



Contents lists available at ScienceDirect

Journal of Molecular Liquids

journal homepage: www.elsevier.com/locate/molliq

Azomethine-functionalized task-specific ionic liquid for diversion of toxic metal ions in the aqueous environment into pharmacological nominates

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ARTICLE INFO

Article history:

Received 11 August 2020

Received in revised form 21 September 2020

Accepted 6 October 2020

Available online xxxx

Keywords:

Task-specific ionic liquid

Toxic metal ions

Remediation by complexation

Antimicrobial

Cytotoxicity

ABSTRACT

By this work, a new azomethine-supported task-specific ionic liquid (ABIIL) was fabricated based on bis-(butylimidazolium hexafluorophosphate) salicylidene Schiff base. The successful formation of ABIIL was confirmed from its microanalytical and spectral results. The excellent aqueous-ethanolic solubility coupled with the high chelating capacity of ABIIL enabled it to act as a multifunctional scavenger for removing of Cu(II) and Fe(III) ions from aqueous environments. Not only has that, but also transformed these toxic ions into promising pharmacological candidates. ABIIL showed an excellent scavenging efficiency, as it can uptake up to 98.9% and 96.7% of Fe(III) and Cu(II) ions, respectively, from aqueous-ethanolic effluents. Besides, it could be easily regenerated and reused for six cycles without significant loss in its efficiency. The scavenging outputs (M-ABIIL) exhibited excellent and broad-spectrum antimicrobial activity, with a slight preference toward fungi than bacteria. Fe-ABIIL is the most potent antimicrobial nominate as revealed from its MIC values ($\mu\text{g/mL}$): 16.8 (*C. albicans*) < 18.9 (*S. aureus*) < 19.5 (*A. flavus*) < 23.1 (*E. coli*). Additionally, this complex is more cytotoxic (IC_{50} $3.40 \pm 0.53 \mu\text{g/mL}$) than the clinical anticancer drug (Vinblastine, VB) (IC_{50} $3.79 \pm 1.01 \mu\text{g/mL}$) toward colon cancer cells (HTC116), whereas it is less cytotoxic (IC_{50} $85.66 \pm 1.44 \mu\text{g/mL}$) than VB (IC_{50} $57.12 \pm 1.95 \mu\text{g/mL}$) to normal human cells (HeLa). Thus, ABIIL has great potential for removing toxic Cu(II) and Fe(III) ions from aqueous environments, converting them into chemotherapeutic agents.

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1. Introduction

Ionic liquids (ILs) have been categorized as eco-friendly alternatives for traditional toxic organic solvents due to their amazing characteristics such as promoted mechanical and thermal stability, high solubilization power, reusability, tunable chemical/ physical properties, non-volatility, non-flammability, and intrinsic conductivity [1]. Moreover, the toxicity study revealed that most of ILs showed relatively low toxic impacts toward several biological systems [2]. These unique features of ILs make them the best choice for many researchers to design and refine their chemical structures (i.e. fine-tuning the cation and/or anion), rendering them inimitably suited for many applications [3]. These functionalized ILs often referred to as task-specific ionic liquids (TSILs) which have been designed, fabricated, and tuned by

modification their chemical constituents according to the user's needs. For instance, they explored their specific efficiency in organic synthesis [4], nanomaterials fabrication [5], catalysis [6], extraction [7], electrochemistry [8], analytical chemistry [9], active pharmaceutical ingredients (API) [10] and so far [3]. Recently, utilized as scavengers for primary amines [11] and a turn-on pH nano-fluorosensor [12].

Previous studies demonstrated that the TSILs could offer promising scavengers for the removal of metal ions from aqueous effluents [13,14]. Despite there are many TAILS that have been investigated for extraction and separation of metal ions from aqueous solutions based upon physical interactions [13–15]; a few trials have been performed to endow coordinating groups to the ionic liquid, rendering them more fiercely for extraction of metal ions. These few trials involve supporting of ionic liquid with thiourea [16], macrocyclic ethers [17], 2-hydroxybenzylamine [18], ethylene glycol [19], carboxylic groups [20], and recently aminothiazoly Schiff bases [21] or porous organic copolymer [22]. The preference of the chelating TSILs over non-chelating ones is due to the availability of the functional group which provides

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them the advantage of the ability to chelate diverse metal ions with pH-controlled selectivity and pH-dependent stability [23].

Amongst toxic heavy metal ions (HMIs) popularly distributed in aqueous environments, cupric (Cu(II)) and ferric (Fe(III)) ions were deemed as potential toxicity sources [24,25] due to their capacity to generate free radicals and reactive oxygen species (ROS) in the live cells [24,26], inducing their programmed death. The major sources of the toxic ferric ions in aqueous environments are; steel tempering, mining, coal coking, and metallurgical industries [27]. On the other hand, cupric ions have emitted in aqueous effluents as a result of many industrial processes, such as electroplating, metal finishing, etching, and plastics [28]. As the Cu(II) and Fe(III) ions are very toxic metal even at low concentrations, therefore, the cupric- and ferric-polluted wastewater should be remediated before discharging into the aquatic environment [27,28].

Few trials have been reported to remove cupric and ferric ions from aqueous solutions using TSILs. For instants, imidazolium-supported TSILs have been applied as scavengers for M(II/III) ions (M = Cu, Co, Ni, Zn, Cd, Fe) from their respective aqueous solutions; by chelating them [20,29]. In addition, ammonium-based TSILs showed moderate to good effect in removing and recovering of several metal ions (Cu (II), Pb(II), Zn(II), Cd(II), Ni(II), Co(II), and Fe(III)) from aqueous effluents [29,30]. Recently, microcapsules composed of styrene and divinylbenzene were used to encapsulate pyridinium TSILs for Zn(II) and Cu(II) recovery from aqueous media [31]. Nevertheless, these reported TSILs are limited and their routes of fabrication are sophisticated and expensive.

All these aforementioned facts prompted us to design and fabricate a new azomethine-supported bis-butylimidazolium chelating task-specific ionic liquid (ABIII) which will be utilized not only for the removal of Cu(II) and Fe(III) ions from the aqueous solutions but also to convert these ions to metals-based pharmacological nominates.

2. Experimental part

2.1. Materials and instrumentation

Chemicals, solvents, and practical procedures utilized in the preparation of salicylaldehyde-supported butylimidazolium ionic liquids (SBIII) (**2a,b**) and 1-(3-aminopropyl)-3-butyl-1H-imidazolium hexafluorophosphate (**5**) coupled with the instrumentation used for the comprehensive characterization of prepared compounds were described in the online electronic supplementary information (ESI†).

2.2. Synthesis of ABIII

Into a 100 mL round-bottomed flask (RBF), a solution of SBIII (**2b**) (1.62 g, 4 mmol) in ethanol (35 mL) was gradually added to an ethanolic solution (25 mL) of the aminopropyl derivative (**5**) (1.31 g, 4 mmol) containing a catalytic amount of glacial acetic acid (three drops) under stirring. Then, the reaction mixture was heated under reflux and continuous stirring for 3–5 h (TLC was used to monitor the completion of reaction). After reaction completion, the solvent was partially evaporated followed by cooling of the content of RBF to ambient temperature. The precipitated product was filtered, washed with cold ethanol and ether, (3 × 5 mL) for each, dried, and then recrystallized from aqueous ethanol to give ABIII as yellow fine crystals. Yield (2.49 g, 89%), mp 98–99 °C. FTIR (KBr, cm⁻¹): 3336 (m, br), 3079 (m, br), 2982 (m, sh), 2959 (m, sh), 2889 (m, sh), 1631 (vs, sh), 1567, 1549, 1466 (s, sh), 1335 (m, sh), 1279 (s, sh), 1150 (s, sh), 965 (m, sh), 836 (vs, sh), 735 (m, sh). ¹H NMR (200 MHz, DMSO-*d*₆) δ (ppm): 10.19 (s, 1H), 9.43 (s, 1H), 9.12 (s, 1H), 9.04 (s, 1H), 8.11–7.96 (m, 2H), 7.82–7.76 (m, 2H), 7.39 (d, 1H, *J* = 7.1 Hz), 7.06–7.01 (m, 2H), 5.69 (s, 2H), 5.10 (t, 2H, *J* = 6.9 Hz), 4.26 (t, 2H, *J* = 7.2 Hz), 4.11 (t, 2H, *J* = 6.9 Hz), 3.99 (t, 2H, *J* = 6.8 Hz), 2.25–2.09 (m, 6H), 1.96–1.81 (m, 4H), 0.93 (t, *J* = 7.2 Hz, 3H), 0.85 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (125 MHz,

DMSO-*d*₆) δ (ppm): 161.33, 158.21, 137.86, 136.22, 133.14, 132.42, 125.02, 123.07, 122.70, 118.74, 117.63, 57.62, 55.91, 54.81, 51.970, 49.02, 36.13, 32.97, 31.61, 31.09, 21.34, 19.17 and 13.61. ³¹P NMR (202 MHz, DMSO-*d*₆): δ -142.95 (hept, *J* = 711.2 Hz). ¹⁹F NMR (471 MHz, DMSO-*d*₆) δ: -70.57 (d, *J* = 711.2 Hz). ESI-MS: in positive mode peaks at *m/z* 568.4 ([C₂₅H₃₇F₆N₅OP]⁺, M - PF₆⁻) a.m.u. and 211.6 ([C₂₅H₃₇N₅O]²⁺, M - 2 PF₆⁻) a.m.u. Anal. Calcd. for C₂₅H₃₇F₁₂N₅OP₂ (M = 713.53 g/mol): C, 42.08; H, 5.23; N, 9.82%. Found: C, 41.99; H, 5.31; N, 9.78%.

2.3. Deprotonation constant (pK_a) of ABIII

The dissociation constant (pK_a) of ABIII was estimated at 298 K in its aqueous solution (10⁻⁴ M) of ionic strength of 0.04 M (KNO₃) by titration with an aqueous NaOH solution (1.6 × 10⁻³ M).

2.4. Scavenging of Cu(II) and Fe(III) ion by ABIII extractant

A general protocol mimic to the common solid-phase extraction (SPE) technique [32] has been utilized for removal of heavy metal ions (HMIs) (Cu(II) and Fe(III)) from aqueous ethanolic solutions by ABIII as an extracting agent. In brief, a solution of metal ion (0.5 mmol, 0.10 g Cu(CH₃CO₂)₂·2H₂O; 0.302 g Fe(NO₃)₃·9H₂O) in distilled water (5 mL) was slowly added to the pale-yellow solution of ABIII (0.71 g, 1.0 mmol) in ethanol (30 mL) containing 100 μL of 0.1 M NaOH_{aq}. Then, the reaction mixture was heated under reflux with stirring for 1 h. After cooling to ambient temperature, the isolated colored solids were collected by filtration, washed with cold ethanol and diethyl ether, (2 × 3 mL) for each, and dried in a desiccator. Samples of the obtained complexes were analyzed as follow;

[Cu(ABIII)₂] (Cu-ABIII): Dark yellowish green crystals, Yield 69%. FTIR (KBr, cm⁻¹): 3125 (w, br), 1647 (m, sh), 1622 (s, sh), 1549 (s, sh), 1457 (m, sh), 1336 (m, sh), 1271 (m, sh), 1150 (m, sh), 837 (vs, sh), 643 (m, br), 525 (w, br), 478 (m, sh). ESI-MS: in positive mode peaks at *m/z* 1315.3 ([C₄₈H₆₈CuF₁₈N₁₀O₂P₃]⁺, M - PF₆⁻) a.m.u., 585.2 ([C₄₈H₆₈CuF₁₂N₁₀O₂P₂]²⁺, M - 2 PF₆⁻) a.m.u., 341.6 ([C₄₈H₆₈CuF₆N₁₀O₂P]³⁺, M - 3PF₆⁻) a.m.u., and 220.0 ([C₄₈H₆₈CuN₁₀O₂]⁴⁺, M - 4 PF₆⁻). Anal. Calcd. for C₄₈H₆₈CuF₂₄N₁₀O₂P₄ (M = 1460.54 g/mol): C, 39.47; H, 4.69; N, 9.59%. Found: C, 39.42; H, 4.71; N, 9.38%.

[Fe(ABIII)₂(H₂O)(NO₃)] (Fe-ABIII): Dark purple powder, Yield 65%. FTIR (KBr, cm⁻¹): 3453 (m, br), 3122 (w, br), 1620 (s, sh), 1549 (s, sh), 1458 (m, sh), 1336 (s, sh), 1272 (m, sh), 1151 (m, sh), 839 (vs, sh), 653 (m, br), 506 (w, br), 458 (w, br). ESI-MS: in positive mode peaks at *m/z* 1387.6 ([C₄₈H₇₀F₁₈FeN₁₁O₆P₃]⁺, M - PF₆⁻) a.m.u., 621.2 ([C₄₈H₇₀F₁₂FeN₁₁O₆P₂]²⁺, M - 2 PF₆⁻) a.m.u., 365.7 ([C₄₈H₇₀F₆FeN₁₁O₆P]³⁺, M - 3 PF₆⁻) a.m.u., and 238.1 ([C₄₈H₇₀FeN₁₁O₆]⁴⁺, M - 4 PF₆⁻). Anal. Calcd. for C₄₈H₇₀F₂₄FeN₁₁O₆P₄ (M = 1532.86 g/mol): C, 37.61; H, 4.60; N, 10.05%. Found: C, 37.52; H, 4.63; N, 9.98.

2.5. Regeneration of ABIII scavenger

An aqueous solution containing a mixture of HCl (1.0 M, 3 mL) and thiourea (4% w/v) was added to the extracting products (M-ABIII complexes) (100 mg). Thereafter, these mixtures have subjected to ultrasonic irradiation for 30 min. The color change of the reaction solution from colorless to yellow color was an indication of the progress of the ABIII-recovering process. After reaction completion, the reaction mixture was neutralized with an aqueous NaHCO₃ solution (1.0 M) which leads to the precipitation of the parent scavenger (ABIII). The recovered ligand (ABIII) was collected by filtration, washed several times with Milli-Q water to remove the undesired byproducts (M-thiourea complexes). Then, this scavenger was dried under reduced pressure and can be used for a further removal process.

2.6. Pharmacological performance study

2.6.1. Antimicrobial activity

The antimicrobial performance for the scavenging products (Cu(II)/Fe(III)-ABILL complexes) was assessed in comparison to the native scavenger (ABILL) and clinical drugs (antibacterial, ampicillin; antifungal, amphotericin B), against different microbial strains including *Staphylococcus aureus* (*S. aureus*, ATCC-25923) as a Gram-positive bacterium, *Escherichia coli* (*E. coli*, ATCC-25922) as a Gram-negative bacterium, and *Aspergillus flavus* (*A. flavus*) & *Candida albicans* (*C. albicans*, NCIM No. 3100) as fungal species. This study was performed according to the usual antimicrobial evaluation protocol followed in our earlier reported works [11,33]. The diameter of zone of inhibition (ZOI, mm) and minimum inhibitory concentration (MIC) were used as antimicrobial activity indices for the tested compounds.

2.6.2. Anticancer activity

The anticancer efficacy of the scavenging products was evaluated in comparison to a clinical anticancer drug (Vinblastine, VB) against two human cell lines; colon carcinoma (HCT116) and normal cells (Hela). Again, this work was carried out using the standard MTT protocol described in our previous study [34]. The cell viability% and IC₅₀ values were utilized as cytotoxicity activity markers for the examined compounds.

3. Results and discussion

3.1. Synthesis of new scavenger (ABILL)

The synthesis of new task-specific ionic liquid (TSIL) based on salicylidene moiety and butylimidazolium ionic liquids (BILs) terminals involves seven chemical reactions. Initially, quaternization of 1-butylimidazole with *N*-(2-bromoethyl)phthalimide (**1**) to generate 1-(2-(phthalimido)ethyl)-3-butylimidazolium bromide (**2a**), which is readily transformed into the corresponding hexafluorophosphate salt through an anion metathesis reaction using an aqueous HPF₆. Then, the 3rd step involves a dephthaloylation of compound **2b** by the traditional hydrazinolysis reaction using hydrazine monohydrate in an ethanolic medium followed by simple extraction with dichloromethane (DCM), filtration, and evaporation of solvent to separate a pure 1-(2-aminoethyl)-3-butylimidazolium hexafluorophosphate (AEBI, **3**). Other sets of reactions were dedicated for the preparation of salicylaldehyde-supported butylimidazolium ionic liquids (SBILs); in the fourth reaction, a bromomethylation process was performed on salicylaldehyde to obtain 5-(bromomethyl) salicylaldehyde (**4**) which is utilized as an alkylating agent for the quaternization of 1-butylimidazole to produce salicylaldehyde-based butylimidazolium bromide (**5a**). Thereafter, a bromide exchange in compound **6a** with hexafluorophosphate anion was performed to prepare compound **5b**. Eventually, a classical Schiff base condensation was carried out between the amino component (**3**) and SBILL (**5b**) in ethanolic solution to fabricate the desired scavenger, 1-butyl-3-((2-(1-butyl-imidazolium-3-yl)ethyl)imino)methyl)-4-hydroxybenzyl)-imidazolium hexafluorophosphate (ABILL). It was obtained in an excellent yield with high purity and structurally characterized based on the elemental analysis and spectroscopic methods (FTIR, UV-Vis, NMR (¹H, ¹³C, ¹⁹F, ³¹P) and ESI-MS).

3.2. Structural characterizations of new scavenger (ABILL)

Regarding solubility, ABILL is insoluble in water (Solubility in water = 9.7 g/L) and methanol but soluble in ethanol and has aqueous stability in basic and neutral solutions (pH = 10–5); while it is readily decomposed, by dissociation of azomethine bond, at low pH values (pH < 4) like the behavior of most Schiff bases [35]. The elemental analysis of new scavenger (ABILL) gave satisfactory results which in good consistent with the suggested molecular formula of it (see experimental part).

The positive mode electrospray ionization mass spectra (ESI-MS) of ABILL displayed four distinctive peaks at *m/z* 568.4 and 211.6 corresponding to the successive departure of hexafluorophosphate counter anions to generate a single-charged [M – PF₆]⁺ and double-charged [M – 2 PF₆]²⁺ cations. Also, these data agree with the molecular formula proposed by the elemental analyses.

The appearance of two sets of absorption bands (characteristic for AEBI and SBILL) and vanishment of others along with the emergence of new ones in the spectrum of the new scavenger (ABILL) (see Fig. S1, ESI) confirms its successful fabrication. Just to name few, the two main peaks centered at 3336 and 1279 cm⁻¹ are assigned to the vibration of hydroxyl and aryl-O fragments of the phenolic group in the SBILL segment. However, absorption bands noticed at 1466, 965, and 836 cm⁻¹ are ascribed to the vibration of imidazolium and PF₆⁻ fragments of ionic liquids terminals in the AEBI and SBILL segments. On the other hand, vanishment of the stretches of the NH₂ group (at 3501 and 3398 cm⁻¹ in AEBI spectrum) and aldehyde moiety (at 2820 and 1658 cm⁻¹ in SBILL spectrum), coupled with the growth of new stretch at 1631 cm⁻¹ (assignable for the vibration of azomethine group) in the spectrum of ABILL; proves the successful formation of the new scavenger (ABILL) by Schiff base condensation of the amino group of AEBI with an aldehyde moiety of SBILL.

The coexistence of two groups of NMR signals in the ¹H NMR spectrum of ABILL (Fig. 1) confirms the successful condensation of AEBI with SBILL to form ABILL. (See Scheme 1.)

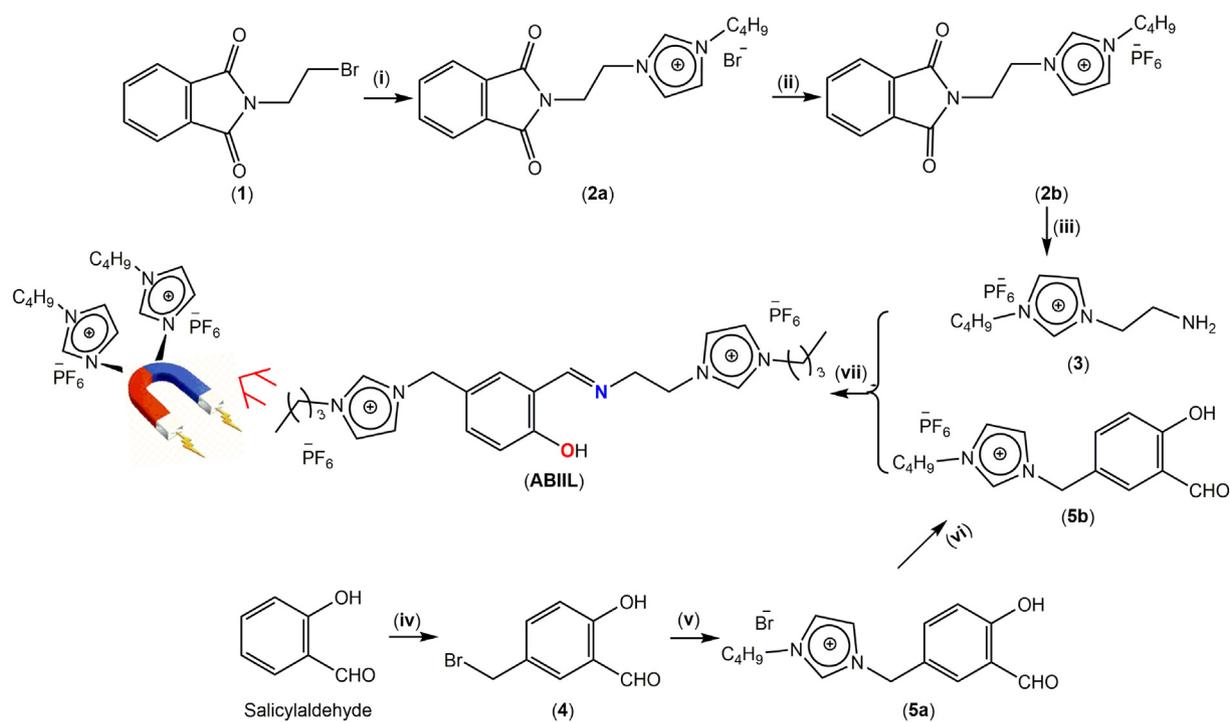
As shown in Fig. 1, the first set of ¹H NMR peaks are centered at chemical shift values of 10.19, 9.04, 8.11–7.96, 5.10, 4.26, and 3.99 ppm (distinctive for the aminoethyl butyl imidazolium (AEBI) part); while the other group which is positioned at 9.12, 7.82–7.01, 5.69, and 4.11 ppm are characteristic for SBILL segment. Interestingly, the emergence of a new singlet at δ 9.43 ppm corresponding to the resonance of azomethine proton, coupled with the downfield shift of the phenolic (–OH) proton to δ 10.19 ppm are ascribed to the intramolecular H-bonding (O–H⋅N) between the azomethine (–CH=N–) and ortho-hydroxyl groups in ABILL.

Additionally, the ¹³C NMR technique provides informative structural insight concerning the carbon-atoms distribution map in the skeleton of ABILL scavenger. The ¹³C NMR spectrum of ABILL revealed the presence of C-atoms distinctive for both AEBI and SBILL segments (see Fig. 2A). Noteworthy, this spectrum exhibited two characteristic peaks at δ 161.33 and 158.21 ppm due to the nuclear resonances of phenolic (C=O) and azomethinic (–CH=N) carbon atoms, respectively. Powerful evidence for anchoring of ionic liquids on the terminals of ABILL is the appearance of a septet signal (centered at δ –142.95 ppm, *J* = 711.2 Hz) and a doublet peak (centered at δ –70.57 ppm, *J* = 711.2 Hz) in the ³¹P (Fig. 2B) and ¹⁹F NMR (Fig. 2C) spectra of ABILL, respectively.

3.3. pH-potentiometry

As the chelating capacity of any protic ligand is strongly correlated to the extent of its ionization and the stability of its ionizable form at different pH values, so, it is of importance to examine the deprotonation process and dissociation constant of ABILL. In this context, the dissociation performance of ABILL was examined by a pH-potentiometric method in an aqueous ethanolic solution at ambient temperature. Moreover, the stability of ABILL in aqueous media was inspected by a pH-potentiometric titration of its solution (10⁻⁴ M) against NaOH (10⁻³ M) (Fig. S2, ESI†). This titration curve showed a single abrupt change corresponding to the departure of one proton from ABILL without losing its identity at a wide pH-range (4.8–10.5). The average number of acidic protons in ABILL (\bar{n}_A) was calculated at serial pH values according to Irving–Rossotti equation (Eq. (1)) [36]:

$$n_A^- = Y - \frac{V_1 N^\circ}{(V_1 + V^\circ) T_1} \quad (1)$$



Scheme 1. The stepwise schematic protocol for the preparation of 1-(2-aminoethyl)-3-butylimidazolium hexafluorophosphate (AEBI, 3), salicylaldehyde-supported butylimidazolium ionic liquids (SBILs), and ABILL scavenger.

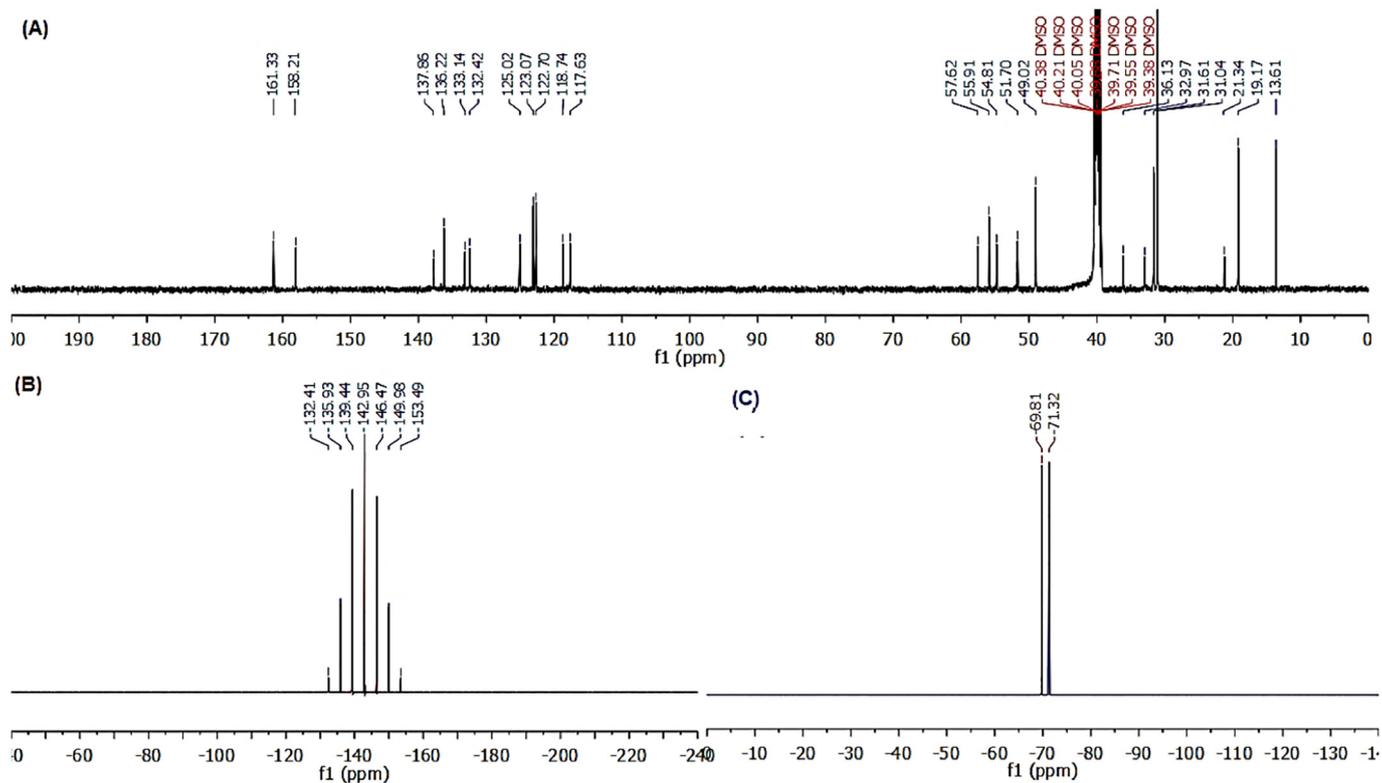
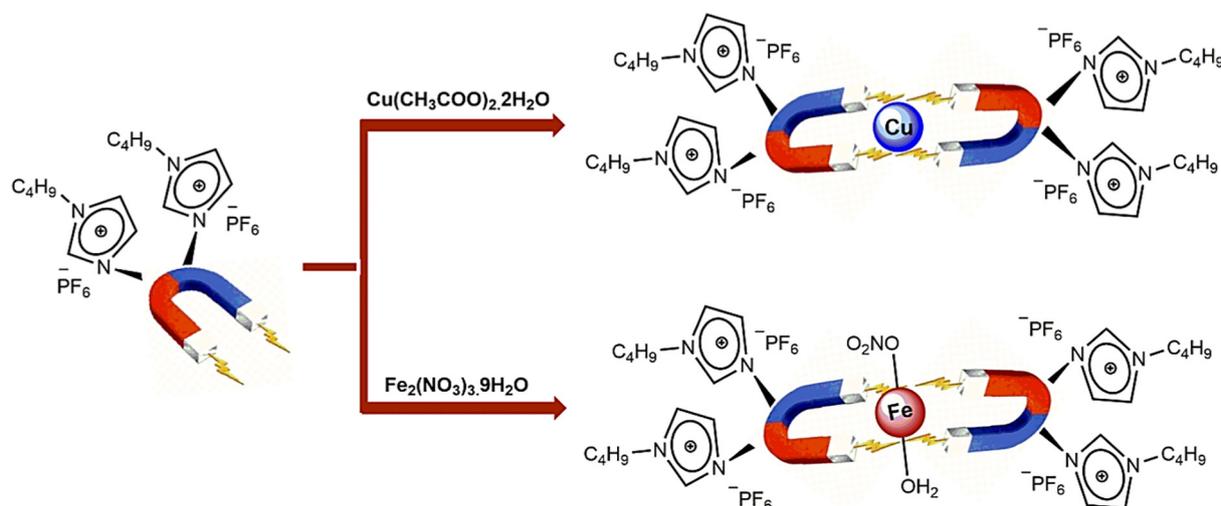


Fig. 2. (A) ¹³C NMR (126 MHz); (B) ³¹P NMR (202 MHz); and (C) ¹⁹F NMR (471 MHz) spectra of the ABILL scavenger in *d*₆-DMSO.



Scheme 2. Scavenging of toxic metal ions (Cu(II) and Fe(III)) by ABIIIL and conversion into the corresponding complexes.

Table 1

Removing of the target metal ions by the new scavenger (ABIIIL); in comparison with reported ones.^a

| Scavenger | Metal ions | Operating conditions | | | Uptake (%) | Ref. |
|-----------------------------------|------------|----------------------|--------|------------|------------|-----------|
| | | pH | T (°C) | Time (min) | | |
| ABIIIL | Cu(II) | 6.4 | 25 | 60 | 96.7 | This work |
| | Fe(III) | 6.4 | 25 | 60 | 98.9 | This work |
| HATPS | Cu(II) | 6–7 | 78 | 120 | 87.2 | [21] |
| | Fe(III) | 6–7 | 78 | 120 | 94.3 | [21] |
| Activated C | Cu(II) | 6–7 | 37 | 40–60 | 90.3–95.6 | [38] |
| | Fe(III) | 1.5 | 20 | 20 | 53 | [39] |
| Fly ash | Cu(II) | 6 | 40 | 90 | 91.4 | [38] |
| | Cu(II) | 6 | 20 | 24 h | 94.6 | [38] |
| GO | Fe(III) | 2–3 | 20 | 10 h | 44.3 | [40] |
| | Cu(II) | 8 | 25 | 5 | 96.6 | [41] |
| GO-NH ₂ | Fe(III) | 8 | 25 | 5 | 95.3 | [41] |
| | Cu(II) | 5.3 | 20 | 24 h | 98.9 | [38] |
| GO/Fe ₂ O ₃ | Cu(II) | 5 | 65 | 120 | 88.1 | [42] |
| | Fe(III) | 5 | 65 | 120 | 74.3 | [42] |
| Zeolite | Cu(II) | 4.5 | 20 | 300 | 91.2 | [38] |
| | Fe(III) | 7 | 37 | 180 | 83.5 | [43] |
| PSCSB | Cu(II) | 8 | 25 | 150 | 94.1 | [44] |
| PISCSB | Cu(II) | 8 | 25 | 30 | 94.5 | [44] |

^a Scavenging conditions for ABIIIL; ABIIIL (0.5×10^{-3})/Metal ion (0.5×10^{-3}) solutions, ethanol (10 mL) containing 100 μ L of 0.1 M NaOH_{aq}, stirring 60 min/25 °C. GO = Graphene oxide; GO-NH₂ = Amine-functionalized graphene oxide; CS = Chitosan; PSCSB = poly(salicylidene) chitosan Schiff base; PISCSB = poly(ionic-salicylidene) chitosan Schiff base.

3.6. Structural features of the scavenging products

The micro-analytical analysis of the scavenging products (Cu(II)/Fe(III)-ABIIIL) agrees with their proposed molecular formulas, [Cu(ABIIIL)₂] and [Fe(ABIIIL)₂(H₂O)(NO₃)], (see [Experimental](#) section). On the other hand, the structural formulas of these complexes could be deduced based on spectral analysis (IR and UV-Vis). FTIR is essential to predict the ABIIIL binding sites with metal ions and the coordination sphere for each metal ion, as well. Synchronous changes in the intensities and/or the positions of the absorption bands recorded in the FTIR spectra of the scavenging products (M-ABIIIL) in comparison to those in the spectrum of scavenger (ABIIIL) (cf. Fig. S1, ESI); coupled with the emergence of new absorptions bands, confirms the success of chelation of metal ion by ABIIIL. Moreover, based on these changes we can propose the most reasonable M-ABIIIL chelation mode. For instance, the phenolic (-OH) absorption band (at 3336 cm⁻¹ in the spectrum of ABIIIL) was disappeared in the spectra of complexes, confirming of its deprotonation to form phenolate anion that chelates metal ions. Additionally, shifting in the wavenumber of the C—O peak ($\Delta\nu = - (9-14)$ cm⁻¹) in the spectra of M-ABIIIL complexes as compared to the native ABIIIL along with the emergence of a weak peak at the rang of 654–547 cm⁻¹ (distinctive for M—O stretching vibration), offer more evidence for sharing of the phenolic (OH) group in chelation of metal ions [45,46]. The band due to the azomethine group (-C=N) was negatively

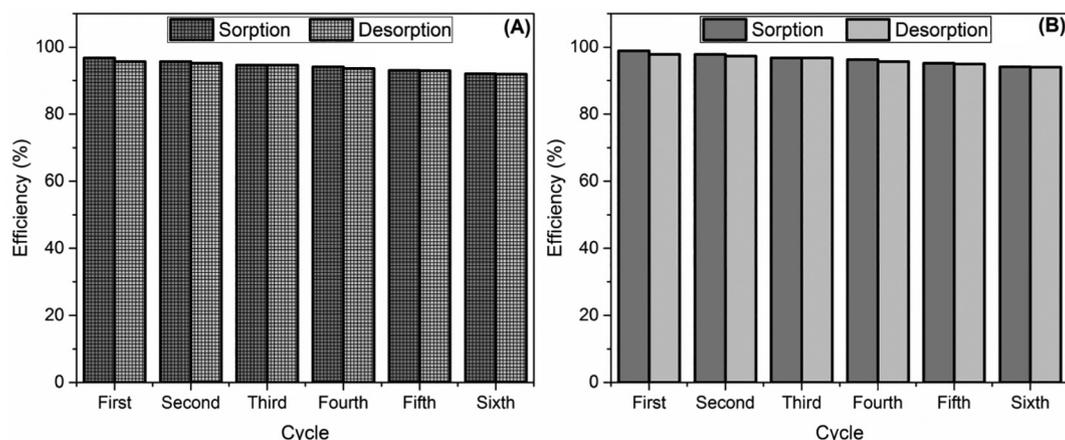


Fig. 3. Scavenging efficiency of the ABIIIL for (A) Cu(II) ions and (B) Fe(III) ions after six consecutive scavenging (sorption)–regeneration (desorption) cycles.

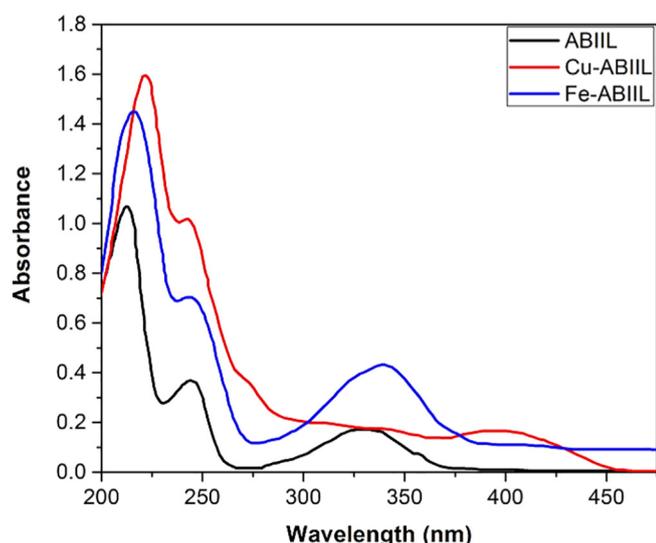


Fig. 4. UV-Vis spectra of ABIIIL scavenger and its scavenging products (Cu-ABIIIL and Fe-ABIIIL).

shifted from 1631 cm^{-1} , in free ligand to $1621 \pm 1\text{ cm}^{-1}$ ($\Delta\nu = -11\text{ cm}^{-1}$), in complexes, as indicative of the coordination of the azomethinic N atom with metal ions [21,46]. This is consistent with the growth of a new absorption band at the rang of $465\text{--}442\text{ cm}^{-1}$ assignable for M-N vibration. Thus, ABIIIL act as a monoanionic bidentate (NO) chelating ligand. The broad band observed at 3453 cm^{-1} in the spectrum of Fe(III)-ABIIIL complex, characteristic for the O-H group, emphasizes the involvement of a water molecule in the coordination sphere for Fe(III) ion [21].

The UV-Vis spectra of the solutions of ABIIIL scavenger and its metal complexes in DMSO (10^{-3} M) were measured within a wavelength range of $200\text{--}500\text{ nm}$ at ambient temperature (Fig. 4). The electronic spectrum of ABIIIL is dominated by three main maxima at 337, 249, and 221 nm attributable for the $n \rightarrow \pi^*$ and $\pi \rightarrow \pi^*$ transitions implicated in the azomethine, phenolic, and imidazolium moieties, respectively [47,48]. UV-visible spectroscopy is a very helpful tool in predicting the geometries of complexes on the basis of the identities of d-d transitions. For example, the electronic spectrum of the Cu-ABIIIL complex showed a new peak at 397 nm which can be assigned as $d \rightarrow \pi^*$ metal-ligand charge-transfer (MLCT) transitions (superimposed ${}^2B_{1g} \rightarrow {}^2B_{2g}$ and ${}^2B_{1g} \rightarrow {}^2E_g$ transitions) that distinctive for the distorted square-planar geometry of Cu(II) complexes [47,49]. The measured magnetic moment for this complex was found to be

1.82 BM which agrees with the presence of odd electron on Cu(II) ion in a square-planar geometry environment [50]. On the other hand, the solution of the Fe-ABIIIL complex displayed a characteristic peak at 343 nm due to the spin-forbidden ${}^6A_{1g} \rightarrow T_{2g}$ transition of a high-spin Fe(III) ion (d^5) in an octahedral field. However, the high-spin octahedral Fe(III) complexes usually show a very weak spin-forbidden d-d transition that hasn't appeared in their UV-Vis spectra. Also, the magnetic moment of the Fe-ABIIIL complex (5.62 BM) proposed the high-spin configuration of the Fe(III) ion (d^5 configuration) in an octahedral environment [51].

The stability of the scavenger (ABIIIL) and its complexes was checked beneath the physiological conditions ($\text{pH} = 7.4$, $T = 36.5\text{--}37.5\text{ }^\circ\text{C}$) by storing a solution (10^{-3} M) of each sample in a PBS-DMSO buffer for 72 h; observing the physicochemical changes and recoding the UV-Vis spectra of each stock solution at time intervals (every 24 h). Noteworthy that no physicochemical changes such as color alteration or formation of precipitates as evidence no aggregation of ABIIIL or its complexes in their respective solutions. Meanwhile, no significant changes were noticed in the UV-Vis spectra of ABIIIL or its complexes during the storing time (Fig. S4, ESI), confirming our earlier suggestion.

3.7. Pharmacological study

3.7.1. Antimicrobial study

To assess the suitability of scavenging products (M-ABIIIL complexes) for application as antibiotics, the capacity of these complexes to suppress the growth of common medically-relevant microbes such as *S. aureus* (ATCC-25923) (Gram-positive (G^+) bacterium), *E. coli* (ATCC-25922) (Gram-negative (G^-) bacterium), *A. flavus*, and *C. albicans* (NCIM No. 3100) (fungi) were in vitro investigated in comparison to the native scavenger (ABIIIL) and clinical drugs (Antibacterial, Streptomycin (ST), and Tetracyclin (TC); Antifungal, Ketoconazole (KCZ)) (see Fig. 5). It is worth that the scavenging products (Cu(II)/Fe(III)-ABIIIL complexes) are more efficient than the native scavenger (ABIIIL) in suppressing the growth of all tested microbial species; with efficacies higher than the clinical drugs, as revealed from the values of ZOI (Fig. 5). Notably, *S. aureus* is more susceptible to the biocidal effects of the M-ABIIIL complexes than *E. coli* (Fig. 5, left). Where the MIC values of Cu(II)/Fe(III) complexes against *S. aureus* and *E. coli* are $25.4/18.9\text{ }\mu\text{g/mL}$ and $41.2/23.1\text{ }\mu\text{g/mL}$, respectively (*i.e.* the activity of the scavenging products against G^+ -bacterium is 1.3–1.6 times higher than G^- -bacterium). This, in turn, means Fe-ABIIIL is more potent than Cu-ABIIIL in fighting the tested bacterial pathogens; with efficacies higher and clinical drugs, streptomycin ($\text{MIC}_{S. aureus/E. coli} = 50.6/27.5\text{ }\mu\text{g/mL}$) and tetracycline ($\text{MIC}_{S. aureus/E. coli} = 37.5/43.2\text{ }\mu\text{g/mL}$). The promoted antibacterial performance of scavenging products (M-ABIIIL) in comparison to the native scavenger (ABIIIL) could be explained

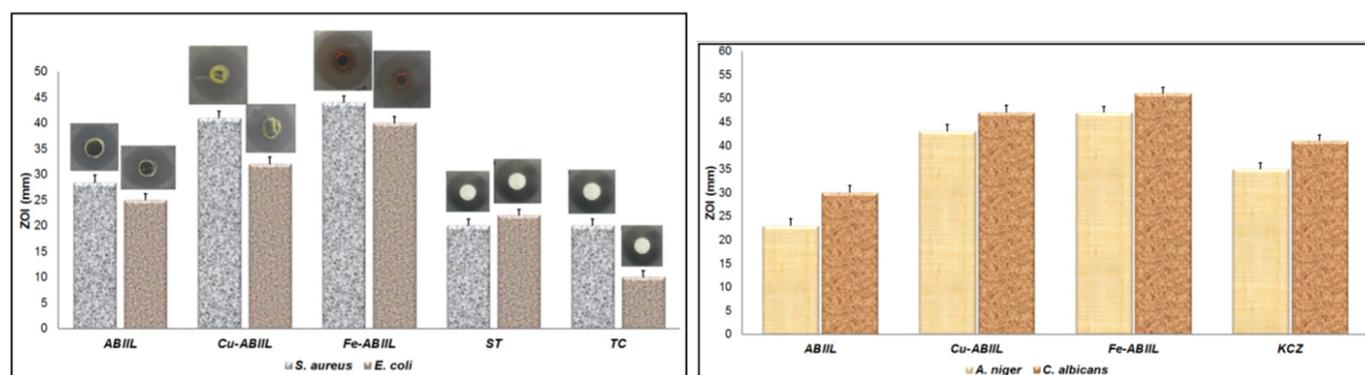


Fig. 5. Graph of the zone of inhibition (ZOI, mm) for the native scavenger (ABIIIL) and scavenging products (Cu-/Fe-ABIIIL complexes) against different medically relevant microbial pathogens. Insets are the micrographs for ZOI of tested compounds against bacterial strains.

based on the chelation theory [52] and Overtone's concept [53]. Generally, the chelating ligands can form highly stable five-/six-membered metallocyclic rings (chelate rings) upon binding to the metal ions. This metallocyclic ring offer template for the delocalization of p-electrons of metal ion, therefore, reduces its polarity and increases the lipophilicity of the whole complex to a larger extent. This increased lipophilicity promotes the penetration of the metal complex into the phospholipid membrane of the microbial cell, causing either collapse of the permeability barrier [54] or inducing malfunction in metal-binding sites of enzymes of this cell [55]. Meanwhile, as the size and/or the number of metallocyclic rings increases; the polarity of metal ion decreases and the lipophilic character of complex increases, which in turn enhances the interaction between complex and the lipid layer in the outer membrane of the microbial cell [56].

As shown in Fig. 6, ABIIL can form two highly stable six-membered metallocyclic rings which significantly diminishes the polarity of Cu (II) and Fe(III) ions and greatly elevates the lipophilic character of their respective complexes, increasing the mutual interactions between complexes and the outer lipid layer of the microbial cell wall to a larger extent. This may lead to a breakdown of the permeability barrier of the cell, resulting in malfunction of normal biological processes of microbial cells. Several structural features can be proposed as the potential reasons for the preferable antimicrobial activity of Fe-ABIIL complex as compared to Cu(II) complex such as; (i) The octahedral geometry and charge distribution around the Fe-ABIIL complex are more compatible with those around pores of the bacterial cell-wall, making it more able to penetrate this wall and induce more toxic reaction within the pores [57]. (ii) Releasing of Fe(III) ions from the Fe-ABIIL complex inside the microbial cell which can highly interact with DNA, more than Cu(II) ions, disordering its functions and replication [58,59]. (iii) Apart from this, the mode of action of the Fe-ABIIL complex may also invoke H-bonding through additional H-bond donor/acceptor (HBD/ HBA) sites (H_2O , ONO_2), absent in the Cu-ABIIL complex, with the active centers of biomolecules in the microbial cell, resulting in dysfunction in their biological processes [60].

As shown in Fig. 5 (right), the M- ABIIL complexes displayed slightly higher fungicidal activity than their bactericidal efficacy, as revealed from the values of their respective inhibition zones against *A. flavus*

and *C. albicans*. Plus, the MIC values ($\mu\text{g}/\text{mL}$) for Fe-ABIIL against microbial species were found to be: 18.9, *S. aureus*; 23.1, *E. coli*; 19.5, *A. flavus*; and 16.8, *C. albicans*.

3.7.2. Cytotoxicity study

The cytotoxic impacts of scavenging products (Cu-ABIIL and Fe-ABIIL) toward two human cell lines (Colon carcinoma cells, HTC116; Normal cells, HeLa) were in vitro assessed as compared to a clinical anticancer drug (Vinblastine, VB) using MTT assay. The cell viability ratio (Fig. 7) and the half-maximal inhibitory concentration (IC_{50}) were used as cytotoxicity indices. Generally, the scavenging products are more cytotoxic than the clinical drug (VB) toward cancer cell lines (HTC116), whereas they are less cytotoxic to the normal human cells (HeLa), as shown in Fig. 7. Just to name a few, remediation of HTC116 and HeLa cell lines with Fe-ABIIL complex ($50.0 \mu\text{g}/\text{mL}$) can reduce their viabilities ratio of HTC116 and HeLa cells by 91.97% and 37.54%, respectively. Whereas, if such treatments were performed using VB, the reduction in cell viability ratios was 87.74% and 51.39%, for HTC116 and HeLa cells respectively. Thus, Fe-ABIIL complex is more active and better safe as anti-colon cancer agent (IC_{50} values: $3.40 \pm 0.53 \mu\text{g}/\text{mL}$ toward HCT116; $85.66 \pm 1.44 \mu\text{g}/\text{mL}$ against HeLa) than the clinical anticancer drug (VB) (IC_{50} values: $3.79 \pm 1.01 \mu\text{g}/\text{mL}$ and $57.12 \pm 1.95 \mu\text{g}/\text{mL}$ for HCT116 and HeLa cell lines, respectively).

Again, Fe-ABIIL complex ($IC_{50} = 3.40 \pm 0.53 \mu\text{g}/\text{mL}$) exhibit better anticancer activity than Cu(II) complex against HCT116 cells ($IC_{50} = 3.58 \pm 0.77 \mu\text{g}/\text{mL}$), and this enhanced cytotoxicity could be ascribed to the same reasons (complex geometry, Fe(III)-DNA interactions, additional HBA and HBD sites) of the promoted antimicrobial activity of Fe-ABIIL complex in comparison to Cu-ABIIL (see Antimicrobial Study section).

4. Conclusion

Aminoethyl-butylimidazolium hexafluorophosphate and salicylaldehyde-butylimidazolium hexafluorophosphate were used as key starting materials for fabrication of a new task-specific ionic liquid (ABIIL) based on bis-(butylimidazolium hexafluorophosphate) supported an azome thine group. The microanalytical and spectral results confirm the

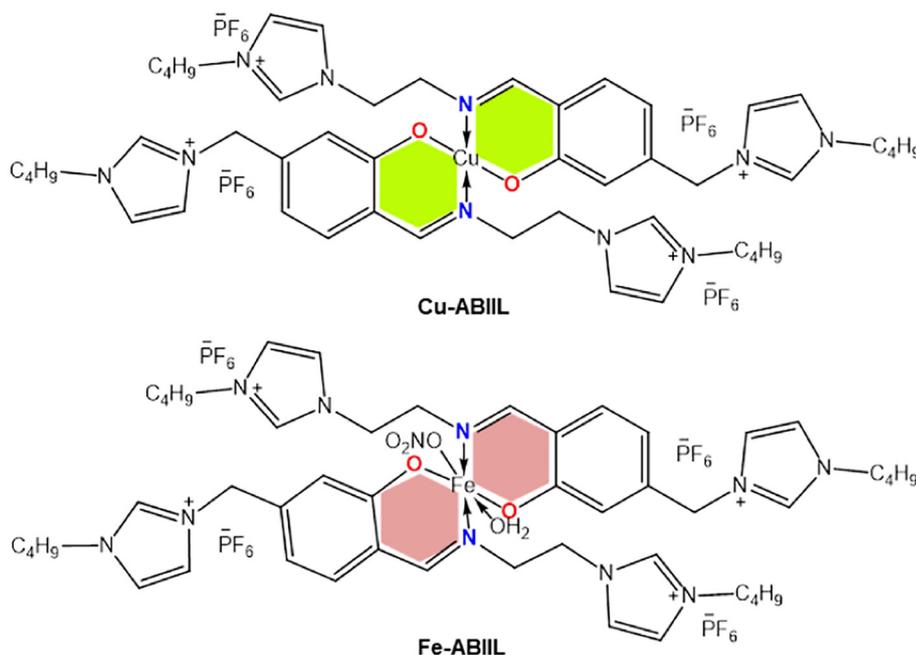


Fig. 6. The proposed structural formula of the scavenging products, $[Cu(II)(ABIIL)_2]$ (Cu-ABIIL) and $[Fe(II)(ABIIL)_2(ONO_2)_2H_2O]$ (Fe-ABIIL), showing the differences in their structural features and additional H-bonding active sites in Fe-ABIIL (HBD/HBA, H_2O ; HBA, ONO_2).

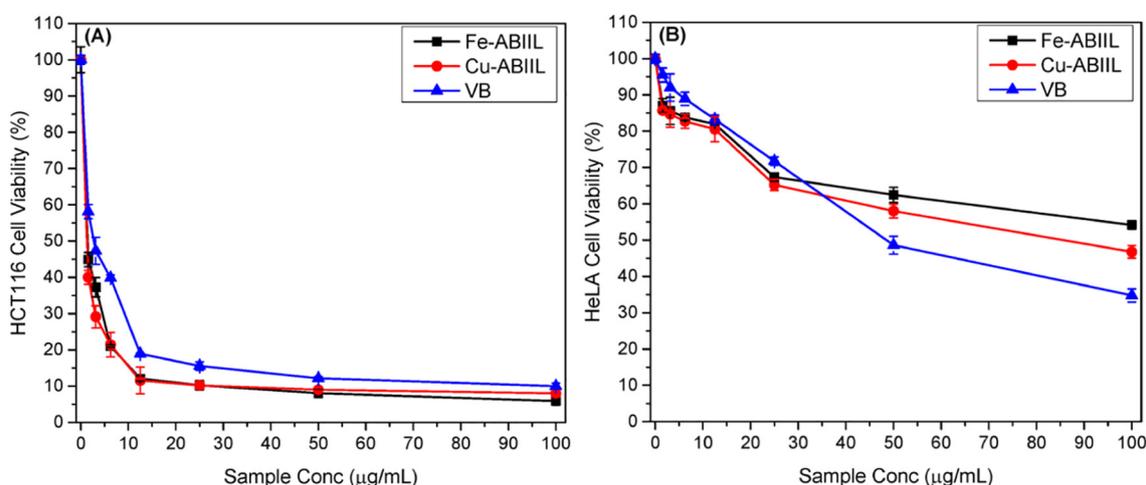


Fig. 7. Cell viability of two human cell lines (A) Colon carcinoma cells (HTC116); (B) Normal cells (HeLa) after 24 h of treatment with serial concentrations of scavenging products (Cu-ABIIL and Fe-ABIIL) and a clinical anticancer drug (Vinblastine, VB).

successful formation of ABIIL. The resultant material (ABIIL) exhibited excellent solubility in aqueous-ethanolic media, which was useful for extracting of heavy metal ions (HMIs) aqueous-ethanol. The new scavenger (ABIIL) displayed excellent scavenging efficiency for tested HMIs, as it can uptake up to 98.9% and 96.7% of Fe(III) and Cu(II) ions, respectively, from aqueous-ethanolic effluents. Besides, it could be easily regenerated and reused without a significant loss of its efficiency even after six consecutive scavenging-regeneration cycles. The physico-chemical features, FTIR, and UV-vis spectral results of the scavenging products indicated that the removal process occurs by the chelation of the anionic form of ABIIL with HMIs. Where the pH-potentiometric studies revealed that ABIIL behaves as a monobasic bidentate (N,O) ligand. The new chelating ionic liquid (ABIIL) underwent a color-change from pale yellow to a yellowish-green and reddish-brown when bound Cu(II) and Fe(III) ions, respectively, to form scavenging products (Cu-/Fe-ABIIL). The pharmacological assessment for the scavenging outputs (M-ABIIL) showed that they have excellent and broad-spectrum antimicrobial activity, with slightly better effectiveness as fungicides than act as bactericides. For instance, the MIC values (μg/mL) for Fe-ABIIL against microbial species were found to be: 16.8, *C. albicans* (fungi) < 18.9, *S. aureus* (G^+ bacterium) < 19.5, *A. flavus* (fungi) < 23.1, *E. coli* (G^- bacterium). Additionally, these complexes possessed more cytotoxic effects than the clinical drug (Vinblastine, VB) toward colon cancer cells (HTC116), whereas they are less cytotoxic than VB to normal human cells (HeLa). Thus, ABIIL has great potential not only for removing toxic Cu(II) and Fe(III) ions from aqueous environments, but also to convert them into promising safe pharmacological nominates for chemotherapeutic applications. Therefore, ABIIL may offer an ideal scavenger for the remediation cupric- and ferric-polluted wastewater produced from many industrial processes, before discharging into the aquatic environment, and consequently averted the environment and human the serious impacts of Cu(II) and Fe(III) ions. Based on these promising findings, still, work on the remaining issues for these ionic liquid-based complexes is continuing and will be presented in future reports.

Author statement

W. N. El-Sayed: Conceptualization, Methodology, Data curation, Validation, Visualization, Writing - Original Draft, Writing - Review & Editing. **J. Alkabli:** Methodology, Conceptualization, Data curation, Validation, Software, Writing - Original Draft. **Khalid Althumayri:** Conceptualization, Methodology, Data curation, Validation, Software, Writing - Original Draft. **Reda F. M. Elshaarawy:** Conceptualization, Methodology, Data curation, Validation, Visualization, Writing - Original Draft, Writing - Review & Editing. **Lamia A. Ismail:** Conceptualization,

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.molliq.2020.114525>.

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