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# Discovery of potent dipeptidyl peptidase IV inhibitors through pharmacophore hybridization and hit-to-lead optimization

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#### ABSTRACT

A novel dipeptidyl peptidase IV inhibitor hit (**5**,  $IC_{50} = 0.86 \,\mu$ M) was structurally derived from our recently disclosed preclinical candidate **4** by replacing the cyanobenzyl with a butynyl based on pharma-cophore hybridization. A hit-to-lead optimization effort was then initiated to improve its potency. Most N-substituted analogs exhibited good in vitro activity, and compound **180** ( $IC_{50} = 1.55 \,n$ M) was identified to be a potent dipeptidyl peptidase IV inhibitor with a significantly improved pharmacokinetic properties (bioavailablity: 41% vs 82.9%;  $T_{1/2}$ : 2 h vs 4.9 h).

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#### 1. Introduction

Type 2 diabetes (T2D) formerly referred to as non-insulindependent or adult-onset diabetes, results from the body's ineffective use of insulin and comprises over 90% of diabetes patients. With more than 371 million people affected worldwide, diabetes is a big social burden with more than 471 billion dollars cost on its healthcare. Developing countries accounts for 80% of diabetic patients. Especially, China has the most population with diabetics of 90 million. And this number is predicted to increase to 129.7 million. Thus, more and persistent effort should be paid to fight against its epidemic damage and the global burden.<sup>1</sup>

Glucose like peptidase-1 (GLP-1) is an important incretin which contributes to the increase of insulin secretion and sensitivity, beta cell mass, and satiety, as well as the reduction of glucagon secretion and gastric emptying, which is helpful for glucose control for type 2 diabetics. However, in the normal physical condition, GLP-1 is rapidly truncated by dipeptidyl peptidase IV (DPP-IV) and lost its function.<sup>2</sup> Thus inhibition of DPP-IV could effectively maintain the GLP-1 function and control glucose level. Compared to conventional anti-diabetic drugs, dipeptidyl peptidase IV (DPP-IV) inhibitors have good patient compliance, reduced risks of hypo-

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glycemia, and less side effect.<sup>3</sup> As a result, DPP-IV inhibitors gradually became the major intervention for type 2 diabetics.

Traditional medicinal strategies for protease inhibitors mostly rely on the direct tight binding to the target, which leads to covalent compounds.<sup>4</sup> However, the emergence of non-covalent compounds to achieve satisfactory binding affinity with lower risk of selectivity issues has been witnessed in recent years.<sup>5</sup> The first non-covalent DPP-IV inhibitor with good selectivity against DPP-8 and DPP-9,<sup>6</sup> Sitagliptin (**1** Fig. 1),<sup>7</sup> was marketed in 2006. Alogliptin (**2** Fig. 1) was the second non-covalent DPP-IV inhibitor approved by the EMA, and it exhibited better efficacy than sitagliptin.<sup>8</sup> The most potent and long lasting drug is Linagliptin (**3** Fig. 1).<sup>9</sup> The market drug of covalent DPP-IV inhibitors are Vildagliptin,<sup>10</sup> and Saxagliptin.<sup>11</sup>

Our recently disclosed compound **4** was derived from Alogliptin by keeping its pharmacophore (3-aminopypiperidinyl region, red box, Fig. 2) while modifying the scaffold.<sup>12</sup> It displayed a better in vivo efficacy than Alogliptin in lean mice and dose-dependent glucose reduction in T2D model ob/ob mice. Although compound **4** acquires a similar pharmacokinetic profile with Alogliptin in rat (bioavailability: 40% and half life: 2 h), it still has a lot of space to improve. Herein, we present how we employed the classic medicinal chemistry strategy to further improve its drug-like properties.

Close comparison of the pharmacophore of Linagliptin and compound **4** revealed that the cyanobenzyl should be freely changeable



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Figure 1. Marketed non-covalent DPP-IV inhibitors.



Figure 2. Generation of hit 5.

with butynyl if the 3-(*R*)-aminopiperidinyl group was still present. Thus, compound **5** was immediately generated based on pharmacophore hybridization. Though its activity was still in the micro molar range (IC<sub>50</sub> = 0.86  $\mu$ M), it could act as a starting point for further optimization. The modification of compound **5** through N-substitution led to a series of novel DPP-IV inhibitors with significantly increased activity. The most active compound, **180** (IC<sub>50</sub> = 1.55 nM), exhibits much better pharmacokinetic profile than compound **4** (bioavailablity: 41% vs 82.9%;  $T_{1/2}$ : 2 h vs 4.9 h), which implies



Scheme 1. Synthesis of compounds 5, 6 and 18a–o. Reagents and conditions: (a) sulfuric acid, fuming nitric acid, rt; (b) dimethylfomamide–dimethyl acetal, DMF, 80 °C then 140 °C; (c) AcOH, Zn, 80 °C; (d) POCl<sub>3</sub>, DIEA, toluene, 70–80 °C; (e) 1 N NaOH/H<sub>2</sub>O, 100 °C; (f) (BoC)<sub>2</sub>O, DMAP, Et<sub>3</sub>N, THF, rt; (g) 1-bromo-2-butyne, DIEA, DMF, rt; (h) HCl/H<sub>2</sub>O, MeOH, rt; (i) NBS, DCM, rt; (j) RX, NaH, DMF, rt; (k) 3-(*R*)-aminopiperidine, NaHCO<sub>3</sub>, 120 °C, ethanol.

potential better and competent in vivo efficacy for glucose control.

### 2. Chemistry

The synthesis of compounds **5**, **6** and **18a–o** is outlined in Scheme 1. Synthesis of compound **13** from **7** was described in our recent report.<sup>9</sup> Alkylation of **13** with 1-bromo-2-butyne provided precursor**14**, which was de-protected to give key intermediate **15**. The bromination of compound **15** with *N*-bromosuccinimide (NBS) provided compound **16**, which was converted to **5** by amination. The direct amination of **15** with 3-(*R*)-aminopiperidine afforded compound **6**. Compounds **18a–o** were obtained by the N-alkylation of **15** followed by the replacement of the chloro group with a 3-(*R*)-aminopiperidinyl group.

## 3. Results and discussion

#### 3.1. Hit identification through pharmacophore hybridization

The similarity of the pharmacophore between Alogliptin, compound **4** and Linagliptin reminded us that the cyanobenzyl group of **4** could be replaced with a butynyl group without sacrificing in vitro activity (Fig. 2). Because Linagliptin is the most potent and longest-lasting DPP-IV inhibitor on the market, we decided to test our hypothesis with a pharmacophore hybridization approach.<sup>10</sup> However, to our disappointment, compound **5** significantly lost activity (IC<sub>50</sub> = 0.86  $\mu$ M). Though this compound did not achieve the desired inhibition against DPP-IV, it might be a hit worthy of further optimization.

#### Table 1

Modifications at the N-5 position



#### 3.2. Hit-to-lead optimization on compound 5

With compound **5** in hand, we immediately initiated a hit-tolead optimization effort. First, compound **6**, without any substituent on the pyrolyl ring, was synthesized and found to have better activity with an IC<sub>50</sub> of 0.46  $\mu$ M. By simply adding a methyl group on the nitrogen (compound **18a**), the activity was further increased. This trend suggested that N-substitution might a powerful way to increase inhibition. Thus, we decided to make a series of Nsubstituted analogs (**18b–o**).

Though the first N-heterocyclic compound (**18b**) afforded further increased activity, it was still relatively inactive compared to parent compound **4** ( $IC_{50}$  = 44.0 nM). Inspired by the discovery of Linagliptin, where it was found that the N-5 position could hold larger steric substituents, a series of compounds with bicyclic rings at the N-position were made (**18c–o**). A few of these compounds displayed activity in the nM range, such as **18f**, **18l**, and **18m** (<10 nM). Surprisingly, the best compound (**18o**,  $IC_{50}$  = 1.55 nM) contained the same N-substituent as Linagliptin (see Table 1)

#### 3.3. Biological evaluation of compound 180

Compound **180** was picked for preliminary evaluation for its high DPP-IV inhibitory activity. It had no inhibition against DPP-8 or 9 up to 10  $\mu$ M. Compound **180** had no CYP 3A inhibition up to 30  $\mu$ M, which is likely to decrease the risk of drug–drug interactions. The pharmacokinetic study of **180** also displayed a good drug–like profile and successfully overcame the insufficiency in the starting compound **4** (Table 2 and Fig. 4). Compared to its parent compound **4**, it had a longer half life in rats (approximately 5 h)

No.	R	DPP-IV <sup>a</sup>	No.	R	DPP-IV <sup>a</sup>
18b		44.0	18i	*F	14.5
18c	* N	44.1	18j	· CH3	52.6
18d	* ~ N	27.6	18k	» CI	11.4
18e	*	48.2	181	* <b>N F</b>	4.67
18f	* N	7.05	18m	» N	5.93
18g	* Br	139.2	18n	* N	15.5
18h	*CI	60.1	180	* N	1.55

<sup>a</sup> nM. Data represent the mean of at least three independent measurements.

Pharmacokinetic	parameters	of	180	in	rat

Dose IV/po	mg/kg	10/30
$AUC_{0-\infty}$ po	$\mu$ g h mL $^{-1}$	25.3 ± 2.6
T <sub>1/2</sub> po	h	$4.9 \pm 2.5 v$
Cl <sub>z</sub> IV	$L h^{-1} kg^{-1}$	$1.0 \pm 0.1$
V <sub>z</sub> IV	$L kg^{-1}$	$4.0 \pm 1.3$
$MRT_{0-\infty}$ po	h	8.3 ± 1.6
F	%	82.9

po: Oral administration. IV: Intravenous injection.



Figure 3. Hit-to-lead optimization strategy.



Figure 4. Concentration-time curve of 180 in rat.

and a better oral bioavailability (82.9%) that was comparable with marketed DPP-IV inhibitors (Alogliptin, 45%; Linagliptin, 50.7%).<sup>9,12,13</sup>

Since compound **180** acquired better half life and bioavailability, it is highly possible to have a better in vivo efficacy than Alogliptin. As the structure is similar with Linagliptin, some evaluation can be done like the peers' work.<sup>14</sup> Currently, there is no more urgent to develop new drugs in DPP-IV inhibitor class, in this paper we mainly report the optimization strategy started from compound **4** and the improved pharmacokinetic profile of compound **180**.

#### 4. Conclusion

Though our recently reported novel DPP-IV inhibitor **4** acquires acceptable drug like properties and has a better in vivo efficacy than

Alogliptin, its bioavailability (40%) and half life (2 h) still have some space to improve. Thus we initiated an optimization toward better PK properties while maintain the necessary in vitro activity. Inspired by the similarity in pharmacophores between Alogliptin, compound 4 and Linagliptin, we identified a novel DPP-IV inhibitor (5) by utilizing a pharmacophore hybridization strategy. Though the in vitro activity was still low, compound **5** ( $IC_{50} = 0.86 \mu M$ ) was a logical hit for optimization and generated a series of DPP-IV inhibitors with significantly improved activity. Compound 180 was finally obtained with the desired in vitro activity (IC<sub>50</sub> = 1.55 nM) and much improved pharmacokinetic profile (bioavailability 82.9%; half life 4.9 h). Thus 180 is highly possible to be more active than compound **4** and Alogliptin in vivo. This paper is mainly to disclose the successful optimization by adopting classical medicinal chemistry strategy and suggests that further optimization within this series may generate more active DPP-IV inhibitors.

#### 5. Experimental section

#### 5.1. Chemistry

All commercially available compounds and solvents were of reagent grade and used without further treatment unless otherwise noted. Reactions were monitored by TLC using Qing Dao Hai Yang GF254 silica gel plates ( $5 \times 10$  cm); zones were detected visually under ultraviolet irradiation (254 nm) and by spraying with an ethanol solution of 2,4-DNP or ninhydrin, or by fuming with iodine steam. Silica gel column chromatography was performed on silica gel (200–300 mesh) from Qing Dao Hai Yang. NMR spectra were recorded on a Bruker NMR AVANCE 400 (400 MHz) or a Bruker NMR AVANCE 500 (500 MHz). Chemical shifts ( $\delta$ ) were recorded in ppm and coupling constants (J) in hertz (Hz). Splitting patterns describe apparent multiplicities and are designated as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) or br (broad). MS data were measured on an Agilent MSD-1200 ESI-MS system.

### 5.1.1. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-3*H*pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (5)

A mixture of 16 (301 mg, 1.0 mmol), 3-(R)-aminopiperidine dihydrochloride (207 mg, 1.2 mmol) and NaHCO<sub>3</sub> (336 mg, 4.0 mmol) in a sealed tube containing 15 mL of ethanol was heated at 120 °C and stirred overnight. The reaction mixture was subsequently cooled to room temperature and filtered. The resulting filtrate was concentrated in vacuo and then purified by flash chromatography to yield compound **5**. Yield: 81.8%. <sup>1</sup>H NMR (400 MHz, MeOD) δ ppm: 7.37 (1H, s), 4.84-4.82 (2H, m), 3.53-3.50 (1H, m), 3.47-3.43 (1H, m), 3.06 (1H, s), 2.92-2.98 (1H, m), 2.03-2.00 (1H, m), 1.98-1.96 (1H, m), 1.89-1.87 (1H, m), 1.81 (3H, s), 1.79–1.73 (1H, m), 1.33–1.27 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD)  $\delta$  ppm: 24.92 (1C), 28.68 (1C), 34.07 (1C), 35.63 (1C), 52.68 (1C), 58.99 (1C), 75.29 (1C), 80.54 (1C), 81.04 (1C), 91.44 (1C), 116.26 (1C), 129.02 (1C), 142.13 (1C), 155.80 (1C), 156.49 (1C); ESI-MS calcd for (C<sub>15</sub>H<sub>19</sub>N<sub>5</sub>O) [M+H]<sup>+</sup>, 363.07, 365.07, found 364.0, 366.0.

Compounds **6** and **18a–o** were prepared in a manner identical to that described for **5**.

### 5.1.2. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-3*H*pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (6)

Yield: 67.8%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 10.87 (1H, s), 7.23 (1H, d, *J* = 2.8 Hz), 6.36 (1H, d, *J* = 2.8 Hz), 4.85 (2H, s), 3.49– 3.46 (1H, m), 3.36–3.33 (1H, m), 3.11 (1H, m), 2.92 (1H, m), 2.80–2.78 (1H, m), 2.27 (2H, s), 1.99–1.97 (1H, m), 1.88–1.86 (1H, m), 1.79 (3H, s), 1.73–1.70 (1H, m), 1.37–1.32 (1H, m); ESI-MS calcd for ( $C_{15}H_{19}N_5O$ ) [M+H]<sup>+</sup>, 286.16, found 286.1.

#### 5.1.3. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5methyl-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18a)

Yield: 52.6%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 6.87 (1H, d, J = 2.8 Hz), 6.17 (1H, d, J = 2.8 Hz), 5.19 (2H, s), 4.77–4.63 (2H, AB q, J = 34.8 Hz, 16.4 Hz), 3.96 (1H, s), 3.50–3.47 (1H, m), 3.32 (2H, m), 3.03–2.98 (1H, m), 2.90 (1H, m), 2.05 (1H, m), 1.84 (1H, m), 1.75 (3H, s), 1.65–1.63 (2H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 22.64 (1C), 28.34 (1C), 30.38 (1C), 34.41 (1C), 35.57 (1C), 47.69 (1C), 51.55 (1C), 55.40 (1C), 74.46 (1C), 79.35 (1C), 101.61 (1C), 115.10 (1C), 131.69 (1C), 142.73 (1C), 153.49 (1C), 155.42 (1C); ESI-MS calcd for (C<sub>16</sub>H<sub>21</sub>N<sub>5</sub>O) [M+H]<sup>+</sup>, 300.17, found 300.1.

## 5.1.4. (*R*)-2-(2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-4oxo-3*H*-pyrrolo[3,2-*d*]pyrimidin-5(4*H*)-yl)acetonitrile (18b)

Yield: 54.2%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 7.14 (1H, d, J = 2.8 Hz), 6.39 (1H, d, J = 2.8 Hz), 5.46 (2H, s), 4.76 (2H, s), 3.49–3.47 (1H, m), 3.38–3.35 (1H, m), 3.15 (1H, m), 2.96–2.91 (1H, m), 2.80 (2H, s), 2.00–2.96 (1H, m), 1.88–1.84 (1H, m), 1.80 (3H, s), 1.75–1.65 (2H, m), 1.42 (1H, m);<sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 22.95 (1C), 28.35 (1C), 32.54 (1C), 34.80 (1C), 35.47 (1C), 47.58 (1C), 50.39 (1C), 51.26 (1C), 58.04 (1C), 73.97 (1C), 79.86 (1C), 104.73 (1C), 114.60 (1C), 130.28 (1C), 143.98 (1C), 154.66 (1C), 155.57 (1C); ESI-MS calcd for (C<sub>17</sub>H<sub>20</sub>N<sub>6</sub>O) [M+H]<sup>+</sup>, 325.17, found 325.1.

### 5.1.5. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-(pyrimidin-2-ylmethyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18b)

Yield: 54.5%. <sup>1</sup>H NMR (400 MHz, MeOD) *δ* ppm: 8.65–8.63 (2H, d, *J* = 4.8 Hz), 7.32 (1H, d, *J* = 2.8 Hz), 7.31–7.28 (1H, t, *J* = 4.8 Hz), 6.31 (1H, d, *J* = 2.8 Hz), 5.81 (2H, s), 4.76–4.67 (2H, q, *J* = 18.8 Hz), 3.46–3.43 (1H, m), 3.36–3.29 (1H, m), 3.12–3.08 (1H, m), 2.90 (1H, m), 2.81 (1H, m), 2.78–2.76 (1H, m), 2.01–1.98 (1H, m), 1.88–1.86 (1H, m), 1.85–1.75 (1H, m), 1.72 (3H, s), 1.43–1.39 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD) *δ* ppm: 22.34 (1C), 28.87 (1C), 32.60 (1C), 35.16 (1C), 49.51 (1C), 50.51 (1C), 52.77 (1C), 54.39 (1C), 58.06 (1C), 75.36 (1C), 80.41 (1C), 103.23 (1C), 116.32 (1C), 121.15 (1C), 134.41 (1C), 145.10 (1C), 155.54 (1C), 156.79 (1C), 158.70 (1C), 167.68 (1C); ESI-MS calcd for ( $C_{20}H_{23}N_7O$ ) [M+H]<sup>+</sup>, 378.20, found 378.1.

### 5.1.6. (*R*)-5-((1*H*-Indol-3-yl)methyl)-2-(3-aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18c)

Yield: 51.7%. <sup>1</sup>H NMR (400 MHz, MeOD)  $\delta$  ppm: 7.46–7.44 (1H, d, *J* = 8.0 Hz), 7.30 (1H, s), 7.28 (1H, s), 7.28 (1H, d, *J* = 2.8 Hz), 7.04–7.00 (1H, m), 6.92–6.88 (1H, m), 6.13 (1H, d, *J* = 2.8 Hz), 5.73 (2H, s), 4.81 (2H, s), 3.57 (1H, m), 3.47–3.44 (1H, m), 3.26–3.25 (1H, m), 3.07–3.01 (1H, m), 2.05 (1H, m), 1.89–1.87 (1H, m), 1.78 (1H, m), 1.77 (3H, s), 1.63–1.61 (1H, m), 1.22 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD)  $\delta$  ppm: 22.47 (1C), 28.92 (1C), 35.44 (1C), 44.25 (1C), 48.49 (1C), 49.51 (1C), 52.92 (1C), 53.85 (1C), 58.06 (1C), 61.44 (1C), 75.36 (1C), 80.41 (1C), 102.73 (1C), 112.44 (1C), 115.73 (1C), 119.23 (1C), 120.29 (1C), 122.77 (1C), 125.81 (1C), 127.80 (1C), 133.07 (1C), 138.14 (1C), 144.66 (1C), 154.56 (1C), 156.82 (1C); ESI-MS calcd for (C<sub>24</sub>H<sub>26</sub>N<sub>6</sub>O) [M+H]<sup>+</sup>, 415.22, found 415.1.

#### 5.1.7. (*R*)-5-((1*H*-Benzo[*d*]imidazol-2-yl)methyl)-2-(3aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-3*H*-pyrrolo[3,2*d*]pyrimidin-4(5*H*)-one (18d)

Yield: 77.1%. <sup>1</sup>H NMR (400 MHz, MeOD)  $\delta$  ppm: 7.47 (2H, m), 7.39 (1H, d, *J* = 2.8 Hz), 7.18–7.15 (2H, m), 6.32 (1H, d, *J* = 2.8 Hz), 5.84 (1H, s), 4.78 (2H, s), 3.46–3.44 (1H, m), 3.43–3.42 (1H, m), 3.41–3.40 (1H, m), 3.03–2.98 (1H, m), 2.08–2.05 (1H, m), 1.91–1.88 (1H, m), 1.82–1.79 (1H, m), 1.77–1.75 (1H, m), 1.73 (3H, s), 1.60–1.57 (1H, m); ESI-MS calcd for ( $C_{23}H_{25}N_7O$ ) [M+H]<sup>+</sup>, 416.21, found 416.2.

#### 5.1.8. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-(quinolin-4-ylmethyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18e)

Yield: 51.6%. <sup>1</sup>H NMR (400 MHz, MeOD) *δ* ppm: 8.67 (1H, d, J = 8.0 Hz), 8.25–8.23 (1H, d, J = 8.0 Hz), 8.08–8.06 (1H, d, J = 8.0 Hz), 7.85–7.81 (1H, m), 7.73–7.69 (1H, m), 7.40 (1H, d, J = 2.8 Hz), 6.62 (1H, d, J = 2.8 Hz), 6.43–6.42 (1H, m), 6.24 (2H, s), 4.79–4.70 (2H, t, J = 19.2 Hz), 3.48–3.45 (1H, m), 3.41–3.38 (1H, m), 3.03–2.98 (1H, m), 2.93–2.87 (1H, m), 2.76–2.70 (1H, m), 2.01–1.98 (1H, m), 1.90–1.87 (1H, m), 1.79–1.76 (1H, m), 1.74 (3H, s), 1.40–1.30 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD) *δ* ppm: 24.52 (1C), 33.74 (1C), 35.42 (1C), 48.53 (1C), 49.55 (1C), 49.58 (1C), 52.76 (1C), 59.35 (1C), 75.28 (1C), 80.52 (1C), 103.96 (1C), 116.09 (1C), 119.10 (1C), 124.29 (1C), 127.06 (1C), 128.60 (1C), 129.89 (1C), 131.22 (1C), 134.10 (1C), 145.54 (1C), 147.43 (1C), 148.48 (1C), 151.30 (1C), 156.14 (1C), 156.94 (1C); ESI-MS calcd for ( $C_{25}H_{26}N_6$ O) [M+H]<sup>+</sup>, 427.22, found 427.1 (see Fig. 3)

### 5.1.9. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-(quinolin-2-ylmethyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18f)

Yield: 54.8%. <sup>1</sup>H NMR (400 MHz, CDCl3)  $\delta$  ppm: 8.20–8.18 (1H, d, *J* = 8.0 Hz), 7.98–7.96 (1H, d, *J* = 8.8 Hz), 7.83–7.81 (1H, d, *J* = 8.0 Hz), 7.85–7.81 (1H, m), 7.73–7.70 (1H, m), 7.54–7.51 (1H, m), 7.40 (1H, d, *J* = 2.8 Hz), 6.35 (1H, d, *J* = 2.8 Hz), 5.87 (2H, s), 4.79–4.69 (2H, t, *J* = 21.6 Hz), 3.44–3.41 (1H, m), 3.36–3.29 (1H, m), 2.99–2.94 (1H, m), 2.88–2.85 (1H, m), 2.71–2.63 (1H, m), 1.99–1.95 (1H, m), 1.86–1.83 (1H, m), 1.73 (1H, m), 1.73 (3H, s), 1.38–1.26 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD)  $\delta$  ppm: 24.48 (1C), 33.69 (1C), 35.32 (1C), 48.48 (1C), 49.51 (1C), 52.69 (1C), 54.44 (1C), 59.33 (1C), 75.36 (1C), 80.43 (1C), 103.79 (1C), 115.92 (1C), 120.19 (1C), 127.82 (1C), 139.13 (1C), 145.41 (1C), 148.50 (1C), 155.96 (1C), 156.92 (1C), 159.53 (1C); ESI-MS calcd for ( $C_{25}H_{26}N_6O$ ) [M+H]<sup>+</sup>, 427.22, found 427.1.

# 5.1.10. (*R*)-2-(3-Aminopiperidin-1-yl)-5-((6-bromoquinolin-2-yl)methyl)-3-(but-2-yn-1-yl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18g)

Yield: 43.8%. <sup>1</sup>H NMR (400 MHz, MeOD) *δ* ppm: 8.14–8.11 (1H, d, *J* = 8.8 Hz), 8.00–7.99 (1H, d, *J* = 2.0 Hz), 7.90–7.87 (1H, m), 7.80–7.77 (1H, m), 7.44 (1H, d, *J* = 2.8 Hz), 7.23–7.21 (1H, d, *J* = 8.8 Hz), 6.42 (1H, d, *J* = 2.8 Hz), 5.91 (2H, s), 4.83–4.81 (2H, m), 3.53–3.50 (1H, m), 3.45–3.42 (1H, m), 3.10–3.04 (1H, m), 2.96–2.91 (1H, m), 2.80–2.75 (1H, m), 2.07–2.03 (1H, m), 1.95–1.90 (1H, m), 1.84 (3H, s), 1.83–1.75 (1H, m), 1.45–1.33 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>+MeOD) *δ* ppm: 24.27 (1C), 33.45 (1C), 35.29 (1C), 48.48 (1C), 49.50 (1C), 49.98 (1C), 52.50 (1C), 54.29 (1C), 59.07 (1C), 75.14 (1C), 80.40 (1C), 103.70 (1C), 115.68 (1C), 121.17 (1C), 129.63 (1C), 130.72 (1C), 131.04 (1C), 133.61 (1C), 134.19 (1C), 137.85 (1C), 145.06 (1C), 146.83 (1C), 155.63 (1C), 156.67 (1C), 159.63 (1C); ESI-MS calcd for ( $C_{25}H_{25}BrN_6O$ ) [M+H]<sup>+</sup>, 505.13, found 505.1.

### 5.1.11. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((6chloroquinolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18h)

Yield: 67.8%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 7.96–7.93 (1H, m), 7.71 (1H, d, J = 2.4 Hz), 7.60–7.57 (1H, dd, J = 2.4 Hz, J = 9.2 Hz), 7.30–7.26 (1H, m), 7.20 (1H, d, J = 2.8 Hz), 6.34 (1H, d, J = 2.8 Hz), 5.88 (2H, s), 4.79 (2H, d, J = 2.0 Hz), 3.48–3.43 (1H, m), 3.06–3.00 (1H, m), 2.90–2.85 (1H, m), 2.72–2.67 (1H, m), 1.98–1.93 (1H, m), 1.89 (2H, s), 1.85–1.82 (1H, m), 1.80 (3H, s), 1.74–1.65 (1H, m), 1.32–1.24 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 23.41 (1C), 33.45 (1C), 34.49 (1C), 47.83 (1C), 51.36 (1C), 53.67 (1C), 59.26 (1C), 74.47 (1C), 77.00 (1C), 79.34 (1C), 103.12

(1C), 114.75 (1C), 120.82 (1C), 126.14 (1C), 127.93 (1C), 130.52 (1C), 130.74 (1C), 131.49 (1C), 132.14 (1C), 136.21 (1C), 143.72 (1C), 145.95 (1C), 154.24 (1C), 155.72 (1C), 158.00 (1C); ESI-MS calcd for  $(C_{25}H_{25}CIN_6O)$  [M+H]<sup>+</sup>, 461.18, found 461.1.

# 5.1.12. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((6-fluoroquinolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18i)

Yield: 62.5%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 8.02–7.97 (2H, m), 7.46–7.41 (1H, m), 7.36–7.33 (1H, m), 7.31–7.29 (1H, d, *J* = 8.4 Hz), 7.20 (1H, d, *J* = 2.8 Hz), 6.33 (1H, d, *J* = 2.8 Hz), 5.89 (2H, s), 4.79 (2H, d, *J* = 2.0 Hz), 3.48–3.43 (1H, m), 3.37–3.34 (1H, m), 3.06–3.00 (1H, m), 2.90–2.85 (1H, m), 1.98–1.93 (1H, m), 1.87–1.84 (1H, m), 1.83 (2H, s), 1.79 (3H, s), 1.74–1.66 (1H, m), 1.32–1.24 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 23.40 (1C), 33.45 (1C), 34.45 (1C), 47.81 (1C), 51.34 (1C), 53.63 (1C), 59.26 (1C), 74.48 (1C), 77.00 (1C), 79.31 (1C), 103.06 (1C), 110.37 (1C), 114.73 (1C), 119.68 (1C), 120.72 (1C), 127.90 (1C), 131.53 (1C), 136.45 (1C), 159.34 (1C), 161.31 (1C); ESI-MS calcd for (C<sub>25</sub>H<sub>25</sub>FN<sub>6</sub>O) [M+H]<sup>+</sup>, 445.21, found 445.1.

# 5.1.13. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((6-methylquinolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18j)

Yield: 69.4%. <sup>1</sup>H NMR (400 MHz, MeOD)  $\delta$  ppm: 8.06–8.04 (1H, d, *J* = 8.4 Hz), 7.86–7.84 (1H, d, *J* = 8.4 Hz), 7.62 (1H, s), 7.61–7.59 (1H, dd, *J* = 8.8 Hz, *J* = 1.6 Hz), 7.39 (1H, d, *J* = 2.8 Hz), 7.30–7.28 (1H, d, *J* = 8.4 Hz), 6.34 (1H, d, *J* = 2.8 Hz), 5.73 (2H, s), 4.80–4.74 (2H, m), 3.45–3.43 (1H, m), 3.35–3.34 (1H, m), 3.01–2.95 (1H, m), 2.87–2.67 (1H, m), 2.65 (3H, s), 2.00–1.96 (1H, m), 1.88–1.84 (1H, m), 1.77 (3H, s), 1.74–1.66 (1H, m), 1.38–1.35 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD)  $\delta$  ppm: 24.63 (1C), 33.73 (1C), 35.33 (1C), 48.49 (1C), 49.51 (1C), 52.19 (1C), 52.26 (1C), 55.50 (1C), 59.40 (1C), 75.49 (1C), 80.41 (1C), 103.56 (1C), 115.64 (1C), 123.60 (1C), 127.07 (1C), 127.79 (1C), 128.74 (1C), 130.34 (1C), 133.41 (1C), 137.70 (1C), 138.21 (1C), 145.28 (1C), 147.72 (1C), 155.81 (1C), 156.77 (1C), 160.52 (1C); ESI-MS calcd for (C<sub>26</sub>H<sub>28</sub>N<sub>6</sub>O) [M+H]<sup>+</sup>, 441.23, found 441.1.

# 5.1.14. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((7-chloroquinolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18k)

Yield: 43.4%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ ppm: 8.03 (1H, d, J = 1.6 Hz), 8.01 (1H, s), 7.69–7.67 (1H, d, J = 8.8 Hz), 7.45–7.43 (1H, dd, J = 8.8 Hz), J = 2.0 Hz), 7.30–7.28 (1H, d, J = 8.8 Hz), 7.22 (1H, d, J = 2.8 Hz), 6.35 (1H, d, J = 2.8 Hz), 5.90 (2H, s), 4.84–4.75 (2H, m), 3.50–3.47 (1H, m), 3.37–3.34 (1H, m), 3.16–3.12 (1H, m), 2.97–2.92 (1H, m), 2.84–2.80 (1H, m), 2.43 (2H, s), 1.99–1.96 (1H, m), 1.89–1.85 (1H, m), 1.80 (3H, t, J = 2.4 Hz), 1.76–1.66 (1H, m), 1.42–1.40 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD) δ ppm: 23.07 (1C), 29.65 (1C), 32.65 (1C), 34.53 (1C), 47.70 (1C), 51.40 (1C), 53.67 (1C), 58.27 (1C), 74.45 (1C), 79.46 (1C), 103.11 (1C), 114.78 (1C), 120.14 (1C), 125.74 (1C), 127.53 (1C), 143.59 (1C), 147.96 (1C), 154.12 (1C), 155.67 (1C), 158.74 (1C); ESI-MS calcd for ( $C_{25}H_{25}ClN_6O$ ) [M+H]<sup>+</sup>, 461.18, found 461.1.

# 5.1.15. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((7-fluoroquinolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18l)

Yield: 45.9%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 8.04–8.02 (1H, d, *J* = 8.4 Hz), 7.75–7.64 (1H, m), 7.67–7.64 (1H, dd, *J* = 5.2 Hz, *J* = 2.0 Hz), 7.30–7.28 (1H, m), 7.24 (1H, s), 7.22 (1H, d, *J* = 2.8 Hz), 6.35 (1H, d, *J* = 2.8 Hz), 5.90 (2H, s), 4.80 (2H, d, *J* = 2.0 Hz), 3.49–3.47 (1H, m), 3.38–3.35 (1H, m), 3.07 (1H, m), 2.93–2.88 (1H, m),

2.76–2.71 (1H, m), 1.98–1.95 (1H, m), 1.88 (1H, m), 1.85 (2H, s), 1.80 (3H, s), 1.76–1.66 (1H, m), 1.33–1.31 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 23.17 (1C), 33.51 (1C), 34.49 (1C), 47.03 (1C), 51.37 (1C), 53.71 (1C), 74.51 (1C), 79.34 (1C), 103.12 (1C), 112.76 (1C), 114.80 (1C), 116.87 (1C), 119.24 (1C), 124.38 (1C), 129.45 (1C), 131.53 (1C), 137.02 (1C), 143.70 (1C), 148.55 (1C), 154.22 (1C), 155.73 (1C), 158.81 (1C), 162.13 (1C), 164.12 (1C); ESI-MS calcd for ( $C_{25}H_{25}FN_{6}O$ ) [M+H]<sup>+</sup>, 445.21, found 445.1.

### 5.1.16. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-(quinoxalin-2-ylmethyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18m)

Yield: 61.5%. <sup>1</sup>H NMR (400 MHz, MeOD)  $\delta$  ppm: 8.67 (1H, s), 7.97–7.90 (2H, m), 7.74–7.69 (2H, m), 7.45 (1H, d, *J* = 2.8 Hz), 6.35 (1H, d, *J* = 2.8 Hz), 5.90 (2H, s), 4.78–4.71 (2H, m), 3.44–3.41 (1H, m), 3.36–3.32 (1H, m), 2.99–2.95 (1H, m), 2.86–2.84 (1H, m), 2.71–2.66 (1H, m), 1.99–1.94 (1H, m), 1.85–1.82 (1H, m), 1.73 (3H, s), 1.69–1.68 (1H, m), 1.37–1.26 (1H, m); <sup>13</sup>C NMR (500 MHz, MeOD)  $\delta$  ppm: 24.46 (1C), 33.68 (1C), 35.33 (1C), 48.48 (1C), 49.98 (1C), 52.64 (1C), 54.40 (1C), 59.33 (1C), 75.35 (1C), 80.43 (1C), 103.71 (1C), 115.75 (1C), 129.67 (1C), 129.98 (1C), 131.16 (1C), 131.58 (1C), 134.10 (1C), 142.64 (1C), 142.93 (1C), 145.16 (1C), 145.45 (1C), 154.23 (1C), 155.90 (1C), 156.81 (1C); ESI-MS calcd for (C<sub>24</sub>H<sub>25</sub>N<sub>7</sub>O) [M+H]<sup>+</sup>, 428.21, found 428.1.

# 5.1.17. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((3-methylquinoxalin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (18n)

Yield: 53.4%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 7.98–7.92 (2H, m), 7.69–7.61 (2H, m), 7.07 (1H, d, *J* = 2.8 Hz), 6.34 (1H, d, *J* = 2.8 Hz), 6.04 (2H, s), 4.78 (2H, d, *J* = 2.0 Hz), 3.49–3.46 (1H, m), 3.38–3.35 (1H, m), 3.06–3.01 (1H, m), 2.91–2.86 (1H, m), 2.72 (3H, s), 2.72 (1H, m), 1.98–1.94 (1H, m), 1.87–1.81 (1H, m), 1.77 (2H, s), 1.77 (3H, s), 1.72–1.68 (1H, m), 1.29–1.27 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  ppm: 22.13 (1C), 23.44 (1C), 29.63 (1C), 33.46 (1C), 34.48 (1C), 47.84 (1C), 50.93 (1C), 51.37 (1C), 59.32 (1C), 74.46 (1C), 76.78 (1C), 79.35 (1C), 103.08 (1C), 115.00 (1C), 128.29 (1C), 128.98 (1C), 129.81 (1C), 131.41 (1C), 140.66 (1C), 141.61 (1C), 143.54 (1C), 150.83 (1C), 152.80 (1C), 154.17 (1C), 155.84 (1C); ESI-MS calcd for (C<sub>25</sub>H<sub>27</sub>N<sub>7</sub>O) [M+H]<sup>+</sup>, 442.23, found 442.2.

# 5.1.18. (*R*)-2-(3-Aminopiperidin-1-yl)-3-(but-2-yn-1-yl)-5-((4-methylquinazolin-2-yl)methyl)-3*H*-pyrrolo[3,2-*d*]pyrimidin-4(5*H*)-one (180)

Yield: 59.0%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) *δ* ppm: 7.97–7.95 (2H, d, *J* = 8.4 Hz), 7.77–7.70 (2H, m), 7.51–7.47 (1H, m), 7.13 (1H, d, *J* = 2.8 Hz), 6.28 (1H, d, *J* = 2.8 Hz), 5.86 (2H, s), 4.68–4.58 (2H, dd, *J* = 33.6 Hz, *J* = 4.0 Hz), 3.85 (2H, s), 3.39–3.37 (1H, m), 3.27–3.24 (1H, m), 3.06 (1H, m), 2.85 (1H, m), 2.79 (3H, s), 2.75–2.73 (1H, m), 1.93–1.90 (1H, m), 1.80–1.76 (1H, m), 1.64 (3H, s), 1.64 (1H, m), 1.36–1.34 (1H, m); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>+MeOD) *δ* ppm: 21.24 (1C), 22.30 (1C), 31.24 (1C), 34.09 (1C), 47.11 (1C), 49.02 (1C), 51.32 (1C), 53.40 (1C), 56.65 (1C), 74.01 (1C), 79.28 (1C), 102.17 (1C), 115.14 (1C), 122.87 (1C), 124.82 (1C), 127.31 (1C), 128.35 (1C), 132.27 (1C), 133.72 (1C), 142.95 (1C), 149.42 (1C), 153.76 (1C), 155.34 (1C), 160.96 (1C), 169.08 (1C); ESI-MS calcd for ( $C_{25}H_{27}N_70$ ) [M+H]<sup>+</sup>, 442.23, found 442.2.

#### 5.2. In vitro inhibition of DPP-IV, DPP-8 and DPP-9

Solutions of test compounds at varying concentrations ( $\leq 10 \text{ mM}$  final concentration) were prepared in dimethyl sulfoxide (DMSO) and diluted into assay buffer containing 20 mM Tris (pH 7.4), 20 mM KCl, and 0.1 mg/mL BSA. Human DPP-IV (0.1 nM final concentration) was added to the dilutions and pre-incubated for

10 min at ambient temperature before the reaction was initiated by the addition of Gly-Pro-AMC (H-glycyl-prolyl-7-amino-4-methylcoumarin, Sigma–Aldrich, 10  $\mu$ M final concentration). The total volume of the reaction mixture was 100  $\mu$ L. The kinetics of the reaction was monitored (excitation at 400 nm, emission at 505 nm) for 5–10 min, or an endpoint was measured after 10 min. Inhibition constants (IC<sub>50</sub>) were calculated from enzyme progress curves using standard mathematical models.

#### 5.3. In vivo pharmacokinetic study

Adult male SD rats (n = 4/group) were administered the test compounds dissolved in distilled water at a single dose of 20 mg/ kg or 25 mg/kg for oral administration and 5 mg/mL by injection. Blood samples of 100–200 µL were collected from the orbit at 11 time points within 24 h. The blood concentration of test compounds was determined by LC–MS/MS. The PK parameters were obtained from the pharmacokinetic software DAS. 2.0.

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