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Syntheses of Hemoprotein Models that can be Covalently Attached onto Electrode Surfaces by Click Chemistry

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Five alkyne-containing hemoprotein models have been synthesized in a convergent manner. Sonogashira coupling was used to introduce the alkyne functional group on the proximal imidazole before or after being attached on the porphyrin. One model was immobilized onto a gold electrode surface via copper-(I)-catalyzed azide—alkyne cycloaddition (Sharpless click chemistry).

Introduction

Cytochrome *c* oxidase (CcO) is the heme/copper oxidase in the respiratory chains of mitochondria and aerobic bacteria that performs the 4e⁻ reduction of dioxygen to water.¹ On the basis of crystal structures,^{2a,b} our group has designed several models that structurally mimic the active site of the enzyme.^{3a,d} Past electrocatalytic studies of O₂ reduction with these models have been conducted on an edge-plane-graphite electrode. Though convenient, this method relies on poorly defined physisorbed catalyst films that are in direct contact with the electrode allowing rapid electron delivery.^{4a,c} To allow a slow and controlled rate of electron transfer as in the enzyme, we proposed to tailor the length and conjugation of azide-terminated alkyl chains that form self-assembled monolayers (SAM) coating the electrode. Such attachment has been achieved by the use of copper(I)-catalyzed azide–alkyne cycloaddition, a form of

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FIGURE 1. Click Chemistry on Surfaces. Reagents and conditions: (a) sodium ascorbate, Cu(I), R-CCH, where R = ferrocene, DNA, a hemoprotein model such as 1–5 (example with 5b in Scheme 7).



FIGURE 2. Alkyne-containing iron porphyrins 1-5 with increasing structural complexity for covalent attachment onto Au surfaces and the corresponding key alkyne-containing imidazole synthons (6a,b) that models 2-5 have in common.

Sharpless "click" chemistry⁵ that we and others have adapted for the functionalization of surfaces (Figure 1).^{6a–e} The conditions required for such reactions are mild and chemoselective, allowing for compatibility with a large number of functional groups.^{5,6a–e} Moreover, this method is a practical alternative to previous immobilization methods on gold surfaces^{6f,g} because it affords quantitative yields and no side products.^{5,6a–e}

This present study is aimed at designing and synthesizing alkyne-containing CcO models and at clicking these models on

azide-functionalized electrodes. The structural complexity is gradually increased from model **1** to model **5** (Figure 2) to revisit, under the regime of slow electron transfer, the role played by each key component of the CcO active site in the reduction of dioxygen. From a simple iron porphyrin (model **1**, that we previously clicked on a SAM),^{6e} we have constructed a built-in proximal base mimicking His376 (model **2**). Then, a series of distal superstructures were added to a $\alpha_3\beta$ -porphyrin atropisomer. Three simple picket fences in model **3** are designed to

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SCHEME 1. Synthesis of TMSAlk-Ester (12)^a



^{*a*} Reagents and conditions: (a) formamide, 155 °C, 4 h, 34%; (b) acrylonitrile, vacuum, 190 °C, 2 h, 74%; (c) 3-bromomethylphenyl-3-methylcarboxylate, MeCN, 4.5 h, 100 °C, 48% (**10**), 80% (**14**); (d) CH₃ONa/CH₃OH, rt, 45 min, 48%; (e) trimethylsilylacetylene, Pd(PPh₃)₄, CuI, Et₃N, THF, rt, 75% (**12**), 88% (**13**), 70% (**14**).

SCHEME 2. Deprotection of Ester and Alkyne Functions in 12: Synthesis of Alk-Acid 16^a



^{*a*} Reagents and conditions: (a) 1.5 N KOH, THF/CH₃OH/H₂O 1:1:2 vol, 1 h; (b) AcOH, excess H₂O, 10 min, 85% (from **12**), 48% (from **14**); (c) KF or K₂CO₃, CH₃OH, rt, 4 h, quantitative; (d) HCl, reflux, quantitative; (e) trimethylsilylacetylene, Pd(PPh₃)₄, CuI, NEt₃, THF, rt, 88%; (f) LiI, acetone, rt, 24 h, 50%; (g) CH₃ONa/CH₃OH/H₂O, rt 1.5 h, 60%; (h) (COCl)₂, MeCN, DMF (cat.), rt, 2-4 h, 80% quantitative.

stabilize the oxygen complex with respect to 2 because of the hydrophobic environment provided by the *t*-Bu groups and the stabilizing hydrogen bond between dioxygen and a proton of

an amide. Then, Cu^{I} in a trisimidazole environment is added (model **4**, mimicking Cu_{B} , His240, His290, and His291) at a biomimetic distance from the Fe center (ca. 5 Å). The final

SCHEME 3. (A) Synthesis of Trisimidazole-alkyne-Tailed Model 4 and (B) Synthesis of Picket-Fence-Tailed Model 3 and "Flat"-Tailed Model 2^a

Α



^{*a*} Reagents and conditions: (a) **6a** or **6b**, MeCN/THF 2:1 vol, rt, 30 min, 70–80%; (b) NH₃-saturated MeOH/CH₂Cl₂ 3:7 vol, 65%; (c) *N*-methyl imidazole acyl chloride, ^{3c} MeCN–THF, 70%; (d) (i) FeBr₂, THF/MeOH, reflux, 1 h (**24b**, **25**) or 10 h (**22**), (ii) EDTA, H₂O (for **Fe-24b** only), ca. 80%; (e) CuPF₆(MeCN)₄, MeCN, THF, rt, 10 min, quantitative; (f) pivaloyl chloride, THF, Et₂NC₆H₅, rt, 1 h, 88%.

model and closest structural analogue of the cytochrome c oxidase active site (model **5a**) was designed to have one imidazole cross-linked to a phenol mimicking Tyr244.

The site chosen for introducing the alkyne function is the para position of the phenyl ring that is grafted on carbon 5 of the proximal imidazole tail. This strategy depends on an imidazole synthon 6 that is crucial for models 2-5 to undergo click chemistry. During the convergent synthesis of models 2-5, the coupling of the acetylene to the imidazole ring can be carried out either before (6b) or after (6a) being attached to the porphyrin. With such a location for the alkyne, the electrons tunneling from the electrode through the alkyl chain are expected to reach the iron center of the immobilized bimetallic catalyst first.

Results and Discussion

The first stage in the synthesis of **12**, the ester precursor of the proximal imidazole tail **6a,b**, involved the preparation of the Michael acceptor I-Mich **7** (Scheme 1, route A). This was prepared by acid-catalyzed bromination of 4-iodoacetophenone

with Br_2 leading to bromoacetophenone I-Br **8**, which subsequently was treated with formamide to give imidazole I-Im **9** in 45% yield. The Horvath protection strategy⁷ conducted on **9** led to I-Mich **7** in a regiospecific manner. Two equally efficient

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⁽⁸⁾ Sonogashira, K.; Tohada, Y.; Hagira, N. Tetrahedron Lett. 1975, 4467 (9) (a) Free acid-sensitive alkyne could no longer sustain harsh HCl refluxing conditions previously used for the imidazole ester hydrolysis.³⁰ Instead, LiI or saponification was more appropriate. (b) This delicate step requires starting from a pure ester precursor, free of Pd contaminants, and also prohibits long reaction times or a high concentration of NaOH solution. (c) No reaction of acyl chloride with alkyne was noticed unlike in ref 9d,e. (d) Brownstein, S.; Morrison, A.; Tan, L. K. J. Org. Chem. 1985, 50, 2796. (e) Martens, H.; Janssens, F.; Hoornaert, G. Tetrahedron 1975, 31, 177. (f) Cu can be removed by acidic treatment but not Pd. (g) Porphyrin 30-mono could also be recycled and used for the synthesis of 30-bis following a method reminiscent of the synthesis of 2 + 1 species **32a,c,d** (not shown). (h) For simplification, only the cis regioisomer of bisimidazolyl-porphyrin synthon 30 is depicted in Scheme 5. The reaction of porphyrin 23b with N-methyl imidazole acyl chloride also leads to the trans regioisomer and to mono- and trisimidazolyl porphyrins (30-mono and 30-tris). (i) No side product was noticed from the metalation of alkyne-containing species with iron(II) and especially Cu(I) salts. (j) The position of the alkyne function allowed a click reaction to proceed.

SCHEME 4. Sonogashira Coupling onto Iodo-porphyrin^a



^{*a*} Reagents and conditions: (a) Zn(OAc)₂, CHCl₃/MeOH, 2:1 vol, rt, 2 h, 92–95%; (b) trimethylsilylacetylene, CuI, Pd(PPPh₃)₄, Et₃N, THF, rt, 18 h, quantitative; (c) HCl, CH₂Cl₂, rt, 15 min, 92% quantitative; (d) NH₃-saturated CH₃OH/CH₂Cl₂, 1:3 vol, rt, 3 days, 65%; (e) CH₂Cl₂, CH₃OH, K₂CO₃, rt, 16 h, quantitative.

SCHEME 5. Synthesis of the CcO Model Having a Tyr244 Mimic Following a 2 + 1 Approach^{*a*}



^{*a*} Reagents and conditions: (a) 2 equiv of *N*-methyl imidazole acyl chloride, ^{3c} MeCN–THF, rt, 20 min, 70%; (b) (i) HCl gas; (ii) **31ac**, MeCN/THF, rt, 70%; (c) (i) 20 equiv of BBr₃, CH₂Cl₂, -78 °C, 30 min, then 0 °C, 1 h, (ii) MeOH, H₂O, -78 °C, 30 min, then 0 °C, 1 h; (d) HCl, CH₂Cl₂, rt, 1 h, 80%; (e) (i) FeBr₂, THF/MeOH, reflux, 1 h, (ii) EDTA, H₂O, rt, 15 min, 80%; (f) Cu(MeCN)₄PF₆, MeCN/THF, rt, 10 min, quantitative.

routes were followed to obtain TMSAlk-Ester **12** from I-Mich **7**. Route A involved the quaternization of **7** (leading to I-q **10**), cyanoethyl deprotection (leading to I-Ester **11**), and Sonogashira alkyne coupling⁸ leading to TMSAlk-Ester **12**.

In route B, Sonogashira coupling is conducted first on I-Mich 7 leading to the alkyne-containing imidazole TMSAlk-Mich 13 in 75% yield which is then quaternized and deprotected leading to TMSAlk-q 14 and TMSAlk-Ester 12, respectively. Sono-gashira coupling was also carried out on imidazole salt I-q 10 despite its low solubility and led to 14.

Ester deprotection (Scheme 2) in the robust imidazoles I-ester 11 was carried out in refluxing acid and led quantitatively to

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I-acid 15. The acid-sensitive alkyne-ester 12^{9a} was deprotected by saponification in 1.5 N NaOH (or K₂CO₃/MeOH) at room temperature followed by neutralization with acetic acid and led to the alkyne-deprotected species Alk-Acid 16.^{9b} The use of milder reagents such as LiI under refluxing conditions also effected both deprotection reactions, whereas K₂CO₃ or KF led to 17 resulting from a selective deprotection of the alkyne. An alternative was to run the alkyne coupling on the poorly soluble iodo-imidazole I-acid 15, leading to TMSAlk-Acid 18, followed by treatment with a nucleophile giving 16. The most efficient method to synthesize 16 was achieved by the simultaneous removal of TMS, ester, and cyanoethyl protective groups in 14.

SCHEME 6. Synthesis of the Imidazole Picket Having a Tyr244 Mimic^{*a*}



^{*a*} Reagents and conditions: (a) 4-methoxybenzyl bromide (for **34b**), trityl bromide (for **34c**), K_2CO_3 , THF, rt, 24 h, 70% (**34bc**); (b) (i) 1.5 N NaOH, MeCN/THF/H₂O, (ii) AcOH, H₂O, 70%; (c) (COCl)₂, MeCN, DMF (cat.), rt, 3 h, 80%.

This triple deprotection reaction was carried out in a CH₃ONa/ CH₃OH/H₂O (1:1:0.1 mol) mixture in 60% yield. Finally, imidazole acyl chlorides **6a,b** were prepared by reaction of oxalyl chloride with their carboxylic acid precursors **15** and **16**.^{3c,9c-e}

The synthesis of models **3**–**5** represents significant modifications to the general synthetic scheme reported earlier for the synthesis of CcO models^{3a–d} because of the presence of the alkyne function. The iodo- or alkyne-tail acyl chlorides **6a** and **6b** reacted with porphyrin $\alpha_3 F\beta A$ **19**^{3b} (Scheme 3A) or trisphenyl *o*-aminophenyl porphyrin **20**^{10a} (Scheme 3B) leading to porphyrins $\alpha_3 F\beta XT$ **21a,b** (X = I, CCH) and **22** (tailed "flat" porphyrin), respectively. The three distal amines were deprotected to give the α_3 -amino synthon $\alpha_3 A\beta XT$ **23a,b** (X = I, CCH). This was used as a platform for reaction with *N*methylimidazole acyl chloride^{3c} or pivaloyl chloride^{10a,b} leading to trisimidazole-tailed porphyrin **24a,b** and picket-fence-tailed porphyrin **25**, respectively.

The introduction of the alkyne function was also carried out on the iodo-imidazole-tailed porphyrins 21a or 24a (Scheme 4). Prior protection of the porphyrin with zinc was mandatory to prevent metalation of the porphyrin by Cu or Pd from occurring during the coupling. After reaction of zinc porphyrins **26a,b** with trimethylsilylacetylene in the presence of Cu^I and Pd⁰ catalysts, zinc was removed by bubbling HCl through the solution of porphyrins 27a,b.9f The TMS protective group was removed in 28b by treatment of the porphyrin with a solution of K₂CO₃ leading to 24b. None of the four steps involved in the Sonogashira coupling on tetrakisimidazolyl porphyrins 24a could be monitored by TLC because the polarities of the reactant 24a and the product 24b are the same. This is unlike the coupling reaction carried out on less-polar zinc tristrifluoroacetamidoporphyrin 26a. Interestingly, once the protected alkyne group had been coupled to the porphyrin $\alpha_3 F\beta IT$ 21a, the TMS protective group remained stable during all subsequent steps of the synthetic scheme (Scheme 4), such as the deprotection of the distal amines in 28a leading to 29. This offered substantial advantages for the purification of porphyrin intermediates (solubility and separation of regioisomers).

The preparation of model **5** having a Tyr244 mimic followed the 2 + 1 strategy previously reported for the CF₃ counterpart (Scheme 5).^{3d} Reaction of the freshly prepared *N*-methylimidazole acyl chloride picket^{3c} with **23b** led to the bisimidazolyl

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porphyrin **30**.^{9g,h} Efficient separation of the regioisomers of this alkyne-containing synthon was achieved on rotating chromatography plates (chromatotron) without decomposition of the porphyrins. However, 30 appeared unstable on preparative TLC plates, unlike CcO models bearing the other imidazole tails reported previously.^{3d} After mandatory reprotection of the imidazole pickets of cis regioisomer 30 by acidification,3b,d reaction with a methyl-protected tyrosine imidazole picket acyl chloride^{3d} yielded the protected 2 + 1 species **32a**. Phenol deprotection of 32a with BBr3 led to a mixture of 32b and a compound that corresponds to a bromoalkene derivative, as shown by mass spectrometry with an additional peak at m/z =1444. Integration of the signal corresponding to the benzylic methylene on the NMR spectrum shows that the contaminant is present in 10-25%. Separation by chromatography could not be achieved because they have similar polarities. Click chemistry on the SAM-coated electrode using a metalated version of this mixture appeared to be the only way to remove the bromoalkene-containing porphyrin: the model containing an alkyne was clicked on the electrode, whereas the bromoalkene side product did not react and remained in solution. To overcome these problems, the syntheses of phenol-protected imidazole synthons 31b,c bearing various protective groups (Scheme 6) and the corresponding 2 + 1 porphyrin intermediates 32c,d (Scheme 5) were undertaken. Trityl (Tr) and *p*-methoxybenzyl (PMB) were chosen as protective groups because they offer a good compromise between robustness and lability under various acidic conditions.^{11a-c} They were chosen because they could possibly be sufficiently acid-resistant during the steps where acid is necessary or generated in small amounts. These steps are the neutralization with 1 equiv of acetic acid after saponification of the imidazole ester leading to 35b,c, the acyl chloride synthesis leading to 31b,c, and the condensation reaction of 31b,c with the bisimidazolyl porphyrin 30 in preacidified media leading to 32c,d. However, the PMB and Tr protective groups display enough acid sensitivity to be removed under mild conditions to prevent side reactions on the proximal alkyne.

Reaction of phenol-imidazole ester 33^{12} with *p*-methoxybenzyl bromide or trityl bromide, in the presence of potassium carbonate, led to imidazole 34b,c, respectively. After saponification and careful neutralization, imidazole acids 35b,c were converted into acyl chlorides **31b,c** under anhydrous conditions. Reaction of **31b**,c with the bisimidazolyl porphyrin **30** proceeded as with OMe-protected phenol 31a (Scheme 6). Both protective groups turned out to be too labile to resist the acidic conditions in the 2 + 1 condensation between **31b.c** and the bisimidazolyl porphyrin 30 (although they were sufficiently robust to achieve purification of the imidazole ester precursors 34b,c by long chromatography). Surprisingly, when this reaction was carried out with a large excess of 31b,c, the phenol deprotected species 32b was isolated in 50% yield. Such a yield was still acceptable considering that two side reactions may occur: an intramolecular reaction in the phenol-deprotected version of **36b**, c leading to a δ -lactone and an intermolecular esterification reaction between phenol-porphyrin 32b and acyl chloride 31b,c (and subsequent

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SCHEME 7. (A) Clicking Model 4 onto an Electrode Surface and (B) Characterization of Immobilized Catalyst 4 by Cyclic Voltammetry^a



^{*a*} Reagents and conditions. (A): (a) (i) DMSO/H₂O, **5b**, CuSO₄-TBTA, sodium ascorbate, 30 min, (ii) rinse with DMSO and buffer. Mixed monolayer was formed from $N_3(CH_2)_{16}SH$ and $CH_3(CH_2)_{15}SH$. (B): Cyclic voltammogram of immobilized CcO model **5b** (surface coverage: 4.4×10^{-11} mol cm⁻²).^{14b} Scan rate: 300 mV/sec. Plain, **Fe-32a**; dashed, **Fe/Cu-32a** (model **5b**).

oligomerization reactions with other phenol-deprotected versions of **31b,c**) that may not be favored sterically. The small amount of the porphyrins **32c,d** that could be isolated by chromatography was easily and quantitatively deprotected by acidic treatment leading to **32b**.

Before metalation and immobilization of the catalysts, the free-base models were characterized and purified according to the standards of porphyrinoid and porphyrin chemistry previously established.^{3a,b,10a,b,13} The metalation of porphyrins **22**, **24b**, **25**, and **32a,b** with FeBr₂ was adapted from previous methods (Schemes 2, 3, and 5)^{3c} and was carried out in refluxing THF/methanol mixtures followed by washing with EDTA to remove the distal Fe atom coordinated to the imidazoles (Schemes 3A and 5). No side reactions were observed during metalation with free-base porphyrins, neither on the alkyne (models **1–5**) nor on the phenol (model **5**).⁹ⁱ Copper insertion

in **Fe-24b**, **Fe-32a**, and **Fe-32b** was achieved by reaction with 1 equiv of CuPF₆(CH₃CN)₄ leading to models **4**, **5a**, and **5b**, respectively (Schemes 3A and 5). NMR characterization of models **2–5** and porphyrin intermediates **Fe-24b**, **Fe-32a**, and **Fe-32b** was made possible by reacting the complexes with CO leading to diamagnetic low-spin Fe–CO adducts.^{3c}

Immobilization of models **1–5** by click chemistry with alkyl–azide-functionalized electrodes was conducted in a water–DMSO mixture in the presence of a Cu^I catalyst for 30 min (Scheme 7A). The surfaces were cleaned by two washes with DMSO. Immobilization of the models by covalent linkage to the alkyl chain through a robust triazole ring has been demonstrated by means of cyclic voltammetry and IR spectroscopy (Scheme 7B), as was previously shown with ferrocene.^{6a,e} Cyclic voltammetry of immobilized species **Fe-32a** and **Fe/Cu-32a** (model **5b**) clearly shows a broad reversible couple assigned to iron and copper centers in these models.¹⁴ Immobilization

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of **5b** monitored by grazing-angle infrared spectrometry shows no evidence of the azide stretch at 2100 cm⁻¹ (data not shown). The surface coverage was electrochemically determined to be approximately 4.4×10^{-11} mol/cm^{2.14}

Conclusions

A general method has been developed to prepare hemoprotein models bearing an alkyne functionality to allow their immobilization by click chemistry. The efficiency of this reaction, particularly for electrode surface functionalization, has been demonstrated with complex molecules such as cytochrome *c* oxidase models. Models **1**–**5** that bear an alkyne function on the proximal side should lead to a better understanding of the contribution of each redox-active component in the CcO active site during the 4e⁻ reduction of dioxygen under a slow regime of electron delivery. Electrocatalytic studies with models **1**–**5** will be reported elsewhere.¹⁴

Experimental Section

General Sonogashira Coupling Procedure. Imidazole (6 mmol. I-Mich, 7, 1.9 g; I-Ester, 11, 2.5 g; I-q, 10, 3.3 g), CuI (22 mol %), Pd(PPh₃)₄ (10 mol %), and 3.0 equiv of Et₃N (2.5 mL) were mixed in THF (100 mL). The resulting suspension was degassed (five pump-thaw cycles), and 7 equiv of trimethylsilylacetylene was added (6 mL). The mixture was stirred for 16 h. The progress of the reaction was monitored by TLC and was indicated by the formation of a large amount of white precipitate (triethylammonium iodide). The solvent was removed under reduced pressure and the residue was subjected to chromatography (SiO₂, eluent CH₂Cl₂/ CH₃OH 195:5 vol). The reaction conducted on iodo-porphyrins (70 μ mol scale) used a 1:0.5:1:3 ratio of zinc-protected iodoporphyrin (26a,26b)/Pd(PPh₃)₄/CuI/Et₃N and a 100 mg/20 mL concentration in porphyrin. Chromatography (SiO₂)/eluents CH₂-Cl₂ (27a), gradient elution of CH₂Cl₂/MeOH (100:2-100:6 vol) (27b). Yield: TMSAlk-Ester, 12, 1.7 g, 75%; TMSAlk-Mich, 13, 1.5 g, 88%; TMSAlk-q, 14, 1.94 g, 73%; porphyrin 27a, 0.097 g, 99%; porphyrin 27b, 0.096 g, 98%.

3-{**5-**[**4-**(**Trimethyl-silanylethynyl**)-**phenyl**]-**imidazol-1-ylmethyl**}-**benzoic Acid Methyl Ester (TMSAlk-Ester, 12):** ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.92 (d, 1H, J = 7.00 Hz), 7.67 (s, 1H), 7.58 (brs, 1H), 7.34 (t, 1H, J = 8.0 Hz), 7.30 (AA'BB', 4H, J = 8.5 Hz), 7.14 (brs, 1H), 7.09 (d, 1H, J = 6.0 Hz), 5.16 (s, 2H), 3.87 (s, 3H), 0.21 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 166.4, 139.0, 137.1, 132.5, 131.2, 131.1, 129.5, 129.4, 128.7, 128.1, 122.9, 104.3, 95.8, 52.6, 48.8, 0.2; MS (ESI⁺) m/z = 389.7 [M]⁺ (calcd for C₂₃H₂₄N₂O₂Si 388.5); TLC (silica, CH₂Cl₂/CH₂Cl₂ 95:5 vol); $R_f = 0.25$; Mp = 136–138 °C.

α₃-(*o*-4-(3-Methylimidazolyl)-amidophenyl)-β-(*o*-3-(1-(5-*p*-trimethylsilyl acetylenyl phenyl)imidazolylmethyl)benzamidophenyl) Zinc Porphyrin. ([Zn(α₃(*N*-MeIm)βTMSAlk-T)], 27b): ¹H NMR (500 MHz, CDCl₃) δ 9.13 (d, 1H, *J* = 8.0 Hz), 8.85 (m, 6H), 8.74 (d, 2H, *J* = 4.5 Hz), 8.57 (d, 2H, *J* = 8.5 Hz), 8.43 (d, 1H, *J* = 7.5 Hz), 8.07 (m, 5H), 7.81 (m, 7H), 7.62 (s, 2H), 7.52 (m, 3H), 7.42 (t, 1H, *J* = 7.5 Hz), 7.08 (t, 1H, *J* = 7.5 Hz), 6.94 (d, 2H, *J* = 8.5 Hz), 6.84 (s, 2H), 6.74 (s, 1H), 6.44 (d, 1H, *J* = 7.5 Hz), 5.86 (d, 2H, *J* = 8.5 Hz), 5.15 (s, 2H), 5.06 (s, 1H), 0.11 (s, 9H); MS (ESI⁻) *m*/*z* = 1416.5 [M]^{•-} (calcd for C₈₁H₆₄N₁₆O₄SiZn 1416.4); UV/vis (CH₂Cl₂) λ_{max} (10⁻³ ϵ , M⁻¹ cm⁻¹) 412 (48), 434 (466), 564 (21); TLC (SiO₂, NH₃-saturated CH₂Cl₂); *R_f* = 0.15.

General Phenol Protection. A solution of phenol-imidazole ester 33 (0.430 g, 1.8 mmol) in THF (60 mL) was cooled at 0 °C for 30

min, then ca. 3 equiv of NaH (0.133 g) was added portionwise. The resulting mixture was stirred at 0 °C for 2 h, then 2.2 equiv of 4-methoxybenzyliodide (ca 1.0 g) or trityl bromide was added at once. The resulting mixture was stirred for 24 h. Two-thirds of the solvent was removed under reduced pressure. Then the vessel was cooled again, and an excess of triethylamine and cool water (20 mL) was added. After extraction with CH₂Cl₂ (3 × 200 mL) and drying (MgSO₄), the solvents were distilled and the residue was subjected to chromatography [SiO₂, 3 × 15 cm, gradient elution, hexane/CH₂Cl₂ (1:2 vol), CH₂Cl₂ 100%, and CH₂Cl₂/CH₃OH (95:5 vol)]. **34b** (0.240 g, 38%) (yellow oil); **34c** (0.367 g, 43%).

3-(2-Trityloxy-phenyl)-*3H***-imidazole-4-carboxylic Acid Ethyl** Ester (Tr-O-Im-Ester, 34c): ¹H NMR (500 MHz, CDCl₃) δ (ppm) 7.87 (d, 1H, J = 1.0 Hz), 7.33 (d, 1H, J = 1.0 Hz), 7.22–7.28 (m, 15H), 7.18 (d, 1H, J = 7.5 Hz, J = 2 Hz), 7.00 (t, 1H, J = 7.0 Hz, J = 1.0 Hz), 6.92 (t, 1H, J = 7.5 Hz, J = 1.0 Hz), 6.63 (d, 1H, J = 8.5 Hz, J = 1.5 Hz), 4.28 (q, 2H, J = 7.0 Hz), 1.25 (t, 3H, J = 7.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ (ppm) 160.0, 151.4, 143.6, 142.3, 136.9, 129.1, 128.74, 128.65, 128.0, 127.64, 127.58, 125.6, 121.4, 120.8, 91.4, 60.7, 14.4; MS (ESI⁺) m/z = 474.9 [M]⁺, MS (ESI⁻) m/z = 473.1 [M – H]^{•–} (calcd for C₃₁H₂₆N₂O₃ 474.5); HPLC-MS (200 μ L/min, mobile phase H₂O/CH₃CN, gradient 2–95%, 35 min); $R_t = 29.1$ min; TLC (SiO₂, hexane/ethyl acetate 6:4 vol); R_f 0.5.

Ester Hydrolysis Procedures for Acid-Sensitive Imidazoles. A solution of imidazole ester (1.4 mmol;TMSAlk-Ester, 12, 0.543 g; PMBO-Ester, 34b, 0.492 g; TrO-Ester, 34c, 0.665 g) in THF/ CH₃OH (2:1 vol, 10 mL) was mixed with 1.5 N NaOH solution, and the resulting mixture was stirred at room temperature for 1 h. Then, only 1.0 equiv of acetic acid (with respect to NaOH, ca. 200 μ L) was slowly added to the mixture containing the acid-sensitive PMBO- or TrO-imidazole pickets to neutralize the mixture, followed by an excess of water (200 mL) causing the imidazole to precipitate. It was isolated by filtration and dried under a vacuum. Yield: Alk-Acid, 16, 0.360 g, 85%; PMBO-Acid, 35b, 0.431 g, 85%; TrO-Acid, 35c, 0.500 g, 80%.

3-[5-(4-Ethynyl-phenyl)imidazol-1-ylmethyl]-benzoic Acid (Alk-Acid, 16): ¹H NMR (500 MHz, CD₃OD) δ (ppm) 7.85 (d, 1H, J = 1.0 Hz), 7.82 (t, 1H, J = 7.5 Hz, J = 1.5 Hz), 7.69 (s, 1H), 7.38 (AA'BB', 4H), 7.26 (t, 1H, J = 7.5 Hz), 7.10 (d, 1H, J = 1.0 Hz), 6.98 (d, 1H, J = 8.0 Hz, J = 1.0 Hz), 5.33 (s, 2H), 3.53 (s, 1H); MS (ESI⁺) m/z = 303.7 [M + H]⁺; HR–MS (m/z) = 303.1136 [M+H]⁺; MS (ESI⁻) m/z = 301.1 [M – H]^{•-} (calcd for C₁₉H₁₄N₂O₂ 302.3267); HPLC-MS (Vydac, 200 mL/min, mobile phase H₂O/CH₃CN, gradient 20–95% in CH₃CN (20 min); R_t 14.6 min; TLC (silica, CH₃OH 100%); R_f 0.65; mp = 262–265 °C; FT–IR (KBr) 1665 cm⁻¹.

General Procedure for the Attachment of a Proximal Imidazole Tail. To a suspension of imidazole tail acyl chloride hydrochloride^{3c} (1.164 mmol; IT–Cl **6a**, 0.472 g; AlkT-Cl **6b**, 0.416 g) in dry MeCN (10 mL) (*sonicated* to brake the aggregates) was slowly added a solution of aminophenylporphyrin $\alpha_3 F\beta A$ **19**^{3b} (up to 1 equiv, 1.12 g) or TPMAPP **20**^{10a} (0.73 g) in MeCN (up to 10 mL). The resulting mixture that progressively turned green was stirred for 20 min. Dichloromethane was added (100 mL), and the mixture was washed with sodium bicarbonate and water (the color went back to purple). The solvent was evaporated, and the residue was subjected to chromatography: SiO₂ gel built with CH₂Cl₂, eluent (1) CH₂Cl₂, (2) gradient of CH₂Cl₂/CH₃OH (up to 97:3 vol). Average yield obtained for $\alpha_3 F\beta$ I-T, **21a** (1.4 g, 90%); $\alpha_3 F\beta$ Alk-T, **21b** (1.3 g, 89%); **TPI-T**, **22b** (0.946 g, 80%); **TPAlk-T**, **22a** (0.851 g, 80%).

α₃-(*o*-Trifluoroacetamidophenyl)-β-(*o*-3-(1-(5-*p*-acetylenylphenyl)imidazolyl methyl) benzamidophenyl)-porphyrin, α₃FβAlk-T (**21b**): ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.91 (d, 2H, J = 4.4Hz), 8.78 (m, 6H), 8.62 (m, 4H), 8.05 (d, 1H, J = Hz), 7.96 (d, 2H, J = 7.2 Hz), 7.90 (m, 5H), 7.61 (m, 4H), 7.50 (s, 2H), 7.44 (s, 2H), 6.92 (d, 2H, J = 8.5 Hz), 6.81 (s, 1H), 6.61 (s, 1H), 6.48 (d, 2H, J = 8.5 Hz), 6.46 (s, 1H), 6.30 (m, 2H), 6.10 (d, 1H, J = 6.5

⁽¹⁴⁾ Collman, J. P.; Devaraj, N. K.; Decréau, R. A.; Yang, Y.; Yan, Y.; Ebina, W.; Eberspacher, T. A.; Chidsey, C. E. D. *Science* **2007**, *315*, 1565.

Hz), 4.16 (s, 2H), 3.00 (s, 1H), -2.68 (s, 2H); ¹⁹F NMR (376 MHz, CDCl₃) δ (ppm) -77.22 (s, 6F), -77.74 (s, 3F); MS (ESI⁺) m/z = 1248.2 [M + H]⁺, (ESI⁻) m/z = 1246.2 [M - H]^{•-} (calcd for C₆₉H₄₃F₉N₁₀O₄ 1247.13); UV-vis. (CH₂Cl₂) λ (10⁻³ × ϵ / mol⁻¹ L cm⁻¹) 290 (15.2), 352 (10.3), 374 (13.0), 402 (sh, 37.0), 420 (141.7), 484 (65.7), 514 (8.0), 548 (2.3), 588 (2.6), 648 (0.8); TLC (silica, CH₂Cl₂/CH₃OH 96:4 vol); R_f 0.35; mp (CH₃CN) 200 °C (decomp.).

General Procedure for the Hydrolysis of Distal Amides. To a solution of $\alpha_3 F \beta X Tail$ porphyrins (0.34 mmol; $\alpha_3 F \beta I T$, 21a, 0.450 g; $\alpha_3 F \beta A lk T$, 21b, 0.423 g; $\alpha_3 F \beta T M S A lk T$, 28a, 0.449 g) in CH₂Cl₂ (24 mL) cooled at 0 °C and protected from light was slowly introduced a cold NH₃-saturated CH₃OH solution (13 mL). The resulting mixture was stirred for 2 to 3 days. After addition of CH₂Cl₂ (100 mL), washings with water (2 × 100 mL), and drying over MgSO₄, the solvents were then distilled. The residue was subjected to chromatography: SiO₂ gel, built with CH₂Cl₂, 20 × 5 cm; loading of porphyrin in solution in CH₂Cl₂; gradient elution of CH₂Cl₂/CH₃OH 200:0–200:2.0 vol. The target porphyrin moves at 200:2.0 vol. Yields in $\alpha_3 A \beta X T$ 23a, b vary according to the initial concentration in NH₃ and the reaction time (50–80%). Average yield found for $\alpha_3 A \beta I$ -T, 23a, 0.202 g, 56%; $\alpha_3 A \beta A lk T$, 23b, 0.192 g, 59%; $\alpha_3 A \beta T M S A lk T$, 29, 0.196 g, 56%.

α₃-(*o*-Aminophenyl)-β-(*o*-3-(1-(5-*p*-acetylenylphenyl)imidazolylmethyl)benzamido-phenyl)-porphyrin, α₃AβAlk-T (23b): ¹H NMR (500 MHz, CDCl₃) δ 8.89 (m, 9H), 8.08 (d, 1H, *J* = 7.5 Hz), 7.87 (t, 1H, *J* = 8.0 Hz), 7.74 (d, 3H, *J* = 7.5 Hz), 7.57 (m, 4H), 7.52 (s, 1H), 7.12 (m, 6H), 6.89 (d, 2H, *J* = 8.0 Hz), 6.84 (s, 1H), 6.79 (s, 1H), 6.48 (d, 2H, *J* = 8.0 Hz), 6.29 (t, 1H, *J* = 8.0 Hz), 6.21 (m, 2H), 6.16 (d, 1H, *J* = 8.0 Hz), 3.93 (s, 2H), 3.54 (s, 2H), 3.50 (s, 4H), 2.99 (s, 1H), -2.66 (s, 2H); MS (ESI⁺) *m/z* = 959.6 [M + H]⁺ (calcd for C₆₃H₄₆N₁₀O 959.4); UV/vis(CH₂Cl₂) λ_{max} (10⁻³ε, M⁻¹ cm⁻¹) 420 (241), 516 (16); HPLC-MS (Vydac, 200 mL/min, mobile phase H₂O/CH₃CN, gradient 20–95% in CH₃-CN (20 min)); *R_t* = 27 min; TLC (SiO₂, NH₃-saturated CH₂Cl₂); *R_f* = 0.67.

General Procedure for the Attachment of Distal Imidazole Pickets. To a suspension of imidazole acyl chloride (105 mg, 0.58 mmol) in acetonitrile (20-35 mL) was slowly added a solution of porphyrin (29, 100 mg, 0.097 mmol; 23a, 212 mg, 0.20 mmol) and diethylaniline (51 µL, 0.32 mmol; for 24a, 95 µL, 0.60 mmol) in CH₃CN/THF (1:1 vol; for the synthesis of 28b, 5 mL; for 24a, 10 mL). After the addition, the reaction mixture was stirred for 1-2 h. The solution was diluted with CH₂Cl₂ (~150-200 mL) and washed with 50% saturated aqueous NaHCO3 and water. The organic layer was dried over MgSO₄, filtered, and evaporated to dryness. The solid residue was subjected to chromatography (SiO₂, 20×3 cm). For **28b**: gradient elution of CH₂Cl₂/MeOH (100:2-100:6 vol). The target compound moves at 100:5 vol. Yield: 0.098 g, 75%. For 24a: gradient elution of CH₂Cl₂/MeOH (100:2-100: 10 vol). The target compound moves at 100:10 vol. Yield: 0.165 g, 60%.

α₃-(*o*-4-(3-Methylimidazolyl)-amidophenyl)-β-(*o*-3-(1-(5-*p*-iodophenyl)imidazolylmethyl)benzamidophenyl)-porphyrin (α₃-(*N*-MeIm)βI-T, 24a: ¹H NMR (500 MHz, CDCl₃) δ 8.85 (m, 9H), 8.53 (d, 2H, J = 8.0 Hz), 8.40 (d, 1H, J = 8.0 Hz), 8.07 (m, 3H), 7.99 (d, 1H, J = 8.0 Hz), 7.84 (m, 4H), 7.78 (s, 2H), 7.60 (t, 1H, J = 7.5 Hz), 7.55 (m, 4H), 7.41 (s, 1H), 6.99 (d, 2H, J = 8.5Hz), 6.92 (m, 4H), 6.82 (s, 1H), 6.68 (s, 1H), 6.31 (t, 1H, J = 8.0Hz), 6.20 (m, 3H), 6.15 (d, 1H, J = 8.5 Hz), 5.40 (s, 2H), 5.35 (s, 1H), 3.96 (s, 2H), 3.58 (s, 3H), 3.56 (s, 6H), -2.54 (s, 2H); MS (ESI⁺) m/z = 1385.6 (calcd for C₇₆H₅₇IN₁₆O₄ 1385.4); UV/vis (CH₂-Cl₂) λ_{max} (10⁻³ε, M⁻¹ cm⁻¹) 424 (274), 518 (9). HPLC-MS (200 µL/min, mobile phase H₂O/CH₃CN, gradient 2–95%, 35 min); $R_t = 21.9$ min; TLC (SiO₂, NH₃-saturated CH₂Cl₂); $R_f = 0.15$; mp (CH₃CN) 240-250 °C (decomp.).

General Procedure for the Metalation of Porphyrins with Iron. A mixture of free base porphyrin $(7.3-7.8 \,\mu$ mol, *ca.* 10 mg) and 3 equiv. FeBr₂ (6 mg) in MeOH/THF (1:4 vol., 8 mL) was heated for 1h at reflux. The volume was reduced to a 1/third then 0.1 M aqueous Na₂. EDTA (1 mL) was introduced and the resulting mixture was stirred for 10 min. Then the volume was expanded with benzene (30 mL), water was added (20 mL) and stirring was continued for 10 more min. The organic phase was then washed with water (4 × 10 mL), dried by addition of Na₂SO₄ with stirring for 10 min, then filtered through a glass wool plug. Solvents were evaporated under reduced pressure. Yields: 80%-quantitative.

cis-α₂-(*o*-4-(3-Methylimidazolyl)-amidophenyl)-α-(*o*-4-(3-(2-methoxyphenyl)imidazole)-5-benzamidophenyl)-β-(*o*-3-(1-(5-*p*-acetylenylphenyl)imidazolylmethyl)benzamidophenyl) Iron(II) Porphyrin, ([Fe(II)α₂(*N*-MeIm)α(*N*-MeOPhIm)βAlk-T)],(Fe-32a): ¹H NMR (500 MHz, CDCl₃/1 atm CO) δ 9.03 (d, 1H, J = 8.4 Hz), 8.68 (m, 9H), 8.43 (d, 1H, J = 8.2 Hz), 8.40 (d, 1H, J = 6.4 Hz), 8.21 (m, 3H), 7.94 (d, 1H, J = 7.3 Hz), 7.80 (m, 7H), 7.69 (m, 3H), 7.65 (s, 2H), 7.51 (m, 3H), 7.47 (t, 2H, J = 7.3 Hz), 7.06 (t, 2H, J = 7.70 Hz), 7.00 (d, 1H, J = 7.1 Hz), 6.91 (m, 5H), 6.48 (d, 1H, J = 7.5 Hz), 5.71 (d, 2H, J = 8.2 Hz), 3.96 (s, 1H), 3.72 (m, 2H), 3.62 (s, 3H), 3.58 (s, 3H), 3.38 (s, 3H), 2.93 (s, 1H), 2.34 (s, 1H), 1.93 (s, 1H), 1.41 (s, 1H); MS (ESI⁺) m/z = 1430.6 [M]⁺ (calcd for C₈₄H₆₀FeN₁₆O₅ 1430.4); UV/vis (CH₂Cl₂) λ_{max} (10⁻³ ϵ , M⁻¹ cm⁻¹) 426 (228), 534 (9).

General Procedure for Distal Metalation with Copper. To a solution of iron trisimidazolyl picket porphyrin (24b, 32a, 32b) in THF (1 mL) was added 1 equiv of Cu(MeCN)4PF6 (from a stock solution of 12 mg of Cu(I) salt in 120 μ L of MeCN). After 10 min (occasionally stirring the solution manually), the solvents were removed.

Copper(I) [cis- α_2 -(o-4-(3-Methylimidazolyl)-amidophenyl)- α -(o-4-(3-(2-methoxyphenyl))imidazolyl)-amidophenyl)- β -(o-3-(1-(5-p-acetylenylphenyl)imidazolylmethyl)benzamidophenyl) Iron **Porphyrin]** Hexafluorophosphate, { $Cu(I)[Fe(II)(\alpha_2(N-MeIm))$ α (*N*-(**PhOMe**)**Im**) β **Alk-T**]}⁺{**PF**₆}⁻, **Fe**/**Cu-32a**, **5b**: ¹H NMR (500 MHz, THF-d₈/MeCN-d₃/CDCl₃ (9:1:3 vol)/1 atm CO) δ 10.3 (brs, 1H), 9.05 (d, 1H, J = 8.4 Hz), 8.78 (d, 1H, 4.0 Hz), 8.72 (t, 2H, J = 5.0 Hz), 8.69–8.67 (m, 3H), 8.58 (m, 2H), 8.47 (d, 1H, J = 8.5 Hz), 8.38 (t, 2H, J = 8.5 Hz), 8.23 (d, 1H, J = 7.0 Hz), 8.14 (d, 1H, J = 7.5 Hz), 8.05 (s, 1H), 8.03 (d, 1H, J = 7.5 Hz), 7.83–7.66 (m, 8H), 7.55 (t, 1H, J = 7.5 Hz), 7.49–7.45 (m, 3H), 7.27 (t, 1H, J = 8.0 Hz), 7.06 (s, 1H), 7.01 (t, 1H, J = 7.5 Hz), 6.98-6.87 (m, 6H), 6.80 (t, 1H, J = 7.5 Hz), 6.57 (d, 1H, J = 7.5Hz), 5.86 (d, 2H, J = 8.0 Hz), 5.54 (s, 1H), 5.30 (s, 1H), 5.19 (s, 1H), 4.05 (s, 1H), 3.95 (s, 2H), 3.57 (s, 3H), 3.42 (s, 3H), 3.39 (s, 3H), 2.20 (s, 1H), 2.15 (s, 1H), 1.41 (s, 1H); ¹⁹F NMR (376 MHz, THF- d_8 /MeCN- d_3 /CDCl₃ (9:1:3 vol)/1 atm CO) δ -73.6 ppm (d, 6F, $J_{\rm FP} = 708$ Hz); HR-MS (ESI⁺) m/z = 1491.3596 [M - PF₆]⁺ (calcd for C₈₄H₆₀CuF₆FeN₁₆O₅ - PF₆ 1491.3678); UV/vis (CH₂-Cl₂) λ_{max} (10⁻³ ϵ , M⁻¹ cm⁻¹) 426 (228), 534 (9).

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Supporting Information Available: Experimental procedures and characterization data for compounds 1-35. This material is available free of charge via the Internet at http://pubs.acs.org.

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