Synthesis, Characterization, and Copper(II) Chelates of 1,11-Dithia-4,8-diazacyclotetradecane

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S Supporting Information

ABSTRACT: Synthesis of 1,11-dithia-4,8-diazacyclotetradecane (L1), a constitutional isomer of the macrocyclic [14]aneN₂S₂ series, is accompanied with reaction and method optimization. Chelation of L1 with copper(II) provided assessment of lattice packing, ring contortion, and evidence of conformational fluxionality in solution through two unique crystal structures: L1Cu(ClO4)₂ and [(L1Cu)₂ μ -Cl](ClO₄)₃. Multiple synthetic approaches are presented, supplemented



with reaction methodology and reagent screening to access [14]ane N_2S_2 L1. Reductive alkylation of bis-tosyl-cystamine was integrated into the synthetic route, eliminating the use and isolation of volatile thiols and streamlining the synthetic scale-up. Late-stage cleavage of protecting sulfonamides was addressed using reductive N-S cleavage to furnish macrocyclic freebase L1.

INTRODUCTION

Mixed N, S-containing macrocyclic ligands continue to be of significant interest due to their metal chelate coordination and redox properties with d-block elements.¹ The property of the [14]aneN₂X₂ series (X = S or O) to chelate both hard and soft metals has been reported throughout the literature² with recently reporting as biological CO₂- and proton-reducing agents,³ MRI contrast agents,⁴ positron emission tomography imaging,⁵ anticancer agents,⁶ nitric oxide sensors,⁷ and as a scaffold for cryptand formation.⁸ Many copper(II) chelates within the [14] ane N_2S_2 series have been previously investigated by both Rorabacher and Siegfried with binding constants and comparisons to other mixed [14]ane systems including S4 and N4.9 Practical pathways leading to the synthesis of [14] ane N_2S_2 $L2^{10}$ and $L3^{11}$ heteroconstitutional isomers (Figure 1) have been previously reported in moderate and reproducible yields.

Although the use and application of heteromacrocycle L1 have been mentioned in the literature,^{3,12} a descriptive synthetic strategy and characterization have been omitted including X-ray diffraction data for copper chelation. This



Figure 1. [14]aneN₂S₂ heteroconstitutional isomers.

prompted the design of a direct synthetic pathway to target macrocycle L1. Chelation of copper(II) in the presence of L1 provided crystallographic information to compare against L3. The copper(I/II) chelates of macrocycle L3 have been analyzed by Walker et al. providing binding constants with corresponding crystallographic structures assessing metal geometry and bond distances.¹³

The simple structural motifs of the macrocyclic isomers, L1-L3, can be synthetically deceptive due to the high variance of possible chemical manipulations. The tendency for macrocyclizations to produce complex reaction mixtures through kinetic oligomerization and polymerization provides an additional challenge for the synthetic optimization of these systems. Structural topology with respect to heteroatom placement was determined to dramatically influence a late-stage reductive N–S cleavage reaction and purification. Three synthetic strategies were implemented to afford [14]aneN₂S₂ L1, accompanied by reaction optimization and topological contrasts between constitutional macrocyclic isomer L3.

DISCUSSION

The strategic pathway toward L1 was initially assumed to be a simple four-step linear sequence, in which the macrocyclization of bis-acid chloride derived from bis-COOH 2 to the corresponding bis-lactam 1 would be the bottleneck reaction (Scheme 1). Reduction with borane followed by hydrolysis of the N–B bond would provide macrocycle L1 efficiently with minimal chemical manipulations (Method 1). A second method was designed around macrocyclic sulfonamide 3,

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Scheme 1. Retrosynthetic Proposals toward the Synthesis of L1



which allowed two different modes for macrocyclization. The first mode (Method 2a) was envisioned to take advantage of sulfur's nucleophilicity for governing the macrocyclization of bis-chloride 4. The second mode (Method 2b) of macrocyclization incorporating bis-sulfonamide 5 proved to be comparable in yield and was much more applicable to scale-up. Removal of sulfonamides (N–S cleavage) through known N–S cleavage reactions would provide pure L1. The use of different pathways to obtain heteromacrocycle L1 provides multiple ways to construct analogs and design larger macrocycles. Overall yields of [14]aneN₂S₂ L1 using methods 1, 2a, and 2b were 7, 48, and 52%, respectively.

The initial approach (Method 1) proceeded through a fourstep sequence, in which the final borane reduction of bislactam 1 afforded macrocycle L1. Synthesis of bis-lactam 1 was synthesized through a tandem N-acylation-macrocyclization from a bis-acid chloride derived from carboxylic acid 2 (Scheme 2). Lactam 1 has been prepared previously by Bradshaw et al. through a tandem bis-alkylation of 1,3propanedithiol and a symmetric α -chloroacetamide in a 31% yield; however, reduction to L1 was not reported.¹⁴

Reaction optimization for the synthesis of bis-lactam 1 from bis-acid chloride 6 was implemented using high dilution, as well as the slow simultaneous addition of the electrophile and nucleophile from separate syringe pumps to provide a maximum yield of 37% (average yield ~ 26%) (Table 1). This method of reactant addition during macrocyclizations has been implemented to thwart oligomerization and provide ample time for the intramolecular cyclization. The use of nonpolar and nonprotic solvents provided 1 in isolable

Table 1. Reaction Optimization of Bis-lactam 1

molarity (1,3-diaminopropane)	molarity 6	solvent	base	% yield
0.05	0.05	CH_2Cl_2	Cs ₂ CO ₃	10
0.05	0.05	CH_2Cl_2	K_2CO_3	18
0.01	0.01	CH_2Cl_2	NEt ₃	22
0.05	0.05	Et ₂ O	K ₂ CO ₃	12
0.05	0.05	Et ₂ O	Cs_2CO_3	7
0.01	0.01	Et ₂ O	K ₂ CO ₃	15
0.01	0.01	Et ₂ O	Cs ₂ CO ₃	12
0.01	0.01	PhMe	K_2CO_3	17
0.05	0.05	PhMe	NEt ₃	20
0.01	0.01	PhMe	NEt ₃	37

quantities. Modulating the temperature during the course of the reaction provided no appreciable yield of lactam 1; higher temperatures (>35 °C) provided oligomers, and at lower temperatures (<5 °C), bis-acid 2 was the major component, even after 72 h.

Reduction of amides to their corresponding amines with borane dimethyl sulfide (BDS) has been previously performed with high yields during the preparation of macrocycle L3.¹¹ When lactam 1 was subjected to a BDS reduction, the majority of the mass was composed of byproducts following a difficult purification of macrocycle L1. Additionally, the cleavage of the nitrogen-boron adducts with methanol/triethanolamine contributed to low yields and long reaction times invoking a change in reaction conditions. Switching to LAH provided no further increase in the yield or purity of L1. With the last two synthetic steps being tedious and low yielding (see Scheme 2), an alternative route was established.

The second method was performed in four sequential steps starting from bis-sulfonamide 7, providing a 48% overall yield of heteromacrocycle L1. Sulfonamide 7 was subjected to bishydroxyethylation through a KOH (20%)/ethylene carbonate melt affording the desired bis-hydroxyethyl sulfonamide 8a along with monoalkylated sulfonamide 8b and overalkylated ether 8c (Scheme 3). This method was optimized (Table 2) through stoichiometric and temperature screening to obtain bis-hydroxyethyl sulfonamide 8a in 89%. Crystal and structure refinement data are located in the Supporting Information for this manuscript.

Interestingly, the bis-hydroxyethylation resulted in a dramatic increase in the yield of hydroxyethyl sulfonamide **8a** between temperatures 100 and 115 °C (Table 2).

Scheme 2. Initial Synthetic Approach to the Synthesis of L1





Table 2. Hydroxyethylation Reaction Optimization (4 h, 10 wt % KOH) Performed with 1 mmol of Bis-sulfonamide 7

ethylene carbonate (Eq)	temperature (°C)	% yield 8a ^a	% yield 8b ^a	% yield 8c ^a
2	190	19	0	30
3	170	20	5	31
4	150	29	5	20
5	130	47	5	12
5	125	47	12	12
5	115	89	5	1
5	110	52	20	0
^{<i>a</i>} Isolated yields.				

Chromatographic separation of sulfonamides 8a-8c was relatively tedious, used large amounts of the mobile phase, and appeared to have high retention to the stationary silica when using traditional ethyl acetate:hexane solutions. Purification using smaller quantities of mobile phase was remedied through the use of toluene:acetone, providing

Scheme 4. Cystamine-Based Synthesis of Macrocycle L1

separation of 8a-8c. The addition of potassium hydroxide above the catalytic 10% did not increase reaction rate or yield. The use of thionyl chloride provided bis-chloroethyl sulfonamide 4 in high yield (Scheme 3). This compound was crystallographically characterized, and the solution is available in the Supporting Information section.

The formation of macrocyclic sulfonamide **3** proceeded through nucleophilic addition followed by intramolecular macrocyclization. This was carried out using the modified dilution/addition conditions outlined by Dietrich and Lehn.¹⁵ To increase reaction precision, reduce byproducts, and increase yields, dichloride **4** and propanedithiol were combined and added simultaneously to cesium carbonate in DMF at ~95 °C to provide macrocyclizations has shown to increase yields through a template effect,¹⁶ serendipitously switching to potassium carbonate furnished macrocycle **3** in similar yields with a clean reaction profile observed in the crude ¹H NMR.

The use of cesium as a counter cation for template-driven macrocyclizations has been reported in the literature and is



observed with oxygen-based heterocycles. When replacing the heteroatom to sulfur, this metal-template effect is diminished, presumably by the tendency for sulfide linkages to orient themselves in an exodentate fashion, directing lone pairs away from the center as shown with other metals.¹⁷ The entropic reordering of sulfides is not energetically favorable and can cause metal ions to bridge the macrocycle instead of chelation within the cavity. Buter and Kellogg demonstrated that a high-yield cyclization of 1, ω -dithiols with 1, and ω -dihalides ruled out any template effect.¹⁸ However, cesium carbonate did not surpass potassium carbonate in cyclization yields in DMF, suggesting that cesium is not critical in this case.

To eliminate the use of volatile thiols as reagents and provide a practical scale-up, Method 2b was pursued (Scheme 4). Starting from tosyl-cystamine (9), a triphenyl phosphine (TPP)-mediated reductive alkylation afforded acyclic bissulfide 5 in moderate yields. Separation of bis-sulfide 5 from triphenyl phosphine oxide (TPPO) byproduct proved difficult and often required two silica-based chromatographic separations. The use of ethanol and zinc chloride precipitated out ~90% of TPPO as $Zn(TPPO)_2Cl_2$ complex, allowing for effective purification and isolation (Table 3).¹⁹

Table 3. Reaction Conditions for the Formation of 5

conditions	yield of 5 (%)
(a) TPP, reflux (b) NaOH, 1,3-dibromopropane (c) ZnCl ₂ , EtOH	82
(a) TPP, reflux (b) K_2CO_3 , 1,3-dibromopropane (c) $ZnCl_2$, EtOH	80
(a) DTT, K ₂ CO ₃ , reflux (b) K ₂ CO ₃ , 1,3-dibromopropane	81
(a) DTT, NaOH, reflux (b) NaOH, 1,3-dibromopropane	72
(a) PBu ₃ , reflux (b) NaOH, 1,3-dibromopropane	88
(a) PBu ₃ , reflux (b) K ₂ CO ₃ , 1,3-dibromopropane	79
(a) TCEP, NaOH, reflux (b) NaOH, 1,3-dibromopropane	91
(a) TCEP, NaOH, reflux (b) NaOH, 1,3-ditosylpropane	68

The major drawback was determined during scale-up using TPP, where ~10% of the TPPO is still present in solution causing contamination during chromatographic separation. Attempts to subject the solution with ~10% TPPO to more zinc chloride did not initially show any precipitation; however, after 72 h at 0 °C, small amounts of zinc-TPPO complex precipitated out allowing for acyclic bis-sulfide **5** purification. In an attempt to eliminate TPPO contamination and use an aqueous workup for purification, tosyl-cystamine (**9**) was subjected to dithiothreitol (DTT) as the reductant.

Issues regarding the use of DTT were observed during the alkylation with 1,3-dibromopropane. Both sodium thiolate molecules derived from DTT and tosyl-cystamine 9 have the potential to perform substitution, lowering the yield of 5. Reducing the equivalents of DTT (1.3 equiv with respect to 9), disulfide cleavage provided an increase in the yield of bissulfide 5 but was difficult to chromatographically separate from byproducts. Mixed S-alkylations were attributed to thiol–disulfide exchange between thiolate derived from tosyl-cystamine and the oxidized cyclic disulfide originated from DTT (Figure 2).²⁰

Switching to tributyl phosphine (PBu₃) as the reductant provided 5 in an 88% yield with simple chromatographic separation (Table 3). Additionally, the disulfide cleavage with PBu₃ was completed within 25 min with no observable disulfide detected via TLC. The use of tris(carboxyethyl)-



Figure 2. Proposed thiol-disulfide exchange modeled after posttranslational modification of cysteine by Gilbert.²⁰

phosphine (TCEP) for reductive alkylation allowed for scaleup, simple aqueous workup, and elimination of column chromatography before macrocyclization. Acyclic, bis-sulfide 5 was subjected to macrocyclization conditions through substitution with the sulfonamide nitrogen atoms. Macrocycle 3 was obtained in a 51% yield through sulfonamide ring closure and then subjected to N–S cleavage conditions to obtain heteromacrocycle L1.

Reductive desulfonylation of sulfonamides to their corresponding free amines is reported throughout the literature, using a multitude of conditions ranging from sonication in methanol/magnesium to HBr/acetic acid.²¹ Many conditions are either too harsh resulting in decomposition/low yields or no reaction with the starting material remaining intact (Table 4). Interestingly, heteromacrocycles L1, L2, and L3 were

Table 4. Desulfonylation Conditions and Isolated Yields

conditions	yield of L1 (%)	yield of 10 (%) ^a	recovery of 3 (%)
Mg/methanol/ sonication	2 ^b	0	93
Na ₂ K/THF/23 °C	2^{b}	0	90
K ₂ Na/THF/23 °C	0	0	89
NaK ₂ stage I/THF/ 23 °C	0	0	91
HBr/HOAc/reflux	5-10	5-10	0
HBr/HOAc/23 °C	0	10	41
Na°/naphthalene/ —50 °C	89	0	0
Red-Al/xylenes/reflux	38 ^b	30	0
SmI ₂ /THF	0	0	95
^a Yield determined throu	gh ¹ H NMR.		

isolated in optimal yields through differing detosylation conditions establishing a reagent bias with respect to heteroatom placement along the $[14]aneN_2S_2$ backbone. Most cleavage conditions attempted on macrocycle 3 failed to produce macrocycle L1. Red-Al (vitride) was the first reagent that provided L1 in ~30% yield. A major limitation of this reduction was the presence of the mono-tosyl 10 (isolated



Figure 3. Detosylation reaction conditions of three heteroconstitutional isomers 3, 11, and 12.

as the HCl salt) that was difficult to remove through chromatographic separation. The use of sodium naphthalenide to perform the N–S bond cleavage afforded [14]aneN₂S₂ L1 cleanly over a 15 min period at -35 °C. Due to the freebase instability in the presence of oxygen and carbon dioxide, macrocyclic salt L1-HCl was initially made for storage purposes and maintained at -20 °C. The analysis of L1-HCl through ¹H and ¹³C NMR provided frequency-resolved spectra, unlike the macrocyclic L1 freebase that is composed of overlapping resonances.

For comparative desulfonylation conditions within the $[14]aneN_2S_2$ family, L2 and L3 were produced following the procedures outlined by Ochrymowycz et al.,¹⁰ Bridger et al.,²² and Walker et al.¹¹ (Figure 3). Two procedural modifications were performed for the synthesis of macrocycle L2 following Bridger's method and are reported in the Experimental Section. This included a bis-Michael addition mediated through sonication affording bis-nitrile 13 (Supporting Information) along with a Red-Al detosylation on substrate 12.

Switching desulfonylation conditions to refluxing Red-Al in xylenes afforded L2 in near-quantitative yields and required no extensive purification, with the exception of acid/base workup. The analysis of $[14]aneN_2S_2$ L2 through NMR spectroscopy proved troublesome as the resonances were overlapping, as seen with $[14]aneN_2S_2$ L1. The hydrogen chloride salt L2-HCl was used to provide frequency-resolved NMR spectra in deuterium oxide.

Heteroatom placement in the backbone of $[14]aneN_2S_2$ macrocycle systems imparts a significant role for the preference of reagents on the cleavage of the N–S system. Although many desulfonation conditions provided the freebase ligands (L1, L2, and L3), the isolated yields were negligible unless the appropriate conditions were applied. Topologic and heteroatom placements intrinsic to each sulfonamide with respect to the adjacent nitrogen and sulfur atoms could explain the variance in reagents with respect to isolated yield.

Due to the instability of L1 freebase, the diprotonated species of L1-HCl was immediately formed and stored at ~0 °C. Crystal structures of L1-HCl were obtained through the slow evaporation of methanol (Figure 4) with significant bond



Figure 4. Thermal ellipsoid diagram of L1-HCl salt with 35% probability displacement ellipsoids. C–H hydrogen atoms are omitted for clarity. The trans-L3-HCl, previously reported, was used for comparison (CCDC 888496).¹¹

lengths and bond angles shown in Table 5. Crystal and structure refinement data for L1-HCl are located in the Supporting Information. Compound L2-HCl failed to produce crystals, and the previously elucidated L3-HCl is shown for comparison. L3-HCl was reported to adopt a rectangular endodentate conformation with heteroatom placement at the four corners of the ring, resulting in the asymmetrical crown.¹¹ The bond length and bond angles of L1-HCl compared to those of L3-HCl are not statistically different; therefore, the structural conformations may be explained by the packing forces and crystal symmetry (see Supporting Information). Ligand L1-HCl was expected to adopt a preferred [3434] conformer (Dale notation)²³ similar to L3-HCl; however, the conformational assessment indicated a [3335] arrangement.

Crystallographic Data and Copper(II) Chelation. Crystallographic elucidation of L1 with copper(II) is lacking. With access to the freebase, reaction with $Cu(ClO4)_2 \cdot 6H_2O$ in CH_3CN at ambient conditions with L1 resulted in a deep purple color, which unlike the reported L3 did not precipitate out. Slow evaporation at 4 °C produced two distinct unit cell

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Table 5. Selected Bond Angles and Bond Lengths for Ligands L1-HCl and L3-HCl

bond lengths, Å	L1-HCl	L3-HCl	bond angles, deg	L1-HCl	L3-HCl
S-C	1.8106(17), 1.8152(17), 1.8194(19), 1.8174(17)	1.807(2), 1.820(3), 1.807(2), 1.817(2)	C-S-C	101.87(8), 101.79(8)	102.5(12), 102.1(12)
N-C	1.4973(19), 1.5024(18), 1.5077(19), 1.4988(19)	1.501(3), 1.497(3), 1.491(3), 1.501(3)	C-N-C	115.09(12), 116.26(11)	117.2(19), 116.3(19)
			C-C-S	111.18(11), 112.04(11), 116.92(12), 114.29(12)	113.3(17), 114.4(17), 112.3(16), 114.9(17)
			C-C-C	112.80(12), 113.30(15)	110.0(2), 109.4(2)
			N-C-C	112.21(13), 111.25(12), 114.73(12), 109.78(12)	111.3(19), 112.8(2), 110.9(19), 112.2(19)



[(L1Cu)₂µ-Cl](ClO₄)₃

L1Cu(ClO₄)₂

L3Cu(ClO₄)₂

Figure 5. Thermal ellipsoid diagrams of two separate conformations of L1 copper complexes, $[(L1Cu)_2\mu$ -Cl](ClO₄)₃, L1Cu(ClO4)₂, and previously reported L3Cu(ClO4)₂, (CCDC 905763)¹¹ at 35% probability displacement ellipsoids. All C–H hydrogen atoms are omitted for clarity, and $[(L1Cu)_2\mu$ -Cl](ClO₄)₃ is shaded for comparison in the conformational structure to L1Cu(ClO4)₂.

Table 6. Selected Bond Angles and Bond Lengths for Copper Complexes

	[(L1Cu) ₂ µ-Cl](ClO ₄) ₃	L1Cu(ClO4) ₂	L3Cu(ClO4) ₂
	Bond Lengths, Å		
S-C	1.822(3), 1.814(3), 1.824(3), 1.822(4), 1.822(3), 1.811(3), 1.818(3), 1.807(4)	1.816(17), 1.806(15)	1.816(2), 1.816(2)
N-C	1.468(4), 1.490(4), 1.488(4), 1.482(4), 1.486(4), 1.480(4), 1.486(4), 1.473(4)	1.44(2), 1.48(2)	1.492(2), 1.500(2)
N-Cu	2.025(3), 2.027(3), 2.028(3), 2.027(3)	2.008(15)	2.014(17)
S-Cu	2.3332(9), 2.3407(9), 2.3324(9), 2.3374(10)	2.315(4)	2.295(8)
	Bond Angles, deg		
C-S-C	102.94(16), 103.21(16), 102.22(16), 102.74(15)	102.6(8)	105.6(9)
C-N-C	110.5(3), 111.2(3), 111.3(3), 110.9(3)	112.6(15)	108.0(14)
C-C-S	112.4(2), 111.5(2), 106.1(2), 107.7(2), 112.4(2), 110.1(2), 106.4(2), 106.7(2)	109.1(12), 111.6(13)	105.6(13), 108.3(13)
C-C-C	116.0(3), 115.1(3), 115.3(3), 115.7(3)	116(2), 117.0(18)	115.1(16)
N-C-C	109.8(3), 111.4(3), 111.6(3), 109.3(3), 109.6(3), 112.6(3), 111.2(3), 109.9(3)	111.4(18), 112.2(15)	110.9(15), 114.9(16)
N-Cu-N	92.54(11), 91.77(11)	101.0(9)	180.0(22)
S-Cu-S	89.93(3), 90.71(3)	85.8(2)	179.9(11)
S-Cu-N	87.77(8), 168.85(8), 167.66(8), 87.40(8), 87.43(8), 168.35(8), 167.57(8), 87.58(8)	86.5(5), 171.6(5)	87.9(5), 92.0(5)

crystals suitable for X-ray diffraction (Figure 5) with crystal and refinement data, reported in the Supporting Information. All three systems exhibit planar coordination environments with respect to the metal center including comparable bond lengths and angles (Table 6).

Chelate $[(L1Cu)_2\mu$ -Cl](ClO₄)₃ has S-Cu-N bond angles averaging 168° due to the bridging three-center, four-electron

chloride atom compared to those of 172 and 180° for L1Cu(ClO4)₂ and L3Cu(ClO4)₂, respectively. More notably, L1Cu(ClO4)₂ differs from [(L1Cu)₂ μ -Cl](ClO₄)₃, in that the N–H bonds orientate opposite the sulfur lone-pair electrons. Both crystal packing and a bridging chloride atom induce changes in the ligand coordination, indicating that the binding

conformation of L1 is perhaps quite flexible and fluxional in solution.

Coordination of ligands L1 and L3 to copper(II) does, however, produce noteworthy structural changes. In both $[(L1Cu)_2\mu$ -Cl](ClO₄)₃ and L1Cu(ClO4)₂, the coordination of copper to both sulfur atoms exhibits a low strain chair conformation as compared to the torsion angles of cyclohexane²⁴ with the connecting five-membered chelate in a favorable equatorial position. However, in the case of L1Cu(ClO4)₂, the coordination of copper to nitrogen induces a conformation similar to a distorted envelope.²⁴

The copper(II)-thioether bonds are quite short (~2.3 Å) in both $[(L1Cu)_2\mu$ -Cl](ClO₄)₃ and L1Cu(ClO4)₂, suggesting that the bond orientation is overlapping with the magnetic orbital of copper(II).²⁵ Qualitative analysis depicted an observable difference with respect to the lone pairs on sulfur and the protons attached to the nitrogen atoms. As the lone pairs on sulfur are diffuse and flexible, L1Cu(ClO4)₂ seemed to perform a pseudo-chair flip with respect to ligand-metal chelate [(L1Cu)_2\mu-Cl](ClO₄)₃, which will be further investigated.²⁶

Ultraviolet-visible spectroscopy of L1Cu(ClO4)₂ provided λ_{max} of 275, 335, and 535 nm in aqueous solution, comparable to the previously reported data.^{12d} A slight blue shift was observed with L1Cu(ClO4)₂ compared to that of L3Cu-(ClO4)₂ (295, 354, and 545 nm) due to heteroatom positioning (Supporting Information). The band at 535 nm corresponds to the (N) \rightarrow Cu(II) ligand-to-metal charge transfer (LMCT), and the most intense absorption at 335 nm results from the (S) \rightarrow Cu(II) LMCT.⁹ Spectrochemical comparison with copper(II) chelates of [14]aneN₄ (cyclam), [14]aneS₄, and [14]aneN₂S₂ isomers (L1 and L3) provided weaker ligand field bands associated with the sulfur-based systems, which corroborates the Kauffman et al. analysis of sulfide donation and configuration.^{12d,27}

CONCLUSIONS

In summary, the synthetic methodology implemented for the synthesis of 1,11-dithia-4,8-diazacyclotetradecane (L1) has been completed and optimized. This is the first detailed synthesis of L1 reported to date, which allows for a multitude of analogs to be tailored for select applications. This study provides a practical approach to macrocycles containing aza/ thia heteroatoms and showcases the difference in reactivity associated with the [14]aneN₂S₂ heteroconstitutional isomers. A unique, efficient disulfide cleavage-alkylation mediated through TCEP was explored and optimized to attain disulfide 5. This study expands upon the pioneering work performed by Lehn and Cram for inter/intratandem macrocyclizations, which implements high-dilution conditions and slow addition of reagents. For detosylation of the resulting sulfonamides, it was determined that some reagents were more amenable to individual constitutional isomers. Currently, computational assessments are underway to assess the fluxionality and conformational bias between macrocyclic chelates L1Cu- $(ClO4)_2$ and $[(L1Cu)_2\mu$ -Cl](ClO₄)₃. The ligands also have the advantage of performing late-stage diversification, potentially leading to biologically compatible motifs used in imaging.

EXPERIMENTAL SECTION

All commercial reagents, unless otherwise stated, were used as received (Sigma-Aldrich, VWR, or Fischer Scientific Ltd.). Dichloromethane and acetonitrile were distilled from calcium hydride under nitrogen. Dimethylformamide was distilled from calcium hydride under vacuum and stored over 3 Å molecular sieves and degassed with argon using a gas dispersion frit. Reactions run at room temperature are in the range of 22-24 °C. ¹H and ¹³C NMR were obtained on a Varian 400 spectrometer as solutions in CDCl₃ with and without TMS and referenced to the CHCl₃ residual protioisomer. For spectra taken in other NMR solvents, D_2O_1 and DMSO- d_{61} the ¹H spectra were referenced to the solvent residual protioisomers, and ¹³C spectra were referenced to the NMR solvent. Chemical shifts are expressed in parts per million values, and coupling constants (1) are reported in Hertz (Hz) and rounded to the nearest 0.5 Hz. The following abbreviations are used to indicate apparent multiplicities: s, singlet; d, doublet; dd, doublet of doublets; ddd, doublet of doublets; t, triplets; q, quartets; m, multiplet. Flash column chromatography on silica gel (60 Å, 230–400 mesh, low acidity, obtained from EMD Millipore Corporation) was performed using reagent-grade solvents. Analytical thin-layer chromatography (TLC) was performed on precoated aluminum-backed silica gel plates (EMD Millipore Corporation) and visualized with a UV lamp (254 nm) or an iodine/silica or potassium molybdic acid solution in ethanol or $KMnO_{4(ag)}$ or *p*-anisaldehyde_(EtOH) or vanillin_(EtOH).

PHYSICAL METHODS

For temperature control in tandem with the reaction, mixing an Ika RCT digital stir plate with a PT 10000.60 temperature probe was used. FTIR spectra were collected on a Jasco 4600 spectrophotometer running the Spectra Manager CFR, where samples were prepared as a neat oil or solid on a ZnSe window using attenuated total reflectance (ATR). Melting points were obtained on a MeltTemp melting point apparatus and are uncorrected.

High-resolution mass spectra (HRMS) were recorded on a time-offlight JMS-T1000LC spectrometer with a DART ion source. ESI-MS spectra were acquired with a Bruker micrOTOF II, in the positive mode (parent ion plus H⁺). ESI samples were prepared for analysis by dissolving samples to 10^{-4} M in acetonitrile and were directly infused into the mass spectrometer at a flow rate of 4 μ L/min. The ESI was operated with a capillary offset of 4500 V and a skimmer potential of 48 V. The samples were calibrated using an internal standard.

X-ray intensity data were measured on a Bruker CCD-based diffractometer with a dual Cu/Mo ImuS microfocus optics (Cu K α radiation, $\lambda = 1.54178$ Å, Mo K α radiation, $\lambda = 0.71073$ Å). Crystals were mounted on a cryoloop using Paratone oil and placed under a steam of nitrogen at 100 K (Oxford Cryosystems). The detector was placed at a distance of 5.00 cm from the crystal. The data were corrected for absorption with the SADABS program. The structures were refined using the Bruker SHELXTL software package (version 6.1) and were solved using direct methods until the final anisotropic full-matrix, least-squares refinement of F2 converged.

UV and visible spectra in solution were run in 1 cm cells using a Perkin Elmer Lambda 365 UV—vis spectrometer at concentrations appropriate for measurements of absorption bands, and the data collection was obtained at ambient temperatures.

The Supporting Information (CCDC 1912553-1912560) contains the supplementary crystallographic data for this article. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

1,11-Dithia-4,8-diazacyclotetradecane-3,9-dione (1). A 250 mL flame-dried three-neck round-bottom flask equipped with a magnetic stir bar, a nitrogen inlet, and three septa was charged with dry toluene (100 mL). A syringe was charged with 1,3-diaminopropane (0.14 mL, 1.60 mmol) and triethylamine (0.47 mL, 3.36 mmol) in toluene (40 mL), and a second syringe was charged with 2,2'-(propane-1,3-diylbis(sulfanediyl))diacetyl chloride (6) (0.428 g, 1.60 mmol) in toluene (40 mL); these two solutions were attached to individual syringe pumps and added to the vigorously stirring toluene at a rate of ~1 drop per 10 s simultaneously. After the addition of the reagents, the solution was allowed to stir for a 12 h period. This solution was then concentrated in vacuo and immediately subjected to column chromatography

(EtOAc) to provide bis-lactam 1 in a 37% yield (155 mg, 0.592 mmol) as an amorphous white residue. $R_{\rm f}$ = 0.3 (EtOAc); ¹H NMR (400 MHz, chloroform-*d*) δ 7.07 (s, 1H), 3.48 (q, J = 5.8, 5.2 Hz, 2H), 3.25 (s, 2H), 2.71 (t, J = 7.0 Hz, 2H), 2.01–1.74 (m, 1H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 169.6, 39.7, 36.9, 32.3, 29.2, 28.9; IR $\nu_{\rm max}$ 3315, 3300, 2923, 2857, 1730, 1638, 1529, 1451, 1054 cm⁻¹; HRMS (EI⁺) m/z [M + Na]⁺ calcd for C₁₀H₁₈O₂N₂NaS₂ 285.0702, found 285.0707.

2,2'-(Propane-1,3-diylbis(sulfanediyl))diacetic acid (2). A 50 mL flame-dried round-bottom flask equipped with a magnetic stir bar and a condenser was charged with 1,3-propanedithiol (1.00 mL, 9.96 mmol), ethanol (4.00 M, 2.49 mL), and sodium borohydride (0.0190 g, 0.498 mmol) in the indicated order. An aqueous potassium hydroxide solution (2.50 mL, 16 M, 39.8 mmol) was added slowly to the mixture (15 min). The addition of chloroacetic acid (1.88 g, 19.9 mmol) in ethanol (5.00 mL) was added dropwise over a period of 15 min, after which the solution was allowed to reflux for 12 h using a heating mantle. The reaction mixture was then concentrated in vacuo and dissolved in water (20 mL), and the impurities were washed out with diethyl ether $(2 \times 3 \text{ mL})$. The aqueous layer was acidified to pH ~1 with concentrated sulfuric acid. The product was extracted with diethyl ether $(3 \times 20 \text{ mL})$. The combined ethereal layers were dried with Na₂SO₄, filtered, and concentrated in vacuo. The residual oil was taken up in toluene (30 mL) and allowed to crystallize over a 12 h period at 0 °C. The bis-carboxylic acid was filtered off affording a white solid (1.653 g, 7.37 mmol) in a 74% yield. Mp = $69-70 \,^{\circ}\text{C}$; ¹H NMR (300 MHz, deuterium oxide) δ 3.45 (s, 3H), 2.80 (t, *J* = 7.2 Hz, 4H), 1.98 (p, J = 7.2 Hz, 2H); ¹³C{¹H} NMR (75 MHz, deuterium oxide) δ 174.8, 33.3, 30.7, 27.6; IR $\nu_{\rm max}$ 2922, 2672, 1707, 1419, 1288, 1205, 1141 cm⁻¹. HRMS (EI⁺) m/z [M + Na]⁺ calcd for C₇H₁₂NaO₄S₂ 247.0069, found 247.0062.

4,8-Ditosyl-1,11-dithia-4,8-diazacyclotetradecane (3). DMF was degassed with nitrogen for ~20 min via gas dispersion frit prior to running the reaction, mitigating the possibility of disulfide formation.

Method A. A 100 mL three-neck round-bottom flask equipped with a septum, a nitrogen inlet, a condenser, and a magnetic stir bar was charged with anhydrous potassium carbonate (0.286 g, 2.07 mmol) and DMF (20 mL). A separate 25 mL pear flask was charged with DMF (10 mL), 1,3-propanedithiol (0.10 mL, 1.03 mmol), and bischloroethyl sulfonamide 6 (0.50 g, 0.985 mmol). This flask was fitted with an inlet line inserted through a septum from a fluid metering pump, and the terminal line was fitted into a septum attached to the three-neck round-bottom flask. The flask containing potassium carbonate and DMF was heated to 90-95 °C via a heating mantle, and the fluid metering pump was set to add the contents of the pear flask into the three-neck round-bottom flask at a rate of 1 drop/8 s. Once all of the contents from the pear flask were added, the reaction was stirred at 90-95 °C for 12 h. After which the reaction was cooled down to room temperature, the DMF was removed in vacuo, and the residue was subjected to column chromatography (19:1; PhMe/ acetone) $R_{f} = 0.4$ (1:1 Hex/EtOAc) affording 3 (0.380 g, 0.670 mmol) in a 68% yield.

Method B. A 1 L two-neck round-bottom flask equipped with a septum, a nitrogen inlet, a condenser, and a magnetic stir bar was charged with anhydrous potassium carbonate (3.22 g, 23.31 mmol) and DMF (155 mL). A separate 250 mL two-neck pear flask was charged with DMF (78 mL), 1,3-dibromopropane (0.79 mL, 7.77 mmol), and N,N'-((propane-1,3-diylbis(sulfanediyl))bis(ethane-2,1diyl))bis(4-methylbenzenesulfonamide) (5) (3.91 g, 7.77 mmol). This flask was fitted with an inlet line inserted through a septum from a fluid metering pump, and the terminal line was fitted into a septum attached to the 1 L two-neck round-bottom flask. The potassium carbonate and DMF were heated via a heating mantle to 100 °C, and the fluid metering pump was set to add the contents of the pear flask into the two-neck round-bottom flask at a rate of 1 drop/8 s. Once all of the contents from the pear flask were added, the reaction was stirred at 100 °C for 24 h. After which the reaction was cooled down to room temperature, the DMF was removed in vacuo, and the residue was subjected to column chromatography (19:1; PhMe/ acetone) affording 3 (2.14 g, 3.96 mmol) in a 51% yield. Mp = 144145 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.67 (d, *J* = 8.5 Hz, 4H), 7.32 (d, *J* = 8.5 Hz, 4H), 3.22 (q, *J* = 6.9 Hz, 8H), 2.81 (t, *J* = 7.4 Hz, 4H), 2.63 (t, *J* = 6.7 Hz, 4H), 2.43 (s, 6H), 2.05 (p, *J* = 7.4 Hz, 2H), 1.91 (p, *J* = 6.6 Hz, 2H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 143.6, 135.3, 129.9, 127.2, 49.8, 48.5, 31.9, 29.7, 29.7, 21.5. IR ν_{max} 2924, 2857, 1337, 1157, 1089, 815, 721, 653 cm⁻¹. HRMS (APCI) *m*/*z* [M + H]⁺ calcd for C₂₄H₃₅O₄N₂S₄ 543.1474, found 543.1477.

N,N'-(Propane-1,3-diyl)bis(N-(2-chloroethyl)-4-methylbenzenesulfonamide) (4)^a. A flame-dried 25 mL round-bottom flask equipped with a magnetic stir bar, a condenser, and a calcium sulfate drying tube was charged with N,N'-(propane-1,3-diyl)bis(N-(2hydroxyethyl)-4-methylbenzenesulfonamide) (8) (1.49 g, 3.20 mmol) and thionyl chloride (5 mL). The reaction was refluxed on a heating mantle for 12 h, at which time the reaction was cooled down to room temperature and toluene (10 mL) was added and removed in vacuo. The toluene (10 mL) washes were performed two additional times to azeotropically remove the excess thionyl chloride. The yellow oil was dissolved in minimal MTBE, and methanol or ethanol was added until a thick white precipitate was observed. This was heated to reflux and allowed to cool at -20 °C for 12 h to precipitate out the final bis-alkyl chloride 4 (0.428 g, 2.94 mmol) in a 92% yield as a white solid. Mp = 83-84 °C. $\tilde{R_f}$ = 0.88 (EtOAc); ¹H NMR (400 MHz, chloroform-d) δ 7.66 (d, J = 8.1 Hz, 4H), 7.31 (d, J = 8.4 Hz, 4H), 3.62 (t, J = 7.0 Hz, 4H), 3.35 (t, J = 7.0 Hz, 4H), 3.17 (t, J = 7.3 Hz, 4H), 2.41 (s, 6H), 1.88 (p, J = 7.3 Hz, 2H); ${}^{13}C{}^{1}H{}$ NMR (101 MHz, chloroform-d) δ 143.9, 135.6, 129.9, 127.2, 50.6, 47.7, 42.2, 28.7, 21.5. IR $\nu_{\rm max}$ 2975, 2933, 2877, 1732, 1597, 1455, 1402, 1337, 1249, 1208, 1154, 1117, 1089, 913, 727 cm⁻¹; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{21}H_{28}O_4N_2S_2NaCl_2$ 529.0760, found 529.0759.

N,N'-((Propane-1,3-divlbis(sulfanedivl))bis(ethane-2,1diyl))bis(4-methylbenzenesulfonamide) (5). Method A. A 250 mL round-bottom flask equipped with a magnetic stir bar, a nitrogen inlet, and a condenser was charged with THF/H2O (5:1, 75 mL) and degassed with nitrogen via gas dispersion frit for 20 min. The addition of tosyl-cystamine (9) (7.60 g, 16.50 mmol) and triphenyl phosphine (5.20 g, 19.83 mmol) followed, and the reaction was refluxed for 12 h with a heating mantle. The solution was then cooled down to room temperature, and degassed 2 M NaOH (17.4 mL, 34.80 mmol) was added in one portion. The light pink solution was stirred at room temperature for 5 min, 1,3-dibromopropane (0.76 mL, 7.43 mmol) was added, and the solution was refluxed via a heating mantle for 12 h. The reaction mixture was diluted with EtOAc (50 mL) and 1 M HCl (40 mL), the aqueous layer was extracted with EtOAc $(2 \times 30 \text{ mL})$, and the organic layers were combined, dried with sodium sulfate, filtered, and concentrated in vacuo. The yellow residue was dissolved in ethanol (200 mL) and warmed to 40 °C followed by the addition of zinc chloride (11.24 g, 82.5 mmol) in ethanol (100 mL). The ethanolic solution was stirred at 40 °C for 0.5 h, then cooled down to room temperature, stirred for 3 h, and the cloudy solution was placed in a 0 °C fridge for 12 h. The white Zn(TPPO)₂Cl₂ was filtered, washed with cold ethanol (2 \times 50 mL), the mother liquor concentrated, and subjected to column chromatography (3:1 \rightarrow 2:1 \rightarrow 1:1; Hex/EtOAc) affording 5 (3.44 g, 13.53 mmol) in an 82% yield as a clear oil.

Method B. A 250 mL round-bottom flask equipped with a magnetic stir bar, a nitrogen inlet, and a condenser was charged with THF/H₂O (5:1, 75 mL) and degassed with nitrogen via gas dispersion frit for 20 min. The addition of tosyl-cystamine (7.60 g, 16.50 mmol), potassium carbonate (6.84 g, 49.50 mmol), and dithiothreitol (DTT) (3.31 g, 21.45 mmol) followed, and the reaction was refluxed with a heating mantle until the disappearance of a starting material via TLC (~1.5 h). The reaction was allowed to cool down to room temperature followed by the dropwise addition of 1,3-dibromopropane (1.85 mL, 18.15 mmol) over 15 min. The solution was refluxed with a heating mantle a final time for 12 h, cooled down to room temperature, and diluted with EtOAc (200 mL). The organic layer was washed with 1 M HCl (2 × 20 mL), water (20 mL), and brine (20 mL), dried with sodium sulfate, filtered, and concentrated in vacuo. The residue was subjected to column chromatography (3:1 \rightarrow

 $2:1 \rightarrow 1:1$; Hex/EtOAc) to obtain 5 (6.72 g, 13.37 mmol) as a clear viscous oil in an 81% yield.

Method C. A 250 mL round-bottom flask equipped with a magnetic stir bar, a nitrogen inlet, and a condenser was charged with THF/H₂O (5:1, 75 mL) and degassed with nitrogen via gas dispersion frit for 20 min. The addition of tosyl-cystamine (7.60 g, 16.50 mmol) and tributyl phosphine (4.89 mL, 19.83 mmol) was followed, and the reaction was refluxed for 2 h via a heating mantle. The solution was then cooled down to room temperature, and degassed 2 M NaOH (17.4 mL, 34.80 mmol) was added in one portion. The solution was stirred at room temperature for 5 min, 1,3dibromopropane (0.76 mL, 7.43 mmol) was added, and the solution was refluxed for 12 h via a heating mantle. The reaction mixture was diluted with EtOAc (150 mL), washed with 1 M HCl (2×20 mL), water (20 mL), and brine (15 mL), dried with sodium sulfate, filtered, concentrated in vacuo, and subjected to column chromatography (3:1 \rightarrow 1:1, Hex/EtOAc). Compound 5 (7.30 g, 14.52 mmol) was obtained in an 88% yield as an opaque oil.

Method D. A 500 mL two-neck round-bottom flask equipped with a magnetic stir bar, a pressure-equalizing drop funnel, a condenser, and a nitrogen inlet was charged with THF/H₂O (5:1, 217 mL) and degassed with nitrogen via gas dispersion frit for 20 min. At which time, NaOH (0.95 g, 23.87 mmol) followed by TCEP·HCl (6.84 g, 23.87 mmol) was added, and the mixture was degassed with nitrogen via gas dispersion frit for 10 min. Tosyl-cystamine (10.0 g, 21.7 mmol) was added in one portion, and the solution was refluxed with a heating mantle until the disappearance of a starting material via TLC $(\sim 15 \text{ min})$. The solution was cooled down to room temperature, at which time, nitrogen-degassed 2 M NaOH (54 mL, 27.00 mmol) was added in one portion, and the pressure-equalizing drop funnel was charged with 1,3-dibromopropane in nitrogen-degassed THF (10 mL). The solution was stirred at room temperature for 5 min followed by the dropwise addition of 1,3-dibromopropane over 15 min and refluxed with a heating mantle for 12 h. The reaction mixture was diluted with EtOAc (300 mL), washed with 1 M HCl (2×30 mL), water (30 mL), and brine (20 mL), dried with sodium sulfate, filtered, concentrated in vacuo, and subjected to column chromatography (3:1 \rightarrow 1:1, Hex/EtOAc) to afford compound 5 (9.82 g, 19.53 mmol) in a 90% yield as an opaque oil. $R_f = 0.5$ (Hex/EtOAc, 1:1). ¹H NMR (400 MHz, chloroform-d) δ 7.70 (d, J = 8.3 Hz, 4H), 7.25 (d, J = 8.2Hz, 4H), 5.54–5.34 (m, 2H), 3.04 (t, J = 7.0 Hz, 4H), 2.52 (t, J = 6.8 Hz, 4H), 2.41 (t, J = 7.1 Hz, 8H), 2.36 (s, 6H), 1.65 (p, J = 7.1 Hz, 2H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 143.6, 136.8, 129.8, 127.0, 42.3, 31.6, 30.2, 28.9, 21.5. IR $\nu_{\rm max}$ 3277, 2921, 2256, 1597, 1494, 1409, 1321, 1185, 1092, 1019, 813 cm⁻¹. HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{21}H_{30}O_4N_2NaS_4$ 525.0981, found 525.0985.

2,2'-(Propane-1,3-diylbis(sulfanediyl))diacetyl Chloride (6). A 50 mL flame-dried round-bottom flask equipped with a magnetic stir bar, a condenser, and a calcium sulfate drying tube was charged with 2,2'-(propane-1,3-diylbis(sulfanediyl))diacetic acid (2) (0.400 g, 1.78 mmol), methylene chloride (17.8 mL), and thionyl chloride (0.320 mL, 1.78 mmol) in the indicated order. This mixture was refluxed with an external heating mantle for 12 h and then concentrated in vacuo. To the yellow oil, trituration from toluene (3 × 10 mL) afforded the final bis-acid chloride 6 (0.428 g, 1.64 mmol) in a 92% yield as a viscous light-yellow oil. ¹H NMR (300 MHz, chloroform-*d*) δ 3.71 (s, 1H), 2.78 (t, *J* = 7.1 Hz, 1H), 1.93 (p, *J* = 7.1 Hz, 1H); ¹³C{¹H} NMR (75 MHz, chloroform-*d*) δ 170.2, 45.2, 31.2, 27.8.

N,N'-(**Propane-1,3-diyl**)**bis**(4-methylbenzenesulfonamide) (7). A 500 mL round-bottom flask equipped with a magnetic stir bar and a pressure-equalizing drop funnel was charged with water (80 mL) and potassium carbonate (13.82 g, 100.0 mmol) followed by 1,3-propanediamine (4.2 mL, 50.0 mmol). The pressure-equalizing drop funnel was charged with THF (85 mL) and *para*-toluenesulfonyl chloride (19.06 g,100.0 mmol) and was added over a 4.5 h period to the carbonate solution. Once added, the solution was stirred for 10 h at 23 °C and poured over water (40 mL) at 0 °C. The white solid was filtered, washed with ethanol (50.0 mL, 0 °C), and dried in vacuo to afford bis-sulfonamide 7 (18.17 g, 47.50 mmol) in a 95% yield. Mp =

145–148 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.73 (d, *J* = 8.3 Hz, 4H), 7.31 (d, *J* = 8.3 Hz, 4H), 4.83 (s, 2H), 3.02 (t, *J* = 6.1 Hz, 6H), 2.43 (s, 4H), 1.67 (p, *J* = 6.2 Hz, 2H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 143.6, 136.8, 129.8, 127.0, 39.7, 29.9, 21.5. HRMS (ESI) *m*/*z* [M + H]⁺ calcd for C₁₇H₂₃O₄N₂S₂ 383.1094, found 383.1097.

N,N'-(Propane-1,3-diyl)bis(N-(2-hydroxyethyl)-4-methylbenzenesulfonamide) (8a). Method A. A 25 mL round-bottom flask equipped with a condenser, a nitrogen inlet, and a magnetic stir bar was charged with N,N'-(propane-1,3-diyl)bis(4-methylbenzenesulfonamide) (7) (0.82 g, 2.14 mmol), potassium carbonate (0.65 g, 4.71 mmol), ethylene carbonate (0.75 g, 8.56 mmol), and DMF (0.5 M, 4.3 mL). This solution was maintained at 110–120 °C for 4 h in an oil bath controlled with a thermocouple and then cooled to 23 °C. The DMF was removed, and the yellow residue was taken up in ethyl acetate (50 mL) and washed with 0.5 M HCl (10 mL), water (10 mL), and brine (10 mL). The organic layer was dried with sodium sulfate, filtered, and concentrated in vacuo to afford a thick yellow oil. The residue was subjected to column chromatography (2:1, ethyl acetate/hexane) to afford 8a (0.473 g, 1.01 mmol) as a clear viscous oil in a 47% yield.

Method B. A 50 mL round-bottom flask was charged with N,N'-(propane-1,3-diyl)bis(4-methylbenzenesulfonamide) (7) (5.00 g, 13.1 mmol), ethylene carbonate (2.88 g, 32.8 mmol), and potassium hydroxide (0.050 g, 0.891 mmol). The mixture was stirred for 4 h at 115 °C in an oil bath controlled with a thermocouple, at which time, the residue was cooled to 0 °C via an external ice bath and diluted with MilliQ water (10 mL). The solution was extracted with ethyl acetate (20 mL \times 3), and the combined organic layers were washed with 1 M HCl (10 mL), water (10 mL), and brine (5 mL), dried with sodium sulfate, filtered, and concentrated in vacuo. The remaining residue was purified via flash chromatography (3:2 EtOAc/Hex) providing 8a (5.47 g, 11.66 mmol) in an 89% yield as a clear oil. $R_f =$ 0.40 (EtOAc); ¹H NMR (400 MHz, chloroform-d) δ 7.63 (d, J = 8.4 Hz, 4H), 7.25 (d, J = 8.1 Hz, 4H), 3.74–3.64 (m, 4H), 3.42–3.33 (m, 4H), 3.20-3.09 (m, 8H), 2.35 (s, 6H), 1.90 (p, J = 7.4 Hz, 2H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 143.6, 135.5, 129.8, 127.2, 61.4, 51.7, 47.9, 28.6, 21.5. IR $\nu_{\rm max}$ 3524, 2927, 1597, 1455, 1331, 1155, 1088, 815, 728, 656 cm⁻¹; HRMS (ESI) m/z [M + H] calcd for C₂₁H₃₁O₆N₂S₂ 471.1618, found 471.1619.

N-(2-Hydroxyethyl)-4-methyl-N-(3-((4-methylphenyl)sulfonamido)propyl)benzenesulfonamide (8b). A 50 mL roundbottom flask was charged with N,N'-(propane-1,3-diyl)bis(4-methylbenzenesulfonamide) (7) (5.00 g, 13.1 mmol), ethylene carbonate (2.88 g, 32.8 mmol), and potassium hydroxide (0.050 g, 0.891 mmol). The mixture was stirred for 2 h at 110 °C in an oil bath controlled with a thermocouple, at which time, the residue was cooled to 0 $^\circ C$ via an external ice bath and diluted with MilliQ water (10 mL). The solution was extracted with ethyl acetate (20 mL \times 3), and the combined organic layers were washed with 1 M HCl (10 mL), water (10 mL), and brine (5 mL), dried with sodium sulfate, filtered, and concentrated in vacuo. The remaining residue was purified via flash chromatography (3:2 EtOAc/Hex) providing 8b (2.18 g, 5.11 mmol) in a 39% yield as a clear oil. $R_f = 0.50$ (EtOAc); ¹H NMR (400 MHz, chloroform-d) δ 7.71 (d, J = 8.3 Hz, 2H), 7.63 (d, J = 8.3 Hz, 2H), 7.31-7.21 (m, 4H), 5.64 (t, J = 6.4 Hz, 1H), 3.68 (q, J = 5.3 Hz, 2H),3.28-3.10 (m, 4H), 2.99 (q, J = 6.3 Hz, 2H), 2.83 (s, 1H), 2.39 (s, 3H), 2.38 (s, 3H), 1.77 (p, J = 6.5 Hz, 2H); ${}^{13}C{}^{1}H{}$ NMR (101 MHz, chloroform-d) δ 143.8, 143.3, 136.9, 135.3, 129.9, 129.7, 127.2, 127.0, 61.5, 51.8, 47.5, 39.9, 28.9, 21.5. IR $\nu_{\rm max}$ 3475, 3217, 2932, 2873, 1597, 1493, 1451, 1326, 1151, 1091, 741 cm⁻¹. HRMS (ESI) $m/z [M + Na]^+$ calcd for $C_{19}H_{26}O_5N_2NaS_2$ 449.1175, found 449.1176.

N-(2-(2-Hydroxyethoxy)ethyl)-*N*-(3-((*N*-(2-hydroxyethyl)-4-methylphenyl)sulfonamido)propyl)-4-methylbenzenesulfonamide (8c). A 50 mL round-bottom flask was charged with $N_{,N'}$ -(propane-1,3-diyl)bis(4-methylbenzenesulfonamide) (7) (5.00 g, 13.1 mmol), ethylene carbonate (2.88 g, 32.8 mmol), and potassium hydroxide (0.050 g, 0.891 mmol). The mixture was stirred for 4 h at 190 °C in an oil bath controlled with a thermocouple, at which time,

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the brownish residue was cooled to 0 °C via an external ice bath and diluted with MilliQ water (10 mL). The solution was extracted with ethyl acetate (20 mL \times 3), and the combined organic layers were washed with 1 M HCl (10 mL), water (10 mL), and brine (5 mL), dried with sodium sulfate, filtered, and concentrated in vacuo. The remaining residue was purified via flash chromatography (3:2, EtOAc/Hex) providing N-(2-(2-hydroxyethoxy)ethyl)-N-(3-((N-(2hydroxyethyl)-4-methylphenyl)sulfonamido)propyl)-4-methylbenzenesulfonamide 8c (3.57 g, 6.94 mmol) in a 53% yield as a clear oil. $R_{\rm f}$ = 0.35 (EtOAc); ¹H NMR (400 MHz, chloroform-d) δ 7.60 (dd, I = 8.3, 2.5 Hz, 4H), 7.38-7.05 (m, 4H), 3.72-3.63 (m, 2H), 3.63-3.58 (m, 2H), 3.52 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.24 (t, J = 5.4 Hz, 2H), 3.45-3.38 (m, 2H), 3.45-3.38 (m, 2H), 3.45-3.38 (m, 2H), 3.45-3.45 (m, 2H), 3.45 (m,Hz, 2H), 3.16 (dd, J = 8.4, 6.4 Hz, 2H), 3.14–3.04 (m, 4H), 2.33 (s, 6H), 1.86 (p, J = 7.4 Hz, 1H); ¹³C{¹H} NMR (101 MHz, chloroformd) δ 143.5, 143.4, 136.2, 135.5, 129.8, 129.7, 127.2, 127.1, 72.4, 70.0, 61.4, 61.2, 51.5, 48.4, 48.0, 47.3, 28.2, 21.4. IR $\nu_{\rm max}$ 3516, 3411, 2926, 2875, 1597, 1328, 1151, 1087, 727 cm⁻¹. HRMS (ESI) m/z [M + Na]⁺ calcd for C₂₂H₃₀O₈N₂NaS₂ 537.1700, found 537.1699.

N,N'-Bis-(p-toluenesulfonyl)cystamine (9). A 300 mL roundbottom flask equipped with a magnetic stir bar, a nitrogen inlet, and a pressure-equalizing addition funnel was charged with cystamine (10.0 g, 44.0 mmol), potassium carbonate (24.32 g, 176.0 mmol), and MilliQ water (70 mL). The addition funnel was charged with THF (75 mL) and p-toluenesulfonyl chloride (16.77 g, 88.0 mmol) and added dropwise over 2 h at room temperature. The mixture was vigorously stirred for 12 h, at which time, the THF was evaporated in vacuo, and the aqueous layer was diluted with 0.5 M HCl (50 mL) and extracted with EtOAc (3 \times 50 mL). The organic layer was washed with 3 M HCl (2 \times 20 mL), water (2 \times 20 mL), and saturated NaCl (1 \times 20 mL), dried over Na₂SO₄, filtered, and concentrated, providing a white solid. The crude product was recrystallized from methanol to give N,N'-bis-(para-toluenesulfonyl)cystamine (19.0 g, 39.6 mmol) in a 90% yield as a white solid. Consistency of spectral data is shown with the lit. data.²⁸ Melting point: 89–90 °C (lit. 77–78 °C)¹; $R_f = 0.10$ (7:3, Hex/EtOAc); IR $\bar{\nu}_{\rm max}$ 3281, 2925, 2865, 1595, 1418, 1324, 1157, 1092, and 814, cm⁻¹; ¹H NMR (chloroform-*d*, 300 MHz): δ 7.77 (d, J = 8.4 Hz, 2H), 7.33 (d, J = 8.4 Hz, 2H), 4.96 (t, J = 6.4 Hz, 1H), 3.26 (q, J = 6.4 Hz, 2H), 2.73 (t, J = 6.4 Hz, 2H), 2.45 (s, 3H); ¹³C{¹H} NMR (chloroform-d, 75 MHz): δ 21.7, 38.0, 41.8, 127.2, 130.0, 136.8, 143.8.

1,11-Dithia-4,8-diazacyclotetradecane (L1). A flame-dried 100 mL round-bottom flask equipped with a septum, a magnetic stir bar, and a nitrogen inlet was charged with small pieces of sodium metal (41.4 mg, 1.80 mmol), freshly distilled dimethoxy ethane (2 mL), and naphthalene (254.0 mg, 1.98 mmol). This mixture was stirred at room temperature under a nitrogen atmosphere for 2 h, affording a dark green solution. A second flame-dried round-bottom flask equipped with a septum, a magnetic stir bar, and a nitrogen inlet was charged with freshly distilled dimethoxy ethane (1 mL), and bis-sulfonamide 3 (100.0 mg, 0.180 mmol) was cooled to -30 °C via an external acetone/liquid nitrogen bath. The sodium naphthalenide solution was added dropwise using a gas-tight syringe to the round-bottom flask at -30 °C, until the solution maintained the dark green color for 5 min. The reaction was then allowed to warm to room temperature over 15 min and cooled to 0 °C via an external ice bath. At this time, water (1 mL) was added, which dissipated the green color immediately, and the solution was concentrated in vacuo. The residue was treated with 3 M HCl (10.0 mL, pH \sim 1) and washed with ethyl acetate (4 \times 5 mL). The aqueous layer was then basified using 6 M NaOH (5.00 mL, pH \sim 11) at 0 °C, and the freebase was extracted with chloroform (5 \times 5 mL), dried with sodium sulfate, filtered, and concentrated in vacuo. The orange oil was then subjected to a Dowex50W-X12 $(H^+)^b$ column (methanol wash, eluted with 6 M $\rm NH_{3(MeOH)})$ to afford L1 (36.70 mg, 0.157 mmol) as a clear oil in an 87% yield. ¹H NMR (400 MHz, chloroform-d) δ 2.88-2.81 (m, 4H), 2.80-2.72 (m, 12H), 1.89 (p, J = 6.5 Hz, 2H), 1.82 (p, J = 5.8 Hz, 2H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 46.9, 46.8, 32.4, 29.0, 28.3, 27.2. IR $\nu_{\rm max}$ 3284, 2912, 2809, 1660, 1445, 1294, 1256, 1129, 742 cm⁻¹. HRMS (ESI) $m/z [M + H]^+$ calcd for $C_{10}H_{23}N_2S_2$ 235.1297, found 235.1297.

1,11-Dithia-4,8-diazacyclotetradecane-4,8-diium Dichloride (L1-HCl). A 25 mL round-bottom flask was charged with 1 M $HCl_{(MeOH)}$ (3 mL), cooled to ~0 °C via an external ice bath, and L1 (36.70 mg, 0.157 mmol) in MeOH (1 mL) was added. The solution was warmed to room temperature and concentrated in vacuo affording 1,11-dithia-4,8-diazacyclotetradecane-4,8-diium dichloride (0.483 mg, 0.157 mmol) as a white residue in quantitative yield. ¹H NMR (400 MHz, deuterium oxide) δ 3.27 (t, J = 6.1 Hz, 4H), 3.23 (t, J = 7.3 Hz, 4H), 2.86 (t, J = 6.1 Hz, 4H), 2.73 (t, J = 6.4 Hz, 4H), 2.16 (p, J = 7.3 Hz, 2H), 1.86 (p, J = 6.3 Hz, 2H); ¹³C{¹H} NMR (101 MHz, deuterium oxide) δ 44.1, 42.5, 29.5, 27.5, 26.3, 20.5; HRMS (ESI) m/z [M-2HCl+H]⁺ calcd for C₁₀H₂₃N₂S₂ 235.1279, found 235.1298.

1,11-Dithia-4,8-diazacyclotetradecane Copper II Perchlorate (L1Cu(ClO4)₂). A 25 mL round-bottom flask equipped with a magnetic stir bar, a nitrogen inlet, and rubber septa was charged with MeCN (5 mL) and L1 (36.70 mg, 0.157 mmol) and cooled to ~0 °C via an external ice bath; in copper(II) perchlorate, MeCN (2 mL) was added. The solution immediately turned a vibrant purple color and was stirred at room temperature for an additional 30 min, followed by concentration in vacuo. Chelate L1Cu(ClO4)₂ was obtained as purple prisms in quantitative yields. [(L1Cu)₂ μ -Cl](ClO₄)₃ was isolated using an identical procedure but as a separate sample. Caution! Perchlorate salts of metal complexes containing organic ligands as solids or in organic solvents are potentially explosive. Although no problems were encountered by us, cautious handling of only small amounts of these compounds should be the rule.

3,3'-(Ethane-1,2-diylbis(sulfanediyl))dipropanenitrile (13). A Pyrex test tube was charged with potassium carbonate (0.304 g, 2.20 mmol), 1,2-ethanedithiol (0.08 mL, 1.00 mmol), and acrylonitrile (0.28 mL, 4.40 mmol). This solution was placed in a sonication bath and sonicated for 30 min at 53 °C. After which, chloroform (5 mL) was added, potassium carbonate was filtered off, and the mother liquor was concentrated in vacuo to provide 0.197 g (98%) of 3,3'-(ethane-1,2-diylbis(sulfanediyl))dipropanenitrile as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 2.83 (s, 2H), 2.82 (t, *J* = 7.0 Hz, 2H), 2.65 (t, *J* = 7.0 Hz, 2H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 118.4, 32.2, 27.8, 19.1. IR ν_{max} 2938, 2917, 2239, 1417, 1279, 1213, 901, 739 cm⁻¹. HRMS (ESI) m/z [M + Na]⁺ calcd for C₈H₁₂N₂NaS₂ 223.0334, found 223.0335.

1,4-Dithia-8,11-diazacyclotetradecane Bis-hydrochloride (L2-HCI). A 25 mL round-bottom flask equipped with a condenser, a nitrogen inlet, and a magnetic stir bar was charged with 8,11-ditosyl-1,4-dithia-8,11-diazacyclotetradecane (12) (0.200 g, 0.368 mmol) and xylene (0.12 M, 3.1 mL). The solution was cooled to 0 °C, Red-Al (3.5 M (toluene), 1.7 mL) was added dropwise, and the mixture was refluxed with a heating mantle for 2 h and then cooled to 23 °C. The solution was cooled to 0 °C, 3 M sodium hydroxide (4 mL) was added dropwise, and the cloudy solution was extracted with chloroform (5 \times 15 mL). The combined organic layers were dried over sodium sulfate, filtered, and concentrated in vacuo to afford a yellow residue. The crude product was treated with methanolic HCl (1 M, 10 mL), concentrated to ~1 mL, and filtered. The filter cake was washed with cold methanol (1 mL) to provide L2-HCl (0.984 g, 0.320 mmol) as a white powder in an 87% yield. ¹H NMR (400 MHz, deuterium oxide) δ 4.65 (s, 4H), 3.51 (s, 4H), 3.28 (t, J = 7.3 Hz, 4H), 2.85 (s, 4H), 2.68 (t, J = 6.7 Hz, 4H), 1.98 (p, J = 6.9 Hz, 4H); $^{13}C{^{1}H}$ NMR (101 MHz, deuterium oxide) δ 47.7, 42.9, 33.7, 30.5, 26.1. IR $\nu_{\rm max}$ 3381, 2935, 1616, 1455, 1015, 805 cm⁻¹. HRMS (ESI) m/z [M-2HCl+H]⁺ calcd for C₁₀H₂₃N₂S₂ 235.1297, found 235.1298.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.9b01682.

¹H, ¹³C, gCOSY, gHSQC, and gHMBC spectra, including X-ray crystallography, and HRMS (PDF) X-ray crystallographic data for compound 7 (CIF) X-ray crystallographic data for compound **8b** (CIF)

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X-ray crystallographic data for compound 4 (CIF) X-ray crystallographic data for compound 3 (CIF) X-ray crystallographic data for compound 10 (CIF) X-ray crystallographic data for compound L1-HCl (CIF) X-ray crystallographic data for compound $[(L1Cu)_2\mu$ -Cl](CIO₄)₃ (CIF) X-ray crystallographic data for compound L1Cu(ClO4)₂

(CIF)

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Notes

The authors declare no competing financial interest.

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ADDITIONAL NOTES

^{*a*}Potential nitrogen mustard derivative, use safe laboratory practices when isolating the product.

^bIf ammonium salts are present after ion-exchange chromatography, dissolve the freebase in chloroform, filter, and concentrate in vacuo to provide L1 free of salts.

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