

# Neighboring group participation Part 16. Stereoselective synthesis and receptor-binding examination of the four stereoisomers of 16-bromomethyl-3,17-estradiols<sup>☆</sup>

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Dedicated to Professor Dr. András Lipták on the occasion of his 70th birthday.

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#### ABSTRACT

The four possible isomers of 3-benzyloxy-16-hydroxymethylestra-1,3,5(10)-trien-17-ol (1a-4a) with proven configurations were converted into the corresponding 3-benzyloxy-16bromomethylestra-1,3,5(10)-triene-3,17-diols (5e-8e). Depending on the reaction conditions the cis isomers of 3-benzyloxy-16-hydroxymethylestra-1,3,5(10)-trien-17-ol (1a and 2a) were transformed into 3-benzyloxy-16-bromomethylestra-1,3,5(10)-trien-17-yl acetate (5b and 6b) or 16-bromomethyl-3-hydroxyestra-1,3,5(10)-trien-17-yl acetate (5c and 6c) on treatment with HBr and acetic acid. The mechanism of the process can be interpreted as involving front-side neighboring group participation. Under similar experimental conditions, the trans isomers (3a and 4a) yielded only 3-benzyloxy-16-acetoxymethylestra-1,3,5(10)-trien-17-yl acetates (3b and 4b) or 16-acetoxymethylestra-1,3,5(10)-triene-3,17-diyl diacetates (3d and 4d). Both the cis (1a and 2a) and the trans (3a, and 4a) isomers were transformed into 16-bromomethylestra-1,3,5(10)-trien-17-ol (5a-8a) by the Appel reaction on treatment with CBr<sub>4</sub>/Ph<sub>3</sub>P. Debenzylation of **5a-8a** was carried out with HBr and acetic acid to yield **5e-8e**. The debenzylation process in the presence of acetic anhydride produces the diacetates 5d-8d. The structures of the compounds were determined by means of MS, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic methods. Compounds **5c–8c** and **5e–8e** were tested in a radioligandbinding assay. Except for the affinity of 7e for the estrogen receptor ( $K_i = 2.55$  nM), the affinities of the eight compounds (5c-8c and 5e-8e) for the estrogen, androgen and progesterone receptors are low ( $K_i > 0.55$ , 0.52 and 0.21  $\mu$ M, respectively).

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# 1. Introduction

On the basis of the observations that more than two-thirds of breast cancers occur in post-menopausal women, and that

<sup>\*</sup> For a preceding paper in this series, see ref. [1].

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their proliferation and survival depend on the estrogens, there are now two major options for the treatment of estrogendependent breast cancers [2]. One is the use of aromatase inhibitors, which reduces the local in situ formation of estrogens. The other is the use of anti-estrogens, which block the interaction of estrogens with their specific receptor (estrogen receptor, ER) [3]. Synthetic non-steroidal compounds are significant anti-estrogens which are routinely used in the endocrine therapy of breast cancer [2]. Tamoxifen behaves as a mixed agonist/antagonist of estrogen action, thus limiting its therapeutic potential [4]. Attempts to improve the therapeutic efficacy have been focused on creating estradiol analogs with anti-estrogenic activity. It is known from the literature that halogen-containing estrogens are of great importance in the diagnosis [5,6] and chemotherapy [4] of estrogen receptor-positive human breast cancers [7]: the strongly electronegative halogens onto a sterane skeleton can enhance the receptor affinity, while at the same time it may reduce the tumorigenicity, too. Earlier examinations suggested that the ER tolerates reasonably large substituents at positions C-7 $\alpha$ , C-11 $\beta$ , C-16 $\alpha$  and C-17 $\alpha$  in estradiol [8,9]. Moreover, Poirier and co-workers reported that  $16\alpha$ -(3-halopropyl)estradiol and 16a-bromoalkylamidoestradiol derivatives inhibit 17βhydroxysteroid dehydrogenase, which is responsible for the interconversion of the less potent estrone into the most potent estrogen, estradiol, in the target tissues [10,11]. On the other hand, Heiman et al. [12] and Katzenellenbogen and co-workers [13] reported that halogenation of ring D of the steroid nucleus, especially at position C-16 $\alpha$ , results in compounds with higher affinities for lamb and rat uterine ERs than that of 17βestradiol itself. In consequence of the above observations, efforts have been directed toward the synthesis of estradiol derivatives with a halomethyl group at C-16, which may be hormonally inactive or at least may have a favorable ratio of tumor inhibitory to hormonal activity.

Herein, we report the synthesis of the four possible 16bromomethyl-3-hydroxyestra-1,3,5(10)-trien-17-yl acetates (5c-8c) and the four possible 16-bromomethylestradiol derivatives (5e-8e) from the corresponding 3-benzyloxy-16hydroxymethylestra-1,3,5(10)-triene-3,17-diols (1a-4a). The starting materials (1a-4a) were synthesized from estrone according to a recently reported method [14]. The receptorbinding examinations on the synthesized new compounds were carried out on the ERs, androgen receptors (ARs) and progesterone receptors (PRs). We wished to obtain answers to the following questions: (1) How do the receptor binding properties/processes of these eight derivatives (5c-8c and 5e-8e) differ? (2) How is the receptor-binding influenced by the bromomethyl group at position C-16 and the acetoxy group at position C-17 in ring D of the estrane skeleton?

# 2. Experimental

# 2.1. General

Melting points were determined with a Kofler hot-stage apparatus and are uncorrected. The reactions were monitored by TLC on Kieselgel-G (Merck Si 254 F) layers (0.25 mm thick); solvent system (ss): (A) ethyl acetate/CHCl<sub>3</sub> (2.5:97.5, v/v); (B) ethyl acetate/CHCl<sub>3</sub> (5:95, v/v); (C) ethyl acetate/CHCl<sub>3</sub> (10:90, v/v). The spots were detected by spraying with 5% phosphomolybdic acid in 50% aqueous phosphoric acid. The R<sub>f</sub> values were determined for the spots observed by illumination at 254 and 365 nm. Flash chromatography: silica gel 60, 40–63 μm (Merck). All solvents were distilled prior to use. Specific rotations were measured in CHCl<sub>3</sub> (c 1.00) or ethyl acetate (c 1.00) at 25  $^{\circ}$ C with a Polamat-A (Zeiss-Jena) polarimeter and are given in units of  $10^{-1\circ}$  cm<sup>2</sup> g<sup>-1</sup>. Elemental analyses were performed with a Perkin-Elmer CHN analyzer model 2400. <sup>1</sup>H NMR spectra were obtained in CDCl<sub>3</sub> or DMSO-d<sub>6</sub> solution at 500 MHz (Bruker DRX 500), and the <sup>13</sup>C NMR spectra were recorded at 125 MHz on the same instrument under the same conditions. Chemical shifts ( $\delta$ ) are reported relative to TMS, and are given in ppm; the coupling constants (J) are given in Hz.  $^{13}\mathrm{C}\,\mathrm{NMR}$  spectra are <sup>1</sup>H-decoupled. Mass spectra were measured on a Varian MAT 311A spectrometer.

#### 2.2. Conversion of 3-benzyloxy-16-

# hydroxymethylestra-1,3,5(10)-trien-17-ol (1a–4a) and 3-benzyloxy-16-bromomethylestra-1,3,5(10)-trien-17-ol isomers (7a and 8a) with HBr in acetic acid

#### 2.2.1. General procedures

2.2.1.1. Method A. A solution of 1.18 g (3 mmol) of 3-benzyloxy-16-hydroxymethylestra-1,3,5(10)-trien-17-ol (**1a–4a**) [14] in 24 ml of anhydrous acetic acid was cooled to 10 °C and 2.4 ml of 33% HBr (in acetic acid) was added. The reaction mixture was diluted with water after stirring for 1 h. The precipitate was collected by filtration and dissolved in  $CH_2Cl_2$ , and the solution was washed with water and dried over  $Na_2SO_4$ . After evaporation in vacuo, the crude product was purified by column chromatography (silica gel, CHCl<sub>3</sub> or ethyl acetate/CHCl<sub>3</sub>: 2.5:97.5).

2.2.1.2. Method B. According to procedure A, 1.18 g(3 mmol) of compound **1a–4a**, or 1.78 g (3 mmol) of compounds **7a–8a** and 2.4 ml of 33% HBr (in acetic acid) were allowed to react for 12 h. The reaction mixture was diluted with water, the precipitate was collected by filtration and dissolved in  $CH_2Cl_2$ , and the solution was washed with water and dried over  $Na_2SO_4$ . After evaporation in vacuo, the crude product was purified by column chromatography (silica gel, ethyl acetate/CHCl<sub>3</sub>: 2.5:97.5).

2.2.1.3. Method C. According to procedure A, 1.18 g (3 mmol) of compounds **1a–4a**, or 1.78 g (3 mmol) **7a–8a** and 2.4 ml of 33% HBr (in acetic acid) were allowed to react for 12 h. Next, 1 ml of acetic anhydride was added, and the reaction mixture was diluted with water after stirring for 6 h. The precipitate was collected by filtration and dissolved in CH<sub>2</sub>Cl<sub>2</sub>, and the solution was washed with water and dried over Na<sub>2</sub>SO<sub>4</sub>. After evaporation in vacuo, the crude product was purified by column chromatography (silica gel, ethyl acetate/CHCl<sub>3</sub>: 2.5:97.5).

2.2.2. Reaction of 3-benzyloxy-16 $\beta$ -hydroxymethylestra-1,3,5(10)-trien-17 $\beta$ -ol (1a) with HBr in acetic acid 2.2.2.1. Method A.

2.2.2.1.1. 3-Benzyloxy-16 $\beta$ -bromomethylestra-1,3,5(10)trien-17 $\beta$ -yl acetate (5b) and 3-benzyloxy-16 $\beta$ -acetoxymethylestra-1,3,5(10)-trien-17 $\beta$ -yl acetate (1b). Compound **5b** (1.24 g, 83%). Mp 144–146 °C;  $R_f = 0.65$  (ss A);  $[\alpha]_D^{20} + 54$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 67.55; H, 6.75. C<sub>28</sub>H<sub>33</sub>O<sub>3</sub>Br requires: C, 67.60; H, 6.69%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.84 (s, 3H, 18-H<sub>3</sub>), 2.10 (s, 3H, OAc-CH<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 3.27 (t, 1H, J = 10.0 Hz) and 3.46 (dd, 1H, J = 10.0 Hz, J = 5.8 Hz): 16a-H<sub>2</sub>, 4.86 (d, 1H, J=10.0 Hz, 17-H), 5.02 (s, 2H, 3-benzyl-CH<sub>2</sub>), 6.71 (d, 1H, J=2.5 Hz, 4-H), 6.76 (dd, 1H, J=8.6 Hz, J=2.5 Hz, 2-H), 7.17 (d, 1H, J=8.6 Hz, 1-H), 7.31 (t, 1H, J=7.3 Hz, 4'-H), 7.36 (t, 2H, J=7.3 Hz, 3',5'-H), 7.41 (d, 2H, J=7.3 Hz, 2',6'-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 13.2 (C-18), 20.9 (OAc-CH<sub>3</sub>), 26.1, 27.3, 29.7, 32.6, 35.2, 37.5, 38.0, 41.5, 43.7, 44.0 (C-13), 48.2, 70.0 (3-benzyl-CH<sub>2</sub>), 82.3 (C-17), 112.4 (C-2), 114.9 (C-4), 126.3 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.4 (C-1'), 137.8 (C-5), 156.9 (C-3), 170.6 (OAc-CO). EI-MS (70 eV) m/z (%): 498 (24) [M<sup>+</sup> + 2], 496 (23) [M<sup>+</sup>], 91 (100), 43 (8). **1b** (157 mg 11%). Mp 100–102 °C ([14], Mp 100–101 °C);  $R_{\rm f}$  = 0.40 (ss A);  $[\alpha]_D^{20} + 37$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 75.48; H, 7.52.  $C_{30}H_{36}O_5$  requires: C, 75.60; H, 7.61%).  $^1H$  NMR (CDCl<sub>3</sub>)  $\delta,$  ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.02 and 2.06 (2  $\times$  s, 2  $\times$  3H, 2  $\times$  OAc-CH<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 4.02 (dd, 1H, J = 11.0 Hz, J = 7.5 Hz) and 4.12 (dd, 1H, J=11.0 Hz, J=6.9 Hz): 16a-H<sub>2</sub>, 4.89 (d, 1H, J=10.1 Hz, 17-H), 5.01 (s, 2H, 3-benzyl-CH<sub>2</sub>), 6.70 (d, 1H, J=2.3Hz, 4-H), 6.76 (dd, 1H, J=8.5 Hz, J=2.3 Hz, 2-H), 7.17 (d, 1H, J=8.5 Hz, 1-H), 7.29 (t, 1H, J=7.3 Hz, 4'-H), 7.35 (t, 2H, J=7.3 Hz, 3',5'-H), 7.41 (d, 2H, J = 7.3 Hz, 2',6'-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.9 (C-18), 20.8 (2C, 2 × OAc-CH<sub>3</sub>), 26.1, 27.3, 29.3, 29.6, 37.4, 37.7, 38.0, 43.6 (C-13), 43.7, 48.8, 65.2 (C-16a), 70.0 (3-benzyl-CH<sub>2</sub>), 81.6 (C-17), 112.4 (C-2), 114.9 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.3 (C-1'), 137.8 (C-5), 156.8 (C-3), 170.7 (2C, 2 × OAc-CO); EI-MS (70 eV) m/z (%): 476 (60) [M<sup>+</sup>], 91 (100), 43 (8).

#### 2.2.2.2. Method B.

2.2.2.2.1. 16β-Bromomethyl-3-hydroxyestra-1,3,5(10)trien-17 $\beta$ -yl acetate (5c) and 16 $\beta$ -acetoxymethyl-3-hydroxyestra-1,3,5(10)-trien-17β-yl acetate (1c). Compound 5c (1.16 g, 75%). Mp 194–198 °C;  $R_f = 0.45$  (ss C);  $[\alpha]_D^{20} + 63$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 62.07; H, 6.54. C<sub>21</sub>H<sub>27</sub>O<sub>3</sub>Br requires: C 61.92; H 6.68%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.84 (s, 3H, 18-H<sub>3</sub>), 2.11 (s, 3H, OAc-CH<sub>3</sub>), 2.81 (m, 2H, 6-H<sub>2</sub>), 3.27 (t, 1H, J = 10.0 Hz) and 3.46 (dd, 1H, J=10.0 Hz, J=5.9 Hz): 16a-H<sub>2</sub>, 4.86 (d, 1H, J=10.0 Hz, 17-H), 6.56 (d, 1H, J=2.4 Hz, 4-H), 6.62 (dd, 1H, J=8.4Hz, J=2.4Hz, 2-H), 7.12 (d, 1H, J=8.4Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 13.2 (C-18), 20.9 (OAc-CH<sub>3</sub>), 26.1, 27.2, 29.5, 32.6, 35.2, 37.4, 38.0, 41.4, 43.7, 44.0 (C-13), 48.2, 82.3 (C-17), 112.8 (C-2), 115.3 (C-4), 126.4 (C-1); 132.3 (C-10), 138.0 (C-5), 153.5 (C-3), 170.8 (OAc-CO); EI-MS (70 eV) m/z (%): 408 (99) [M<sup>+</sup>+2], 406 (100) [M<sup>+</sup>], 267 (18), 213 (14), 172 (16), 159 (28), 133 (28), 43 (52). 1c (157 mg, 12%). Mp 184  $^{\circ}$ C; R<sub>f</sub> = 0.40 (ss C);  $[\alpha]_D^{20}$  + 46 (c 1.00 in CHCl<sub>3</sub>). (Found: C, 71.54; H, 7.65. C<sub>23</sub>H<sub>30</sub>O<sub>5</sub> requires: C 71.48; H 7.82%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.84 (s, 3H, 18-H<sub>3</sub>), 2.04 and 2.08 (2 × s, 2 × 3H, 2 × OAc-CH<sub>3</sub>), 2.81 (m, 2H, 6-H<sub>2</sub>), 4.02 (dd, 1H, J=11.0 Hz, J=7.6 Hz) and 4.13 (dd, 1H, J=11.0Hz, J=6.9Hz): 16a-H<sub>2</sub>, 4.89 (d, 1H, J=10.0Hz, 17-H), 6.57 (d, 1H, J=2.2 Hz, 4-H), 6.64 (dd, 1H, J=8.4 Hz, J=2.2 Hz, 2-H), 7.12 (d, 1H, J = 8.4 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.9 (C-18), 21.0 (2C, 2 × OAc-CH<sub>3</sub>), 26.1, 27.3, 29.5 (2C), 37.4, 37.6, 38.0, 43.6 (C-13), 43.7, 48.7, 65.4 (C-16a), 81.8 (C-17), 112.8 (C-2), 115.3 (C-4), 126.4 (C-1); 132.0 (C-10), 138.0 (C-5), 153.8 (C-3), 171.3 (2C, 2 × OAc-CO); EI-MS (70 eV) m/z (%): 386 (100) [M<sup>+</sup>], 266 (4), 213 (12), 172 (12), 159 (16), 133 (12), 106 (13), 43 (48).

#### 2.2.2.3. Method C.

2.2.2.3.1. 16β-Bromomethylestra-1,3,5(10)-triene-3,17βdiyl diacetate (5d) and  $16\beta$ -acetoxymethylestra-1,3,5(10)triene-3,17β-diyl diacetate (1d). Compound 5d (1.02 g, 76%). Mp 122–123.5 °C;  $R_f = 0.45$  (ss A);  $[\alpha]_D^{20} + 58$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 61.58; H, 6.67. C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>Br requires: C, 61.47; H, 6.50%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.84 (s, 3H, 18-H<sub>3</sub>), 2.10 (s, 3H, 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.86 (m, 2H, 6-H<sub>2</sub>), 3.26 (t, 1H, J = 10.0 Hz) and 3.46 (dd, 1H, J = 10.0 Hz, J = 5.8 Hz): 16a-H<sub>2</sub>, 4.86 (d, 1H, J=10.1 Hz, 17-H), 6.79 (d, 1H, J=2.2 Hz, 4-H), 6.84 (dd, 1H, J=8.5 Hz, J=2.2 Hz, 2-H), 7.25 (t, 1H, J=8.5 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 13.2 (C-18), 20.8 (17-OAc-CH<sub>3</sub>), 21.0 (3-OAc-CH<sub>3</sub>), 25.9, 27.1, 29.4, 32.6, 35.1, 37.4, 37.6, 41.5, 43.9, 44.0 (C-13), 48.2, 82.2 (C-17), 118.6 (C-2), 121.5 (C-4), 126.3 (C-1); 137.6 (C-10), 138.0 (C-5), 148.6 (C-3), 169.6 (3-OAc-CO), 170.6 (17-OAc-CO); EI-MS (70 eV) m/z (%): 450 (8) [M<sup>+</sup> + 2], 448 (8) [M<sup>+</sup>], 408 (100), 406 (100), 267 (12), 172 (9), 159 (14), 133 (14), 43 (41). **1d** (103 mg, 8%). Mp 150–152 °C;  $R_f = 0.50$  (ss B);  $[\alpha]_D^{20} + 44$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 69.95; H, 7.45. C<sub>25</sub>H<sub>32</sub>O<sub>6</sub> requires: C, 70.07; H, 7.53%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.02 and 2.07 ( $2 \times s$ ,  $2 \times 3H$ , 16a- and 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.86 (m, 2H, 6-H<sub>2</sub>), 4.02 (dd, 1H, J=11.0 Hz, J = 7.5 Hz) and 4.12 (dd, 1H, J = 11.0 Hz, J = 6.9 Hz): 16a-H<sub>2</sub>, 4.89 (d, 1H, J = 10.1 Hz, 17-H), 6.79 (d, 1H, J = 1.8 Hz, 4-H), 6.83 (dd, 1H, J=8.3Hz, J=1.8Hz, 2-H), 7.26 (d, 1H, J=8.3Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.8 (C-18), 20.8 (2C, 16a- and 17-OAc-CH<sub>3</sub>), 21.0 (3-OAc-CH<sub>3</sub>), 25.9, 27.1, 29.3, 29.4, 37.4, 37.6, 37.7, 43.5 (C-13), 43.8, 48.8, 65.1 (C-16a), 81.5 (C-17), 118.6 (C-2), 121.4 (C-4), 126.3 (C-1); 137.6 (C-10), 137.9 (C-5), 148.5 (C-3), 169.6 (3-OAc-CO), 170.7 (2C, 16a- and 17-OAc-CO); EI-MS (70 eV) m/z (%): 428 (11) [M<sup>+</sup>], 386 (100), 172 (4), 159 (6), 133 (5), 43 (16).

2.2.3. Conversion of 3-benzyloxy-16 $\alpha$ -hydroxymethylestra-1,3,5(10)-trien-17 $\alpha$ -ol (2*a*) with HBr in acetic acid 2.2.3.1. Method A.

2.2.3.1.1. 3-Benzyloxy-16α-bromomethylestra-1,3,5(10)trien-17α-yl acetate (6b) and 3-benzyloxy-16α-acetoxymethylestra-1,3,5(10)-trien-17α-yl acetate (2b). Compound 6b was eluted with CHCl<sub>3</sub> (1.27 g, 85%). Mp 107–108 °C; R<sub>f</sub> = 0.65 (ss A);  $[\alpha]_D^{20} + 75$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 67.52; H, 6.75. C<sub>28</sub>H<sub>33</sub>O<sub>3</sub>Br requires: C, 67.60; H, 6.69%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.11 (s, 3H, 17-OAc-H<sub>3</sub>), 2.85 (m, 2H, 6-H<sub>2</sub>), 3.36 (dd, 1H, J = 9.3 Hz, J = 7.6 Hz) and 3.44 (t, 1H, J = 9.3 Hz): 16a-H<sub>2</sub>, 5.02 (s, 2H, 3-benzyl-CH<sub>2</sub>), 5.08 (d, 1H, J = 5,5 Hz, 17-H), 6.70 (d, 1H, J=2.4Hz, 4-H), 6.77 (dd, 1H, J=8.5Hz, J=2.4Hz, 2-H), 7.17 (d, 1H, J = 8.5 Hz, 1-H), 7.30 (t, 1H, J = 7.4 Hz, 4'-H), 7.37 (t, 2H, J = 7.4 Hz, 3',5'-H), 7.41 (d, 2H, J = 7.4 Hz, 2',6'-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 16.7 (C-18), 20.8 (17-OAc-CH<sub>3</sub>), 25.9, 27.9, 29.8, 31.2, 31.9, 33.4, 38.9, 42.7, 43.5, 46.3 (C-13), 48.3, 70.0 (3-benzyl-CH<sub>2</sub>), 81.5 (C-17), 112.4 (C-2), 114.9 (C-4), 126.3 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.3 (C-1'), 137.8 (C-5), 156.8 (C-3), 170.3 (OAc-CO). EI-MS (70 eV) m/z (%): 498 (28) [M<sup>+</sup> + 2], 496 (28) [M<sup>+</sup>], 91 (100), 43 (7). Continued elution with ethyl acetate/CHCl<sub>3</sub> (2.5:97.5) yielded 2b (170 mg, 12%). Mp 106–108 °C ([14], Mp 106–108); R<sub>f</sub> = 0.30 (ss A);  $[\alpha]_{D}^{20}$  + 67 (c 1.00 in CHCl<sub>3</sub>). (Found: C, 75.42; H, 7.56. C<sub>30</sub>H<sub>36</sub>O<sub>5</sub> requires: C 75.60%, H 7.61%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.85 (s,

3H, 18-H<sub>3</sub>), 2.01 and 2.07 (2 × s, 2 × 3H, 2 × OAc-H<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 4.03–4.12 (overlapping multiplets, 2H, 16a-H<sub>2</sub>), 5.02 (d, 2H, 3-benzyl-CH<sub>2</sub>), 5.09 (d, 1H, *J* = 5.7 Hz, 17-H), 6.71 (d, 1H, *J* = 2.5 Hz, 4-H), 6.77 (dd, 1H, *J* = 8.6 Hz, 1-H), 7.30 (t, 1H, *J* = 7.3 Hz, 4'-H), 7.36 (t, 2H, *J* = 7.3 Hz, 3',5'-H), 7.41 (d, 2H, *J* = 7.3 Hz, 2',6'-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 16.8 (C-18), 20.8 (2C, 2 × OAc-CH<sub>3</sub>), 25.9, 27.9, 28.1, 29.7, 31.6, 38.6, 38.9, 43.5, 46.0 (C-13), 48.1, 64.0 (C-16a), 70.0 (3-benzyl-CH<sub>2</sub>), 80.7 (C-17), 112.3 (C-2), 114.9 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.3 (C-1'), 137.8 (C-5), 156.8 (C-3), 170.2 and 170.9 (2C, 2 × OAc-CO); EI-MS (70 eV) *m*/z (%): 476 (16) [M<sup>+</sup>], 91 (100), 43 (74).

#### 2.2.3.2. Method B.

2.2.3.2.1. 16α-Bromomethyl-3-hydroxyestra-1,3,5(10)trien-17 $\alpha$ -yl acetate (6c) and 16 $\alpha$ -acetoxymethyl-3-hydroxyestra-1,3,5(10)-trien-17α-yl acetate (2c). Compound 6c (953 mg, 78%). Mp 201–203 °C;  $R_f = 0.30$  (ss A)  $[\alpha]_D^{20} + 85$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 62.05; H, 6.72. C<sub>21</sub>H<sub>27</sub>O<sub>3</sub>Br requires: C, 61.92; H, 6.68%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.12 (s, 3H, OAc-CH<sub>3</sub>), 2.81 (m, 2H, 6-H<sub>2</sub>), 3.36 (dd, 1H, J = 9.4 Hz, J = 7.5 Hz) and 3.44 (t, 1H, J = 9.4 Hz): 16a-H<sub>2</sub>, 4.76 (s, 1H, 3-OH), 5.08 (d, 1H, J=5.6 Hz, 17-H), 6.56 (d, 1H, J=2.5 Hz, 4-H), 6.62 (dd, 1H, J=8.4 Hz, J=2.5 Hz, 2-H), 7.12 (d, 1H, J=8.4 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 16.8 (C-18), 20.9 (OAc-CH<sub>3</sub>), 25.9, 27.8, 29.6, 31.2, 31.9, 33.4, 38.9, 42.8, 43.5, 46.3 (C-13), 48.3, 81.6 (C-17), 112.8 (C-2), 115.3 (C-4), 126.5 (C-1); 132.4 (C-10), 138.1 (C-5), 153.5 (C-3), 170.5 (OAc-CO); EI-MS (70 eV) m/z (%): 408.1 (69) [M<sup>+</sup>+2], 406 (67) [M<sup>+</sup>], 267 (16), 172 (26), 160 (100), 146 (47), 133 (50), 107 (22), 43 (87). 2c (115 mg, 10%) as a colorless oil.  $R_f = 0.45$  (ss C);  $[\alpha]_D^{20} + 57$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 71.36; H, 7.95. C<sub>23</sub>H<sub>30</sub>O<sub>5</sub> requires: C, 71.48; H, 7.82%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.01 and 2.08 (2 × s, 2 × 3H, 2 × OAc-CH<sub>3</sub>), 2.81 (m, 2H, 6-H<sub>2</sub>), 4.04–4.13 (overlapping multiplets, 2H, 16a-H<sub>2</sub>), 5.10 (d, 1H, J=5.7 Hz, 17-H), 6.56 (d, 1H, J=2.4 Hz, 4-H), 6.63 (dd, 1H, J=8.5 Hz, J=2.4 Hz, 2-H), 7.13 (d, 1H, J = 8.5 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 16.8 (C-18), 20.8 (2C, 2 × OAc-CH<sub>3</sub>), 25.9, 27.9, 28.2, 29.6, 31.7, 38.6, 38.9, 43.5, 46.0 (C-13), 48.2,64.1 (C-16a), 80.8 (C-17), 112.8 (C-2), 115.3 (C-4), 126.5 (C-1); 132.4 (C-10), 138.1 (C-5), 153.6 (C-3), 170.4 and 171.2 (2C,  $2 \times \text{OAc-CO}$ ); EI-MS (70 eV) m/z (%): 386 (100) [M<sup>+</sup>], 266 (12), 213 (7), 172 (12), 159 (26), 133 (16), 106 (16), 43 (74).

# 2.2.3.3. Method C.

2.2.3.3.1. 16α-Bromomethylestra-1,3,5(10)-triene-3,17αdiyl diacetate (6d) and  $16\alpha$ -acetoxymethylestra-1,3,5(10)triene-3,17 $\alpha$ -diyl diacetate (2d). Compound 6d (990 mg, 74%). Mp 120–122 °C;  $R_f = 0.50$  (ss A);  $[\alpha]_D^{20} + 72$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 61.66; H, 6.35. C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>Br requires: C 61.47; H 6.50%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.12 (s, 3H, 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.87 (m, 2H, 6-H<sub>2</sub>), 3.36 (dd, 1H, J = 9.2 Hz, J = 7.5 Hz) and 3.44 (t, 1H, J = 9.2 Hz): 16a-H<sub>2</sub>, 5.08 (d, 1H, J=5.6 Hz, 17-H), 6.79 (d, 1H, J=2.1 Hz, 4-H), 6.84 (dd, 1H, J=8.4Hz, J=2.1Hz, 2-H), 7.26 (d, 1H, J=8.4Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 16.7 (C-18), 20.8 (17-OAc-CH<sub>3</sub>), 21.1 (3-OAc-CH<sub>3</sub>), 25.7, 27.7, 29.5, 31.2, 31.8, 33.3, 38.5, 42.7, 43.6, 46.2 (C-13), 48.4, 81.4 (C-17), 118.6 (C-2), 121.5 (C-4), 126.3 (C-1), 137.6 (C-10), 138.0 (C-5), 148.5 (C-3), 169.7 (3-OAc-CO) and 170.3 (17-OAc-CO); EI-MS (70 eV) m/z (%): 450 (10) [M<sup>+</sup> + 2], 448 (10) [M<sup>+</sup>], 408 (96), 406 (100), 267 (87), 172 (7), 160 (26), 133 (12), 83 (14), 43 (39). 2d (102 mg, 8%). Mp 67–68 °C;  $R_f = 0.45$  (ss B);  $[\alpha]_D^{20} + 64$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 69.92; H, 7.61.  $C_{25}H_{32}O_6$ requires: C 70.07; H 7.53%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.01 and 2.07 (2 × s, 2 × 3H, 16a- and 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.86 (m, 2H, 6-H<sub>2</sub>), 4.03–4.13 (m, 2H, 16a-H<sub>2</sub>), 5.10 (d, 1H, *J* = 5.2 Hz, 17-H), 6.79 (d, 1H, *J* = 2.3 Hz, 4-H), 6.83 (dd, 1H, *J* = 8.1 Hz, *J* = 2.3 Hz, 2-H), 7.26 (d, 1H, *J* = 8.1 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 16.7 (C-18), 20.8 (2C, 16a- and 17-OAc-CH<sub>3</sub>), 21.0 (3-OAc-CH<sub>3</sub>), 25.7, 27.7, 28.1, 29.4, 31.6, 38.5, 38.6, 43.6, 45.9 (C-13), 48.1, 64.0 (C-16a), 80.6 (C-17), 118.6 (C-2), 121.4 (C-4), 126.2 (C-1); 137.6 (C-10), 138.0 (C-5), 148.5 (C-3), 169.6 (3-OAc-CO), 170.2 and 170.9 (2C, 16a- and 17-OAc-CO); EI-MS (70 eV) *m*/z (%): 428 (14) [M<sup>+</sup>], 386 (100), 172 (5), 159 (12), 133 (8), 43 (41).

# 2.2.4. Reaction of 3-benzyloxy-16 $\alpha$ -hydroxymethylestra-1,3,5(10)-trien-17 $\beta$ -ol (**3a**) with HBr in acetic acid 2.2.4.1. Method A.

2.2.4.1.1.3-Benzyloxy-16α-acetoxymethylestra-1,3,5(10)trien-17β-yl acetate (3b). Compound 3b (1.36 mg, 95%). Mp 132–134 °C ([14] Mp 131–132 C);  $R_f = 0.30$  (ss A);  $[\alpha]_D^{20} + 7$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 75.58; H, 7.78. C<sub>30</sub>H<sub>36</sub>O<sub>5</sub> requires: C, 75.60; H, 7.61%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.03 and 2.06 (2 × s, 2 × 3H, 2 × OAc-CH<sub>3</sub>), 2.83 (m, 2H, 6-H<sub>2</sub>), 4.06 (dd, 1H, J = 10.9 Hz, J = 6.9 Hz) and 4.12 (dd, 1H, J = 10.9 Hz, J = 6.4 Hz): 16a-H<sub>2</sub>, 4.72 (d, 1H, J=7.7 Hz, 17-H), 5.01 (s, 2H, 3-benzyl-H<sub>2</sub>), 6.70 (d, 1H, J=2.4 Hz, 4-H), 6.76 (dd, 1H, J=8.6 Hz, J=2.4 Hz, 2-H), 7.17 (d, 1H, J = 8.6 Hz, 1-H), 7.29 (t, 1H, J = 7.3 Hz, 4'-H), 7.35  $(t, 2H, J = 7.3 Hz, 3', 5'-H), 7.41 (d, 2H, J = 7.3 Hz, 2', 6'-H); {}^{13}C NMR$ (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.7 (C-18), 20.8 and 21.0 (2C, 2 × OAc-CH<sub>3</sub>), 26.1, 27.1, 27.4, 29.6, 36.9, 38.4, 40.3, 43.7, 44.3 (C-13), 48.7, 66.7 (C-16a), 70.0 (3-benzyl-CH<sub>2</sub>), 83.8 (C-17), 112.4 (C-2), 114.9 (C-4), 126.3 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.4 (C-1'), 137.8 (C-5), 156.8 (C-3), 170.7 and 170.9 (2C, 2 × OAc-CO); EI-MS (70 eV) m/z (%): 476 (48), 91 (100), 84 (20), 49 (52), 43 (11).

#### 2.2.4.2. Method B.

2.2.4.2.1. 16α-Acetoxymethyl-3-hydroxyestra-1,3,5(10)trien-17β-yl acetate (3c). Compound 3c (1.00 g, 87%). Mp 145–147 °C;  $R_f = 0.40$  (ss C);  $[\alpha]_D^{20} - 3$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 71.57; H, 7.88.  $C_{23}H_{30}O_5$  requires: C, 71.48; H, 7.82%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.05 and 2.08 (2 × s, 2 × 3H, 2 × OAc-CH<sub>3</sub>), 2.80 (m, 2H, 6-H<sub>2</sub>), 4.08 (dd, 1H, *J* = 10.9 Hz, *J* = 6.8 Hz) and 4.12 (dd, 1H, *J* = 10.9 Hz, *J* = 6.5 Hz): 16a-H<sub>2</sub>, 4.74 (d, 1H, *J* = 7.7 Hz, 17-H), 5.74 (s, 1H, 3-OH), 6.56 (d, 1H, *J* = 2.4 Hz, 4-H), 6.63 (dd, 1H, *J* = 8.4 Hz, *J* = 2.4 Hz, 2-H), 7.11 (d, 1H, *J* = 8.4 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.6 (C-18), 20.8 and 21.0 (2C, 2 × OAc-CH<sub>3</sub>), 26.0, 27.1, 27.4, 29.4, 36.8, 38.4, 40.2, 43.6, 44.3 (C-13), 48.7, 66.9 (C-16a), 84.0 (C-17), 112.8 (C-2), 115.3 (C-4), 126.4 (C-1); 132.0 (C-10), 137.9 (C-5), 153.7 (C-3), 171.0 and 171.3 (2C, 2 × OAc-CO); EI-MS (70 eV) *m*/z (%): 386 (95) [M<sup>+</sup>], 266 (12), 213 (12), 172 (19), 159 (34), 159 (34), 133 (27), 10 (8), 43 (100).

#### 2.2.4.3. Method C.

# 2.2.4.3.1. 16α-Acetoxymethylestra-1,3,5(10)-triene-3,17β-diyl diacetate (3d). Compound 3d (1.25 g, 98%). Mp 85–88 °C; $R_f = 0.40$ (ss B); $[\alpha]_D^{20} - 6.0$ (c 1.00 in CHCl<sub>3</sub>). (Found: C, 69.95; H, 7.65. $C_{25}H_{32}O_6$ requires: C, 70.07; H, 7.53%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.04 and 2.07 (2 × s,

2 × 3H, 16a- and 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.85 (m, 2H, 6-H<sub>2</sub>), 4.06 (dd, 1H, J = 10.8 Hz, J = 6.8 Hz) and 4.12 (dd, 1H, J = 10.8 Hz, J = 6.4 Hz): 16a-H<sub>2</sub>, 4.74 (d, 1H, J = 7.7 Hz, 17-H), 6.79 (d, 1H, J = 2.2 Hz, 4-H), 6.83 (dd, 1H, J = 8.3 Hz, J = 2.2 Hz, 2-H), 7.25 (d, 1H, J = 8.3 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.6 (C-18), 20.8, 20.9 and 21.0 (3C, 3 × OAc-CH<sub>3</sub>), 25.9, 26.9, 27.3, 29.3, 36.8, 38.0, 40.2, 43.8, 44.2 (C-13), 48.7, 66.7 (C-16a), 83.7 (C-17), 118.6 (C-2), 121.4 (C-4), 126.3 (C-1); 137.6 (C-10), 137.9 (C-5), 148.5 (C-3), 169.6 (3-OAc-CO), 170.6 and 170.9 (2C, 16a-and 17-OAc-CO); EI-MS (70 eV) m/z (%): 428 (11) [M<sup>+</sup>], 386 (100), 172 (8), 159 (14), 133 (10), 43 (54).

2.2.5. Conversion of 3-benzyloxy- $16\beta$ hydroxymethylestra-1,3,5(10)-trien- $17\alpha$ -ol (4a) with HBr in acetic acid 2.2.5.1. Method A.

2.2.5.1.1. 16β-Acetoxymethyl-3-benzyloxy-17-methyl-18-nor-estra-1,3,5(10),13(17)-tetraene (16b), 3-benzyloxy-16β-hydroxymethylestra-1,3,5(10),13(17)-tetraene (16a) and 16β-acetoxymethyl-3-benzyloxyestra-1,3,5(10)-trien-17α-yl acetate (4b). Compound 16b (290 mg, 23%) as colorless oil;  $R_f = 0.80$  (ss A);  $[\alpha]_D^{20} + 85$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 80.56; H, 7.83. C<sub>28</sub>H<sub>32</sub>O<sub>3</sub> requires: C, 80.73; H, 7.74%). <sup>1</sup>H-NMR δ, ppm 1.61 (s, 3H, 18-H<sub>3</sub>), 2.03 (s, 3H, Ac-Me), 2.81 (m, 2H,  $6-H_2$ ), 3.98 (dd, 1H, J=10.8Hz, J=6.9Hz) and 4.22 (dd, 1H, J = 10.8 Hz, J = 4.8 Hz): 16a-H<sub>2</sub>, 5.02 (s, 2H, benzyl-H<sub>2</sub>), 6.70 (d, 1H, J=2.3 Hz, 4-H), 6.78 (dd, 1H, J=8.5 Hz, J=2.3 Hz, 2-H), 7.24 (d, 1H, J=8.5 Hz, 1-H), 7.30 (t, 1H, J=7.5 Hz, 4'-H), 7.37 (t, 2H, J=7.5 Hz, 3'-, 5'-H), 7.42 (d, 2H, J=7.5 Hz, 2'-, 6'-H). <sup>13</sup>C-NMR δ, ppm: 12.3 (C-18), 21.4 (Ac-Me), 26.4, 28.1, 30.8, 31.8, 32.8, 42.8, 48.1, 49.6, 51.3, 68.0, 70.4, 113.0, 115.1, 127.6, 127.9, 128.2, 128.9, 132.8, 137.8, 138.8, 139.3, 157.1 (C-3), 171.7 (Ac-CO).

The white oily 16b (104 mg, 0.25 mmol) obtained in the chromatographic separation was dissolved in methanol (10 ml) containing NaOCH<sub>3</sub> (5 mg, 0.092 mmol) and the solution was allowed to stand for 24 h. It was diluted with water, and the white precipitate that separated out was filtered off and recrystallized from a mixture of acetone/hexane; 16a (76 mg, 81%). Mp 128–129 °C;  $R_f$  = 0.65 (ss A);  $[\alpha]_D^{20}$  + 52 (c 1.00 in CHCl<sub>3</sub>). (Found: C, 83.57; H, 7.92. C<sub>26</sub>H<sub>30</sub>O<sub>2</sub> requires: C, 83.38; H, 8.07%). <sup>1</sup>H-NMR δ, ppm: 1.67 (s, 3H, 18-H<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 3.72 (m, 2H, 16a-H<sub>2</sub>), 5.06 (s, 2H, benzyl-H<sub>2</sub>), 6.74 (d, 1H, J=2.6 Hz, 4-H), 6.82 (dd, 1H, J=8.6 Hz, J=2.6 Hz, 2-H), 7.27 (d, 1H, J=8.6 Hz, 1-H), 7.35 (t, 1H, J=7.0 Hz, 4'-H), 7.41 (t, 2H, J = 7.0 Hz, 3'-, 5'-H), 7.46 (d, 2H, J = 7.0 Hz, 2'-, 6'-H). <sup>13</sup>C-NMR  $\delta$ , ppm: 12.14 (C-18), 26.5, 28.0, 28.1, 28.2, 30.8, 31.7, 31.9, 42.8, 49.5, 51.3, 51.4, 65.2, 70.4, 113.0, 115.2, 127.6, 127.9, 128.3, 129.0, 132.9, 137.8, 138.9, 140.4, 157.1.

**4b** (850 mg, 59%). Mp 80–82 °C ([14], Mp 79–82 °C);  $R_f = 0.35$  (ss A);  $[α]_D^{20} + 51$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 75.82; H, 7.51. C<sub>30</sub>H<sub>36</sub>O<sub>5</sub> requires: C, 75.60; H, 7.61%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.06 and 2.07 (2 × s, 2 × 3H, 2 × OAc-CH<sub>3</sub>), 2.85 (m, 2H, 6-H<sub>2</sub>), 4.17 (dd, 1H, *J* = 10.9 Hz, *J* = 7.1 Hz) and 4.22 (dd, 1H, *J* = 10.9 Hz, *J* = 7.4 Hz): 16a-H<sub>2</sub>, 4.71 (d, 1H, *J* = 1.7 Hz, 17-H), 5.03 (s, 2H, 3-benzyl-H<sub>2</sub>), 6.71 (d, 1H, *J* = 2.2 Hz, 4-H), 6.77 (dd, 1H, *J* = 8.5 Hz, *J* = 2.2 Hz, 2-H), 7.19 (d, 1H, *J* = 8.5 Hz, 1-H), 7.30 (t, 1H, *J* = 7.3 Hz, 4'-H), 7.37 (t, 2H, *J* = 7.3 Hz, 3',5'-H), 7.42 (d, 2H, *J* = 7.3 Hz, 2',6'-H); <sup>13</sup>C-NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.1 (C-18), 20.9 and 21.1 (2C, 2 × OAc-CH<sub>3</sub>), 25.9, 27.9, 29.3, 29.6, 32.2, 38.5, 43.4, 44.6 (C-13), 45.8, 50.0, 66.6 (C-16a), 70.0 (3-benzyl-CH<sub>2</sub>), 83.6 (C-

17), 112.4 (C-2), 114.9 (C-4), 126.3 (C-1), 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.7 (C-10), 137.4 (C-1'), 137.9 (C-5), 156.8 (C-3), 170.3 and 171.0 (2C,  $2 \times \text{OAc-CO}$ ); EI-MS (70 eV) *m*/z (%): 476 (39) [M<sup>+</sup>], 91 (100), 43 (9).

#### 2.2.5.2. Method B.

2.2.5.2.1. 16β-Acetoxymethyl-3-hydroxyestra-1,3,5(10)trien-17a-yl acetate (4c). Compound 4c (560 mg, 48%). Mp 155–157 °C;  $R_f = 0.40$  (ss C);  $[\alpha]_D^{20} + 57$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 71.55; H, 7.96. C<sub>23</sub>H<sub>30</sub>O<sub>5</sub> requires: C, 71.48; H, 7.82%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ, ppm: 0.84 (s, 3H, 18-H<sub>3</sub>), 2.07 and 2.08  $(2 \times s, 2 \times 3H, 2 \times OAc-CH_3)$ , 2.81 (m, 2H, 6-H<sub>2</sub>), 4.18 (dd, 1H, J = 11.0 Hz, J = 7.2 Hz) and 4.22 (dd, 1H, J = 11.0 Hz, J = 7.5 Hz): 16a-H<sub>2</sub>, 4.72 (d, 1H, J = 1.9 Hz, 17-H), 5.70 (s, 1H, 3-OH), 6.57 (d, 1H, J=2.4 Hz, 4-H), 6.64 (dd, 1H, J=8.4 Hz, J=2.4 Hz, 2-H), 7.13 (d, 1H, J = 8.5 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.0 (C-18), 20.8 and 21.0 (2C, 2 × OAc-CH<sub>3</sub>), 25.9, 27.8, 29.2, 29.5, 32.1, 38.5, 43.3, 44.5 (C-13), 45.6, 49.9, 66.6 (C-16a), 83.7 (C-17), 112.8 (C-2), 115.2 (C-4), 126.3 (C-1); 132.0 (C-10), 137.9 (C-5), 153.7 (C-3), 170.6 and 171.3 (2C,  $2 \times \text{OAc-CO}$ ); EI-MS (70 eV) m/z (%): 386 (100) [M<sup>+</sup>], 266 (10), 213 (6), 172 (10), 159 (23), 133 (15), 106 (13), 43 (10).

#### 2.2.5.3. Method C.

#### 2.2.5.3.1. 16β-Acetoxymethylestra-1,3,5(10)-triene-

**3,17α-diyl diacetate (4d).** Compound **4d** (530 mg, 41%) as a colorless oil;  $R_f = 0.30$  (ss A);  $[α]_D^{20} + 45$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 69.91; H, 7.45.  $C_{25}H_{32}O_6$  requires: C, 70.07; H, 7.53%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.06 and 2.07 (2 × s, 2 × 3H, 16a- and 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.87 (m, 2H, 6-H<sub>2</sub>), 4.17 (dd, 1H, *J* = 10.9 Hz, *J* = 7.1 Hz) and 4.23 (dd, 1H, *J* = 10.9 Hz, *J* = 7.4 Hz): 16a-H<sub>2</sub>, 4.71 (d, 1H, *J* = 1.64 Hz, 17-H), 6.79 (d, 1H, *J* = 1.9 Hz, 4-H), 6.84 (dd, 1H, *J* = 8.4 Hz, *J* = 1.9 Hz, 2-H), 7.27 (d, 1H, *J* = 8.4 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 16.9 (C-18), 20.8 (3-OAc-CH<sub>3</sub>) and 21.0 (2C, 16a- and 17-OAc-CH<sub>3</sub>), 25.7, 27.7, 29.2, 29.4, 32.1, 38.1, 43.5, 44.5 (C-13), 45.7, 50.0, 66.4 (C-16a), 83.5 (C-17), 118.6 (C-2), 121.4 (C-4), 126.2 (C-1); 137.6 (C-10), 138.0 (C-5), 148.5 (C-3), 169.6 (3-OAc-CO), 170.3 and 171.0 (2C, 16a- and 17-OAc-CO); EI-MS (70 eV) *m/z* (%): 428 (12) [M<sup>+</sup>], 386 (100), 172 (6), 159 (12), 133 (9), 43 (57).

# 2.3. Conversion of 3-benzyloxy-16hydroxymethylestra-1,3,5(10)-trien-17-ol isomers (1a-4a) to 3-benzyloxy-16-bromomethylestra-1,3,5(10)-trien-17-ol (5a-8a) by Appel bromination

# 2.3.1. Method D., General procedure

Compounds **1a–3a** or **4a** (1.18 g, 3 mmol) was suspended in 40 ml of anhydrous  $CH_2Cl_2$ , and 4.97 g (15 mmol) of  $CBr_4$  was added. After cooling the suspension to 0 °C, 3.94 g (15 mmol) of Ph<sub>3</sub>P in 20 ml anhydrous  $CH_2Cl_2$  was added dropwise and the reaction mixture was allowed to become warm and stirred until it became a yellow solution. After evaporation in vacuo the crude product was purified on a silica gel column with ethyl acetate/CHCl<sub>3</sub> 2.5:97.5.

**5a** (1.13 g, 83%). Mp 109–110 °C;  $R_f = 0.60$  (ss A);  $[\alpha]_D^{20} + 66$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 68.35; H, 6.92.  $C_{26}H_{31}O_2Br$  requires: C, 68.57; H, 6.86%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.79 (s, 3H, 18-H<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 3.33 (t, 1H, *J*=9.7 Hz) and 3.72 (dd, 1H, *J*=9.7 Hz, *J*=6.4 Hz): 16a-H<sub>2</sub>, 3.82 (d, 1H, *J*=9.8 Hz, 17-H), 5.01

(s, 2H, benzyl-H<sub>2</sub>), 6.71 (d, 1H, J=2.3Hz, 4-H), 6.77 (dd, 1H, J=8.6Hz, J=2.3Hz, 2-H), 7.18 (d, 1H, J=8.6Hz, 1-H), 7.30 (t, 1H, J=7.3Hz, 4'-H), 7.36 (t, 2H, J=7.3Hz, 3',5'-H), 7.41 (d, 2H, J=7.3Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.3 (C-18), 26.2, 27.4, 29.7, 32.6, 36.4, 37.6, 38.1, 43.3, 43.9, 44.5 (C-13), 48.2, 70.0 (benzyl-CH<sub>2</sub>), 81.9 (C-17), 112.4 (C-2), 114.9 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.8 (C-10), 137.4 (C-1'), 137.9 (C-5), 156.8 (C-3); EI-MS (70 eV) m/z (%): 456 (42) [M<sup>+</sup> + 2], 454 (41) [M<sup>+</sup>], 91 (100).

**6a** (1.16 g, 85%). Mp 119–120 °C;  $R_f = 0.80$  (ss A);  $[\alpha]_D^{20} + 89$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 68.67; H, 6.79.  $C_{26}H_{31}O_2Br$  requires: C, 68.57; H, 6.86%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.78 (s, 3H, 18-H<sub>3</sub>), 2.83 (m, 2H, 6-H<sub>2</sub>), 3.48 (dd, 1H, J = 9.4 Hz, J = 6.5 Hz) and 3.61 (t, 1H, J = 9.4 Hz): 16a-H<sub>2</sub>, 3.85 (t, 1H, J = 4.4 Hz, 17-H), 5.02 (s, 2H, benzyl-H<sub>2</sub>), 6.70 (d, 1H, J = 2.5 Hz, 4-H), 6.77 (dd, 1H, J = 8.5 Hz, J = 2.5 Hz, 2-H), 7.19 (d, 1H, J = 8.5 Hz, 1-H), 7.30 (t, 1H, J = 7.3 Hz, 4'-H), 7.36 (t, 2H, J = 7.3 Hz, 3',5'-H), 7.41 (d, 2H, J = 7.3 Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.2 (C-18), 26.0, 27.9, 29.8, 31.1, 31.2, 34.9 (C-16a), 39.0, 43.5, 44.0, 46.3 (C-13), 47.3, 70.0 (benzyl-CH<sub>2</sub>), 79.9 (C-17), 112.3 (C-2), 114.9 (C-4), 126.3 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.9 (C-10), 137.4 (C-1'), 137.9 (C-5), 156.8 (C-3); EI-MS (70 eV) m/z (%): 456 (23) [M<sup>+</sup> + 2], 454 (23) [M<sup>+</sup>], 91 (100).

**7a** (1.08 g, 79%). Mp 90–91 °C;  $R_f = 0.30$  (ss A);  $[\alpha]_D^{20} + 50$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 68.63; H, 6.95.  $C_{26}H_{31}O_2Br$  requires: C, 68.57; H, 6.86%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.83 (s, 3H, 18-H<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 3.47 (d, 1H, *J* = 7.6 Hz, 17-H), 3.54–3.61 (overlapping multiplets, 2H, 16a-H<sub>2</sub>, 5.02 (s, 2H, benzyl-H<sub>2</sub>), 6.71 (d, 1H, *J* = 2.3 Hz, 4-H), 6.77 (dd, 1H, *J* = 8.6 Hz, *J* = 2.3 Hz, 2-H), 7.18 (d, 1H, *J* = 8.6 Hz, 1-H), 7.30 (t, 1H, *J* = 7.3 Hz, 4'-H), 7.36 (t, 2H, *J* = 7.3 Hz, 3',5'-H), 7.41 (d, 2H, *J* = 7.3 Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.0 (C-18), 26.1, 27.2, 29.7, 29.8, 36.6, 38.3, 38.9 (C-16a), 43.9, 44.6 (C-13), 45.8, 48.3, 69.9 (benzyl-CH<sub>2</sub>), 85.8 (C-17), 112.3 (C-2), 114.8 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.7 (C-10), 137.3 (C-1'), 137.9 (C-5), 156.7 (C-3); EI-MS (70 eV) *m*/z (%): 456 (24) [M<sup>+</sup> + 2], 454 (25) [M<sup>+</sup>], 91 (100).

**8a** (1.09 g, 80%). Mp 51–53 °C;  $R_f = 0.35$  (ss A);  $[\alpha]_D^{20} + 35$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 68.62; H, 6.94.  $C_{26}H_{31}O_2Br$  requires: C, 68.57; H, 6.86%. <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.76 (s, 3H, 18-H<sub>3</sub>), 2.84 (m, 2H, 6-H<sub>2</sub>), 3.51 (m, 2H, 16a-H<sub>2</sub>),), 3.64 (s, 1H, 17-H), 5.02 (s, 2H, benzyl-H<sub>2</sub>), 6.71 (d, 1H, J = 2.4Hz, 4-H), 6.77 (dd, 1H, J = 8.6Hz, J = 2.4Hz, 2-H), 7.19 (d, 1H, J = 8.6Hz, 1-H), 7.30 (t, 1H, J = 7.3Hz, 4'-H), 7.36 (t, 2H, J = 7.3Hz, 3',5'-H), 7.41 (d, 2H, J = 7.3Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.9 (C-18), 25.9, 27.9, 29.7, 31.9, 32.1, 37.3 (C-16a), 38.6, 43.3, 45.1 (C-13), 49.3, 52.2, 70.0 (benzyl-CH<sub>2</sub>), 84.1 (C-17), 112.3 (C-2), 114.9 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.8 (C-10), 137.3 (C-1'), 137.9 (C-5), 156.8 (C-3); EI-MS (70 eV) *m/z* (%): 456 (6) [M<sup>+</sup> + 2], 454 (6) [M<sup>+</sup>], 91 (100).

# 2.4. Debenzylation of 3-benzyloxy-16bromomethylestra-1,3,5(10)-trien-17-ol isomers (5a-8a) with Pd/C to 16-bromomethylestra-1,3,5(10)triene-3,17-diols (5e-8e)

#### 2.4.1. General procedure

Compounds 5a-7a or 8a (700 mg, 1.54 mmol) was dissolved in 50 ml of ethyl acetate and 250 mg of Pd/C was added to the solution, which was stirred at room temperature for 6 h at 30 bar H<sub>2</sub> pressure. The reaction mixture was filtered over celite. After evaporation of the filtrate, the crude product was purified by column chromatography with ethyl acetate/CHCl<sub>3</sub> 10:90).

**5e** (542 mg, 96%). Mp 168–169 °C;  $R_f = 0.40$  (ss B);  $[\alpha]_D^{20} + 114$  (c 1.00 in ethyl acetate). (Found: C, 62.55; H, 7.02.  $C_{19}H_{25}O_2Br$  requires: C 62.47; H 6.90%). <sup>1</sup>H NMR (DMSO-d<sub>6</sub>)  $\delta$ , ppm: 0.68 (s, 3H, 18-H<sub>3</sub>), 2.70 (m, 2H, 6-H<sub>2</sub>), 3.28 (t, 1H, *J* = 10.9 Hz, 17-H), 3.63 (m, 1H) and 3.76 (dd, 1H, *J* = 9.6 Hz, *J* = 4.2 Hz): 16a-H<sub>2</sub>, 4.84 (d, 1H, *J* = 5.5 Hz, 17-OH), 6.43 (s, 1H, 4-H), 6.50 (d, 1H, *J* = 7.9 Hz, 2-H), 7.02 (d, 1H, *J* = 8.5 Hz, 1-H), 8.96 (s, 1H, 3-OH). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>)  $\delta$ , ppm: 12.4 (C-18), 25.9, 27.0, 29.1, 32.4, 37.1, 37.9, 39.3, 43.1, 43.4, 43.8, 47.4, 80.6 (C-17), 112.7 (C-2), 114.9 (C-4), 125.9 (C-1); 130.3 (C-10), 137.0 (C-5), 154.8 (C-3); EI-MS (70 eV) *m*/z (%): 366 (34) [M<sup>+</sup> + 2], 364 (36) [M<sup>+</sup>], 267 (23), 213 (12), 185 (9), 159 (17), 145 (10), 133 (24), 83 (11), 43 (100).

**6e** (550 mg, 98%). Mp 181–183 °C;  $R_f = 0.50$  (ss B);  $[\alpha]_D^{20} + 70$  (c 1.00 in ethyl acetate). (Found: C, 62.55; H, 7.02. C19H<sub>25</sub>O<sub>2</sub>Br requires: C 62.47; H 6.90%). <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta$ , ppm: 0.68 (s, 3H, 18-H<sub>3</sub>), 2.70 (m, 2H, 6-H<sub>2</sub>), 3.41 (t, 1H, *J* = 8.6 Hz) and 3.63 (t, 1H, *J* = 8.6 Hz): 16a-H<sub>2</sub>, 3.60 (d, 1H, *J* = 5.0 Hz, 17-H), 6.43 (d, 1H, *J* = 2.0 Hz, 4-H), 6.50 (dd, 1H, *J* = 8.4 Hz, *J* = 2.0 Hz, 2-H), 7.04 (d, 1H, *J* = 8.4 Hz, 1-H), 8.96 (s, 1H, 3-OH). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>)  $\delta$ , ppm: 17.0 (C-18), 25.8, 27.6, 29.2, 30.6, 31.2, 36.7, 38.8, 43.2, 43.6, 46.1, 46.3, 78.5 (C-17), 112.7 (C-2), 114.8 (C-4), 125.9 (C-1); 130.4 (C-10), 137.1 (C-5), 154.8 (C-3); EI-MS (70 eV) *m*/*z* (%): 366 (99) [M<sup>+</sup> + 2], 364 (100) [M<sup>+</sup>], 267 (15), 213 (21), 198 (15), 185 (11), 172 (22), 159 (35), 146 (20), 133 (37), 107 (13).

**7e** (547 mg, 97%). Mp 188–189 °C;  $R_f = 0.30$  (ss B);  $[\alpha]_D^{20} + 37$  (c 1.00 in ethyl acetate). (Found: C, 62.36; H, 6.84. C19H<sub>25</sub>O<sub>2</sub>Br requires: C 62.47; H 6.90%). <sup>1</sup>H NMR (DMSO-d<sub>6</sub>) δ, ppm: 0.71 (s, 3H, 18-H<sub>3</sub>), 2.70 (m, 2H, 6-H<sub>2</sub>), 3.24 (m, 1H, 17-H), 3.55 (t, 1H, J = 9.6 Hz) and 3.73 (dd, 1H, J = 9.6 Hz, J = 3.4 Hz): 16a-H<sub>2</sub>, 4.87 (d, 1H, J = 4.8 Hz, 17-OH), 6.43 (s, 1H, 4-H), 6.50 (d, 1H, J = 8.4 Hz, 2-H), 7.01 (d, 1H, J = 8.4 Hz, 1-H), 9.00 (s, 1H, 3-OH). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ, ppm: 12.0 (C-18), 25.9, 26.8, 28.9, 29.1, 36.5, 38.3, 40.6, 43.4, 44.0, 45.6, 47.7, 83.4 (C-17), 112.7 (C-2), 114.9 (C-4), 125.9 (C-1); 130.3 (C-10), 137.1 (C-5), 154.8 (C-3); EI-MS (70 eV) *m/z* (%): 366 (87) [M<sup>+</sup> + 2], 364 (89) [M<sup>+</sup>], 267 (31), 213 (34), 198 (43), 185 (32), 172 (72), 159 (89), 145 (44), 133 (100), 107 (27), 43 (19).

**8e** (540 mg, 96%). Mp 186–188 °C;  $R_f 0.30$  (ss B);  $[\alpha]_D^{20} + 61$  (c 1.00 in ethyl acetate). (Found: C, 62.55; H, 7.08.  $C_{19}H_{25}O_2Br$  requires: C 62.47, H 6.90%). <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$ , ppm: 0.67 (s, 3H, 18-H<sub>3</sub>), 2.69 (m, 2H, 6-H<sub>2</sub>), 3.38 (d, 1H, J = 4.2 Hz, 17-H), 3.50 (t, 1H, J = 9.2 Hz) and 3.66 (t, 1H, J = 9.2 Hz): 16a-H<sub>2</sub>, 4.67 (d, 1H, J = 4.6 Hz, 17-OH), 6.43 (s, 1H, 4-H), 6.50 (d, 1H, J = 8.4 Hz, 2-H), 7.03 (d, 1H, J = 8.4 Hz, 1-H), 8.95 (s, 1H, 3-OH). <sup>13</sup>C NMR (DMSO- $d_6$ )  $\delta$ , ppm: 17.8 (C-18), 25.8, 27.7, 29.2, 31.6, 32.0, 38.4, 38.9, 43.0, 44.6, 48.7, 51.7, 82.6 (C-17), 112.7 (C-2), 114.9 (C-4), 125.9 (C-1); 130.4 (C-10), 137.0 (C-5), 154.8 (C-3); EI-MS (70 eV) m/z (%): 366 (100) [M<sup>+</sup> + 2], 364 (98) [M<sup>+</sup>], 267 (23), 213 (17), 198 (11), 172 (18), 159 (21), 146 (12), 133 (23).

### 2.5. Reaction of 3-benzyloxy-16 $\alpha$ bromomethylestra-1,3,5(10)-trien-17 $\beta$ -ol (7a) with HBr in acetic acid

#### 2.5.1. Method B

2.5.1.1.  $16\alpha$ -Bromomethyl-3-hydroxyestra-1,3,5(10)-trien- $17\beta$ -yl acetate (**7c**). Compound **7c** (950 mg, 78%). Mp 181–182°C,

[ $\alpha$ ]<sub>20</sub><sup>20</sup> – 3 (c 1.00 in CHCl<sub>3</sub>); R<sub>f</sub> = 0.60 (ss C). (Found: C, 62.05; H, 6.50. C<sub>21</sub>H<sub>27</sub>O<sub>3</sub>Br requires: C, 61.92; H, 6.68%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ, ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.09 (s, 3H, OAc-CH<sub>3</sub>), 2.81 (m, 2H, 6-H<sub>2</sub>), 3.43 (dd, 1H, *J* = 10.0 Hz, *J* = 7.6 Hz) and 3.57 (dd, 1H, *J* = 10.0 Hz, *J* = 5.0 Hz): 16a-H<sub>2</sub>, 4.71 (d, 1H, *J* = 7.7 Hz, 17-H), 6.56 (d, 1H, *J* = 2.4 Hz, 4-H), 6.63 (dd, 1H, *J* = 8.4 Hz, *J* = 2.5 Hz, 2-H), 7.11 (d, 1H, *J* = 8.4 Hz, 1-H). <sup>13</sup>C-NMR (CDCl<sub>3</sub>) δ, ppm: 12.9 (C-18), 21.1 (OAc-CH<sub>3</sub>), 26.0, 27.0, 29.5, 29.8, 36.8, 37.6, 38.3, 43.6 (2C), 44.8 (C-13), 48.4, 85.3 (C-17), 112.8 (C-2), 115.3 (C-4), 126.4 (C-1); 132.2 (C-10), 138.0 (C-5), 153.5 (C-3), 171.4 (OAc-CO); EI-MS (7 eV) *m*/z (%): 408 (100) [M<sup>+</sup> + 2], 406 (99) [M<sup>+</sup>], 267 (9), 172 (16), 159 (18), 146 (11), 133 (16), 107 (4), 43 (11).

#### 2.5.2. Method C

2.5.2.1. 16α-Bromomethylestra-1,3,5(10)-triene-3,17β-diyl diacetate (**7d**). Compound **7d** (1.01 g, 75%) as a colorless oil.  $[\alpha]_{20}^{20} - 1$  (c 1.00 in CHCl<sub>3</sub>);  $R_f = 0.50$  (ss A). (Found: C, 61.32; H, 6.68. C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>Br requires: C 61.47, H 6.50%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)

δ, ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.09 (s, 3H, 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.86 (dd, 2H, *J* = 7.6 Hz, *J* = 3.2 Hz, 6-H<sub>2</sub>), 3.43 (dd, 1H, *J* = 10.0 Hz, *J* = 7.7 Hz) and 3.58 (dd, 1H, *J* = 10.0 Hz, *J* = 4.8 Hz): 16a-H<sub>2</sub>, 4.70 (d, 1H, *J* = 7.7 Hz, 17-H), 6.78 (d, 1H, *J* = 1.8 Hz, 4-H), 6.83 (dd, 1H, *J* = 8.4 Hz, *J* = 2.2 Hz, 2-H), 7.25 (d, 1H, *J* = 8.4 Hz, 1-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ, ppm: 12.8 (C-18), 21.1 (2C, OAc-CH<sub>3</sub>), 25.8, 26.8, 29.3, 29.7, 36.7, 37.7, 37.9, 43.6, 43.8, 44.7 (C-13), 48.4, 84.9 (C-17), 118.6 (C-2), 121.4 (C-4), 126.3 (C-1), 137.6 (C-10), 138.0 (C-5), 148.4 (C-3), 169.7 (3-OAc-CO), 171.0 (17-OAc-CO); EI-MS (7 eV) *m*/*z* (%): 450 (8) [M<sup>+</sup> + 2], 448 (8) [M<sup>+</sup>], 408 (100), 406 (98), 267 (9), 172 (11), 159 (12), 133 (10), 83 (30), 57.1 (39), 43 (27).

# 2.5.3. Reaction of 3-benzyloxy-16 $\beta$ -bromomethylestra-1,3,5(10)-trien-17 $\alpha$ -ol (**8a**) with HBr in acetic acid 2.5.3.1. Method B.

2.5.3.1.1. 16β-Bromomethyl-3-hydroxyestra-1,3,5(10)trien-17α-yl acetate (8c). Compound 8c (940 mg, 77%). Mp 210–212 °C;  $R_f = 0.60$  (ss C);  $[\alpha]_D^{20} + 38$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 61.25; H, 6.81.  $C_{21}H_{27}O_3Br$  requires: C 61.92, H 6.68%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.09 (s, 3H, OAc-CH<sub>3</sub>), 2.83 (m, 2H, 6-H<sub>2</sub>), 3.43 (t, 1H, J = 9.7 Hz) and 3.74 (dd, 1H, J = 9.7 Hz, J = 6.6 Hz): 16a-H<sub>2</sub>, 4.63 (d, 1H, J = 2.0 Hz, 17-H), 4.75 (s, 1H, 3-OH), 6.57 (d, 1H, J = 2.5 Hz, 4-H), 6.63 (dd, 1H, J = 8.4 Hz, J = 2.5 Hz, 2-H), 7.14 (d, 1H, J = 8.4 Hz, 1-H). <sup>13</sup>C-NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.5 (C-18), 21.1 (OAc-CH<sub>3</sub>), 25.9, 27.8, 29.6, 32.2 (2C), 36.7, 38.5, 43.3, 44.8 (C-13), 49.5, 50.1, 85.2 (C-17), 112.7 (C-2), 115.2 (C-4), 126.5 (C-1); 132.4 (C-10), 138.1 (C-5), 153.5 (C-3), 170.6 (OAc-CO); EI-MS (7 eV) m/z (%): 408 (96) [M<sup>+</sup> + 2], 406 (100) [M<sup>+</sup>], 267 (17), 172 (21), 160 (78), 146 (39), 133 (50), 43 (100).

### 2.5.3.2. Method C.

2.5.3.2.1. 16β-Bromomethylestra-1,3,5(10)-triene-3,17αdiyl diacetate (8d). Compound 8d (1.03 g, 76%) as a colorless oil;  $R_f = 0.50$  (ss A);  $[α]_D^{20} + 36$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 61.58; H, 6.82.C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>Br requires: C, 61.47; H, 6.50%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.08 (s, 3H, 17-OAc-CH<sub>3</sub>), 2.27 (s, 3H, 3-OAc-CH<sub>3</sub>), 2.86 (m, 2H, 6-H<sub>2</sub>), 3.43 (t, 1H, J = 9.6 Hz) and 3.74 (dd, 1H, J = 9.6 Hz, J = 6.6 Hz): 16a-H<sub>2</sub>, 4.62 (d, 1H, J = 1.8 Hz, 17-H), 6.79 (d, 1H, J = 1.8 Hz, 4-H), 6.83 (dd, 1H, J = 8.4 Hz, J = 1.8 Hz, 2-H), 7.27 (d, 1H, J = 8.4 Hz, 1-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.4 (C-18), 21.1 (2C, OAc-CH<sub>3</sub>), 25.6, 27.6, 29.4, 32.2 (2C), 36.6, 38.1, 43.4, 44.7 (C-13), 49.5, 50.1, 85.1 (C-17), 118.6 (C-2), 121.5 (C-4), 126.3 (C-1), 137.6 (C-10), 138.0 (C-5), 148.4 (C-3), 169.8 (3-OAc-CO), 170.4 (17-OAc-CO); EI-MS (7 eV) *m*/z (%): 450 (16) [M<sup>+</sup>+2], 448 (16) [M<sup>+</sup>], 408 (17), 406 (19), 160 (6), 85 (15), 83 (26), 57 (10), 43 (100).

# 2.6. 3-Benzyloxy-16-bromomethylestra-1,3,5(10)-trien-17-yl acetate (7b and 8b)

### 2.6.1. General procedure

Compound **7a** or **8a** (455 mg, 1 mmol) was dissolved in 2 ml of pyridine and 2 ml of acetic anhydride. The solution was allowed to stand at room temperature for 6 h. The mixture was then diluted with water. The precipitate was collected by filtration and dissolved in  $CH_2Cl_2$ , and the solution was washed with water and dried over  $Na_2SO_4$ . After evaporation in vacuo, the product was chromatographed on silica gel with ethyl acetate/CHCl<sub>3</sub> (2.5:97.5).

**7b** (483 mg, 97%). Mp 120–121 °C;  $R_f = 0.65$  (ss A);  $[\alpha]_D^{20} + 12$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 67.51; H, 6.75.  $C_{28}H_33O_3Br$  requires: C, 67.60; H, 6.69%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.85 (s, 3H, 18-H<sub>3</sub>), 2.08 (s, 3H, OAc-CH<sub>3</sub>), 2.83 (m, 2H, 6-H<sub>2</sub>), 3.42 (dd, 1H, *J* = 9.8 Hz, *J* = 7.7 Hz) and 3.57 (dd, 1H, *J* = 9.8 Hz, *J* = 5.1 Hz): 16a-H<sub>2</sub>, 4.70 (d, 1H, *J* = 7.7 Hz, 17-H), 5.01 (s, 2H, Bzl-H<sub>2</sub>), 6.70 (d, 1H, *J* = 2.2 Hz, 4-H), 6.76 (dd, 1H, *J* = 8.5 Hz, *J* = 2.5 Hz, 2-H), 7.17 (d, 1H, *J* = 8.6 Hz, 1-H), 7.31 (t, 1H, *J* = 7.3 Hz, 4'-H), 7.36 (t, 2H, *J* = 7.3 Hz, 3',5'-H), 7.41 (d, 2H, *J* = 7.3 Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 12.8 (C-18), 21.1 (Ac-CH<sub>3</sub>), 26.0, 27.1, 29.6, 29.8, 36.8, 37.7, 38.3, 43.6 (2C), 44.7 (C-13), 48.4, 69.9 (C-Bzl-CH<sub>2</sub>), 85.0 (C-17), 112.3 (C-2), 114.8 (C-4), 126.2 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.5 (C-10), 137.3 (C-1'), 137.8 (C-5), 156.7 (C-3), 171.0 (OAc-CO); EI-MS (7 eV) *m*/z (%): 498 (25) [M<sup>+</sup> + 2], 496 (25) [M<sup>+</sup>], 91 (100), 43 (11).

**8b** (478 mg, 96%) as a colorless oil;  $R_f = 0.70$  (ss A);  $[\alpha]_D^{20} + 37$  (c 1.00 in CHCl<sub>3</sub>). (Found: C, 67.48; H, 6.75. C<sub>28</sub>H<sub>33</sub>O<sub>3</sub>Br requires: C, 67.60; H, 6.69%). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 0.86 (s, 3H, 18-H<sub>3</sub>), 2.08 (s, 3H, OAc-CH<sub>3</sub>), 2.85 (m, 2H, 6-H<sub>2</sub>), 3.42 (t, 1H, *J* = 9.7 Hz) and 3.74 (dd, 1H, *J* = 9.7 Hz, *J* = 6.5 Hz): 16a-H<sub>2</sub>, 4.62 (d, 1H, *J* = 2.1 Hz, 17-H), 5.02 (s, 2H, Bzl-H<sub>2</sub>), 6.71 (d, 1H, *J* = 2.5 Hz, 4-H), 6.77 (dd, 1H, *J* = 8.6 Hz, *J* = 2.5 Hz, 2-H), 7.19 (d, 1H, *J* = 8.6 Hz, 1-H), 7.31 (t, 1H, *J* = 7.3 Hz, 4'-H), 7.37 (t, 2H, *J* = 7.3 Hz, 3',5'-H), 7.42 (d, 2H, *J* = 7.3 Hz, 2',6'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ , ppm: 17.44 (C-18), 21.1 (OAc-CH<sub>3</sub>), 25.8, 27.8, 29.7, 32.2 (2C), 36.7, 38.5, 43.3, 44.8 (C-13), 49.5, 50.1, 69.9 (Bzl-CH<sub>2</sub>), 85.1 (C-17), 112.3 (C-2), 114.8 (C-4), 126.3 (C-1); 127.4 (2C, C-2',6'), 127.8 (C-4'), 128.5 (2C, C-3',5'), 132.6 (C-10), 137.3 (C-1'), 137.8 (C-5), 156.8 (C-3), 170.5 (OAc-CO); EI-MS (7 eV) *m*/z (%): 498 (9) [M<sup>+</sup> + 2], 496 (13) [M<sup>+</sup>], 91 (100), 43 (10).

### 2.7. Receptor-binding assay

Eight synthesized compounds (**5c–8c** and **5e–8e**) were evaluated for their ability to inhibit competitively the binding of [<sup>3</sup>H]estra-1,3,5(10)-triene-3,17 $\beta$ -diol ([<sup>3</sup>H]E2; 110 Ci mmol<sup>-1</sup>) to ERs, of [<sup>3</sup>H]ORG2058 (34 Ci mmol<sup>-1</sup>) to PRs in cytosol prepared from ovariectomized rat uteri, and of [<sup>3</sup>H]dihydrotestosterone ([<sup>3</sup>H]DHT; 110 Ci mmol<sup>-1</sup>) to ARs in castrated rat prostate cytosol [15].

Mature female (200–300 g) and male (300–400 g) rats were gonadectomized under general anesthesia (sodium

pentobarbital 1 g/kg i.p.) 48 or 24 h before the tissue preparation, respectively.

The tritiated ligands were purchased from Perkin-Elmer, USA, except for [3H]ORG2058, which was obtained from Amersham, UK. All subsequent steps were carried out at 4°C. The minced uterine tissues were homogenized in 10 volume (w/v) of buffer A (25 mM Tris-HCl, 1.5 mM EDTA disodium salt, 10 mM α-monothioglycerol, 10% glycerol and 10 mM sodium molybdate, pH 7.4) to measure the abilities of the compounds to bind to the ERs or PRs [16]. The ventral prostates tissues were homogenized in 3 volume (v/v) of buffer I (20 mM Tris-HCl, 1 mM EDTA disodium salt, 1 mM dithiothreitol and 10% glycerol, pH 7.5) to measure the abilities of the compounds to bind to the ARs [17]. The homogenates were centrifuged at  $105,000 \times g$  for 60 min and the supernatants (cytosol) were used for competitive binding. Aliquots of  $100\,\mu l$  of cytosol were incubated with 5 nM [<sup>3</sup>H]E2, 4 nM [<sup>3</sup>H]ORG2058 or 5 nM [<sup>3</sup>H]DHT in the presence of synthesized molecules in 11 different concentrations (from  $10^{-12}$  to  $10^{-5}$  M) for 3 h at 25  $^\circ C$  for ER, for 24 h at 4 °C for PRs, and for 4 h at 4 °C for ARs. To determine the total bound radiolabeled ligand, we incubated  $100 \,\mu$ l of cytosol with  $100\,\mu$ l of isotope and  $100\,\mu$ l of assay buffer. Unbound steroids were separated by incubation for 15 min at 4 °C with dextran-coated charcoal (0.5% Norit-A and 0.05% dextran T-70 in buffer B (1.5 mM EDTA disodium salt, 10 mM  $\alpha$ -monothioglycerol and 10 mM Tris–HCl, pH 7.4) for ERs and PRs, and for 30 min at 4 °C with 1.25% Norit-A and 0.625% dextran T-70 in buffer II (10 mM Tris-HCl and 1 mM EDTA disodium salt, pH 7.5) for ARs. After incubation, the samples were centrifuged at 3000 rpm for 10 min, and the radioactivity of the supernatant was determined in a Wallac 1409 liquid scintillation counter. The percentage of the bound radioligand in the presence of competitor was plotted against the concentration of unlabeled steroid. The molar concentration of the steroid competitor that reduced the radioligand binding by 50% (IC<sub>50</sub>) and the  $K_i$  value (inhibition constant) were calculated with GraphPad 2.0 software. To compare the ER-binding abilities of the investigated molecules with the  $K_i$  value of the reference compound, we calculated the relative binding affinity (RBA) according to the following equation: RBA = ( $K_{i,E2}/K_{i,x}$ ) × 100. All assays were carried out at least three times.

# 3. Results and discussion

## 3.1. Synthetic studies

In preliminary experiments, the four possible isomers of 3-benzyloxy-16-hydroxymethylestra-1,3,5(10)-trien-17-ol isomers (1a–4a) were synthesized via a multistep pathway [14] in order to obtain the 16-bromomethyl-3-hydroxyestra-1,3,5(10)-trien-17-yl acetates (5c–8c) and the parent 16-bromomethyl-diols (5e–8e) derivatives.

First we studied the behavior of the four starting materials **1a–4a** in the presence of HBr and acetic acid. From the cis diols **1a**, and **2a**, we obtained different 16-bromomethyl-17-acetates (**5b–d** and **6b–d**) which, depending on the reaction time or conditions, differed from each other in the substituent at position C-3 (Schemes 1 and 2). Reactions for 1 h led to **5b** and **6b**, which have benzyl protecting groups at position C-3. After 12 h, the bromo substitution was at C-16a and deben-zylation also occured. By this method, the cis diols furnished



A: HBF / acetic acid 1n B: HBr / acetic acid 12h C: HBr / acetic acid 12 h + acetic anhydride 6h D: CBr<sub>4</sub> / (Ph)<sub>3</sub>P



3-deprotected 16-bromomethyl derivatives (**5c** and **6c**), which were subjected to receptor-binding examinations directly. In the following experiments, the 3-hydroxy function was transformed into an acetoxy group by adding acetic anhydride to the reaction mixture, yielding **5d** and **6d**. This procedure allowed us to replace the benzyl protecting group with the acetate group, which is easily removable and convenient. The 16-bromomethylestradiol derivatives were accompanied by 16-acetoxymethylestradiol derivatives (**1b–d** and **2b–d**) as byproducts in low yields.

In contrast, from the trans diols (**3a** and **4a**), 16bromomethyl products were not formed (Schemes 3 and 4). Instead of bromination, acetylation occurred, yielding different diacetoxyestradiol derivatives (**3b-d** and **4b-d**), depending on the reaction conditions. In the case of 3-benzyloxy-16βhydroxymethylestra-1,3,5(10)-trien-17 $\alpha$ -ol (**4a**), compound **16b** was formed as a by-product via Wagner–Meerwein rearrangement.

The experimental results reveal that the four diols (1a–4a) take part in stereoselective substitution in the presence of HBr and acetic acid. The formation of 16-bromomethyl-estradiol derivatives **5b** and **6b** can easily be explained in terms of front-side neighboring group participation [18]. According to the earlier observations the 16-hydroxymethyl group of **1a** and **2a** undergoes acetylation and generates **9** and **12**, which are formed under the appropriate reaction conditions. These monoacetates (**9** and **12**) cyclized into an *ortho* acid–*ortho* ester (**10** and **13**) in the acid medium, which are transformed via the ambidentate 2'-methyl-1',3'-dioxenium ions **11** and **14** into **5b** and **6b**. During the formation of **5b** and **6b**, no change in config-

uration occurs, since ring cleavage in the ambidentate cations 11 and 14 takes place at the sterically favored site C-16a [19]. Furthermore, the absence of 16-bromomethylestradiol derivatives in the case of the trans diols **3a** and **4a** can be explained by the rigidity of the estrane skeleton and the unfavorable configuration of the substituents, which should take part in a formation of six-membered acyloxonium cation intermediates. The formation of 16b from 4a can be explained by a Wagner-Meerwein rearrangement: the secondary hydroxy function at C-17 is protonated and a carbocation 15 is generated after the elimination of water under the appropriate reaction conditions. This carbocation (15) furnishes 16b through migration of the angular methyl group and subsequent deprotonation. The isolated double bond of the tetraene of type 16b during a prolonged reaction time with HBr addition produced very sensitive undefined compounds.

The above experimental findings led us to develop a convenient method for the synthesis of trans 16-bromomethyl-17-acetate derivatives (7 and 8). First we brominated the trans diols 3a and 4a by the Appel reaction and obtained the trans 3-benzyloxy-16-bromomethylestra-1,3,5(10)-trien-17-ols (7a and 8a). These compounds (7a and 8a) permitted direct synthesis of the trans 16-bromomethyl-3,17-diacetates 7d and 8d by conversion of the substituent at C-3. The acetylation of 7a and 8a with acetic anhydride yielded 17acetoxy compounds 7b and 8b in good yields. The transprotected derivatives 7d and 8d were synthesized from 7b and 8b by debenzylation with HBr and the subsequent addition of acetic anhydride to the reaction mixture in one step.



After the synthesis of 17-acetoxy-16-bromomethyl-3hydroxy compounds (**5c-8c**) we set out to produce the appropriate 17-hydroxy derivatives, too. This was reasonable in view of their potential biological activities, while 17-hydroxyestradiol derivatives presumably have higher RBAs than their 17-acetoxy counterparts. The cis 16bromomethylestradiol derivatives **5a** and **6a** could also be obtained by application of the Appel reaction. The 16bromomethyl-17-hydroxy compounds **5a-8a** were deprotected by hydrogenolysis, which yielded the corresponding estradiols (**5e-8e**). The synthesized products could be subjected to receptor-binding examinations directly.

The NMR spectra demonstrated certain interesting features. In the <sup>1</sup>H NMR spectra of the 16-bromomethyl-17acetates (**5b–8b**, **d**) and 16-bromomethylestradiol derivatives (**5e–8e**), the double doublet or triplet of the 16a-H<sub>2</sub> appears at  $\delta$ =3.26–3.76 ppm. The analogous signals of the 16-acetoxymethyl-17-acetates (**1b–4b**, **d**) are found at higher chemical shifts, around 4.0–4.2 ppm). They also occur at higher ppm values than the similar signals of the cis isomers.

Further, the doublet assigned to 17-H in 16-bromomethyl,17-acetates (**5b–8b**, **d**) and 16-acetoxymethyl-17acetates (**1b–4b**, **d**) appears at  $\delta$ =4.62–5.10 ppm. For 16bromomethylestradiol derivatives (**5e–8e**), the signal of 17-H is observed at higher frequencies,  $\delta$ =3.47–3.82 ppm, as compared with the 17-acetates. As opposed to the 16a-H signals, the doublets of 17-H of the corresponding cis isomers lie at higher ppm values than in the spectra of the trans isomers. In agreement with the literature [20], the coupling constants  $J_{16,17}$  display the following sequence:  $J_{16\alpha H,17\beta H} < J_{16\beta H,17\beta H} < J_{16\beta H,17\alpha H} < J_{16\alpha H,17\alpha H}$ . The <sup>13</sup>C NMR resonance of C-17 is usually observed at  $\delta$ =79.9–85.8 ppm and this signal of the corresponding trans isomer is found at higher ppm values than for the analogous signals of the cis isomers.

For the products containing a benzyl protecting group at position C-3 (**1a–8a**, **b**, **d**), the singlet of the benzyl-CH<sub>2</sub> appears in the <sup>1</sup>H NMR spectra at  $\delta$  = 5.02 ppm and in the <sup>13</sup>C NMR spectra at  $\delta$  = 70.0 ppm. The resonance of C-3 in the <sup>13</sup>C NMR spectra is observed at  $\delta$  = 148.5–156.8 ppm. The singlet at  $\delta$  = 156.8 ppm can be assigned to bearing the C-3 benzyloxy group, that at  $\delta$  = 153.5 ppm to the C-3 hydroxy function, and that at  $\delta$  = 148.5 ppm to the C-3 acetoxy group. Additionally, the OAc-CH<sub>3</sub> at position C-3 in **1d–8d** resonates at  $\delta$  = 2.27 ppm in <sup>1</sup>H NMR spectra and at 5 = 21.0 ppm in the <sup>13</sup>C NMR spectra. Finally, the signal from C=O (3-OAc) is found at  $\delta$  = 169.9 ppm.

#### 3.2. Radioligand-binding assay

The RBAs of 16-bromomethyl-3-hydroxyestra-1,3,5(10)-trien-17-yl 17-acetates (**5c–8c**) and 16-bromomethylestra-1,3,5(10)triene-3,17-diols (**5e–8e**) and their derivatives for the ERs, PRs and ARs were determined by radioligand-binding assay.

Our reference molecule for the ERs was estra-1,3,5(10)-triene-3,17 $\beta$ -diol (E2), the inhibition constant of which under our experimental conditions ( $K_i = 2.31 \pm 0.76$  nM) was in good agreement with the literature  $K_i$  value (2.48 nM) [16].

Specifically bound [<sup>3</sup>H]E2 was displaced from the ovariectomized rat uterine ERs by **5c** and its stereoisomers (**6c–8c**).  $16\alpha$ -Bromomethylestra-1,3,5(10)-triene-3,17-diyl 17\beta-acetate (**7c**) displays the highest binding affinity to the ERs in this series, though its  $K_i$  is two orders of magnitude lower than that of **E2**, the endogenous ligand of the ER. The other three molecules (**5c**, **6c** and **8c**) have  $K_i$  values one order of magnitude higher than that for **7c**, and there is no major difference between the RBAs of the compounds. The  $K_i$  values are listed



Scheme 4

in Table 1, and representative curves from the ER displacement analysis are depicted in Fig. 1.

The reference molecule for the PR was **ORG2058**, with  $K_i = 7.40 \pm 0.98$  nM under our experimental circumstances. All

of the 17-acetate derivatives of estradiol (5c-8c) bind weakly to the PRs, with  $K_i$  values in excess of 3000 nM, and thus this receptor does not recognize them. Three compounds in this series exhibit a higher affinity to the ARs than to the ERs. The

Table 1 – Inhibition constants ( $K_i$ ) of estra-1,3,5(10)-triene-3,17 $\beta$ -diol (E2), ORG2058 and dihydrotestosterone (DHT) as reference compounds and currently synthesized derivatives for the uterine ERs and PRs and for the prostate ARs

Investigated compounds	K <sub>i</sub> (nM) ER	RBA (%)	K <sub>i</sub> (nM) PR	K <sub>i</sub> (nM) AR
E2	2.31 ± 0.76	100	-	-
ORG2058	-	-	7.40±0.98	-
DHT	-	-	-	9.17±0.71
5c	$\begin{array}{c} 3855 \pm 2350 \\ 553.8 \pm 198.3 \\ 5254 \pm 1040 \\ 4143 \pm 2269 \end{array}$	0.06	≥3000	Not measurable <sup>a</sup>
7c		0.42	≥3000	475.5 ± 7.28
8c		0.04	≥3000	2963 ± 1528
6c		0.09	≥3000	237.2 ± 23.76
5e	$\begin{array}{c} 461.4 \pm 145.6 \\ 2.55 \pm 0.64 \\ 781.1 \pm 149.1 \\ 684.4 \pm 154.0 \end{array}$	0.50	≥3000	≥3000
7e		90.59	525.2±142.7	≥3000
8e		0.29	≥3000	≥3000
6e		0.34	≥3000	214.7 ± 10.96

The following radioligands were used: 5 nM [<sup>3</sup>H]estra-1,3,5(10)-triene-3,17 $\beta$ -diol for the ERs, 4 nM [<sup>3</sup>H]ORG2058 for the PRs and 5 nM [<sup>3</sup>H]dihydrotestosterone for the ARs. The relative binding affinities (RBAs) on the ERs were calculated according to the following equation: RBA = ( $K_{i;E2}/K_{i;x}$ ) × 100.

<sup>a</sup>  $K_i > 10,000$  nM; the ligand is not able to displace the radioligand from the receptor-binding site.



Fig. 1 – Competitive inhibition of  $[{}^{3}H]$ estra-1,3,5(10)-triene-3,17 $\beta$ -diol binding in rat uterine cytosol by 16-bromomethyl-3-hydroxyestra-1,3,5(10)-trien-17-yl 17-acetate (A) and 16-bromomethylestra-1,3,5(10)-triene-3,17-diol (B) and their stereoisomers. The reference molecule is estra-1,3,5(10)-triene-3,17b-diol (E2).

most marked difference is observed in the case of **6c**, which binds one order of magnitude more strongly to the ARs than to the ERs. Its  $K_i$  value for the ARs is the lowest in this group. The isomer (**5c**) could not displace specifically bound [<sup>3</sup>H]DHT from the castrated rat prostate ARs.

The 16-bromomethylestrane derivatives of 3,17-estradiol (**5e–8e**) recognize the ERs well. Compound **7e** exhibits the highest affinity to this receptor ( $K_i = 2.55 \pm 0.64$  nM), and this compound therefore binds to the ERs as strongly as **E2** does. The  $K_i$  values of the other molecules (**5e, 6e** and **8e**) are at least 200-fold higher than that for **7e**.

Compound **7e** displays the highest affinity for the PRs, but this interaction is 200-fold weaker than that with the ERs. The remaining three derivatives have  $K_i$  values higher than 3000 nM. The members of this series display weak affinity for the ARs ( $K_i > 3000$  nM), except for the 16 $\alpha$ -bromomethylestra-1,3,5(10)-triene-3,17 $\alpha$ -diol substituent (**6e**). This latter exhibits the strongest RBA to the ARs. It binds 3-fold more strongly to the ARs than to the ERs, but its interactions with the ARs are 23-fold weaker than that of to the reference compound DHT ( $K_i = 9.17 \pm 0.71$  nM).

#### 3.3. Discussion

In the present study, the binding affinities of eight estradiol derivatives containing substituents at positions C- 16 and C-17 to the ERs, PRs and ARs have been characterized. The molecules belong in two different substituted stereoisomer series: 16-bromomethylestra-1,3,5(10)triene-3,17-diyl 17-acetates (**5c–8c**) and 16-bromomethylestra-1,3,5(10)-triene-3,17-diols (**5e–8e**).

Examination of the binding abilities of these compounds to the ERs proved that this receptor recognizes them. Considerable differences are observed between the K<sub>i</sub> values, which can be explained by the different modes of receptor binding. Tanenbaum et al. [21] suggest that estrogens adopt a welldefined orientation in the ligand-binding domain (LBD) of the ERs: ring D is in contact with helix-11, the 17β-hydroxy group is H-bonded to His 524, ring A is projected toward helix-3, the 3-phenolic hydroxy forms a water-mediated H-bond with Glu 353, and the molecule is fixed by van der Waals contact around the binding pocket. The biggest divergence in the K<sub>i</sub> values is exhibited between the members of the 17-acetate epimers and the 3,17-diols, suggesting that a hydroxy group at position  $17\beta$ of the estradiol ring is necessary to form the H-bond with the His 524 residue. Highly polar and bulky groups, such as acetate, are poorly tolerated by ER LBD at positions  $17\alpha$  and  $\beta$  [22].

Several research groups have investigated the effects of C16 substitution on the affinity of estradiol derivatives for the ERs [13,23]. They concluded that ER tolerance for substituents at position  $16\alpha$  is mixed, although groups of moderate size and polarity, such as halogens or haloalkyl, are generally tolerated, but position  $16\beta$  is sterically less permissive, even the smallest functional group (e.g. fluorine) decreasing the RBA substantially. Our experimental work with the four estradiol epimer pairs lend support to these findings, because the tested  $17\beta$  derivatives with  $16\alpha$  substituents (7c and 7e) possess  $K_i$ values at least one order of magnitude lower than those of  $16\beta$ -substituted molecules. The RBA of 7e is 90.95%, and this epimer has a two-orders of magnitude lower K<sub>i</sub> than that of its cis counterpart 5e. Ring D of estradiol derivatives makes nonpolar contacts with the Ile 424, Gly 521 and Leu 525 residues, and the Ile 424 residue can be found close to C-16 of the ligand in the ERLBD [24]. Our results suggest that the position  $16\beta$  is sterically unfavorable because of the side-chain of the Ile 424 residue, which is close to the ligand.

The investigated molecules are able to bind to the ARs and PRs, because the C-3 hydroxy group can act not only as an H-donor, but also as an H-acceptor [22], and in the AR and PR there is a good H-donor Glu residue instead of the Gln 353 in the ERs [21,25].

In summary, four substituted estradiol epimer pairs were tested in the radioligand-binding assay. The ER recognizes them, but only **7e** exhibits a really high binding affinity for the ERs. The selectivities of the investigated compounds are variable: only three derivatives (**5e**, **7e** and **8e**) are considered ER-selective. The others, (**7c**, **8c** and **6c** and **6e**) bind slightly more strongly to the ARs than to the ERs, and **5c** does not show any affinity to the ARs.

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