Computer-guided design of a Cu(II) receptor and sensor

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The computer program CAVEAT was used to design a linker structure to connect four imidazole 4-carboxamide groups in a distorted square planar arrangement optimal for binding Cu(II) ions. The resulting macrocyclic structure was prepared by the synthesis of a protected monomer followed by amide coupling reactions to form the cyclic tetramer. Binding to Cu(II) ion was evaluated using the inherent fluorescence of the receptor and its quenching upon binding to Cu(II) ion. Much higher concentrations of Zn(II), Co(II), and Ni(II) ions had little or no effect on the fluorescence and did not significantly compete with the fluorescence response to Cu(II) ions.

Introduction

The design and synthesis of specific ligands and receptors for metal ions is a field of longstanding and continuing interest. This work is of importance for a variety of reasons including the development of sensors for the detection and quantitation of metal ions in biological systems and environmental samples.¹⁻⁴ Success with a Ca²⁺ sensor and its application to studies of cell calcium demonstrates the value of sensors for biologically important metal ions.⁵ Metal receptors have also been designed as models or mimics for metalloproteins.⁶ Metal ion receptors have been based on a variety of structural motifs including synthetic acyclic and macrocyclic polydentate ligands as well as synthetic peptides.⁷⁻¹² The Cu²⁺ ion is somewhat unique in its preference for distorted square planar coordination and this geometry is observed in a number of interesting Cu²⁺ binding proteins.^{13,14} This preference offers the potential for development of selective receptors for Cu²⁺ based on ligand groups organized in a distorted square planar arrangement.¹⁵ A number of Cu²⁺ sensors have been recently developed based on both new and old Cu²⁺-binding motifs.10,11,16-18

Recent structural and computational analysis have illustrated that most complexes between metal ions and multidentate ligands are far from optimal when one considers all of the bond angles and dihedral angles, in addition to the more obvious issues of coordination geometry and bond lengths about the metal ion.^{15,19} While metal-binding proteins might be expected to achieve the ideal metal ligand structure, if this has been the direction of evolutionary pressure, more sophisticated design approaches are clearly needed for the design of more optimal man-made multidentate ligands that approximate the ideal metal coordination environments. Recently computer-based design approaches to this objective have begun to be pursued.²⁰ We have recently begun a research program in the application of the computer program CAVEAT to the design of receptors for molecule and ion recognition.²¹⁻²³ CAVEAT searches electronic databases of 3-dimensional chemical structures to identify structures having bonds matching a defined pair of vectors.^{24,25} Reported here is the computer-assisted design of a Cu^{2+} receptor, using CAVEAT to identify a linker structure to connect individual ligand groups in the orientation that matches a computational model of the complex with unlinked ligands. Also reported is the synthesis of the receptor based on this design as well as initial studies of Cu^{2+} affinity and selectivity and the utility of this receptor as a fluorescence-based Cu^{2+} sensor.

Results and discussion

Design

The design of the Cu(II) receptor was based on the premise that a distorted square planar arrangement of imidazole ligands could exhibit selectivity for Cu²⁺ over other metal ions, and that CAVEAT could be used to design a connector moiety to organize the imidazole groups into such an arrangement. The design process is illustrated in Fig. 1. The complex of Cu²⁺ with four modified imidazole amide moieties 1, analogous to type 2 Cu(II) centers in proteins¹³ was optimized using PM3 calculations. The initial distorted square planar structure was not precisely symmetrical, with the near planar N-Cu-N angles ranging from 94.45-95.38°. The structure was re-optimized in PM3 with the "in plane" N-Cu-N angles constrained at 95°. The resulting highly symmetrical structure 2 had Cu-N bond lengths of 1.93 Å, in fair agreement with the average distance of 1.98 Å observed in Cu-imidazole complexes.²⁶ The symmetry assures that one appropriately designed linker structure can be used for linking each consecutive imidazole pair in a tetrameric structure. To design the linker structure, the amide N-methyl and imidazole N-methyl bonds of adjacent ligands of 2 (represented by the bonds in bold in the upper right portion of structure 1) were used to define the vectors for CAVEAT. A CAVEAT search was conducted of the TRIAD and ILLIAD databases, both being collections of computergenerated structures developed specifically for use with CAVEAT.²⁴ Several possible structures to serve as linkers between ligand moieties were thereby identified. Of these, structure 3 from ILLIAD was chosen for development, in which the $C-H_a$ and $C-H_b$ bonds match the vectors defined by

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Fig. 1 Structures in the design of the macrocyclic Cu(II) receptor 4.

the imidazole–methyl and amide–methyl C–N bonds, respectively. Incorporation of this linker structure between consecutive monomeric groups in the copper complex 1 gives the macrocyclic structure 4. Computational modeling of the Cu^{2+} complex of 4 further supported this design, including the absence of significant strain introduced at the points of connection between linker and ligand. The receptor has a fair degree of flexibility, but modeling indicates it is sufficiently constrained that tetrahedral coordination to a metal ion would impart significant strain relative to the designed distorted square planar coordination.

Synthesis

The preparation of the macrocyclic tetramer 4 began with synthesis of a protected monomer unit. The initial steps in the synthesis are shown in Scheme 1. The thioether 7 was prepared by copper-mediated coupling of 3-mercaptobenzyl alcohol 5²⁷ with 3-bromoiodobenzene 6^{28} Conversion of the benzylic alcohol to the bromide via the mesylate followed by reaction with methyl-4-imidazolecarboxylate 8 and base formed the desired intermediate 9 along with the isomer 10 resulting from alkylation of the wrong imidazole nitrogen. The isomers were readily separated by column chromatography. The assignment of isomers was confirmed by synthesis of 9 by N-alklyation of a glycine ester with the mesylate derived from 7 followed by formation of the imidazole ring using Gold's salt, a known route for the synthesis of imidazole derivatives.²⁹ However, this reaction gave a low yield and was difficult to reproduce so the synthetic route of Scheme 1 was preferred. The remainder of the synthesis of the protected monomer is shown in Scheme 2. *t*-Boc protection of *p*-iodo-*N*-methylaniline 11^{30} followed by conversion of the iodide to the boronic ester by the Miyaura



method formed 12.³¹ Suzuki coupling of 9 with 12 formed the protected monomer unit $13.^{32,33}$

The monomer 13 was converted to the cyclic tetramer 4 as shown in Scheme 3. Separate samples of 13 were deprotected to form the free carboxylic acid 14 and the free amine 15. 14 was converted to the acid chloride followed by reaction with 15 to form the protected dimer 16.^{34,35} Separate samples of 16 were deprotected to the free acid 18 and the free amine 19 as with the monomer 13. The dimer units were again coupled via the acid chloride to form the protected tetramer 17. The amine was deprotected with HCl to avoid introduction of trifluoroacetic acid, which was difficult to separate from the deprotected tetramer and when present caused problems in the coupling step. The ester was hydrolyzed with sodium hydroxide as in the previous deprotections. Attempts to form the cyclic tetramer via the acid chloride and using several other coupling agents were unsuccessful. Cyclization was successfully accomplished using dichlorotriphenylphosphorane in dilute (0.4 mM) solution at reflux for three days to form the desired macrocycle 4. Formation of the macrocycle resulted in a marked simplification of the ¹H NMR spectrum as expected for this highly symmetrical product. Most notably, the four closely spaced singlets between 3.26 and 3.33 for the four *N*-methyl groups of the protected linear tetramer **17** collapsed



to a single peak at 3.26 integrating to 12 hydrogens in 4. Another interesting observation was the shifting of H-4 of the imidazole ring to about 6.4 ppm in the imidazole 5-carbox-amide group, apparently due to substantial loss of conjugation of the amide carbonyl with the imidazole ring due to twisting caused by steric effects of the tertiary amide. A slightly smaller shift was reported though not discussed for a morpholine amide of imidazole-5-carboxylic acid.³⁶ As predicted from this analysis 13, 16, 17, and 4 exhibited the diagnostic 0, 1, 3, and 4 upfield shifted imidazole protons respectively, further supporting the structure of 4.

Binding studies

The macrocycle **4** was studied as a receptor for Cu^{2+} and other metal ions using fluorescence. The integral phenyl biphenyl thioether moiety of the receptor provided a good fluorophore for observation of $Cu(\pi)$ -induced fluorescence quenching. **4** is insoluble in water, so these experiments were conducted in 1 : 1 methanol : tetrahydrofuran. **4** exhibited a maximum absorbance at 290 nm and an emission maximum at 357 nm, which were used as the excitation and emission wavelengths in binding experiments. Fig. 2 shows the fluorescence emission spectra for **4** with increasing concentration of $CuCl_2$. As expected, the paramagnetic $Cu(\pi)$ ion substantially quenched the fluorescence of the receptor, with slightly more



Fig. 2 Decreasing fluorescence of 4 (initial conc. $32.6 \ \mu\text{M}$) with increasing [CuCl₂] (0, 7.7, 15.3, 23.7, 32.1, 40.4, 48.7, 56.9, 65.0, and 86.9 $\ \mu\text{M}$).

than 50% quenching observed at saturation. A plot of fluorescence intensity vs. CuCl₂ concentration gave a dissociation constant for the Cu²⁺-4 complex of 3.2×10^{-6} M. This binding constant in organic solvent is influenced by the counter ion, as the equivalent titration with copper(II) triflate gave a linear decrease in fluorescence with increasing copper salt until one equivalent was reached after which the fluorescence was unchanged, with insufficient curvature in the fluorescence intensity vs. [Cu²⁺] plot to accurately deduce a binding constant. The titration was also performed with Zn(II) triflate. Ni(II) chloride and Co(II) chloride. Zn²⁺ and Ni²⁺ showed no quenching or other change in the fluorescence spectrum at concentrations up to 2.2×10^{-4} and 4.5×10^{-4} M, respectively, while Co²⁺ gave about 5% quenching at a concentration of 2.8×10^{-4} M. As binding of these metal ions could not be directly evaluated using fluorescence, selectivity for Cu(II) ions was assessed by competition. Stepwise addition of Cu(II) chloride to each of the solutions of 4 containing Ni^{2+} , Co^{2+} , or Zn^{2+} resulted in a fluorescence quenching curve that was indistinguishable from the titrations with no other metal salt present, with significant quenching observed even at the first concentration of 7.7 \times 10⁻⁶ M Cu²⁺. This indicates that Ni^{2+} , Co^{2+} , and Zn^{2+} ions do not compete significantly with binding of Cu²⁺ and that the selectivity for Cu²⁺ is at least 100-fold vs. each of the other three metals. The selectivity vs. Ni²⁺ is noteworthy as Ni²⁺ exhibits similar or even greater affinity than Cu²⁺ toward some simple ligands such as bipyridine, and selectivity vs. Ni²⁺ has been an issue with some Cu²⁺ sensors.^{10,37} The aqueous insolubility of **4** limits the more complete affinity and selectivity analysis of this initial design.

Conclusion

A novel computer-guided approach was demonstrated for designing a multidentate ligand for optimal positioning of individual ligand groups for metal ion binding. This approach with incorporation of appropriate solubility and fluorescence signaling properties may be valuable as metal ion sensors. This work further demonstrates the broad utility of the computer program CAVEAT in molecular design. Experimental General

PM3 calculations were performed using PC Spartan Pro from Wavefunction, Inc. Chemicals were obtained from commercial suppliers and used without further purification. Column chromatography was performed on silica gel 60 (40–63 μ m). NMR spectra were obtained on a Varian Gemini-2300 instrument.

may be valuable in designing selective metal ion receptors that

Synthesis

[3-(3-Bromo-phenylsulfanyl)-phenyl]-methanol (7). To a solution of $\mathbf{5}^{27}$ (4.21 g, 0.030 mol) and sodium ethoxide (0.090 mol) in ethanol (45 mL) was added copper powder (0.2 g, 3.13 mmol) and 3-iodobromobenzene **6** (3.84 ml, 8.51 g, 0.03 mol). The mixture was stirred at reflux for 36 h under nitrogen. After cooling to room temperature, the solution was filtered and concentrated to dryness. The residue was dissolved in ethyl acetate (100 mL) and washed with water (3 × 30 mL), dried (MgSO₄), and concentrated. The residue was purified by column chromatography (silica gel, hexanes : ethyl acetate 8 : 1 to 4 : 1) to give 7 (5.05 g, 17 mmol, 43%) as a brown solid. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 4.67 (2H, s, CH₂O), 7.14–7.43 (8H, m, aromatic).

.Methanesulfonic acid 3-(3-bromo-phenylsulfanyl)-benzyl ester. To a solution of 7 (6.67 g, 22.6 mmol) and Et₃N (6.3 ml, 4.57 g, 45.2 mmol) in dichloromethane (80 mL) was added dropwise a solution of methanesulfonyl chloride (2.63 ml, 3.89 g, 34.0 mmol) in dichloromethane (20 mL) at 0 °C over 30 min under nitrogen. The mixture was stirred at 0 °C for 3 h and the reaction was terminated by the addition of ice-cold water (30 mL). The organic layer was washed with ice-cold water (2×20 mL), dried (MgSO₄) and concentrated to give the product (8.32 g, 22.3 mmol, 99%) as a yellow solid, which was used directly in the next step. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 2.94 (3H, s, MeSO₂), 5.19 (2H, s, CH₂O), 7.17–7.44 (8H, m, aromatic).

[3-(3-Bromo-phenylsulfanyl)-benzyl]-bromide. To a solution of methanesulfonic acid 3-(3-bromo-phenylsulfanyl)-benzyl ester (1.2 g, 3.22 mmol) in DMF (20 mL) was added sodium bromide (0.40 g, 3.89 mmol). The solution was stirred for 4 h at room temperature under nitrogen. Then the solution was diluted with ethyl acetate (80 mL) and water (40 mL). The organic layer was washed with water (2 × 30 mL), dried (MgSO₄) and concentrated to give the product (1.14 g, 3.18 mmol, 99%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 4.44 (2H, s, CH₂Br) 7.16-7.45 (8H, m, aromatic).

3-[3-(3-Bromo-phenylsulfanyl)-benzyl]-3H-imidazole-4-carboxylic acid methyl ester (9). To a solution of [3-(3-bromophenylsulfanyl)-benzyl]-bromide (1.56 g, 4.36 mmol) in DMF (30 mL) was added methyl-4-imidazolecarboxylate **8** (0.6 g, 4.76 mmol) and K₂CO₃ (3.01 g, 21.8 mmol). The solution was stirred at room temperature overnight under nitrogen. The solution was diluted with ethyl acetate (100 mL), washed with water (3 × 30 mL), dried (MgSO₄), and concentrated. The residue was purified by the column chromatography (silica gel, hexanes : ethyl acetate 4 : 1 to 2 : 3) to give the pure product **9** (0.8 g, 2.0 mmol, 46%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 3.80 (3H, s, OMe), 5.49 (2H, s, CH₂), 7.05–7.40 (8H, m, aromatic), 7.66 (1H, s, imidazole), 7.76 (1H, s, imidazole). $\delta_{\rm C}$ (75.6 MHz; CDCl₃; Me₄Si) 49.34, 51.41, 122.19, 122.89, 126.24, 129.01, 129.75, 129.82, 130.04, 130.88, 132.91, 135.39, 137.64, 137.73, 137.99, 142.06, 160.35.

(4-Iodophenyl)-methyl-carbamic acid *tert*-butyl ester. To a solution of (4-iodophenyl)-methylamine 11^{25} (0.99 g, 4.25 mmol) in dichloromethane (25 mL) was added triethylamine (1.2 ml, 0.86 g, 8.5 mmol), DMAP (0.052 g, 0.43 mmol) and di*tert*-butyl dicarbonate (1.40 g, 6.4 mmol). The solution was stirred overnight at reflux under nitrogen. The solution was diluted with dichloromethane (40 mL), washed with aq. HCl (1 N, 2 × 20 mL), dried (MgSO₄) and concentrated. The residue was purified by column chromatography on silica gel (hexanes : ethyl acetate 8 : 1) to give (4-iodophenyl)-methyl-carbamic acid *tert*-butyl ester (1.18 g, 3.54 mmol, 83%) as a dark oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.44 (9H, s, OCMe₃), 3.23 (3H, s, NMe), 7.00 (2H, d, *J* 8.7, H2,6), 7.62 (2H, d, *J* 8.7, H3,5). $\delta_{\rm C}$ (75.6 MHz; CDCl₃; Me₄Si) 28.19, 36.96, 77.42, 89.34, 127.19, 137.44, 143.48, 154.19.

Methyl-[4-(4,4,5,5-tetramethyl-[1,3,2]dioxaborolan-2-yl)phenyll-carbamic acid tert-butyl ester (12). To a mixture of bis(pinacolato)diboron (0.34 g, 1.34 mmol), PdCl₂ (dppf) · CH₂Cl₂ (0.029 g, 35.5 µmol) and potassium acetate (0.35 g, 3.57 mmol) was added a solution of (4-iodophenyl)methyl-carbamic acid tert-butyl ester (0.4 g, 1.2 mmol) in DMSO (6 mL) under nitrogen. The solution was stirred at 80 °C for 3.5 h under nitrogen. After cooling to room temperature, the solution was diluted with ethyl acetate (40 mL), washed with water (4 \times 20 mL), dried (MgSO₄) and concentrated. The resulting residue was purified by column chromatography on silica gel (hexanes : ethyl acetate 8 : 1) to give 12 (0.25 g, 7.51 mmol, 63%) as a yellow solid. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.34 (12H, s, CMe₂), 1.44 (9H, s, OCMe₃), 3.27 (3H, s, NMe), 7.24 (2H, d, J 8.7, H2,6), 7.76 (2H, d, J 8.7, H3.5).

3-{3-[4'-(*tert*-Butoxycarbonyl-methyl-amino)-biphenyl-3-ylsulfanyl]-benzyl}-3*H*-imidazole-4-carboxylic acid methyl ester (13). To a mixture of 12 (0.86 g, 2.58 mmol), PdCl₂ (dppf) \cdot CH₂Cl₂ (0.113 g, mmol) and K₂CO₃ (1.37 g, 6.21 mmol) was added a solution of 9 (0.8 g, 1.98 mmol) in dimethoxyethane (15 mL) under nitrogen. The solution was stirred at 80 °C for 24 h under nitrogen. After cooling to room temperature, the solution was concentrated to dryness. Ethyl acetate (80 mL) was added and the solution was washed with water (2 × 30 mL). The organic layer was dried (MgSO₄) and concentrated. The resulting residue was purified by column chromatography (silica gel, hexanes : ethyl acetate 4 : 1 to 2 : 3) to give 13 (0.66 g, 1.25 mmol, 63%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.47 (9H, s, OCMe₃), 3.28 (3H, s, NMe), 3.76 (3H, s, CO₂Me), 5.46 (2H, s, CH₂), 6.95–7.50 (12H, m, aromatic), 7.60 (1H, s, imidazole), 7.74 (1H, s, imidazole). $\delta_{\rm C}$ (75.6 MHz; CDCl₃; Me₄Si) 24.86, 28.15, 36.86, 83.28, 83.56, 124.13, 134.97, 146.32, 154.26.

3-{3-[4'-(*tert***-Butoxycarbonyl-methyl-amino)-biphenyl-3-yl-sulfanyl]-benzyl}-3***H***-imidazole-4-carboxylic acid 14. To a solution of 13 (0.91 g, 1.72 mmol) in methanol (18 mL) was added a solution of NaOH (0.8 g, 0.02 mol) in water (2 mL). The solution was stirred at room temperature for 24 h. The solution was concentrated to dryness and water (20 mL) was added. Aq. HCl (1 N) was added dropwise until the pH reached 5. The mixture was extracted with dichloromethane (3 × 60 mL). The combined organic layers were dried (MgSO₄) and concentrated to give 14 (0.82 g, 1.60 mmol, 93%) as an orange oil. \delta_{\rm H} (300 MHz; CDCl₃; Me₄Si) 1.47 (9H, s, OCMe₃), 3.27 (3H, s, NMe), 5.50 (2H, s, CH₂), 7.00–7.50 (12H, m, aromatic), 7.74 (1H, s, imidazole), 7.78 (1H, s, imidazole).**

3-[3-(4'-Methylamino-biphenyl-3-ylsulfanyl)-benzyl]-3*H*-imidazole-4-carboxylic acid methyl ester (15). To a solution of 13 (0.92 g, 1.74 mmol) in dichloromethane (6 mL) was added dropwise trifluoroacetic acid (1.25 mL, 0.85 g, 7.46 mmol). The solution was stirred for 2 h at room temperature. The reaction mixture was added dropwise to an aq. saturated NaHCO₃ solution (40 mL). The solution was extracted with dichloromethane (70 mL). The organic layer was washed with saturated aq. NaHCO₃ (2 × 30 mL), and brine (20 mL), dried (MgSO₄) and concentrated to give 15 (0.74 g, 1.72 mmol, 99%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 2.86 (3H, s, NMe), 3.76 (3H, s, CO₂Me), 5.45 (2H, s, CH₂), 6.66 (2H, d, *J* 8.4, aromatic *ortho* to N), 6.90–7.54 (10H, m, aromatic), 7.60 (1H, s, imidazole), 7.74 (1H, s, imidazole).

3-(3-{4'-[(3-{3-[4'-(tert-Butoxycarbonyl-methyl-amino)-biphenyl-3-ylsulfanyl]-benzyl}-3H-imidazole-4-carbonyl)-methylamino]-biphenyl-3-ylsulfanyl}-benzyl)-3H-imidazole-4-carboxylic acid methyl ester (16). To a solution of 14 (0.82 g, 1.60 mmol) in dichloromethane (10 mL) and toluene (10 mL) with 1 drop of dimethylformamide at 0 °C under nitrogen was added dropwise oxalyl chloride (1.2 mL, 0.83 g, 6.5 mmol). The solution was stirred for 1 h at 0 °C under nitrogen and the reaction mixture was concentrated to dryness. Anhydrous dichloromethane (5 mL) was added and the resulting solution was added dropwise to a solution of 15 (0.74 g, 1.72 mmol) in pyridine (5 mL) at 0 °C under nitrogen. The reaction mixture was allowed to warm to room temperature over 0.5 h and was stirred overnight at room temperature under nitrogen. The mixture was then concentrated to dryness. The resulting residue was dissolved in dichloromethane (60 mL), washed with aqueous saturated NaHCO₃ (2×20 mL), brine (20 mL), dried (MgSO₄), and concentrated. The resulting residue was purified by column chromatography on silica gel (hexanes : ethyl acetate 1:4 to ethyl acetate : methanol 18:1) to give 16 (0.94 g, 1.0 mmol, 63%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.45 (9H, s, OCMe₃), 3.25 (3H, s, NMe), 3.31 (3H, s, NMe), 3.76 (3H, s, CO₂Me), 5.45 (2H, s, CH₂), 5.49 (2H, s, CH₂), 6.36 (1H, s, imidazole), 6.87 (2H, d, J 8.1, aromatic ortho to amide N), 6.90-7.74 (25H, m, aromatic, 3H from imidazole).

3-(3-{4'-[(3-{3-[4'-(*tert*-Butoxycarbonyl-methyl-amino)-biphenyl-3-ylsulfanyl]-benzyl}-3*H*-imidazole-4-carbonyl)-methylamino]-biphenyl-3-ylsulfanyl}-benzyl)-3*H*-imidazole-4-carboxylic acid (18). The procedure for the synthesis of 14 was followed to give 18 (94%). $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.47 (9H, s, OCMe₃), 3.26 (3H, s, NMe), 3.34 (3H, s, NMe), 5.47 (2H, s, CH₂), 5.52 (2H, s, CH₂), 6.13 (1H, s, imidazole), 6.70–7.75 (27H, m, aromatic, 3H from imidazole).

3-{3-[4'-(Methyl-{3-[3-(4'-methylamino-biphenyl-3-ylsulfanyl)-benzyl]-3*H*-imidazole-4-carbonyl}-amino)-biphenyl-3-ylsulfanyl]-benzyl}-3*H*-imidazole-4-carboxylic acid methyl ester (19). The procedure for the synthesis of 15 was followed to give 19 (99%). $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 2.82 (3H, s, amine NMe), 3.32 (3H, s, amide NMe), 3.77 (3H, s, CO₂Me), 5.45 (2H, s, CH₂), 5.49 (2H, s, CH₂), 6.39 (1H, s, imidazole), 6.60 (2H, d, *J* 8.7, aromatic *ortho* to amine N), 6.87 (2H, d, *J* 8.7, aromatic *ortho* to amide N), 6.90–7.80 (23H, m, aromatic, 3H from imidazole).

Linear tetramer (17). The procedure for the synthesis of 16 was followed, but the product was purified by column chromatography on silica gel (ethyl acetate : methanol : Et₃N, 19 : 1 : 1) to give 17 (60%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 1.46 (9H, s, OCMe₃), 3.24 (3H, s, NMe), 3.28 (3H, s, NMe), 3.29 (3H, s, NMe), 3.31 (3H, s, NMe), 3.77 (3H, s, CO₂Me). 5.44 (2H, s, CH₂), 5.49 (6H, s, 3 unresolved CH₂), 6.36 (2H, s, unresolved imidazole H), 6.38 (1H, s, imidazole) 6.86-6.91 (6H, m, aromatic ortho to amide N), 6.94-7.74 (47H, m, aromatic, 7H from imidazole). $\delta_{\rm C}$ (75.6 MHz; CDCl₃; Me₄Si) 28.24, 37.08, 37.81, 37.84, 37.86, 45.27, 49.50, 49.72, 51.38, 80.38, 122.24, 125.00, 125.46, 125.65, 126.05, 126.19, 126.23, 126.28, 127.02, 127.27, 128.12, 128.84, 129.35, 129.39, 129.56, 129.62, 129.68, 129.72, 129.86, 129.93, 130.08, 130.12, 130.17, 130.20, 130.51, 130.54, 130.72, 134.22, 135.27, 135.36, 135.42, 135.46, 136.77, 136.80, 136.83, 136.92, 137.35, 137.81, 137.86, 139.44, 139.49, 139.60, 140.82, 140.86, 141.65, 142.05, 143.33, 143.49, 154.49, 160.35, 160.65.

Cyclic tetramer (4). To a solution of 17 (0.124 g, 72.0 µmol) in dioxane (2 mL) was added dropwise a solution of HCl in dioxane (4 M, 2 mL). The solution was stirred overnight at room temperature. The reaction was quenched by the addition of triethylamine (2 mL). The resulting mixture was concentrated to dryness and then dissolved in dichloromethane (40 mL). The solution was washed with saturated NaHCO₃ (2 \times 20 mL), dried (MgSO₄) and concentrated to give the free amine of 17 (0.117 g, 72.1 μ mol, 99%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 2.83 (3H, s, amine NMe), 3.29-3.33 (9H, 3s, amide NMe), 3.77 (3H, s, OMe), 5.47 (8H, 4s, CH₂), 6.36 (3H, 3s, imidazole), 6.60 (2H, d, J 8.7, aromatic ortho to NHMe), 6.82–6.91 (6H, m, aromatic ortho to amide NMe), 6.92–7.80 (45H, m, aromatic). The amine (0.117 g, 72.1 μmol) was dissolved in methanol (5 mL) and THF (8 mL), and a solution of NaOH (52.6 mg, 1.32 mmol) in water (1 mL) was added. The solution was stirred for 2 days at room temperature. The mixture was concentrated to dryness and dissolved in water (15 mL). An aq. solution of HCl (1 N) was added dropwise until the pH reached 6. The mixture was extracted with dichloromethane (3 \times 50 mL). The combined organic layers were dried (MgSO₄) and concentrated to give the fully deprotected linear tetramer (0.1 g, 62.2 µmol, 86%) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 2.82 (3H, s, amine NMe), 3.28-3.33 (9H, 3s, amide NMe), 5.44-5.50 (8H, 4s, CH₂), 6.26 (1H, s, imidazole), 6.36 (2H, s, imidazole), 6.60 (2H, d, J 8.1), 6.61-7.80 (51 H, m, aromatic). A portion of this product (0.0475 g, 29.5 µmol) was dissolved in dichloromethane (60 mL) and triphenylphosphine dichloride (0.06 g, 18.0 mmol) was added. The solution was stirred for 3 days at reflux under nitrogen. After cooling to room temperature, the solution was washed with saturated aq. NaHCO₃ (20 mL), dried (MgSO₄) and concentrated. The resulting residue was purified by column chromatography (ethyl acetate : methanol : Et₃N, 18 : 1:1) to give 4 (0.031 g, 19.5 µmol, 66% for the final step) as a brown oil. $\delta_{\rm H}$ (300 MHz; CDCl₃; Me₄Si) 3.26 (12H, s, NMe), 5.47 (8H, s, CH₂), 6.36 (4H, s, imidazole), 6.79 (8H, d, J 8.7, aromatic ortho to N), 7.00–7.60 (44H, m, aromatic). m/z (ES) 1589.46 (MH⁺. C₉₆H₇₇N₁₂O₄S₄ requires 1589.51).

Fluorescence titrations

The fluorescence titrations were performed using an Aminco-Bowman Series2 Spectrometer at 20 °C. The emission spectrum was scanned from 325-390 nm with excitation at 290 nm. Spectra were obtained for the solution of 4 (3.26×10^{-5} M, 3.1 mL) in 1 : 1 methanol : tetrahydrofuran and after two successive additions of 9 μ L and six additions of 10 μ L of a 2.65×10^{-3} M CuCl₂ solution and of 10 µL of a 7.04 × 10⁻³ M CuCl₂ solution, all in 1 : 1 methanol : tetrahydrofuran. For data fitting, the fluorescence intensity at 357 nm was used. NiCl₂ solution (17.4 mM) was added in six 5 µL portions followed by five 10 µL portions to a solution of 4 (3.0 mL, 32.6 μM) in 1 : 1 methanol : tetrahydrofuran. CoCl₂ titrations were performed by addition to a solution of 4 (2.09×10^{-5} M, 2.9 mL) of 1.04×10^{-3} M CoCl₂ solution (6 × 20 µL, 4 × 40 µL) followed by 1.04×10^{-2} M CoCl_2 solution (2 \times 10 $\mu L,$ 2 \times 20 μ L). Zinc triflate titrations were performed by addition to a solution of 4 (2.09 \times 10⁻⁵ M, 3.0 mL) of 8.6 \times 10⁻³ M Zn(OTf)₂ solution (6 \times 5 µL, 3 \times 10 µL, 20 µL).

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References

- 1 (a) A. W. Czarnik, Acc. Chem. Res., 1994, **27**, 302–308; (b) A. W. Czarnik, Chem. Biol., 1995, **2**, 423–428.
- 2 L. Prodi, F. Bolletta, M. Montalti and N. Zaccheroni, *Coord. Chem. Rev.*, 2000, **205**, 59–83.
- 3 K. R. Gee, Z.-L. Zhou, D. TonThat, S. L. Sensi and J. H. Weiss, *Cell Calcium*, 2002, **31**, 245–251.
- 4 A. C. Benniston, A. Harriman, D. J. Lawrie, A. Mayeux, K. Rafferty and O. D. Russell, *Dalton Trans.*, 2003, 4762–4769.

- 5 (a) R. Y. Tsien, in *Fluorescent Chemosensors for Ion and Molecule Recognition*, ed. A. W. Czarnik, American Chemical Society, Washington, DC, 1993, pp. 130–146; (b) R. Y. Tsien, *Chem. Eng. News*, July 18, 1994, **72**, 34–44.
- 6 R. H. Holm and E. I. Solomon, Chem. Rev., 2004, 104, 347-348.
- 7 G. Xue, P. B. Savage, J. S. Bradshaw, X. X. Zhang and R. M. Izatt, *Adv. Supramol. Chem.*, 2000, **7**, 99–137.
- 8 A. Ikeda and S. Shinkai, Chem. Rev., 1997, 97, 1713-1734.
- 9 R. P. Cheng, S. L. Fisher and B. Imperiali, J. Am. Chem. Soc., 1996, 118, 11349–11356.
- 10 A. Torrado, G. K. Walkup and B. Imperiali, J. Am. Chem. Soc., 1998, 120, 609–610.
- 11 S. Bhattacharya and M. Thomas, *Tetrahedron Lett.*, 2000, 41, 10313–10317.
- 12 N. Jotterand, D. A. Pearce and B. Imperiali, J. Org. Chem., 2001, 66, 3224–3228.
- 13 K. A. Koch, M. Marjorette, O. Peña and D. J. Thiele, *Chem. Biol.*, 1997, 4, 549–560.
- 14 L. Rulíšek and Z. Havlas, J. Am. Chem. Soc., 2000, 122, 10428–10439.
- 15 B. P. Hay and R. D. Hancock, *Coord. Chem. Rev.*, 2001, **212**, 61–78.
- 16 R. Kramer, Angew. Chem., Int. Ed., 1998, 37, 772-773.
- 17 M. Royzen, Z. Dai and J. W. Canary, J. Am. Chem. Soc., 2005, 127, 1612–1613.
- 18 Z. Xu, X. Qian and J. Cui, Org. Lett., 2005, 7, 3029-3032.
- 19 P. Comba and W. Schiek, Coord. Chem. Rev., 2003, 238, 21–29.
- 20 B. P. Hay, A. A. Oliferenko, J. Uddin, C. Zhang and T. K. Firman, J. Am. Chem. Soc., 2005, 127, 17043–17053.
- 21 W. Yang, H. He and D. G. Drueckhammer, Angew. Chem., Int. Ed., 2001, 40, 1714–1718.
- 22 H. Huang and D. G. Drueckhammer, Chem. Commun., 2005, 5196–5198.
- 23 H. Huang and D. G. Drueckhammer, *Chem. Commun.*, 2006, 2995–2997.
- 24 G. Lauri and P. A. Bartlett, J. Comput. Aided Mol. Des., 1994, 8, 51–66.
- 25 (a) P. A. Bartlett, G. T. Shea, S. J. Telfer and S. Waterman, in Molecular Recognition: Chemical and Biological Problems, ed. S. M. Roberts, Royal Society of Chemistry, London, 1989, pp. 182–196; (b) P. A. Bartlett, F. A. Etzkorn, T. Guo, G. Lauri, K. Liu, M. Lipton, B. P. Morgan and G. T. Shea, in Chemistry at the Frontiers of Medicine, Proceedings of the Robert A. Welch Foundation Conference on Chemical Research XXXV, The Robert A. Welch Foundation, Houston TX, 1992, pp. 45–68.
- 26 A. G. Orpen, L. Brammer, F. H. Allen, O. Kennard, D. G. Watson and R. Taylor, J. Chem. Soc., Dalton Trans., 1989, S1–S83.
- 27 T. C. Thurber, A. Prince and O. Halpern, J. Heterocycl. Chem., 1982, 19, 961–965.
- 28 H. D. H. Showalter, M. M. Angelo, E. M. Berman, G. D. Kanter, D. F. Ortwine, S. G. Ross-Kesten, A. D. Sercel, W. R. Turner, L. M. Werbel, D. F. Worth, E. F. Elslager, W. R. Leopold and J. L. Shillis, J. Med. Chem., 1988, **31**, 1527–1539.
- 29 R. Kirchlechner, M. Casutt, U. Heywang and M. W. Schwarz, Synthesis, 1994, 247–248.
- 30 D. V. Kosynkin and J. M. Tour, Org. Lett., 2001, 3, 991–992.
- 31 T. Ishiyama, T. M. Murata and N. Miyaura, J. Org. Chem., 1995, 60, 7508–7510.
- 32 N. Miyaura and A. Suzuki, Chem. Rev., 1995, 95, 2457–2483.
- 33 F. Firooznia, C. Gude, K. Chan, N. Marcopulos and Y. Satoh,
- Tetrahedron Lett., 1999, 40, 213–216.
 34 I. Azumaya, T. Okamoto, F. Imabeppu and H. Takayanagi, Tetrahedron, 2003, 59, 2325–2331.
- 35 I. Azumaya, T. Okamoto, F. Imabeppu and H. Takayanagi, *Heterocycles*, 2003, **60**, 1419–1424.
- 36 E. Belgodere, R. Bossio, V. Parrini and R. Pepino, J. Heterocycl. Chem., 1982, 19, 561–566.
- 37 R. M. Smith and A. E. Martell, *Critical Stability Constants, Amines*, Plenum Press, New York, 1975, vol. 2, p. 235.