion probe of an AEI MS902 S mass spectrometer or a g.l.c.-m.s. combination (glass jet separator).

N.m.r. spectra (CDCl₃, internal Me₄Si) were recorded with a Jeol 100-MHz or a Varian XL 100 spectrometer.

Allyl 2,3,4-tri-O-allyl-D-glucopyranuronate (7). — Compound 1 (5 g) was converted¹⁰ into the glycosyl bromide 2, to a solution of which in acetone (50 ml) were added water (0.35 ml) and silver carbonate (6 g). The mixture was stirred for 24 h at 22°, and t.l.c. (9:1, chloroform-acetonitrile) then showed quantitative conversion of 2 ($R_{\rm F} \sim 0.7$) into 3 ($R_{\rm F} \sim 0.3$). The mixture was filtered, and concentrated *in vacuo* to yield crude, oily 3.

To a solution of 3 in tetrahydrofuran (30 ml) at 0° were added ethyl vinyl ether (10 ml) and trifluoroacetic acid (1 ml). The formation of 4 was incomplete after 1 day at 0° and nearly complete after 2 more days at 22°, as shown by g.l.c. and t.l.c. of the neutralised reaction mixture. Dichloromethane (50 ml), triethylamine (5 ml), and ice-cold water (50 ml) were added, and the organic layer was washed four times with water, dried over molecular sieves, and concentrated, to yield crude 4. T.l.c. (15:1 chloroform-acetonitrile) showed two closely moving spots with $R_F \sim 0.7$ (anomers or diastereoisomers of 4). G.l.c.-m.s. (180°, isothermal) of an acetylated (2:1 pyridine-acetic anhydride) sample of 4 showed 98–99% conversion of 3 into 4 (1-2% of unreacted 3 was detected as 1). The retention time of 4 relative to that of 1 was 1.55.

The mass spectrum of 4 (mol. wt. 406) showed peaks at m/z 347, 304, and 317, corresponding to losses of CO₂CH₃, HOAc + CH₂CO, and OCHMeOEt, respectively,

Hydrolysis of 4 with methanol-triethylamine-water (2:1:1, 60 ml), first at 0° for 3 h and then at 22° for 2 days, gave a major product with $R_F \sim 0.3$ (t.l.c., 9:2 1-propanol-water). Concentration of the hydrolysate and drying of the residue *in vacuo* yielded the crude triethylammonium salt of 5, $\lceil \alpha \rceil_D + 38^\circ$ (c 1, water).

A solution of the salt in dry N,N-dimethylformamide (30 ml) was stirred at 22° under nitrogen with sodium hydride (3 g) added in portions. Allyl bromide (20 g) was added dropwise during ~30 min and the reaction was monitored by g.l.c.-m.s. $(150^{\circ} \rightarrow, 4^{\circ}/\text{min})$. Samples of the reaction mixture (0.2 ml) were dissolved in dichloromethane (1 ml), internal standard (0.2 ml of a 1% solution of 1) was added, and the solution was washed with water. An optimum yield of the allyl ether 6 (eluted at 188°) was observed after a reaction time of 2.5-3 h. Minor side-products were eluted at 180° (probably the methyl ester of 6) and 183° (fully allylated glucuronic acid). Dichloromethane (50 ml) was then added to the reaction mixture, and the organic layer was washed four times with water and concentrated, to give crude 6, $[\alpha]_D + 50^{\circ}$ (c 1, chloroform).

A solution of crude 6 in acetic acid (30 ml) and 0.1M hydrochloric acid (10 ml) was kept at 22° for 2 h and then at 90° for 30 min. T.I.c. (9:1 chloroform-acetonitrile) showed quantitative conversion of 6 ($R_{\rm F} \sim 0.6$) into 7 ($R_{\rm F} \sim 0.3$). The reaction mixture was cooled, diluted with water, and extracted with chloroform. Column chromatography (200 g of silica gel) with 15:1 chloroform-acetonitrile yielded 7 (1.75 g) as a colorless liquid, $[\alpha]_{\rm D} + 42^{\circ}$ (c 1, chloroform). The purity of 7 was checked by t.l.c., and by g.l.c. of the α,β -acetate (see below).

The mass spectrum of 7 (mol. wt. 354) showed peaks at m/z 336, 313, 295, 251, and 239, corresponding to losses of H₂O, allyl, H₂O + allyl, H₂O + allyl-CO₂, and allyl-OH + allyl-O, respectively. Intense peaks were observed at m/z 153 (allyl-O⁺=CH-CH=CH-O-allyl) and 41 (allyl⁺). The n.m.r. spectrum showed signals at δ 3.20-4.10 (m, 3 H, H-2,3,4), 4.10-4.40 (m, 6 H, CH₂-O ether groups), 4.43 (d, $J_{4,5}$ 9 Hz, H-5), 4.68 (d, 2 H, J 6 Hz, CH₂-O ester group), 4.80 (d, 0.2 H, $J_{1,2}$ 7 Hz, H-1 β), 5.00-5.50 (m, 8 H, CH₂=C), and 5.60-6.20 (m, 4 H, CH=C).

Anal. Calc. for C₁₈H₂₆O₇: C, 61.0; H, 7.4. Found: C, 60.7; H, 7.4.

Allyl 1-O-acyl-2,3,4-tri-O-allyl-D-glucopyranuronates. — The 1-O-acetyl derivatives were prepared by the reaction of 7 with an excess of pyridine-acetic anhydride (2:1) at 22° for 2 h, followed by evaporation of reagents *in vacuo* and column chromatography of the residue (chloroform-acetonitrile, 49:1). The product had $\lceil \alpha \rceil_{\rm P} + 53^{\circ}$ (c 1, chloroform).

Anal. Calc. for C₂₀H₂₈O₈: C, 60.6; H, 7.1. Found: C, 60.4; H, 7.0.

The 1-O-methacryloyl derivatives were prepared by dissolving 7 (5 mg) in dichloromethane (2 ml) and pyridine (0.2 ml) and treatment with methacryloyl chloride (0.1 ml) at 0° for 3 h. The solution was washed thoroughly (aqueous sodium hydrogencarbonate, 0.3M hydrochloric acid, and water) and concentrated *in vacuo*.

Column chromatography (chloroform-acetonitrile, 49:1) yielded an $\alpha\beta$ -mixture, $[\alpha]_{\rm D}$ +49° (c 1, chloroform).

Anal. Calc. for C₂₂H₃₀O₈: C, 62.55; H, 7.2. Found: C, 62.4; H, 7.1.

The 1-O-phenylacetyl derivatives were prepared by the reaction of 7 (50 mg) with 1-phenylacetylimidazole (100 mg) in acetonitrile (2 ml) in the presence of triethylamine (0.2 ml) at 22° for 24 h. Alternatively, a solution of 7 and 1-phenylacetylimidazole in chloroform was concentrated *in vacuo* and the residue was heated under argon at 100° for 30 min. The 1-phenylacetylimidazole reagent was prepared by the reaction of phenylacetic acid (70 mg) with carbonyldi-imidazole (100 mg) in ethanol-free chloroform (10 ml) at 22° for 1 h. The chloroform solution was washed four times with water and dried with molecular sieves. Column chromatography of the product prepared at 100° gave an $\alpha\beta$ -mixture, $[\alpha]_D + 35°$ (c 1, chloroform).

Anal. Calc. for C₂₆H₃₂O₈: C, 66.1; H, 6.8. Found: C, 66.1; H, 6.9.

G.l.c.-m.s. of the esters generally revealed a 2:1 $\alpha\beta$ -mixture. For the 1-Ophenylacetyl ester prepared at 100°, a 1:1 $\alpha\beta$ -mixture was obtained. G.l.c. was performed from 150° at 4°/min. The elution temperatures were 175 and 177° (α - and β -1-O-acetyl esters, respectively), 180 and 184° (α - and β -1-O-methacryloyl esters), and 209 and 216° (α - and β -1-O-phenylacetyl esters). The mass spectra of the 1-O-acyl esters were very similar to the spectrum for 7. A series of weak-intensity peaks in the higher mass region were due to the molecular ions and associated fragment ions corresponding to loss of allyl, allyl-O, allyl-OH, allyl-O + allene, allyl-OH + allene, and allyl-CO₂.

The n.m.r. spectrum of the 1-O-phenylacetyl ester showed signals at δ 3.2–3.8 (m, 3 H, H-2,3,4), 3.70 and 3.72 (2 H, Ph-CH₂ for α and β anomers), 3.8–4.4 (m, 6 H, CH₂-O ether), 4.28 (d, $J_{4,5}$ 7 Hz, H-5), 4.66 (d, J 6 Hz, CH₂-O ester), 5.0–5.48 (m, 8 H, CH₂=C), 5.55 (d, $J_{1,2}$ 7 Hz, H-1 β), 5.6–6.2 (m, 4 H, CH=C), 6.30 (d, $J_{1,2}$ 3 Hz, H-1 α), and 7.28 (s, 5 H, Ph).

1-O-Acyl-D-glucopyranuronic acids. — A solution of the allyl 1-O-acyl-2,3,4tri-O-allyl-p-glucopyranuronate (20 mg) in methanol (2 ml) and 0.1M hydrochloric acid (0.5 ml) was stirred under argon with 10% palladium-on-carbon (40 mg). The reaction was monitored by t.l.c. (9:2 1-propanol-water) and usually was complete within 48 h (sometimes catalyst poisoning¹³ was observed); R_F values: ~0.35, 0.40, and 0.45 for the 1-O-acetyl, 1-O-methacryloyl, and 1-O-phenylacetyl-D-glucopyranuronic acids, respectively. Some less polar impurities were also observed. The reaction mixtures were filtered, and concentrated in vacuo, and the residues {1-Oacetyl, $[\alpha]_{\rm D}$ + 30° (c 1, water); 1-O-methacryloyl, $[\alpha]_{\rm D}$ + 50° (c 1, water); and 1-Ophenylacetyl, $\lceil \alpha \rceil_{\rm D}$ +46° (c 1, water) were treated with N,O-bis(trimethylsilyl)trifluoroacetamide (50 μ l) and pyridine (100 μ l). G.l.c.-m.s. (from 150°, 4°/min) of the silylated 1-O-acetyl- and 1-O-methacryloyl-D-glucopyranuronic acids revealed 2:1 $\alpha\beta$ -mixtures; elution temperatures were 176.5° and 178° for the α - and β -1-Oacetyl-D-glucopyranuronic acids, and 179° and 180° for the α - and β -1-O-methacryloyl-D-glucopyranuronic acids. The 1-O-phenylacetyl-D-glucopyranuronic acids were not eluted from the column, and were analysed directly by mass spectrometry.

The mass spectra of the α and β anomers of the silvlated 1-O-acyl-D-glucopyranuronic acids showed weak-intensity molecular ions at m/z 524, 550, and 600 for the 1-Oacetyl, 1-O-methacryloyl, and 1-O-phenylacetyl esters, respectively. Characteristic ions were due to loss of methyl, methyl + Me₃SiOH, RCO₂ (m/z 465), RCO₂H (m/z 464), RCO₂H + Me (m/z 449), and RCO₂ + Me₃SiOH (m/z 375). Commonly observed fragment ions were found at m/z 305, 217, 204, 147, and 73.

Bilirubin and biliverdin 1-O-acyl derivatives of 7. — The di-imidazole derivative of bilirubin was prepared⁴ from bilirubin (100 mg) and carbonyldi-imidazole (200 mg) in dry N,N-dimethylformamide (5 ml). The solution was stirred at 22° for 24 h in the dark and the excess of reagent was decomposed with water (20 μ l). After 15 min, a solution of 7 (200 mg) in dichloromethane (1 ml) was added and the solution was concentrated *in vacuo* at 50°. The residue was heated under argon at 85° in the dark for 40 min, cooled, and dissolved in dichloromethane. T.l.c. (97:2:1 chloroformmethanol-acetic acid) showed the absence of bilirubin ($R_F \sim 0.7$), and formation of the di-D-glucosyluronic acid derivative ($R_F \sim 0.6$).

Allyl-protected 1-O-bilirubin-D-glucuronic acids were oxidised to the corresponding biliverdin compounds by shaking a dichloromethane solution (5 ml, containing the equivalent of 10 mg of bilirubin) with a fresh solution of 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (10 mg) in water (5 ml) for \sim 3 min. The blue solution was washed thoroughly with water. T.l.c. (9:1 chloroform-methanol) showed the allyl-protected biliverdin di-D-glucosyluronic acid derivative as the major compound (R_F 0.77) and the mono-D-glucosyluronic acid derivative as the minor compound (R_F 0.45). Free biliverdin (R_F 0.03) was not observed.

Ethyl anthranilate azopigments were prepared¹⁴ from the bilirubin derivatives by using N,N-dimethylformamide instead of ethanol as an accelerator of the diazonium reaction (to prevent formation of ethyl ester from possibly unreacted bilirubinimidazole). After reaction at pH 2.7 for 30 min, water was added and the azopigments were extracted with dichloromethane. N,N-Dimethylformamide was removed by washing with water. T.l.c. revealed azodipyrrole free acid (more-polar azopigment derived from bilirubin mono-D-glucosyluronic acid derivatives) and, mainly, allylprotected 1-O-acyl-D-glucopyranuronic acids (less-polar azopigments, derived from mono- and di-D-glucosyluronic acid derivatives). A first development (19:1 benzeneethyl acetate) fractionated the 1:1 mixture of *exo*-vinyl (more labile) and *endo*-vinyl (less mobile) ester isomers. A second development (17:3 benzeneethyl acetate) gave a further fractionation of the anomeric pairs. For each pair, the slower-moving azopigment was the major compound. Isolation of the *exo*- and *endo*-vinyl isomers and partial separation of the anomers was also effected by column chromatography (19:1 benzene-ethyl acetate).

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REFERENCES

- 1 F. COMPERNOLLE, G. P. VAN HEES, N. BLANCKAERT, AND K. P. M. HEIRWEGH, Biochem. J., 171 (1978) 185-201.
- 2 E. R. GORDON, T. H. CHAN, K. SAMODAI, AND C. A. GORESKY, Biochem. J., 167 (1977) 1-8.
- 3 K. P. M. HEIRWEGH, J. FEVERY, R. MICHIELS, G. P. VAN HEES, AND F. COMPERNOLLE, Biochem. J., 145 (1975) 185–199.
- 4 R. P. H. THOMPSON AND A. H. HOFMANN, Biochim. Biophys. Acta, 451 (1976) 267-277.
- 5 A. F. McDonagh and F. Assisi, Chem. Commun., (1972) 117-119.
- 6 D. KEGLEVIC, N. PRAVDIC, AND J. TOMASIK, J. Chem. Soc., C, (1968) 511-514.
- 7 N. PRAVDIC AND D. KEGLEVIC, Tetrahedron, 21 (1965) 1897-1901.
- 8 D. KEGLEVIC AND D. LJEVAKOVIC, Carbohydr. Res., 64 (1978) 319-322.
- 9 N. ROY AND C. P. J. GLAUDEMANS, Carbohydr. Res., 45 (1975) 299-301.
- 10 G. N. BOLLENBACK, J. W. LONG, D. G. BENJAMIN, AND J. A. LINDQUIST, J. Am. Chem. Soc., 77 (1955) 3310-3315.
- 11 E. J. COREY AND J. W. SUGGS, J. Org. Chem., 38 (1973) 3224.
- 12 P. A. GENT AND R. GIGG, Chem. Commun., (1974) 277-278.
- 13 R. BOSS AND R. SCHEFFOLD, Angew. Chem., 88 (1976) 578-579.
- 14 F. P. VAN ROY AND K. P. M. HEIRWEGH, Biochem. J., 107 (1968) 507-518.
- 15 F. COMPERNOLLE, F. H. JANSEN, AND K. P. M. HEIRWEGH, Biochem. J., 120 (1970) 891-894.
- 16 F. H. JANSEN AND M. S. STOLL, Biochem. J., 125 (1971) 585-597.
- 17 M. SALMON, E. DIAZ, M. C. ROCK. AND C. FENSELAU, Org. Magn. Reson., 8 (1976) 126-128.
- 18 M. SALMON, J. Heterocycl. Chem., 14 (1977) 1101-1102.