Chem. Pharm. Bull. 36(8)2968—2976(1988)

# Thromboxane A<sub>2</sub> Synthetase Inhibitors. I. Syntheses and Activities of Various N-Heteroaromatic Derivatives

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(Received January 19, 1988)

Basic N-heteroaromatic derivatives (1,2,4-triazole, thiazole and pyrimidine derivatives) having a 2-[4-(carboxy)phenoxy]ethyl moiety or a 4-[2-(carboxy)vinyl]benzyl moiety were prepared, and evaluated for ability to inhibit thromboxane  $A_2$  (TXA<sub>2</sub>) synthesis. Among the compounds prepared in this study, the 5-substituted thiazole derivatives 14d, 27 and 28 were more potent inhibitors of TXA<sub>2</sub> production than the corresponding imidazole derivatives, and the 1-substituted 1H-1,2,4-triazole derivatives 14a and 15a were almost as potent as the corresponding imidazole derivatives.

**Keywords**—thiazole; 1H-1,2,4-triazole; pyrimidine; imidazole; thromboxane  $A_2$ ;  $TXA_2$  synthetase inhibitor

Thromboxane A<sub>2</sub> (TXA<sub>2</sub>) is a kind of prostaglandin endoperoxide metabolite, and possesses potent vascular constricting and platelet aggregating activities. Prostacyclin (PGI<sub>2</sub>) is another such compound, and has potent vasodilating and platelet antiaggregating activities. TXA<sub>2</sub> and PGI<sub>2</sub> are produced from prostaglandin G<sub>2</sub> (PGG<sub>2</sub>) and/or prostaglandin H<sub>2</sub> (PGH<sub>2</sub>) by TXA<sub>2</sub> synthetase and PGI<sub>2</sub> synthetase, respectively. It was postulated that, under normal conditions, there is a balance between the biological effects of TXA2 and PGI2, but an increased plasma TXA<sub>2</sub> level has been observed among patients with ischemic heart diseases.<sup>1)</sup> Reduction of TXA2 level or increase of PGI2 level may be one approach to the treatment of these diseases. Thus, many selective TXA2 synthetase inhibitors without inhibitory effects on PGI<sub>2</sub> synthetase and cyclooxygenase have been synthesized,<sup>2-7)</sup> and several of these compounds have been subjected to clinical trials for a variety of diseases such as ischemic heart disease, thromboembolic disorders and cerebral circulatory disorders. Most of the compounds possessing a potent selective inhibitory effect on TXA2 synthesis are 1-substituted imidazole, 3-substituted pyridine and imidazo[1,5-a]pyridine derivatives. Several investigators<sup>2-7)</sup> reported that introduction of a carboxyl moiety at the terminal of the side chain on the imidazole or pyridine ring increased the inhibitory activity, and the optimum distance between the carboxyl group and the heterocycle was between 8.5 and 10 Å.2) This distance appears to correspond to that between the carboxyl group and the endoperoxide moiety of PGG<sub>2</sub> or PGH<sub>2</sub>. Thus, it has been assumed that compounds having a weakly basic Nheteroaromatic ring at one end of the molecule and a carboxyl group at the other possess a potent activity, and the most suitable distance between the two is about 9 Å.

On the basis of these hypotheses, some series of compounds having a weakly basic N-heterocyclic ring were synthesized and tested for  $TXA_2$  synthetase inhibitory activity. This paper deals with the syntheses and activities of a series of 1-substituted 1H-1,2,4-triazole, 4-substituted 4H-1,2,4-triazole, 5-substituted pyrimidine and 5-substituted thiazole derivatives.

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## Chemistry

The compounds prepared in this paper are shown in Table I, and the synthetic pathways employed are outlined in Charts 1, 2 and 3.

Preparation of 1-substituted 1H-1,2,4-triazoles was carried out by alkylation of 1H-1,2,4-triazole with the halide (1, X=I) to give a 1-substituted derivative (2a) as a main product.

Synthesis of a 4-substituted analog (2b) was carried out by another route from ethyl 4-(2-aminoethoxy)benzoate (3)<sup>8)</sup> via the thiosemicarbazide (4), because only a small amount of 2b was formed by the alkylation described above. Compound 4, prepared from 3, was treated with formic acid, and then heated in ethanol in the presence of pyridine to give 5. Treatment of 5 with hydrogen peroxide in acetic acid gave the desired 2b.

A pyrimidine derivative (2c) was prepared from 1 (X=Br). Condensation of compound

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6, prepared from 1 (X = Br), with formamidine acetate in a solution of sodium ethoxide in ethanol gave a 5-substituted-4,6-dihydroxypyrimidine (7). Chlorination of 7, followed by hydrogenation gave a 5-substituted pyrimidine derivative (2c).

A thiazole derivative substituted with a 2-[4-(carboethoxy)phenoxy]ethyl moiety at the 5 position on the thiazole ring was prepared from ethyl 4-(4-hydroxybutoxy)benzoate (9). Oxidation of 9 with pyridinium chlorochromate (PCC), followed by bromination, and treatment with thiourea afforded a 2-aminothiazole derivative (12). Sandmeyer reaction of 12

Chart 3

gave the crude chloride 13, which was reduced with zinc powder in acetic acid to give the desired thiazole compound (2d).

The esters (2a-d) were hydrolyzed in aqueous sodium hydroxide solution to give the corresponding carboxylic acids (14a-d).

Alkylation of 1*H*-1,2,4-triazole with ethyl 3-[4-(bromomethyl)phenyl]-2-propenoate gave **15a** as a main product. However, alkylation with ethyl 3-[4-(bromomethyl)phenyl]-2-methyl-2-propenoate afforded a mixture of the 1-substituted isomer (**16a**) and 4-substituted isomer (**16b**). Compound **16a** was transformed to **17a** by hydrolysis.

Reaction of the chloroaldehyde derivative (19), prepared from 18 by a method similar to that described by Dombrovskii et al., with thiourea gave the 5-substituted-2-aminothiazole (20). Compound 20 was treated with cupric chloride and tertiary butyl nitrite to give the 2-chlorothiazole 21, which was reduced by a similar method to that described for the preparation of 2d to give 22. Reduction of 22 with lithium aluminum hydride, followed by oxidation with N,N-dicyclohexylcarbodiimide in dimethylsulfoxide afforded 4-(5-thiazolylmethyl)benzaldehyde (24). Wittig reaction of 24 with 25, followed by hydrolysis gave 28. The cinnamic acid derivative (27) was prepared by condensation of 24 with malonic acid.

### **Biological Activities**

The activities of the compounds were expressed as the IC<sub>50</sub> values for the inhibition of TXA<sub>2</sub> synthesis in rat platelet-rich plasma (PRP). Usually the potency of a TXA<sub>2</sub> synthesis inhibitory compound is indicated as its inhibitory activity against the TXA<sub>2</sub> synthesis reaction mediated by a partially purified enzyme fraction such as platelet microsomes.<sup>2-7)</sup> However, we think that the value of the inhibitory activity obtained from a less purified assay system using PRP may be more practical than that obtained from the more purified enzyme assay system, because the former reflects not only the inhibitory activity against the enzyme itself but also the ability of the compound to permeate through the plasma membrane of platelets and the effects of its interaction with proteins, lipids and many other substances in blood plasma and the cytoplasm of platelets.

Table I shows the *in vitro* inhibitory activities of the compounds obtained here on TXA<sub>2</sub> production in rat PRP. 1-Substituted-1*H*-1,2,4-triazole derivatives (**14a**, **15a**, **17a**) exhibited potent inhibitory activities against TXA<sub>2</sub> synthesis, and were as potent as the corresponding imidazole derivatives (Dazoxiben and OKY-046). On the other hand, introduction of a 4*H*-1,2,4-triazol-4-yl or 5-pyrimidinyl moiety diminished the activity (compare **14b** and **14c** with Dazoxiben). Replacement of the imidazole ring by a thiazole ring increased the activities (compare **14d** and **27** with Dazoxiben and OKY-046, respectively).

In some kinds of cells, tissues and organs, some selective TXA<sub>2</sub> synthetase inhibitors have been found to increase the production of particular prostaglandins (PGs) such as PGE<sub>2</sub>, PGF<sub>2</sub>, PGD<sub>2</sub> and PGI<sub>2</sub> along with the inhibition of TXA<sub>2</sub> production.<sup>10)</sup> On the other hand, cyclooxygenase inhibitors inhibit the production of not only TXA<sub>2</sub>, but also PGs synthesized through the reaction catalyzed by fatty acid cyclooxygenase. The suppression of the production of certain PGs, especially PGI<sub>2</sub> and PGD<sub>2</sub> is undesirable for the treatment of ischemic heart disease, because the former has a potent vasodilating activity and both have potent antiaggregative activities against platelets.<sup>6,10)</sup> In rat platelets the selective inhibition of TXA<sub>2</sub> synthetase results in a marked increase in PGE<sub>2</sub> production and in a much smaller increase in other PGs such as PGF<sub>2</sub>, PGD<sub>2</sub> and PGI<sub>2</sub>. Therefore in the assay system using rat PRP, the selectivity of the inhibitory activity of a compound to TXA<sub>2</sub> synthetase as against cyclooxygenase can be estimated by measuring the increase in PGE<sub>2</sub> production along with the inhibition of TXA<sub>2</sub> production. In this paper, the selectivities of the tested compounds are indicated as the "conversion ratio (PGE<sub>2</sub>/TXA<sub>2</sub>)," obtained by dividing the individual amounts of the increase in PGE<sub>2</sub> production by those of the decrease in TXA<sub>2</sub> production at

TABLE I. Preparation of N-Heteroaromatic Derivatives and Inhibitory Activities of TXA<sub>2</sub> Production

$$A \sim O - \bigcirc CO_2 R$$
 $I$ 
 $A \sim O - \bigcirc R$ 
 $R'$ 

No.	Struc- ture	A	R	R′	Yield (%)	mp (°C)	Formula Calcd (Found)			Inhibition of TXA <sub>2</sub> production in vitro (rat PRP)	
							С	Н	N	IC <sub>50</sub> (μм)	Converting ratio
2a	I	N	Et		55	118—131 <sup>a)</sup>	52.44 5	5.42	4.11	22	0.53
2b	I	N	Et		75	125—127 <sup>b)</sup>	C <sub>13</sub> H <sub>15</sub> N 59.76 5	N <sub>3</sub> O <sub>3</sub> 5.79 1	(4.03) (6.08	1300	
2c	I	$\langle \hat{Q} \rangle$	Et	_	51	93—95 <sup>b)</sup>	C <sub>15</sub> H <sub>16</sub> N 66.16 5	N <sub>2</sub> O <sub>3</sub> 5.92 1	.0.29	> 50	_
<b>2d</b>	I	N S	Et		57	116—126 <sup>a)</sup>	C <sub>14</sub> H <sub>15</sub> N 53.59 5	NO <sub>3</sub> S · I 5.14	4.46	39	_
14a	I	N N-	Na		39	$> 280^{a}$	$C_{11}H_{10}N_{51.77}$	N₃NaO₃ 3.95 1	6.47	11	0.43
14b	I	N^N—	Na	_	56	$> 280^{a}$	$C_{11}H_{10}N_{51.77}$	N <sub>3</sub> NaO <sub>3</sub> 3.95 l	16.37) 3 16.47 16.33)	>1000	_
14c	I	$\bigvee_{N}$	Na		76	$> 280^{a}$	C <sub>13</sub> H <sub>11</sub> N 58.65 4	N <sub>2</sub> NaO <sub>3</sub> 1.16 l	,	65	0.61
14d	I	N S	Na	_	92	$> 280^{a}$	$C_{12}H_{10}N_{53.13}$			3.8	0.84
Dazoxiben <sup>d)</sup>	I	N^N-	Н	_			(33.41 3	7.00	3.01)	11	0.62
15a	II	N	Et	Н	48	139—145 <sup>a)</sup>	57.24 5	5.49 1	ICl  4.31  1.49)	6.6	0.53
16a	II	N	Et	Me	54	135—147 <sup>a)</sup>	C <sub>15</sub> H <sub>17</sub> N 58.54 5	N <sub>3</sub> O <sub>2</sub> · H 5.89 1		16	1.04
16b	II	N	Et	Me	10	182—195 <sup>a)</sup>	C <sub>15</sub> H <sub>17</sub> N 58.54 5	N <sub>3</sub> O <sub>2</sub> · H 5.89 1	,	200	
17a	II	<u></u>	Na	Me	82	$> 280^{a}$	$C_{13}H_{12}N_{58.87}$	N <sub>3</sub> NaO 1.56 1		5.8	0.36
27	II	N S	Н	Н	54	183—185°)	C <sub>13</sub> H <sub>11</sub> N 63.65 4		5.71 5.76)	1.5	0.68
28	II	N S	Н	Me	48	182—184 <sup>c)</sup>	C <sub>14</sub> H <sub>13</sub> N 64.84 5		5.40 5.31)	0.22	1.22
OKY-046 <sup>d</sup> )	II	N^N− 	Н	Н			(07.01 )	1	J.J1)	4.5	0.54
OKY-1581 <sup>d</sup>	II		Na	Me						0.15	0.60

a) Recrystallized from EtOH-ether. b) Recrystallized from EtOAc-petr. ether. c) Recrystallized from 50% EtOH. d) These compounds were prepared in our institute for experimental use. See references 2 and 3.

the concentrations of the tested compounds that gave about 90% inhibition of TXA<sub>2</sub> production in rat PRP stimulated by arachidonic acid. Theoretically the ratios will be high (about 0.5 to above 1.0) for highly selective TXA<sub>2</sub> synthetase inhibitors but low (near to 0) for the less selective ones or cyclooxygenase inhibitors.

Dazoxiben and OKY-046 (both are imidazole derivatives), which were reported<sup>2)</sup> to be highly selective TXA<sub>2</sub> synthetase inhibitors, showed conversion ratios of 0.62 and 0.54, respectively, in our experiment.

The conversion ratios of thiazole derivatives (14d, 27, 28) were 0.84, 0.68 and 1.22, respectively, being higher than those for the corresponding imidazole derivatives. However, those for triazole derivatives (14a and 17a) were 0.43 and 0.36, respectively, being lower than those for Dazoxiben and OKY-046.

From these results, it can be assumed that the inhibitory activities of the thiazole derivatives against the  $TXA_2$  production in PRP are due to the selective inhibition of  $TXA_2$  synthetase.

#### **Experimental**

Melting points are uncorrected. Infrared (IR) spectra were taken on a Hitachi 285 spectrometer. Proton nuclear magnetic resonance ( $^{1}$ H-NMR) spectra were recorded with Hitachi R40 and JEOL JNM-FX90Q spectrometers (Me<sub>4</sub>Si as an internal standard). For column chromatography, silica gel (Merck, Kieselgel 60, 0.05—0.2 mm) was used.

1-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]-1H-1,2,4-triazole Hydrochloride (2a)—A mixture of ethyl 4-(2-iodoethoxy)benzoate (1, X=I)<sup>11)</sup> (2.24 g, 7.0 mmol), 1H-1,2,4-triazole (0.49 g, 7.0 mmol) and NaOEt (0.5 g, 7.4 mmol) in EtOH (40 ml) was refluxed for 5 h, and concentrated *in vacuo*. The residue was extracted with CHCl<sub>3</sub>. The extract was washed with water, dried, and concentrated *in vacuo*. The residue was purified by silica gel (30 g) chromatography using CHCl<sub>3</sub>-MeOH (98:2) as an eluent to give the free base of 2a (1.01 g, 55%) as colorless needles; mp 92—93 °C. ¹H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.37 (3H, t), 4.24—4.74 (6H, m), 6.87 (2H, d, J=9 Hz), 7.89 (2H, d, J=9 Hz). *Anal*. Calcd for C<sub>13</sub>H<sub>15</sub>N<sub>3</sub>O<sub>3</sub>: C, 59.76; H, 5.79; N, 16.08. Found: C, 59.91; H, 5.91; N, 16.01.

The free base of 2a obtained above was treated with 23% HCl-EtOH solution to give 2a as a colorless powder. 4-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]thiosemicarbazide (4)——A mixture of CS<sub>2</sub> (2.92 g, 38.3 mmol) and CHCl<sub>3</sub> (4 ml) was added dropwise to a suspension of ethyl 4-(2-aminoethoxy)benzoate (3)<sup>8)</sup> (9.42 g, 38.3 mmol) and Et<sub>3</sub>N (7.76 g, 76.7 mmol) in CHCl<sub>3</sub> 70 ml at -13—-15 °C, and the whole was stirred at 10 °C for 10 min. After addition of a solution of ethyl chloroformate (4.1 g, 38.3 mmol) in CHCl<sub>3</sub> (7 ml), stirring was continued at room temperature for 0.5 h. After addition of ice-cold water, the organic layer was separated, washed with water, dried, and concentrated *in vacuo* to give a pale yellow oil. A solution of this oil in EtOH (100 ml) was added to a solution of hydrazine hydrate (1.93 g, 38.3 mmol) in water (2 ml), and the mixture was stirred at 5 °C for 0.5 h. The precipitate was collected by filtration, and washed with a small volume of EtOH to give 4 (5.77 g, 53%) as a colorless powder; mp 125—128 °C (EtOH). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.39 (3H, t, J=7 Hz), 3.80 (2H, br), 4.0—4.5 (6H, m), 6.93 (2H, d, J=9 Hz), 7.4—8.2 (2H, m), 7.98 (2H, d, J=9 Hz).

**4-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]-2,3-dihydro-4***H***-1,2,4-triazole-3-thione** (5)——A mixture of **4** (1.0 g, 3.5 mmol) and 80% HCOOH (3 ml) was heated at 60—70 °C for 25 min to give a solid, which was heated under reflux in pyridine (380 mg) and EtOH (20 ml) for 19 h. The mixture was concentrated *in vacuo*, and the residue was extracted with CHCl<sub>3</sub>. The extract was washed, dried, and concentrated *in vacuo* to give **5** (460 mg, 98%) as a colorless powder; mp 149—151 °C (EtOAc–petr. ether). <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 1.30 (3H, t, J = 9 Hz), 4.27 (2H, q, J = 7 Hz), 4.39 (2H, s), 7.05 (2H, d, J = 9 Hz), 7.88 (2H, d, J = 9 Hz), 8.47 (1H, s), 13.4—14.0 (1H, br).

Ethyl 4-[2-(4H-1,2,4-Triazol-4-yl)ethoxy] benzoate (2b)— $H_2O_2$  (35%, 0.25 ml) was added dropwise to a suspension of 5 (182 mg, 0.62 mmol) in AcOH (3 ml). The mixture was refluxed for 1 h, and concentrated *in vacuo*. The extract was washed with water, dried, and concentrated *in vacuo* to dryness to give 2b (122 mg, 75%) as colorless prisms.  $^1H$ -NMR (CDCl<sub>3</sub>)  $\delta$ : 1.37 (3H, t, J=7 Hz), 4.22—4.56 (6H, m), 6.88 (2H, d, J=9 Hz), 7.98 (2H, d, J=9 Hz), 8.32 (2H, s).

**Diethyl 2-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]malonate (6)**—Diethyl malonate (6.41 g, 40 mmol) was added dropwise to a solution of Na (920 mg, 40 mmol) in EtOH (80 ml). The mixture was stirred at room temperature for 0.5 h, the bromide (1, X=Br) (10.9 g, 40 mmol) was added, and the whole was refluxed for 4 h. The mixture was concentrated *in vacuo*, and the residue was extracted with benzene. The extract was washed, dried, and concentrated *in vacuo*. The residual oil was purified by silica gel chromatography to give 6 as an oil (8.85 g, 63%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.38 (3H, t, J=7 Hz), 3.42 (2H, t, J=7 Hz), 4.30 (2H, t, J=7 Hz), 4.33 (2H, q, J=7 Hz), 6.91 (2H, d, J=9 Hz), 7.98 (2H, t, J=7 Hz).

5-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]pyrimidine-4,6-dione (7)——Compound 6 (16.5 g, 4.7 mmol) was added

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to a cold solution of formamidine acetate (4.87 g, 4.7 mmol) in NaOEt solution [prepared from Na (3.23 g) and EtOH (70 ml)], and the mixture was stirred at room temperature for 24 h. After cooling, the mixture was neutralized with concentrated HCl. The precipitate was collected by filtration, and washed with EtOH to give a colorless powder (7.46 g, 52%); mp 261—267 °C.  $^{1}$ H-NMR (DMSO- $^{2}$ G)  $\delta$ : 1.30 (3H, t,  $^{2}$ G) = 6 Hz), 2.77 (2H, t,  $^{2}$ G), 4.10 (2H, t,  $^{2}$ G) = 6 Hz), 7.03 (2H, d,  $^{2}$ G) + 129 Hz), 7.88 (2H, d,  $^{2}$ G) + 129 Hz), 7.96 (1H, s).

This compound was used for the next reaction without further purification.

**4,6-Dichloro-5-[2-[4-(ethoxycarbonyl)phenoxy]ethyl]pyrimidine (8)**—A mixture of 7 (3.04 g, 10 mmol) and POCl<sub>3</sub> (3 ml) was refluxed for 1 h and concentrated *in vacuo*. The residue was poured into ice-water. The mixture was neutralized with Na<sub>2</sub>CO<sub>3</sub>, and extracted with CHCl<sub>3</sub>. The extract was washed, dried, and concentrated *in vacuo*. The residue was purified by silica gel chromatography using CHCl<sub>3</sub> as an eluent to give colorless needles (2.5 g, 73%); mp 99—102 °C. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.30 (3H, t, J = 6 Hz), 3.36 (2H, t, J = 6 Hz), 4.33 (2H, t, J = 6 Hz), 4.37 (2H, q, J = 6 Hz), 7.91 (2H, d, J = 9 Hz), 7.99 (2H, d, J = 9 Hz), 8.68 (1H, s).

This compound was used for the next reaction without further purification.

5-[2-[4-(Ethoxycarbonyl)phenoxy]ethyl]pyrimidine (2c)—A suspension of 8 (0.44 g, 1.3 mmol) and MgO (0.12 g) in 33% aqueous EtOH (12 ml) was treated with H<sub>2</sub> in the presence of 10% Pd-carbon (0.22 g). After absorption of the theoretical amount of H<sub>2</sub>, the catalyst was filtered off, and the filtrate was concentrated *in vacuo*. The residue was purified by silica gel chromatography to give colorless needles (0.18 g, 51%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.34 (3H, t, J=6 Hz), 3.08 (2H, d, J=6 Hz), 4.23 (2H, t, J=9 Hz), 4.32 (2H, q, J=6 Hz), 6.86 (2H, d, J=9 Hz), 7.98 (2H, d, J=9 Hz), 8.72 (2H, s), 9.13 (1H, s).

Ethyl 4-(3-Formylpropoxy)benzoate (10)—A solution of ethyl 4-(3-hydroxybutoxy)benzoate (9)<sup>12</sup>) (26.5 g, 0.11 mol) in CH<sub>2</sub>Cl<sub>2</sub> (22 ml) was added to a suspension of PCC (35.6 g, 0.17 mol) in CH<sub>2</sub>Cl<sub>2</sub> (220 ml). After being stirred at room temperature for 1.5 h, the mixture was treated with ether (220 ml). The ether layer was washed with water, dried, and concentrated *in vacuo* to give a colorless oil (21.5 g, 82%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.36 (3H, t, J=6 Hz), 1.98—2.27 (2H, m), 2.66 (2H, t, J=6 Hz), 4.04 (2H, t, J=6 Hz), 4.33 (2H, q, J=6 Hz), 6.87 (2H, d, J=9 Hz), 7.96 (2H, d, J=9 Hz), 9.82 (1H, s).

This oil was used for the next step without further purification.

Ethyl 4-[2-(2-Aminothiazol-5-yl)ethoxy]benzoate Hydrochloride (12)—Br<sub>2</sub> (1 ml) was added dropwise to dioxane (3 ml), and the mixture was stirred for 10 min, and diluted with  $CH_2Cl_2$  (10 ml). The solution was added dropwise to an ice-cooled solution of 10 (4.72 g, 20 mmol) in  $CH_2Cl_2$  (10 ml) over 3 h under  $N_2$  atmosphere. The mixture was stirred for 0.5 h, then a solution of  $Na_2CO_3$  (1.5 g) in waster (6 ml) was added. After being stirred for 1 h, the reaction mixture was extracted with  $CHCl_3$ . The extract was washed with water, dried, and concentrated *in vacuo* to give the crude 11 as an oil. The crude 11 was stirred in a solution of thiourea (1.47 g, 19 mmol) in EtOH (40 ml) for 13 h, and then refluxed for 24 h. The mixture was neutralized with 2 N NaOH, and concentrated *in vacuo*. The residue was extracted with  $CHCl_3$ . The extract was washed, dried, and concentrated *in vacuo*. The residue was purified by silica gel chromatography to give colorless prisms of the free base of 12; mp 171—177 °C (EtOH-ether). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.36 (3H, t, J=7 Hz), 3.10 (2H, t, J=6 Hz), 4.14 (2H, t, J=6 Hz), 4.33 (2H, q, J=7 Hz), 5.20 (2H, br), 6.83 (1H, s), 6.88 (2H, d, J=9 Hz), 7.97 (2H, d, J=9 Hz).

The free base obtained above was treated with HCl–EtOH solution by the usual method to give 12 (2.67 g, 31%) as colorless prisms; mp 174—182 °C. Anal. Calcd for  $C_{14}H_{16}N_2O_3S$ ·HCl: C, 51.14; H, 5.21; N, 8.52; Cl, 10.78. Found: C, 50.76; H, 5.03; N, 8.52; Cl, 10.87.

Ethyl 4-[2-(5-Thiazolyl)ethoxy]benzoate Hydrochloride (2d) — A solution of NaNO<sub>2</sub> (1.52 g, 0.22 mol) in H<sub>2</sub>O (3 ml) was added dropwise to a mixture of the free base of 12 (5.84 g, 0.02 mol) in H<sub>3</sub>PO<sub>4</sub> (70 ml) and concentrated HNO<sub>3</sub> (35 ml) under -5 °C. After being stirred at -5 °C for 1 h, the mixture was added to an ice-cooled solution of CuCl (15.9 g, 0.16 mol) in concentrated HCl (20 ml), and the whole was stirred at the same temperature for 3 h, neutralized with Na<sub>2</sub>CO<sub>3</sub>, and extracted with CHCl<sub>3</sub>. The extract was washed, dried, and concentrated *in vacuo*. The residue was purified by silica gel chromatography to give the crude 13 as an oil. Zn (2.17 g) was added to a solution of the crude 13 in HOAc (40 ml), and the mixture was refluxed for 2 h. The insoluble material was filtered off, and the filtrate was concentrated *in vacuo*. The residue was dissolved in CHCl<sub>3</sub>, and the solution was washed with water, dried, and concentrated *in vacuo*. The residue was treated with HCl-EtOH solution to give 2d (3.03 g, 48%) as a colorless powder. <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 1.30 (3H, t, J=7 Hz), 3.39 (3H, t, J=6 Hz), 4.28 (2H, q, J=7 Hz), 4.31 (2H, t, J=6 Hz), 7.08 (2H, d, J=9 Hz), 7.92 (2H, d, J=9 Hz), 8.03 (1H, s), 9.40 (1H, s).

Sodium 4-[2-(1*H*-1,2,4-Triazol-1-yl)ethoxy]benzoate (14a)——A mixture of 2a (0.77 g, 2.95 mmol) and NaOH (10 ml) was stirred at room temperature for 8 h, and then concentrated *in vacuo*. The residue was dissolved in water (15 ml), and the pH was ajusted to 6 with 1 N HCl. The precipitate was collected by filltration to give the free acid (312 mg) of 14a as colorless needles; mp 198—200 °C. A suspension of this material in water was treated with 5% NaOH (1.1 ml), and concentrated to dryness *in vacuo* to give 14a (0.92 g, 39%) as colorless needles. IR (KBr) cm<sup>-1</sup>: 3425, 1600, 1545, 1395.

Compounds 14b—d were prepared from 2b—d, respectively, in a fashion analogous to that used for 14a. The results are shown in Table I.

Ethyl (E)-2-Methyl-3-[4-(1H-1,2,4-triazol-1-ylmethyl)phenyl]-2-propenoate Hydrochloride (16a)——A solution

of 1H-1,2,4-triazole (0.68 g, 9.8 mmol) in NaOEt solution [prepared from Na (0.33 g, 10 mmol) and EtOH (20 ml)] was stirred at room temperature. Ethyl (E)-2-methyl-3-[4-(bromomethyl)phenyl]-2-propenoate<sup>2)</sup> (2.77 g, 9.8 mmol) was added, and after being refluxed for 14 h, the reaction mixture was concentrated *in vacuo*. The residue was dissolved in benzene. This solution was washed, dried, and concentrated *in vacuo*. The residual oil was chromatographed on silica gel (20 g).

The oily material obtained from the first elution with CHCl<sub>3</sub> was treated with HCl-EtOH solution to give **16a** (1.62 g, 48%) as a colorless powder. <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 1.26 (3H, t, J=7 Hz), 2.03 (3H, s), 4.18 (2H, q, J=7 Hz), 5.48 (2H, s), 7.25—7.55 (4H, m), 7.54 (1H, s), 8.23 (1H, s), 9.04 (1H, s).

Further elution with CHCl<sub>3</sub>-MeOH (98:2) gave the free base of **16b**, which was treated with HCl-EtOH solution to give **16b** (0.31 g, 10%) as a colorless powder. <sup>1</sup>H-NMR (DMSO- $d_6$ )  $\delta$ : 1.26 (3H, t, J=7 Hz), 2.04 (3H, s), 4.18 (2H, q, J=7 Hz), 5.53 (2H, s), 7.48 (4H, s), 7.56 (1H, s), 9.59 (1H, s).

Compound 15a was prepared in the same manner as described above for 16a.

Sodium (E)-3-[4-(1H-1,2,4-triazol-1-ylmethyl)phenyl]-2-methyl-2-propenoate (17a) was synthesized from 16a in the same manner as described above for 14a.

17a: Yield 82%, colorless powder; mp > 280 °C.

**2-Chloro-3-[4-(ethoxycarbonyl)phenyl]propionaldehyde (19)**—A solution of Na<sub>2</sub>NO<sub>2</sub> (6.0 g, 87 mmol) in water (27 ml) was added dropwise to a solution of **18** (11.9 g, 72 mmol) in 20% HCl (36 ml) at -5—0 °C. After being stirred at 0 °C for 20 min, the mixture was neutralized with NaHCO<sub>3</sub>, and added to a cooled mixture of acrolein (11.7 ml, 224 mmol), CuCl<sub>2</sub> (4.49 g, 33.4 mmol) and CaO (1.8 g, 32.1 mmol) in acetone (90 ml). The whole was stirred at 5 °C for 1.7 h, and the acetone was evaporated off. The insoluble materials were filtered off, and the filtrate was extracted with benzene. The extract was washed with water, dried, and concentrated *in vacuo* to give an oil (15.1 g, 87.1%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.39 (3H, t, J=7 Hz), 3.18 (1H, dd, J=8, 14 Hz), 3.45 (1H, dd, J=5, 14 Hz), 4.25—4.50 (1H, m), 4.37 (2H, q, J=7 Hz), 7.31 (2H, d, J=8 Hz), 8.01 (2H, d, J=8 Hz), 9.55 (1H, d, J=2 Hz).

Ethyl 4-(2-aminothiazol-5-ylmethyl)benzoate (20) was synthesized from 19 in the same manner as described above for 12. Yield, quantitative, colorless prisms; mp  $108-110\,^{\circ}\text{C}$  (benzene-hexane). H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.38 (3H, t, J=7 Hz), 4.01 (2H, s), 4.36 (2H, q, J=7 Hz), 4.50-5.00 (2H, br), 6.81 (1H, s), 7.28 (2H, d, J=8 Hz), 7.90 (2H, d, J=8 Hz). Anal. Calcd for C<sub>13</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>S: C, 59.52; H, 5.38; N, 10.68. Found: C, 59.52; H, 5.41; N, 10.07.

Ethyl 4-(2-Chlorothiazol-5-ylmethyl)benzoate (21)—A solution of 20 (18.8 g) in MeCN (80 ml) was added dropwise to a mixture of CuCl<sub>2</sub> (11.6 g, 86.1 mmol) and *tert*-butyl nitrite (10.35 g, 101 mmol) in MeCN (200 ml) at 55—65 °C. The mixture was stirred for 20 min, 15% HCl (150 ml) was added under ice-cooling, and the mixture was extracted with CHCl<sub>3</sub>. The extract was washed, dried, and concentrated *in vacuo*. The residue was purified by silica gel chromatography to give 21 (12.1 g, 68.5%) as an oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.39 (3H, t, J=7 Hz), 4.13 (2H, s), 4.37 (2H, q, J=7 Hz), 7.31 (2H, d, J=8 Hz), 8.01 (2H, d, J=8 Hz).

Compound 22 was obtained in the same manner as described above for 2d. Yield 81.8%, a pale yellow oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.38 (3H, t, J=7 Hz), 4.23 (2H, s), 4.37 (2H, q, J=7 Hz), 7.28 (2H, d, J=8 Hz), 7.65 (1H, s), 8.00 (2H, d, J=8 Hz), 8.69 (1H, s).

5-[4-(Hydroxymethyl)benzyl]thiazole (23)—A solution of 22 (8.49 g, 34.3 mmol) in tetrahydrofuran (THF) (60 ml) was added to a suspension of LiAlH<sub>4</sub> (1.3 g, 34.3 mmol) in THF (30 ml). The mixture was stirred at room temperature for 5 h, then water (2 ml), 15% NaOH (2 ml) and water (6 ml) were added successively. The insoluble material was filtered off, and the filtrate was concentrated *in vacuo*. The residue was extracted with CHCl<sub>3</sub>. The extract was washed, dried, and concentrated *in vacuo* to give 23 as an oil (6.3 g, 89.4%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 3.25—3.75 (1H, br), 4.15 (2H, s), 4.64 (2H, s), 7.17 (2H, d, J=8 Hz), 7.30 (2H, d, J=8 Hz), 7.58 (1H, s), 8.60 (1H, s).

The crude 23 obtained above was used for the next reaction without further purification.

4-(Thiazol-5-ylmethyl)benzaldehyde (24) was prepared from 23 in a fashion analogous to that used for 10.

Ethyl (*E*)-2-Methyl-3-[4-(thiazol-5-ylmethyl)phenyl]-2-propenoate (26)—A mixture of 24 (1.0 g, 5.0 mmol) and 25<sup>13)</sup> (1.51 g, 2.78 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (25 ml) was stirred at room temperature for 2 h. The mixture was concentrated *in vacuo*. The residue was purified by silica gel chromatography to give 26 as a pale yellow oil (0.79 g, 66%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 1.34 (3H, t, J=7 Hz), 2.11 (3H, d, J=1.3 Hz), 4.20 (2H, s), 4.27 (2H, q, J=7 Hz), 7.22 (2H, d, J=8 Hz), 7.37 (2H, d, J=8 Hz), 7.67 (2H, s), 8.68 (1H, s).

3-[4-(Thiazol-5-ylmethyl)phenyl]-2-propenoic Acid (27)—A mixture of 24 (0.2 g, 1.0 mmol) and malonic acid (0.104 g, 1.0 mmol) in pyridine (0.3 ml) was heated at 100 °C for 4 h. After cooling of the mixture, dilute NH<sub>4</sub>OH was added, and insoluble material was filtered off. The filtrate was made weakly acidic with concentrated HCl. The precipitate was filtered, washed with water, and recrystallized from 50% EtOH to give 27 (0.13 g, 53.9%) as a colorless powder.  $^{1}$ H-NMR (CDCl<sub>3</sub>)  $\delta$ : 4.21 (2H, s), 6.43 (1H, d, J=16 Hz), 7.25 (2H, d, J=8 Hz), 7.52 (2H, d, J=8 Hz), 7.67 (1H, s), 7.76 (1H, d, J=16 Hz), 8.72 (1H, s).

2-Methyl-3-[4-(thiazol-5-ylmethyl)phenyl]-2-propenoic acid (28) was prepared from 26 in the same manner as described for 14a. Yield: 47.7%; colorless powder.  $^1$ H-NMR (CDCl<sub>3</sub>)  $\delta$ : 2.14 (3H, d, J=1.3 Hz), 4.21 (2H, s), 7.26 (2H, d, J=8 Hz), 7.41 (2H, d, J=8 Hz), 7.69 (1H, s), 7.79 (1H, s), 8.73 (1H, s).

**Biological Assays for Inhibition of TXA<sub>2</sub> Synthesis**—(a) In Vitro Assay of the Inhibition of TXA<sub>2</sub> Production in PRP: Citrated PRP  $(4 \times 10^7 \text{ platelets})$  from rats was preincubated with the test compound for 1 min at room

temperature with gentle shaking, and then sodium arachidonate (final concentration was  $0.5\,\mathrm{mm}$ ) was added to initiate the reaction. The reaction mixture (total volume  $0.1\,\mathrm{ml}$ ) was incubated for  $5\,\mathrm{min}$  at room temperature with vigorous shaking and indomethacin (final concentration  $0.1\,\mathrm{mm}$ ) was added to stop the reaction. Then the mixture was centrifuged at  $1000\times g$  for  $5\,\mathrm{min}$  and the supernatant was subjected to the measurement of  $TXB_2$  (the stable breakdown product of  $TXA_2$ ) and  $PGE_2$  by the radioimmunoassay method.

(b) Calculation of the Conversion Ratio ( $PGE_2/TXA_2$ ): The conversion ratio ( $PGE_2/TXA_2$ ) was used to estimate the selectivity of the inhibition of  $TXA_2$  synthetase. The conversion ratio was calculated at a concentration of the tested compound that gave about 90% inhibition of the  $TXB_2$  production in (a) according to the following equation.

the conversion ratio (PGE<sub>2</sub>/TXA<sub>2</sub>) = (the amount of the increase in PGE<sub>2</sub> production compared to the control)/ (the amount of the decrease in TXB<sub>2</sub> production compared to the control)

#### References and Notes

- 1) M. Tada, T. Kuzuya, M. Inoue, K. Kodama, M. Mishima, M. Yamada, M. Inui, and H. Abe, *Circulation*, 64, 1107 (1981).
- 2) K. Iizuka, K. Akahane, D. Momose, M. Nakazawa, T. Tanouchi, M. Kawamura, I. Ohyama, I. Kajiwara, Y. Iguchi, T. Okada, K. Taniguchi, T. Miyamoto, and M. Hayashi, J. Med. Chem., 24, 1139 (1981).
- 3) K. Iizuka, K. Akahane, D. Momose, M. Nakazawa, T. Tanouchi, M. Kawamura, I. Ohyama, I. Kajiwara, Y. Iguchi, T. Okada, K. Taniguchi, T. Miyamoto, and M. Hayashi, J. Med. Chem., 24, 1149 (1981).
- 4) P. E. Cross, R. P. Dickinson, M. J. Parry, and M. J. Randall, J. Med. Chem., 28, 1427 (1985).
- 5) P. E. Cross, R. P. Dickinson, M. J. Parry, and M. J. Randall, J. Med. Chem., 29, 342 (1986).
- 6) N. F. Ford, L. J. Browne, T. Cambell, C. Gemenden, R. Goldstein, C. Gude, and J. W. F. Wasley, J. Med. Chem., 28, 164 (1985).
- 7) K. Kato, S. Ohkawa, S. Terao, Z. Terashita, and K. Nishizaki, J. Med. Chem., 28, 287 (1985).
- 8) R. N. Prasad, H. H. Stein, and K. R. Tietje, Ger. Patent 2615406 (1976) [Chem. Abstr., 86, 5333w (1977)].
- 9) A. V. Dombrovskii, A. M. Yurkevich, and A. Terentiev, Zh. Obshch. Khim., 27, 3047 (1957) [Chem. Abstr., 52, 8087b (1958)].
- 10) a) P. Needleman, A. Wyche, and A. Raz, J. Clin. Invest., 63, 345 (1979); b) M. J. Randall, J. Michael, M. J. Parry, E. Hawkeswood, P. E. Cross, and R. P. Dickinson, Thromb. Res., 23, 145 (1981); c) K. Yamaki and S. Ohishi, Chem. Pharm. Bull., 34, 3526 (1986); d) N. Kayama, K. Sakaguchi, S. Kaneko, T. Kubota, T. Fukuzawa, S. Kawamura, and T. Yoshimoto, Prostaglandins, 21, 543 (1981).
- 11) G. Bruno and J. Cahn, Fr. Patent 2555581 (1985) [Chem. Abstr., 104, P68860b (1986)].
- 12) H. Demarne, R. Filhol, and M. Mosse, Fr. Demande Patent 2550785 (1985) [Chem. Abstr., 103, P123165k (1985)].
- 13) O. Isler, H. Gutmann, M. Montavon, R. Ryegg, G. Ryser, and P. Zeller, Helv. Chim. Acta, 40, 1242 (1957).