

\$0957-4166(96)00004-3

## First Enantiospecific Syntheses of Crotepoxide and *iso*-Crotepoxide from (-)-Quinic Acid

Tony K. M. Shing\* and Eric K. W. Tam

Department of Chemistry, The Chinese University of Hong Kong, Shatin, Hong Kong.

Abstract: The optically active crotepoxide 1 and *iso*-crotepoxide 2 have been constructed from quinic acid involving a singlet oxygen photooxygentation as the key step.

Recently, the chemotherapeutic potential of glycosidase inhibitors<sup>1</sup> as anti-HIV,<sup>2</sup> anti-metastasis,<sup>3</sup> and antihyperglycemic agents<sup>4</sup> has aroused considerable attention from the synthetic chemists. Crotepoxide 1, a cyclohexane oxide<sup>5</sup> isolated from the fruits of *Croton macrostachys*<sup>6</sup> and of *Piper futokadzura*,<sup>7</sup> has been shown to display significant tumour-inhibitory activity against Lewis lung carcinoma in mice (LL) and Walker intramuscular carcinosarcoma in rats (WM).<sup>6a</sup> The structures of crotepoxide 1 and its diastereoisomer, *iso*crotepoxide 2, are related to those of (1*R*,6*S*)-cyclophellitol 3 and cyclophellitol 4, respectively. (1*R*,6*S*)cyclophellitol 3<sup>8,3b</sup> and cyclophellitol 4<sup>9</sup> have been shown to be potent  $\beta$ -D- and  $\alpha$ -D-glucosidase inhibitors, respectively, presumably attributable to the structural resemblance with  $\beta$ -D- and  $\alpha$ -D-glucose. Along the same vein of reasoning, tumour inhibitor crotepoxide 1 might reveal  $\alpha$ -D-glucosidase inhibition and *iso*-crotepoxide 2 might inhibit  $\beta$ -D-mannosidase. The presence of ester groupings in 1 and in 2 may be advantageous because the anti-HIV activity of  $\alpha$ -D-glucosidase inhibitors such as castanospermine and 1-deoxynojirimycin improved significantly by increasing the lipophilicity of the compounds *via* esterification of the hydroxy groups.<sup>2b,10</sup>



Since the glycosidase inhibitory activity of crotepoxide 1 has not been studied, a successful construction of enantiopure crotepoxide and related cyclohexane oxides would permit extensive biological evaluation. Hitherto there has been only one report on the synthesis of optically pure crotepoxide from a chemically resolved Diels-Alder adduct of furan and acrylic acid.<sup>11</sup> Our endeavours in natural and non-natural product synthesis from (-)-quinic acid 5 have already furnished anti-tumour agent 2-crotonyloxymethyl-(4R, 5R, 6R)-4, 5, 6-trihydroxycyclohex-2-enone (COTC),<sup>12</sup> pseudo- $\beta$ -D-mannopyranose, pseudo- $\beta$ -D-fructopyranose,<sup>13</sup> pseudo- $\alpha$ -D-glucopyranose, pseudo- $\alpha$ -D-mannopyranose,<sup>14</sup> glycosidase inhibitors cyclophellitol and its diastereoisomers,<sup>15</sup> and moreover valiolamine and its diastereoisomers.<sup>16</sup> In continuation with our investigation on the preparation of potential glycosidase inhibitors, we now disclose the first enantiospecific syntheses of 1 and 2, and hence further demonstrate the versatility of quinic acid in the fabrication of heavily oxygenated cyclohexanoid natural products.

Our previous work<sup>13,15</sup> has shown that quinic acid **5** could be readily converted into the diol **6** in four steps with an overall yield of 64.5% (Scheme 1). Regioselective benzoylation<sup>17</sup> at the primary hydroxy group in **6** followed by silylation of the remaining alcohol afforded the silyl benzoate **7**.<sup>18</sup> The double bond in **7** was subjected to our recently developed ruthenium catalyzed flash dihydroxylation<sup>19</sup> protocol at the less hindered  $\beta$ -face to give, exclusively, the desired  $\beta$ -diol **8**.<sup>18</sup> Selective acetylation of the secondary hydroxy group in **8** gave the monoacetate **9**. Thionyl chloride<sup>20</sup> mediated elimination of the tertiary alcohol in **9** followed by selective hydrolysis of the cyclohexylidene ketal furnished the ene-diol **10**. Corey-Winter<sup>21</sup> deoxygenation of the vicinal diol moiety in **10** provided the diene **11** that now possessed the functionality required for the bisoxirane formation *via* a singlet oxygen oxidation reaction.<sup>22</sup>



Scheme 1. Syntheses of dienes 11, 12, and 13: a) 4 steps, (64.5%), see ref. 13, 15; b) benzoyl chloride, collidine, CH<sub>2</sub>Cl<sub>2</sub>, room temp. (82%); c) *tert*-butylchlorodimethylsilane (*t*BuMe<sub>2</sub>SiCl), imidazole, 4-dimethylaminopyridine (DMAP), CH<sub>2</sub>Cl<sub>2</sub>, room temp. (97%); d) RuO4, NaIO4, H<sub>2</sub>O : CH<sub>3</sub>CN : EtOAc (1:3:3), 0 °C (75%); e) Ac<sub>2</sub>O, pyridine (pyr), DMAP, CH<sub>2</sub>Cl<sub>2</sub>, room temp. (97%); f) SOCl<sub>2</sub>, pyr, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to room

temp. (81%); g) 50% aqueous CF<sub>3</sub>CO<sub>2</sub>H, CH<sub>2</sub>Cl<sub>2</sub>, room temp. (80%); h) 1,1'-thiocarbonyldiimidazole, toluene, reflux, then P(OMe)<sub>3</sub>, reflux (68%); i) 48% aqueous HF, CH<sub>3</sub>CN (80%).

Photo-oxygenation of the diene moiety in 11 proceeded at the sterically less hindered  $\beta$ -face, giving the  $\beta$ -endoperoxide 14 as the preponderant product ( $\beta$ -endoperoxide 14 :  $\alpha$ -endoperoxide 15 = 54 : 1) as shown in Scheme 2. The use of *tert*-butyldimethyl silyl ether as the stereodirecting group in singlet oxygen oxidation reaction was therefore highly efficient. The isomeric endoperoxides were easily separable on silica chromatography. The photo-oxygenation reaction<sup>22</sup> of the desilylated 11, i.e., the diene-alcohol 12, was also examined and the ratio of  $\beta$ - 16 to  $\alpha$ -endoperoxide 17 was 3 : 2. It is noteworthy that the diacetate 13 did not afford any observable reaction with singlet oxygen. Subjection of the silylated  $\beta$ -endoperoxide 18 in essentially quantitative yield.<sup>18</sup> Desilylation of 18 afforded alcohol 20 that was identical to the product from the same rearrangement reaction<sup>23</sup> of  $\beta$ -1,4-endoperoxide 16. Acetylation of the free alcohol in 20 then yielded the target molecule crotepoxide 1,<sup>24</sup> m.p. 146–148 °C (Et<sub>2</sub>O/hexanes) (lit.<sup>6a</sup> m.p. 150–151 °C);  $[\alpha]^{26}D = + 71.9 (c = 0.6, CHCl_3) {lit.<sup>6a</sup> [<math>\alpha$ ]<sup>25</sup>D = + 74 (c = 1.7, CHCl<sub>3</sub>)}. Likewise reactions of  $\alpha$ -endoperoxide 15 ( $\rightarrow$  19 $\rightarrow$  21 $\rightarrow$  2) or 17 ( $\rightarrow$  21 $\rightarrow$  2) furnished, for the first time, optically active *iso*-crotepoxide 2, oil;  $[\alpha]^{27}D = - 35.8 (c = 0.67, CHCl_3).<sup>18,24</sup>$ 



Scheme 2. Syntheses of 1 and 2: a) O<sub>2</sub>, hv, tetraphenylporphyrin (TPP), CCl4, 0 °C (14 : 15, *ca.* 54 : 1, 80% from 11), (16 : 17, *ca.* 3 : 2; 80% from 12); b) cobalt-*meso*-tetraphenylporphyrin (CoTPP), CHCl<sub>3</sub>, 0 °C (98%); c) HF-pyridine, THF, (85%); d) Ac<sub>2</sub>O, pyr, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, room temp. (98%).

In summary, we have described facile and efficient syntheses of enantiopure crotepoxide 1 and its diastereoisomer 2 from (–)-quinic acid and application of this flexible strategy to the fabrication of other cyclohexanoid natural products including boesenoxide, senepoxide,  $\beta$ -senepoxide, pipoxide, and tingtanoxide is underway.<sup>5</sup>

We thank the Croucher Foundation for financial support.

## REFERENCES

- a) B. Winchester, G. W. J. Fleet, *Glycobiology*, **1992**, 2, 199; b) G. W. J. Fleet, L. E. Fellows in *Natural Product Isolation* (Eds.: G. H. Wagman, R. Cooper), Elsevier, Amsterdam, **1988**, p. 540; c) G. Legler, *Adv. Carbohydr. Chem. Biochem.* **1990**, 48, 319.
- a) M. J. Humphries, K. Matsumoto, S. L. White, K. Olden, Proc. Natl. Acad. Sci. U.S.A. 1986, 83, 1752; b) G. W. J. Fleet, A. Karpas, R. A. Dwek, L. E. Fellows, A. S. Tyms, S. Petursson, S. K. Namgoong, N. G. Ramsden, P. W. Smith, J. C. Son, F. Wilson, D. R. Witty, G. S. Jacob, T. W. Rademacher, FEBS Lett. 1988, 237, 128 and references cited therein; c) D. C. Montefoiori, W. E. Robinson, W. M. Mitchell, Proc. Natl. Acad. Sci. U.S.A. 1988, 85, 9248.
- a) M. J. Humphries, K. Matsumoto, S. L. White, K. Olden, *Cancer Res.* 1986, 46, 5215; b) S. Atsumi, C. Nosaka, Y. Ochi, H. Iinuma, K. Umezawa, *Cancer Res.* 1993, 53, 4896.
- 4. a) J. Arends, B. H. L. Willms, Horm. Metab. Res. 1986, 18, 761; b) S. Horii, H. Fukase, T. Matsuo, Y Kameda, N. Asano, K. Matsui, J. Med. Chem. 1986, 29, 1038.
- 5. a) For an excellent review on natural cyclohexane oxides, see C. Thebtaranonth and Y. Thebtaranonth, *Acc. Chem. Res.* **1986**, *19*, 84; b) M. Balcì, Y. Sütbeyaz, H. Secen, *Tetrahedron* **1990**, *46*, 3715.
- a) S. M. Kupchan, R. J. Hemingway, P. Coggon, A. T. Mcphail, J. Am. Chem. Soc. 1968, 90, 2982;
  b) S. M. Kupchan, R. J. Hemingway, R. M. Smith, J. Org. Chem. 1969, 34, 3898.
- 7. S. Takahashi, Phytochemistry 1969, 8, 321.
- 8. K. Tatsuta, Y. Niwata, K. Umezawa, K. Toshima, M. Nakata, J. Antibiot. 1991, 44, 912.
- 9. S. Atsumi, K. Umezawa, H. Iinuma, H. Naganawa, H. Nakamura, Y. Iitaka, T. Takeucki, J. Antibiot. 1990, 43, 49.
- a) P. S. Sunkara, D. L. Taylor, M. S. Kang, T. L. Bowlin, P. S. Liu, A. S. Tyms, A. Sjoerdsma, Lancet, 1989, 1206; b) A. Karpas, G. W. J. Fleet, R. A. Dwek, S. Petursson, S. K. Namgoong, N. G. Ramsden, G. S. Jacob, T. W. Rademacher, Proc. Natl. Acad. Sci. U.S.A. 1988, 85, 9229; J. R. Behling, A. L. Campbell, K. A. Babiak, J. S. Ng, J. Medich, P. Farid, G. W. J. Fleet, Tetrahedron, 1993, 49, 3359.
- 11. S. Ogawa, T. Takagaki, Bull. Chem. Soc. Jpn. 1987, 60, 800.
- 12. T. K. M. Shing, Y. Tang, J. Chem. Soc. Chem. Commun. 1990, 312; Tetrahedron 1990, 46, 6575.
- 13. T. K. M. Shing, Y. Tang, J. Chem. Soc. Chem. Commun. 1990, 748; Tetrahedron 1991, 47, 4571.
- 14. T. K. M. Shing, Y.-X. Čui, Y. Tang, J. Chem. Soc. Chem. Commun. 1991, 756; Tetrahedron 1992, 48, 2349.
- 15. T. K. M. Shing, V. W.-F. Tai, J. Chem. Soc. Chem. Commun. 1993, 995; J. Chem. Soc. Perkin Trans. 1 1994, 2017.
- 16. T. K. M. Shing, L. H. Wan, Angew. Chem. Int. Ed. Engl. 1995, 34, 1643.
- 17. K. Ishihara, H. Kurihara, H. Yamamoto, J. Org. Chem. 1993, 58, 3791.
- 18. All new compounds gave satisfactory analytical and spectral data.
- 19. T. K. M. Shing, V. W.-F. Tai, E. K. W. Tam, Angew. Chem. Int. Ed. Engl. 1994, 33, 2312.
- 20. A. Schwartz, P. Madan, J. Org. Chem. 1986, 51, 5463.
- 21. E. J. Corey, R. A. E. Winter, J. Am. Chem. Soc. 1963, 85, 2677.
- 22. Y. Sütbeyaz, H. Secen, M. Balci, J. Chem. Soc. Chem. Commun. 1988, 1330.
- 23. J. D. Boyd, C. S. Foote, D. K. Imagawa, J. Am. Chem. Soc. 1980, 102, 3641.
- 24. Selected spectral data: for 1, <sup>1</sup>H-NMR (250 Hz, CDCl<sub>3</sub>)  $\delta$  = 8.02 (d, 2H, *J* = 7.0 Hz), 7.57 (t, 1H, *J* = 7.6 Hz), 7.47 (t, 2H, *J* = 7.1 Hz), 5.71 (d, 1H, *J* = 9.0 Hz), 4.99 (dd, 1H, *J* = 1.6, 9.0 Hz), 4.58 (d, 1H, *J* = 12.1 Hz), 4.24 (d, 1H, *J* = 12.1 Hz), 3.67 (d, 1H, *J* = 2.7 Hz), 3.46 (dd, 1H, *J* = 2.7, 3.9 Hz), 3.11 (dd, 1H, *J* = 1.6, 3.9 Hz), 2.13 (3, 3H), 2.03 (s, 3H). For 2, <sup>1</sup>H-NMR (250 Hz, CDCl<sub>3</sub>)  $\delta$  = 8.02 (d, 2H, *J* = 7.05 Hz), 7.60 (t, 1H, *J* = 7.1 Hz), 7.47 (t, 2H, *J* = 6.4 Hz), 5.36 (s, 2H), 4.39 (ABq, 2H, *J* = 12.3 Hz), 3.57 (dd, 1H, *J* = 2.7, 4.0 Hz), 3.50 (d, 1H, *J* = 2.7 Hz), 3.42 (d, 1H, *J* = 4.0 Hz), 2.11 (3, 3H), 2.00 (s, 3H).

(Received in Japan 9 November 1995)