A Flexible Three-Component Synthesis of Novel δ-Amino Acids Incorporating an Imidazo[1,2-*a*]pyridine Backbone

Ivana Veljkovic, Reinhold Zimmer, Hans-Ulrich Reissig,* Irene Brüdgam, Hans Hartl

Freie Universität Berlin, Institut für Chemie und Biochemie, Takustrasse 3, 14195 Berlin, Germany Fax +49(30)83855367; E-mail: hans.reissig@chemie.fu-berlin.de *Received 3 March 2006; revised 18 April 2006* In Memory of Professor Ivar Ugi

Abstract: Methyl 2-trimethylsiloxycyclopropanecarboxylates 1, 2aminopyridine (2), and isonitriles 3 combine in a one-pot reaction to provide a series of novel δ -amino acids 4 incorporating an imidazo[1,2-*a*]pyridine backbone. Scope and limitations of this new three-component synthesis were investigated. Several reactions of compounds 4 affording suitably protected derivatives such as 5, 6 and 9 were performed to allow couplings with L-alanine derivatives. The expected peptides 11 and 12 were obtained by standard coupling reactions with excellent or moderate yield. Cyanide-catalyzed reactions converted compounds 4e and 4h into tricyclic δ -lactams 10e and 10h in very good yields.

Key words: amino acids, cyclopropanes, pyridines, lactams, peptides

Synthesis as well as structural and biological evaluation of amino acids and peptides derived thereof with modified in particular restricted – conformations are recent topics in bioorganic and medicinal chemistry.¹ Simple, efficient and highly flexible methods to generate new 'unnatural' amino acids are of particular importance, and among other strategies multicomponent reactions have found numerous applications to approach this goal.² In many of these reactions aldehydes are crucial components in the proceeding reaction cascades. In several examples we could demonstrate that methyl 2-siloxycyclopropanecarboxylates are equivalents of functionalized and substituted carbonyl compounds.³ They are prepared from the trimethylsiloxycyclopropane derivatives under very mild conditions (either neutral, with fluoride or slightly acidic or basic media) and thus these cyclopropane derivatives can directly be introduced into multicomponent reactions.^{4,5} Recent reports on the synthesis of imidazo[1,2a]pyridines by an Ugi-type reaction of 2-aminopyridine with aldehydes and isonitriles attracted our attention,⁶ since a related process with methyl 2-siloxycyclopropanecarboxylates as aldehyde equivalents should directly provide compounds A. The expected products are δ -amino acids incorporating an imidazo[1,2-*a*]pyridine backbone, which can be regarded as a conformationally altered dipeptide **B** (Figure 1). In addition, imidazo[1,2-a]pyridine derivatives are of general interest because of their

SYNTHESIS 2006, No. 16, pp 2677–2684 Advanced online publication: 19.07.2006 DOI: 10.1055/s-2006-942506; Art ID: T03306SS © Georg Thieme Verlag Stuttgart · New York

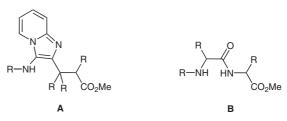


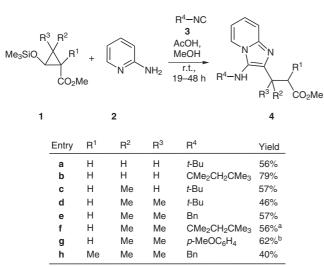
Figure 1 Equivalence of methyl (3-aminoimidazo[1,2-*a*]pyridin-2-yl)propanoates **A** to **B**

broad range of physiological effects (e.g. antibacterial, antiviral, anti-inflammatory, analgesic activities).⁷

In this paper we report on scope and limitations of the synthesis of compounds **A** starting from siloxycyclopropanes and we also reveal first model reactions to couple these novel δ -amino acids with L-alanine derivatives to produce small peptides.

Methyl 2-siloxycyclopropanecarboxylates 1 (1 to 1.1 equiv), 2-aminopyridine (2) (1 equiv), and isonitriles 3 (1 equiv) were stirred in a mixture of acetic acid (2 equiv) and methanol for 19-48 hours at room temperature. After acidic work up (which destroys remaining isonitrile) and chromatographic purification we obtained methyl (3-aminoimidazo[1,2-a]pyridin-2-yl)propanoates 4 in moderate to good yields (Scheme 1). With respect to the cyclopropanes 1, the parent compound (entries **a**, **b**), monomethyl substituted (entry c), dimethyl substituted (entries d, e, f, g), and trimethyl substituted (entry h) derivatives were also suitable aldehyde equivalents. The isonitrile component can bear a tertiary alkyl, a benzyl, or an aryl substituent. Although not examined, other 2-aminopyridine derivatives may also be used. Hence, this one-pot threecomponent reaction leading to δ -amino acid derivatives 4 with an imidazo [1,2-a] pyridine backbone seems to allow a fairly broad substitution pattern. For the synthesis of compound 4g (entry g) the yield was 62% when the mixture was exposed to microwave irradiation,⁸ whereas only 43% yield was obtained under the standard conditions applied for all other entries of Scheme 1.

For heterocycle **4g**, we could also obtain suitable crystals for an X-ray analysis, which not only proved the constitution of the product but also showed that the propanoate side chain is spatially in close proximity to the arylamino group (Figure 2).



^a In addition of 30% of compound 7

^b Under microwave conditions.

Scheme 1 Synthesis of methyl (3-aminoimidazo[1,2-*a*]pyridin-2yl)propanoates 4 by one-pot reaction of siloxycyclopropanes 1, 2aminopyridine 2 and isonitriles 3

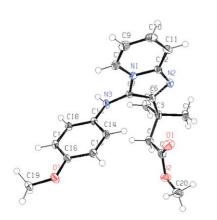
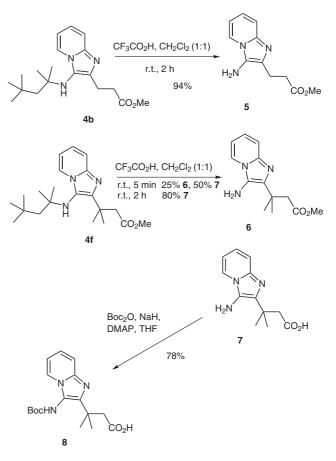


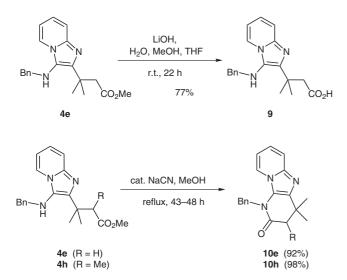
Figure 2 X-ray crystal structure of compound 4g

Since we wanted to prepare small model peptides including δ -amino acids derived from 4, we had to convert these compounds into suitably protected intermediates. Treatment of compound 4b with trifluoroacetic acid for two hours at room temperature furnished the expected amino ester 5 in excellent yield (Scheme 2).⁹ Surprisingly, when identical conditions were applied to the higher substituted analog 4f not only the tetramethylbutyl group was displaced, but in addition the methyl ester was cleaved providing the amino acid 7 in 80% yield. Precursor 4f seems to be particular sensitive to acid since amino acid 7 was isolated as a by-product in 30% yield during synthesis of 4f. We therefore shortened the reaction time of 4f with trifluoroacetic acid to five minutes at room temperature and obtained a mixture of the required amino ester 6 (25%) yield) and of the amino acid 7 (50% yield). The deprotection reactions of 4f thus occur under milder acidic conditions. We speculate that the particular conformation of the ammonium salt derived from **4f** facilitates the cleavage of the methyl ester. According to the X-ray crystal structure of the related dimethyl substituted compound **4g**, the methoxycarbonyl is in close proximity of the amino substituents (Figure 2) which might exhibit a neighboring group effect. No further attempts have been made to selectively deprotect compound **4f**. Employing sodium hydride, DMAP and Boc-anhydride, compound **7** was converted into the N-protected δ -amino acid **8** with reasonable efficacy (Scheme 2). We had to recognize already here that the amino groups of imidazo[1,2-*a*]pyridine derivatives are fairly unreactive under acylation conditions.



Scheme 2 Trifluoroacetic acid promoted deprotections of 4b and 4f leading to amino esters 5, 6 and amino acid 7 and Boc-protection of 7 to δ -amino acid 8

Saponification of the amino ester **4e** proceeded without problem and afforded the expected *N*-benzyl-protected amino acid **9** in 77% yield (Scheme 3). It should be mentioned here that attempts to debenzylate compounds such as **4e** and **4h** were not successful and in general led to decomposition of the heterocycles. However, we discovered a smooth cyclization method to transform δ -amino acid derivatives into tricyclic δ -lactam derivatives. By cyanide catalysis in refluxing methanol¹⁰ (most likely via the corresponding acyl cyanides¹¹) precursors **4e** and **4h** delivered the dipyrido[1,2-*a*;3',2'-*d*]imidazol-2-ones **10e** and **10h** in excellent yields (Scheme 3).

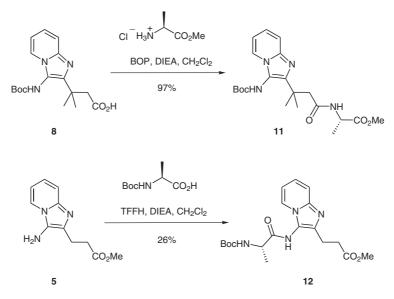


Scheme 3 Preparation of δ -amino acid **9** and of dipyrido[1,2-a;3',2'-d]imidazol-2-ones **10e** and **10h**

With amino acid derivatives 8 and 5 we tried model reactions to generate small peptides. The peptide coupling of acid $\mathbf{8}$ with L-alanine methyl ester was conventionally performed with benzotriazol-1-yloxytris(dimethylamino)phosphonium hexafluorophosphate¹² (BOP) as activating agent and furnished almost quantitatively compound 11, which may be regarded as a tripeptide equivalent (Scheme 4). On the other hand, reaction at the amino group of ester 5 with N-Boc-protected L-alanine required the very powerful coupling reagent tetramethylfluoroformamidinium hexafluorophosphate¹³ (TFFH) and still only 26% of the desired product 12 was obtained. It should be mentioned that experiments employing the dimethyl substituted compound 6 did not result in peptide formation. This may again be the effect of the conformations of these compounds (see discussion above and Figure 2). Further investigations are required to overcome this problem and take profit of the apparently restricted conformation of this particular skeleton.

In conclusion, we have demonstrated that the novel threecomponent synthesis of siloxycyclopropanes 1 with 2aminopyridine (2) and isonitriles 3 efficiently provided new δ -amino acid derivatives 4 incorporating a 2-imidazo[1,2-a] pyridine backbone. This transformation again proved the equivalence of siloxycyclopropanes 1 to substituted and functionalized aldehydes, which are difficult to prepare by alternative methods. In various model reactions the heterocyclic amino acid derivatives 4 were suitably protected and coupled with L-alanine derivatives to afford the expected peptides 11 or 12. Furthermore, a mild method was found to convert compounds 4 into the tricyclic δ -lactams 10. Future studies should be devoted to the synthesis of longer peptide chains - optionally under multiple inclusion of derivative 4 - and to the elucidation of the influence of the 2-imidazo[1,2-a]pyridine backbone on the conformation of the resulting constructs.

All reactions were performed under argon in flame-dried flasks, and the components were added by means of syringes. All solvents were dried by standard methods. TLC was carried out on commercial Polygram Sil G/UV₂₅₄ or Polygram Alox N/UV₂₅₄ (Macherey & Nagel). Column chromatography was performed using 70-230 mesh silica gel (Merck) or neutral aluminum oxide (activity grade-III; Fluka or Merck). Nucleosil 50-5 (Macherey & Nagel) was used for HPLC. Unless stated otherwise, ¹H NMR and ¹³C NMR spectra were determined on Bruker (AM-270, AC-500) or JEOL (Eclipse 500) instruments. The chemical shifts are related to TMS or to the CDCl₃ signal ($\delta_{\rm H}$ = 7.26, $\delta_{\rm C}$ = 77.0). Higher order NMR spectra were approximately interpreted as first-order spectra, if possible. Missing signals could not be unambiguously identified due to low intensity. IR spectra were measured on Nicolet spectrometer FTIR 5 SXC. MS and HRMS analyses were performed on Finnigan MAT 711 (EI, 80 eV, 8 kV), MAT CH7A (EI, 80 eV, 3 kV) and CH5DF (FAB, 80 eV, 3 kV) instruments. Elemental analyses were performed on a Perkin Elmer CHN analyzer 2400. Melting points were determined with a Büchi MP 510 apparatus and are uncorrected.



Scheme 4 Couplings of L-alanine derivatives with amino acid 8 to peptide 11 and with amino ester 5 to peptide 12

Boiling points of compounds obtained in small-scale experiments refer to the temperature in a Büchi Kugelrohrofen. Starting materials 1^{14} and *p*-methoxyphenylisocyanide¹⁵ were prepared using known procedures. All other chemicals were commercially available and were used as received.

Ugi-Type Reaction of Methyl Siloxycyclopropanecarboxylates 1 and Isonitriles 3 with 2-Aminopyridine (2), General Procedure

Methyl siloxycyclopropanecarboxylate 1 (1–1.1 equiv), 2-aminopyridine (2; 1 equiv) and the corresponding isonitrile 3 (1 equiv) were dissolved in anhyd MeOH (2.5–3 mL/mmol of 2) under argon. After addition of AcOH (2 equiv), the mixture was stirred at r.t. for the time indicated in the individual experiment. Then 1 M HCl solution was added to adjust pH 1, the mixture was stirred for 30 min to destroy unreacted isocyanide 3 and then evaporated to dryness. The residue was taken up in sat. aq NaHCO₃ solution (2 mL/mmol of 2) and extracted with EtOAc (3×10 mL/mmol of 2). The combined organic phases were dried (Na₂SO₄) and the solvent was removed under reduced pressure. The resulting crude product was purified by column chromatography on silica gel (EtOAc– hexane, 2:1) and in some cases followed by HPLC (20% *i*-PrOH– hexane).

Methyl 3-(3-*tert*-Butylaminoimidazo[1,2-*a*]pyridin-2-yl)propanoate (4a)

According to general procedure, a mixture of methyl 2-trimethylsiloxycyclopropanecarboxylate (0.510 g, 2.71 mmol), 2-aminopyridine (0.258 g, 2.74 mmol), *tert*-butyl isocyanide (0.225 g, 2.67 mmol) and AcOH (0.325 g, 5.41 mmol) in MeOH (8 mL) gave after 12 h at r.t., 0.420 g (56%) of **4a** as a foamy resin.

IR (neat): 3330–3220 (N–H), 3060–2800 (=C–H, C–H), 1730 (C=O), 1630 (C=N) $\rm cm^{-1}.$

¹H NMR (CDCl₃, 270 MHz): δ = 1.20 (s, 9 H, *t*-C₄H₉), 2.75 (t, *J* = 7 Hz, 2 H, CH₂), 3.00 (t, *J* = 7 Hz, 2 H, CH₂), 3.40 (s, 1 H, NH), 3.55 (s, 3 H, OCH₃), 6.60 (dd, *J* = 7, 7.5 Hz, 1 H, 6-H), 7.05 (dd, *J* = 7.5, 9 Hz, 1 H, 7-H), 7.45 (d, *J* = 9 Hz, 1 H, 8-H), 8.10 (d, *J* = 7 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 67.9 MHz): δ = 22.6 (t, CH₂), 30.2, 55.4 (q, s, *t*-C₄H₉), 33.3 (t, CH₂), 51.5 (q, OCH₃), 110.7 (d, C-6), 116.5 (d, C-8), 123.4 (d, 2 C, C-5, C-7), 124.1 (s, C-3), 138.8 (s, C-2), 141.9 (s, C-9), 174.1 (s, C=O).

 $\begin{array}{l} MS \ (EI): \ m/z \ (\%) = 275 \ (41, \ [M]^+), \ 260 \ (2, \ [M-CH_3]^+), \ 244 \ (9, \ [M-OMe]^+), \ 218 \ (100, \ [M-C_4H_9]^+), \ 191 \ (12), \ 186 \ (10), \ 160 \ (28), \ 159 \ (13), \ 78 \ (45, \ [C_5H_4N]^+), \ 57 \ (7, \ [t-Bu]^+), \ 41 \ (5, \ [C_3H_5]^+), \ 29 \ (5, \ [C_2H_5]^+). \end{array}$

HRMS: *m*/*z* calcd for C₁₅H₂₁N₃O₂: 275.1638; found: 275.1656.

Methyl 3-{3-(1,1,3,3-Tetramethylbutyl)aminoimidazo[1,2-*a*]py-ridin-2-yl}propanoate (4b)

According to general procedure, a mixture of methyl 2-trimethylsiloxycyclopropanecarboxylate (0.827 g, 4.40 mmol), 2-aminopyridine (0.376 g, 4.00 mmol), 1,1,3,3-tetramethylbutyl isocyanide (0.558 g, 4.00 mmol) and AcOH (0.500 g, 8.00 mmol) in MeOH (12 mL) gave after 48 h at r.t., 1.07 g (79%) of **4b** as a yellowish oil.

IR (neat): 3335 (N–H), 3090–2870 (C–H), 1735 (C=O), 1630 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 270 MHz): $\delta = 1.05$, 1.12 (2 s, 9 H, 6 H, CH₃), 1.64 (s, 2 H, CH₂), 2.76 (t, J = 6.8 Hz, 2 H, CH₂), 3.01 (t, J = 6.8 Hz, 2 H, CH₂), 3.21 (br s, 1 H, NH), 3.56 (s, 3 H, OCH₃), 6.63 (td, J = 6.8, 1.2 Hz, 1 H, 6-H), 6.86 (ddd, J = 9.0, 6.8, 1.2 Hz, 1 H, 7-H), 7.34 (dt, J = 9.0, 1.2 Hz, 1 H, 8-H), 8.11 (dt, J = 6.8, 1.2 Hz, 1 H, 5-H). ¹³C NMR (CDCl₃, 125.8 MHz): δ = 22.8 (t, CH₂), 29.1 (q, CH₃), 31.7 (s, *C*Me₃), 31.9 (q, CH₃), 33.3 (t, CH₂), 51.4 (q, OCH₃), 56.8 (t, CH₂), 59.5 (s, N*C*Me₂), 110.6 (d, C-6), 116.6 (d, C-8), 123.2 (d, C-7), 123.5 (d, C-5), 123.8 (s, C-3), 139.2 (s, C-2), 142.0 (s, C-9), 173.9 (s, C=O).

 $\begin{array}{l} MS \ (EI): \ m/z \ (\%) = 331 \ (27, \ [M]^+), \ 220 \ (14), \ 219 \ (100), \ 218 \ (79), \\ 186 \ (22), \ 161 \ (12), \ 160 \ (71), \ 159 \ (16), \ 158 \ (12), \ 146 \ (21), \ 131 \ (16), \\ 119 \ (11), \ 85 \ (53), \ 84 \ (71), \ 79 \ (11), \ 78 \ (40), \ 57 \ (42), \ 55 \ (18), \ 49 \ (14), \\ 47 \ (18), \ 43 \ (20), \ 41 \ (23), \ 29 \ (15), \ 28 \ (34). \end{array}$

HRMS: *m*/*z* calcd for C₁₉H₂₉N₃O₂: 331.2260; found: 331.2264.

Methyl 3-(3-*tert*-Butylaminoimidazo[1,2-*a*]pyridin-2-yl)butanoate (4c)

According to general procedure, a mixture of methyl 3-methyl-2-trimethylsiloxycyclopropanecarboxylate (0.202 g, 1.00 mmol), 2-aminopyridine (0.094 g, 1.00 mmol), *tert*-butyl isocyanide (0.084 g, 1.00 mmol) and AcOH (0.120 g, 2.00 mmol) in MeOH (3 mL) gave after 12 h at r.t., 0.166 g (57%) of **4c** as amber crystals; mp 63–65 °C.

IR (KBr): 3320–3220 (N–H), 3060–2850 (=C–H, C–H), 1730 (C=O), 1635 (C=N) $\rm cm^{-1}.$

¹H NMR (CDCl₃, 270 MHz): δ = 1.20 (s, 9 H, *t*-C₄H₉), 1.45 (d, J = 7 Hz, 3 H, CH₃), 1.65 (s, 1 H, NH), 2.60 (d, J = 7 Hz, 2 H, CH₂), 2.80 (d, J = 7 Hz, 2 H, CH₂), 3.45 (br s, 1 H, NH), 3.48–3.53 (m, 1 H, CH), 3.55 (s, 3 H, CO₂CH₃), 6.65 (dd, J = 7, 7.5 Hz, 1 H, 6-H), 7.05 (dd, J = 7, 5, 9 Hz, 1 H, 7-H), 7.45 (d, J = 9 Hz, 1 H, 8-H), 8.15 (d, J = 7 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 125.8 MHz): δ = 20.1 (q, CH₃), 27.5 (d, CH), 30.1, 54.9 (q, s, *t*-C₄H₉), 41.2 (t, CH₂), 51.3 (q, OCH₃), 110.5 (d, C-6), 116.6 (d, C-8), 123.2 (d, C-5), 123.3 (s, C-3), 123.5 (d, C-7), 142.1, 142.9 (2 s, C-2, C-9), 173.6 (s, C=O).

MS (EI): m/z (%) = 289 (37, [M]⁺), 274 (10, [M - CH₃]⁺), 258 (7, [M - OMe]⁺), 233 (37, [M - C₄H₈]⁺), 232 (100, [M - C₄H₉]⁺), 160 (28), 79 (16, [C₅H₅N]⁺), 78 (46, [C₅H₄N]⁺), 57 (13, [*t*-Bu]⁺), 41 (12, [C₃H₅]⁺), 29 (8, [C₂H₅]⁺).

HRMS: *m*/*z* calcd for C₁₆H₂₃N₃O₂: 289.1790; found: 289.1772.

Methyl 3-(3-*tert*-Butylaminoimidazo[1,2-*a*]pyridin-2-yl)-3methylbutanoate (4d)

According to general procedure, a mixture of methyl 3,3-dimethyl-2-trimethylsiloxycyclopropanecarboxylate (0.648 g, 3.00 mmol), 2aminopyridine (0.283 g, 3.00 mmol), *tert*-butyl isocyanide (0.253 g, 3.00 mmol) and AcOH (0.361 g, 6.00 mmol) in MeOH (9 mL) gave after 12 h at r.t., 0.419 g (46%) of **4d** as colorless crystals; mp 183– 185 °C.

IR (KBr): 3300–3230 (N–H), 3080–2860 (=C–H, C–H), 1730 (C=O), 1630 (C=N) $\rm cm^{-1}.$

¹H NMR (CDCl₃, 270 MHz): $\delta = 1.25$ (s, 9 H, *t*-C₄H₉), 1.55 (br s, 1 H, NH), 1.65 (s, 6 H, CH₃), 3.05 (s, 2 H, CH₂), 3.55 (s, 3 H, OCH₃), 7.15 (dd, J = 7.5, 8.0 Hz, 1 H, 6-H), 7.60 (dd, J = 7.5, 9.0 Hz, 1 H, 7-H), 8.35 (d, J = 9.0 Hz, 1 H, 8-H), 8.50 (d, J = 8.0 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 67.9 MHz): δ = 28.7 (q, CH₃), 31.2, 54.8 (q, s, *t*-Bu), 34.8 (s, *C*Me₂), 45.5 (t, CH₂), 51.6 (q, OCH₃), 113.4 (d, C-6), 115.5 (d, C-8), 124.3 (s, C-3), 124.7 (d, C-5), 131.2 (d, C-7), 133.8, 137.2 (2 s, C-2, C-9), 171.9 (s, C=O).

 $\begin{array}{l} MS \ (EI): {\it m/z} \ (\%) = 303 \ (35, [M]^+), 287 \ (3, [M-CH_4]^+), 272 \ (6, [M-OMe]^+), 247 \ (27, [M-C_4H_8]^+), 246 \ (100, [M-t-Bu]^+), 214 \ (29), \\ 198 \ (10), 78 \ (29, [C_5H_4N]^+), 73 \ (18, [C_4H_{11}N]^+), 57 \ (85, [t-Bu]^+), 43 \ (65, [C_3H_7]^+), 29 \ (19, [C_2H_5]^+). \end{array}$

HRMS: *m*/*z* calcd for C₁₇H₂₅N₃O₂: 303.1947; found: 303.1911.

Methyl 3-(Benzylaminoimidazo[1,2-*a*]pyridin-2-yl)-3-methylbutanoate (4e)

According to general procedure, a mixture of methyl 3,3-dimethyl-2-siloxycyclopropanecarboxylate (0.216 g, 1.00 mmol), 2-aminopyridine (0.094 g, 1.00 mmol), benzyl isocyanide (0.118 g, 1.00 mmol) and AcOH (0.120 g, 2.00 mmol) in MeOH (3 mL) gave after 19 h at r.t., 0.201 g (57%) of **4e** as a pale yellow oil.

IR (KBr): 3375 (N–H), 3085–2950 (=C–H, C–H), 1735 (C=O), 1630 (C=N) $\rm cm^{-1}.$

¹H NMR (CDCl₃, 250 MHz): δ = 1.58 (s, 6 H, CH₃), 2.82 (s, 2 H, CH₂), 3.50 (s, 3 H, OCH₃), 3.62 (br s, 1 H, NH), 4.13 (s, 2 H, CH₂Ph), 6.69 (td, *J* = 6.8, 1.2 Hz, 1 H, 6-H), 7.06 (ddd, *J* = 9.0, 6.8, 1.2 Hz, 1 H, 7-H), 7.25–7.49 (m, 6 H, 8-H, C₆H₃), 7.94 (dt, *J* = 6.8, 1.2 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 67.9 MHz): δ = 28.4 (q, CH₃), 35.3 (s, *C*Me₂), 46.5 (t, CH₂), 51.1 (q, OCH₃), 52.9 (t, *C*H₂Ph), 111.1 (d, C-6), 117.1 (d, C-8), 122.2 (d, C-5), 123.2 (d, C-7), 124.8 (s, C-3), 127.5, 128.0, 128.6 (3 d, C₆H₅), 139.2 (s, C₆H₅), 140.4 (s, C-2), 143.1 (s, C-9), 172.6 (s, C=O).

MS (EI): m/z (%) = 337 (32, [M]⁺), 247 (16), 246 (100), 219 (22), 214 (21), 105 (32), 78 (29), 73 (13).

HRMS: *m*/*z* calcd for C₂₀H₂₃N₃O₂: 337.1790; found: 337.1784.

Methyl 3-{3-(1,1,3,3-Tetramethylbutyl)imidazo[1,2-*a*]pyridin-2-yl}-3-methylbutanoate (4f)

According to general procedure, a mixture of methyl 3,3-dimethyl-2-trimethylsiloxycyclopropanecarboxylate (0.951 g, 4.40 mmol), 2aminopyridine (0.376 g, 4.00 mmol), 1,1,3,3-tetramethylbutyl isocyanide (0.558 g, 4.00 mmol) and AcOH (0.480 g, 8.00 mmol) in MeOH (12 mL) gave after 48 h at r.t., 0.797 g (56%) of **4f** as a pale yellow oil and 0.280 g (30%) of **7** as colorless crystals (for analytical and spectral data of **7**, see below).

IR (neat): 3370 (N–H), 3075–2870 (=C–H, C–H), 1735 (C=O), 1630 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 270 MHz): δ = 1.08, 1.22, 1.58 (3 s, 9 H, 3 H, 3 H, CH₃), 1.69, 2.91 (2 s, 2 H, 2 H, CH₂), 3.11 (s, 1 H, NH), 3.52 (s, 3 H, OCH₃), 6.63 (td, *J* = 6.8, 1.2 Hz, 1 H, 6-H), 6.99 (ddd, *J* = 9.0, 6.8, 1.2 Hz, 1 H, 7-H), 7.38 (dt, *J* = 9.0, 1.2 Hz, 1 H, 8-H), 8.16 (dt, *J* = 6.8, 1.2 Hz, 1 H, 5-H).

 ^{13}C NMR (CDCl₃, 67.9 MHz): δ = 29.0, 30.3, 31.9 (3 q, CH₃), 32.0, 35.9 (2 s, *C*Me₃), *C*Me₂), 46.6 (t, CH₂), 51.1 (q, OCH₃), 57.8 (t, CH₂), 58.8 (s, N*C*Me₂), 110.3 (d, C-6), 116.8 (d, C-8), 122.5 (d, C-5), 123.1 (d, C-7), 123.8 (s, C-3), 140.8 (s, C-2), 144.9 (s, C-9), 172.6 (s, C=O).

MS (EI): m/z (%) = 359 (66, [M]⁺), 328 (11), 248 (18), 247 (86), 246 (100), 219 (29), 215 (13), 214 (51), 188 (17), 174 (44), 105 (17).

HRMS: *m*/*z* calcd for C₂₁H₃₃N₃O₂: 359.2573; found: 359.2554.

Anal. Calcd for $C_{21}H_{33}N_3O_2$ (359.5): C, 70.16; H, 9.25; N, 11.69. Found: C, 69.90; H, 9.14; N, 11.70.

Methyl 3-{(4-Methoxyphenyl)aminoimidazo[1,2-*a*]pyridin-2-yl}-3-methylbutanoate (4g)

According to general procedure, a mixture of methyl 3,3-dimethyl-2-trimethylsiloxycyclopropanecarboxylate (0.238 g, 1.10 mmol), 2aminopyridine (0.094 g, 1.00 mmol), 4-methoxyphenyl isocyanide (0.133 g, 1.00 mmol) and AcOH (0.120 g, 2.00 mmol) in MeOH (3 mL) gave after 60 min under microwave conditions (200 W, T_{max} = 30 °C), 0.218 g (62%) of **4g** as pale brownish crystals; mp 140–143 °C.

IR (KBr): 3270 (N–H), 3105–2825 (=C–H, C–H), 1730 (C=O), 1675 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 250 MHz): δ = 1.50 (s, 6 H, CH₃), 2.82 (s, 2 H, CH₂), 3.55, 3.72 (2 s, 3 H, 3 H, OCH₃), 5.79 (s, 1 H, NH), 6.20–6.26 (m, 2 H, Ar), 6.50 (td, *J* = 6.8, 1.2 Hz, 1 H, 6-H), 6.54–6.60 (m, 2 H, Ar), 6.94 (ddd, *J* = 9.1, 6.8, 1.2 Hz, 1 H, 7-H), 7.34 (dt, *J* = 9.1, 1.2 Hz, 1 H, 8-H), 7.58 (dt, *J* = 6.8, 1.2 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 67.9 MHz): δ = 28.3 (q, CH₃), 35.3 (s, CMe₂), 46.0 (t, CH₂), 51.1 (q, OCH₃), 55.6 (q, ArOCH₃), 111.4 (d, C-6), 114.0 (d, C-8), 115.1 (d, C-5), 117.2 (d, C-7), 118.6 (s, C-3), 122.6, 124.0 (2 d, Ar), 139.2 (s, Ar), 141.4 (s, C-2), 146.3 (s, C-9), 153.1 (s, Ar), 172.9 (s, C=O).

MS (EI): *m/z* (%) = 354 (100, [M]⁺), 294 (30), 281 (38), 151 (11), 147 (25), 112 (15), 95 (12), 94 (42), 86 (21), 85 (16), 84 (42), 83 (17), 78 (28), 73 (21), 71 (23), 70 (20), 69 (32), 67 (15), 60 (22), 57 (46), 56 (19), 55 (35), 45 (37), 44 (37), 43 (71), 42 (14), 39 (19).

Anal. Calcd for $C_{20}H_{23}N_3O_3$ (353.4): C, 67.97; H, 6.56; N, 11.89. Found: C, 67.83; H, 6.47; N, 11.44.

Crystal Data

C₂₀H₂₃N₃O₃, M_r = 353.4; T = 173(2) K; crystal size: $0.5 \times 0.2 \times 0.1$ mm; monoclinic, space group *P*2(1)/*c*, *a* = 9.870(4), *b* = 10.817(4), *c* = 17.218(7) A; *Z* = 4; D_c = 1.290 Mg/m³; F(000) = 752; μ(Mo-K) = 0.088 mm⁻¹. Θ Range for data collection: 2.08–30.55°, index ranges: $-12 \le h \le 14$, $-15 \le k \le 14$, $-22 \le l \le 24$; reflections collected/ unique: 13376/5119 (R_{int} = 0.0296); final R [I>2σ(I)]: *R*1 = 0.0526, *wR*2 = 0.1400. For structure solution and refinement, the programs SHELXS97 and SHELXL97 were used.^{16,17}

Methyl 3-(Benzylaminoimidazo[1,2-*a*]pyridin-2-yl)-2,3-dimethylbutanoate (4h)

According to general procedure, a mixture of methyl 1,3,3-trimethyl-2-trimethylsiloxycyclopropanecarboxylate (0.277 g, 1.20 mmol), 2-aminopyridine (0.094 g, 1.00 mmol), benzyl isocyanide (0.118 g, 1.00 mmol) and AcOH (0.120 g, 2.00 mmol) in MeOH (3 mL) gave after 17 h at r.t., 0.140 g (40%) of **4h** as a pale yellow oil.

IR (KBr): 3375 (N–H), 3085–2950 (=C–H, C–H), 1735 (C=O), 1630 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 250 MHz): $\delta = 1.06$ (d, J = 7.2 Hz, 3 H, CH₃), 1.54 (s, 6 H, CH₃), 3.17 (q, J = 7.2 Hz, 1 H, CH), 3.44 (t, J = 6.4 Hz, 1 H, NH), 3.50 (s, 3 H, OCH₃), 4.07 (d, J = 6.4 Hz, 2 H, CH₂Ph), 6.66 (td, J = 6.8, 1.1 Hz, 1 H, 6-H), 7.04 (ddd, J = 9.0, 6.8, 1.1 Hz, 1 H, 7-H), 7.24–7.55 (m, 6 H, 8-H, C₆H₅), 7.91 (dt, J = 6.8, 1.1 Hz, 1 H, 5-H).

 $\label{eq:constraint} \begin{array}{l} ^{13}\text{C NMR} \ (\text{CDCl}_3, 62.9 \ \text{MHz}); \ \delta = 13.1, 24.0, 26.0 \ (3 \ q, \text{CH}_3), 38.5 \\ (\text{s}, C\text{Me}_2), 48.5 \ (\text{d}, \text{CH}), 51.0 \ (\text{q}, \text{OCH}_3), 52.8 \ (\text{t}, C\text{H}_2\text{Ph}), 111.2 \ (\text{d}, \text{C-6}), 117.0 \ (\text{d}, \text{C-8}), 122.1 \ (\text{d}, \text{C-5}), 123.3 \ (\text{d}, \text{C-7}), 125.3 \ (\text{s}, \text{C-3}), \\ 127.5, 127.9, 128.6 \ (3 \ \text{d}, \text{C}_6\text{H}_5), 139.0 \ (\text{s}, \text{C}_6\text{H}_5), 140.2 \ (\text{s}, \text{C-2}), \\ 142.3 \ (\text{s}, \text{C-9}), 176.2 \ (\text{s}, \text{C=0}). \end{array}$

MS (EI): m/z (%) = 351 (34, [M]⁺), 264 (14), 261 (20), 260 (100), 233 (16), 229 (12), 228 (66), 201 (10), 174 (16), 176 (15), 158 (14), 121 (21) 105 (19), 91 (10, [Bn]⁺), 78 (13).

HRMS: *m*/*z* calcd for C₂₁H₂₅N₃O₂: 351.1947; found: 351.1963.

Methyl 3-(3-Aminoimidazo[1,2-*a*]pyridin-2-yl)propanoate (5)

Compound **4b** (0.643 g, 1.94 mmol) was dissolved in trifluoroacetic acid–CH₂Cl₂ solution (1:1; 5 mL) and stirred for 2 h at r.t. The mixture was loaded on a Dowex cation exchange resin, washed with MeOH until it was acid-free and then eluted with MeOH saturated with ammonia. The solvent was removed under reduced pressure and the residue was purified by column chromatography (silica gel, CH₂Cl₂–MeOH, 19:1); yield: 0.399 g (94%) of **5** as yellowish oil.

IR (KBr): 3320–3180 (N–H), 2950–2850 (=C–H, C–H), 1730 (C=O), 1670 (C=N) $\rm cm^{-1}.$

¹H NMR (CDCl₃, 500 MHz): $\delta = 2.78$ (t, J = 6.7 Hz, 2 H, CH₂), 3.02 (t, J = 6.7 Hz, 2 H, CH₂), 3.52 (s, 3 H, OCH₃), 6.05 (br s, 2 H, NH₂), 6.76 (t, J = 6.8 Hz, 1 H, 6-H), 7.03–7.19 (m, 1 H, 7-H), 7.42 (d, J = 9.1 Hz, 1 H, 8-H), 8.00 (d, J = 6.8 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 125.8 MHz): δ = 21.6, 33.2 (2 t, CH₂), 51.6 (q, OCH₃), 111.6 (d, C-6), 116.7 (d, C-8), 122.1 (d, C-5), 123.3 (d, C-7), 123.4 (s, C-3), 132.7 (s, C-2), 140.2 (s, C-9), 174.1 (s, C=O).

MS (EI): *m/z* (%) = 219 (58, [M]⁺), 188 (16), 161 (14), 160 (100), 159 (19), 146 (52), 133 (14), 121 (16), 119 (25), 115 (14), 80 (24), 79 (82), 55 (15), 51 (17).

HRMS: *m*/*z* calcd for C₁₁H₁₃N₃O₂: 219.1008; found: 219.1016.

Methyl 3-(3-Aminoimidazo[1,2-*a*]pyridin-2-yl)-3-methylbutanoate (6) and 3-(3-Aminoimidazo[1,2-*a*]pyridin-2-yl)-3-methylbutanoic Acid (7)

Compound **4f** (0.116 g, 0.32 mmol) was dissolved in trifluoroacetic acid– CH_2Cl_2 solution (1:1; 2 mL) and stirred for 5 min at r.t. The mixture was loaded on a Dowex cation exchange resin, washed with MeOH until it was acid-free and then eluted with MeOH saturated with ammonia. The solvent was removed under reduced pressure and the residue was purified by column chromatography (silica gel, CH_2Cl_2 –MeOH, 19:1); yield: 0.020 g (25%) of **6** as a yellow oil and 0.037 g (50%) of **7** as colorless crystals; mp 250–251 °C.

According to the preparation described above, compound **4f** (0.861 g, 2.40 mmol) was dissolved in trifluoroacetic acid–CH₂Cl₂ solution (1:1; 5 mL) and stirred for 2 h at r.t. Purification by column chromatography (silica gel, CH₂Cl₂–MeOH, 19:1) gave 0.450 g (80%) of **7** as colorless crystals.

Compound 6

IR (KBr): 3380–3085 (N–H, O–H), 2950–2875 (=C–H, C–H), 1720 (C=O), 1610 (C=N) cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): $\delta = 1.35$ (s, 6 H, CH₃), 2.54 (s, 2 H, CH₂), 3.91 (s, 3 H, OCH₃), 6.75 (td, J = 6.7, 1.2 Hz, 1 H, 6-H), 7.02 (ddd, J = 9.0, 6.7, 1.2 Hz, 1 H, 7-H), 7.49 (dt, J = 9.0, 1.2 Hz, 1 H, 8-H), 8.09 (dt, J = 6.7, 1.2 Hz, 1 H, 5-H); the NH₂ signal could not be detected.

¹³C NMR (CDCl₃, 125.8 MHz): δ = 27.0 (q, CH₃), 32.2 (s, CMe₂), 42.8 (t, CH₂), 53.7 (q, OCH₃), 111.8 (d, C-6), 117.1 (d, C-8), 121.5 (d, C-5), 122.4 (d, C-7), 127.5 (s, C-3), 135.8 (s, C-2), 141.9 (s, C-9), 165.9 (s, C=O).

MS (EI): *m/z* (%) = 229 (100, [M – NH₄]⁺), 215 (14), 214 (95), 200 (15), 199 (14), 91 (12, [Bn]⁺), 85 (21), 83 (21), 78 (21), 55 (40), 45 (25), 43 (74), 41 (46).

HRMS: m/z calcd for $C_{13}H_{13}N_2O_2$ [M – NH₄]⁺: 229.0978; found: 229.0982.

Compound 7

IR (KBr): 3335–2865 (O–H, N–H, =C–H, C–H), 1680 (C=O), 1570 (C=N) cm⁻¹.

¹H NMR (CD₃OD, 500 MHz): δ = 1.47 (s, 6 H, CH₃), 2.78 (s, 2 H, CH₂), 6.88 (td, *J* = 6.8, 1.2 Hz, 1 H, 6-H), 7.13 (ddd, *J* = 9.2, 6.8, 1.2 Hz, 1 H, 7-H), 7.44 (dt, *J* = 9.2, 1.2 Hz, 1 H, 8-H), 8.06 (dt, *J* = 6.8, 1.2 Hz, 1 H, 5-H).

 ^{13}C NMR (CD₃OD, 125.8 MHz): δ = 27.2 (q, CH₃), 33.1 (s, CMe₂), 48.0 (t, CH₂), 113.8 (d, C-6), 117.5 (d, C-8), 121.3 (d, C-5), 122.5 (d, C-7), 123.9 (d, C-3), 139.3 (s, C-2), 142.1 (s, C-9), 172.4 (s, C=O).

 $\begin{array}{l} MS \ (EI): \textit{m/z} \ (\%) = 215 \ (87, \ [M-H_2O]^+), \ 201 \ (29), \ 200 \ (100), \ 173 \\ (13), \ 172 \ (11), \ 158 \ (12), \ 145 \ (20), \ 79 \ (19), \ 78 \ (57), \ 52 \ (10), \ 51 \ (17), \\ 41 \ (12), \ 39 \ (12), \ 28 \ (16), \ 27 \ (10). \end{array}$

HRMS: m/z calcd for $C_{12}H_{13}N_3O [M - H_2O]^+$: 215.1059; found: 215.1063.

Synthesis 2006, No. 16, 2677–2684 © Thieme Stuttgart · New York

3-{3-[(*tert*-Butoxycarbonyl)amino]imidazo[1,2-*a*]pyridin-2-yl}-3-methylbutanoic Acid (8)

A stirred solution of amino acid **5** (0.090 g, 0.386 mmol) in THF (6 mL) was treated with NaH (0.016 g, 0.038 mmol; 60% suspension in oil), and further stirred for 1 h at r.t. Boc_2O (0.168 g, 0.772 mmol) was then added to the mixture, stirred for 1 h, and then treated with a catalytic amount of DMAP followed by stirring for 1 h. The mixture was partitioned between EtOAc and sat. aq NH₄Cl solution and the organic phase was separated and dried (Na₂SO₄). The solvent was removed under reduced pressure and the resulting crude product was purified by column chromatography (silica gel; 10% *i*-PrOH–hexane) to give 0.100 g (78%) of **8** as brownish oil.

IR (KBr): 3445 (N–H), 3050–2875 (=C–H, C–H), 1785, 1740 (C=O), 1635 (C=N), 1290 (C–O) $cm^{-1}.$

¹H NMR (CDCl₃, 250 MHz): δ = 1.36 (s, 6 H, CH₃), 1.49 (s, 9 H, *t*-C₄H₉), 2.57 (s, 2 H, CH₂), 6.75 (t, *J* = 6.9 Hz, 1 H, 6-H), 7.07 (t, *J* = 6.9 Hz, 1 H, 7-H), 7.43–7.53 (m, 2 H, 8-H, 5-H).

¹³C NMR (CDCl₃, 62.9 MHz): δ = 26.0 (q, CH₃), 27.7, 85.2 [q, s, C(CH₃)₃], 31.6 (s, *C*Me₂), 50.4 (t, CH₂), 111.8 (d, C-6), 117.5 (d, C-8), 118.0 (s, C-3), 122.7 (d, C-5), 123.0 (d, C-7), 140.4 (s, C-2), 142.3 (s, C-9), 148.5, 172.4 (2 s, C=O).

MS (EI): m/z (%) = 315 (2, $[M - H_2O]^+$), 215 (62), 200 (100), 78 (15), 57 (27).

HRMS: m/z calcd for $C_{17}H_{21}N_3O_3$ [M – H_2O]⁺: 315.1583; found: 315.1573.

3-[3-(Benzylamino)imidazo[1,2-*a*]pyridin-2-yl]-3-methylbutanoic Acid (9)

To a solution of compound **4e** (0.109 g, 0.320 mmol) in MeOH– THF (1 mL/3 mL), was added a solution of LiOH·H₂O (0.041 g, 0.096 mmol) in H₂O (1 mL), and the resulting mixture was stirred for 22 h at r.t. Then aq 2 M HCl was added to adjust the pH to 7, followed by Et₂O and the layers were separated. The aqueous layer was extracted with Et₂O, the combined organic phases were dried (MgSO₄) and the solvent was evaporated. Purification of the crude product by column chromatography (silica gel, hexane–EtOAc, 1:2) gave 0.080 g (77%) of **9** as a pale yellow solid; mp 111–115 °C.

IR (KBr): 3230–3030 (O–H, N–H), 2960–2870 (=C–H, C–H), 1680 (C=O), 1235 (C–O) cm⁻¹.

¹H NMR (CDCl₃, 250 MHz): $\delta = 1.38$ (s, 6 H, CH₃), 2.74 (s, 2 H, CH₂), 5.23 (s, 2 H, CH₂Ph), 6.65 (td, J = 6.9, 1.2 Hz, 1 H, 6-H), 7.01 (ddd, J = 9.1, 6.9, 1.2 Hz, 1 H, 7-H), 7.24–7.40 (m, 5 H, C₆H₅), 7.56 (dt, J = 9.1, 1.2 Hz, 1 H, 8-H), 7.76 (dt, J = 6.9, 1.2 Hz, 1 H, 5-H); the NH signal could not be detected.

¹³C NMR (CDCl₃, 62.9 MHz): δ = 26.4 (q, CH₃), 31.6 (s, CMe₂), 46.6 (t, CH₂), 48.5 (t, CH₂Ph), 112.8 (d, C-6), 118.2 (d, C-8), 121.8 (d, C-5), 121.9 (d, C-7), 122.1 (s, C-3), 126.8, 127.8, 129.0, 136.7 (3 d, s, C₆H₅), 138.6 (s, C-2), 141.5 (s, C-9), 171.0 (s, C=O).

MS (EI): m/z (%) = 323 (31, [M]⁺), 305 (32), 233 (16), 232 (100, [M - Bn]⁺), 215 (14), 214 (94), 205 (42), 121 (31), 105 (17), 91 (39, [Bn]⁺), 83 (11), 79 (14), 78 (59), 41 (11).

HRMS: *m/z* calcd for C₁₉H₂₁N₃O₂: 323.1634; found: 323.1663.

1-Benzyl-4,4-dimethyl-3,4-dihydro-1*H*-dipyrido[1,2-*a*;3',2'-*d*]imidazol-2-one (10e)

Compound **4e** (0.060 g, 0.178 mmol) was dissolved in anhyd MeOH (2 mL) under argon, followed by addition of NaCN (1 mg, 0.02 mmol). The mixture was heated at 85 °C for 43 h, and then the solvent was evaporated to dryness. EtOAc and H_2O were added and the layers were separated. The aqueous layer was extracted with EtOAc, the combined organic layers were dried (Na₂SO₄) and the solvent was removed under reduced pressure. The resulting crude product was purified by column chromatography (alumina, activity

III; hexane–EtOAc, 1:1) leading to 0.050 g (92%) of 10e as colorless crystals; mp 138–140 °C.

IR (KBr): 3065–2865 (=C–H, C–H), 1680 (C=O), 1630 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 250 MHz): δ = 1.38 (s, 6 H, CH₃), 2.74 (s, 2 H, CH₂), 5.23 (s, 2 H, CH₂Ph), 6.64 (td, *J* = 6.9, 1.2 Hz, 1 H, 6-H), 7.01 (ddd, *J* = 9.1, 6.9, 1.2 Hz, 1 H, 7-H), 7.23–7.37 (m, 5 H, C₆H₅), 7.55 (dt, *J* = 9.1, 1.2 Hz, 1 H, 8-H), 7.77 (dt, *J* = 6.9, 1.2 Hz, 1 H, 5-H).

¹³C NMR (CDCl₃, 62.9 MHz): δ = 26.3 (q, CH₃), 31.5 (s, *C*Me₂), 46.5, 48.4 (2 t, CH₂), 112.6 (d, C-6), 118.2 (d, C-8), 121.6 (d, C-5), 121.8 (d, C-7), 122.0 (s, C-3), 126.7, 127.7, 128.9, 136.6 (3 d, s, C₆H₅), 138.6 (s, C-2), 141.4 (s, C-9), 170.9 (s, C=O).

MS (EI): *m/z* (%) = 305 (32, [M]⁺), 215 (14), 214 (100, [M – Bn]⁺), 91 (13, [Bn]⁺), 78 (18).

HRMS: *m/z* calcd for C₁₉H₁₉N₃O: 305.1528; found: 305.1536.

1-Benzyl-3,4,4-trimethyl-3,4-dihydro-1*H*-dipyrido[1,2-*a*;3',2'-*d*]imidazol-2-one (10h)

According to the preparation of **10e**, compound **4h** (0.414 g, 1.12 mmol), NaCN (6 mg, 0.12 mmol) and MeOH (14 mL) gave after 48 h at 85 °C and purification by column chromatography (alumina, activity III; hexane–EtOAc, 1:1), 0.350 g (98%) of **10h** as colorless crystals; mp 114–115 °C.

IR (KBr): 3085–2870 (=C–H, C–H), 1680 (C=O), 1635 (C=N) cm^{-1} .

¹H NMR (CDCl₃, 500 MHz): δ = 1.11 (s, 3 H, CH₃), 1.19 (d, J = 7.1 Hz, 3 H, CH₃), 1.42 (s, 3 H, CH₃), 2.66 (q, J = 7.1 Hz, 1 H, 10-H), 5.16, 5.23 (2 dd, J = 16.3 Hz, 2 H, CH₂), 6.62 (td, J = 6.9, 1.2 Hz, 1 H, 6-H), 6.98 (ddd, J = 9.1, 6.9, 1.2 Hz, 1 H, 7-H), 7.19–7.33 (m, 5 H, C₆H₅), 7.54 (dt, J = 9.1, 1.2 Hz, 1 H, 8-H), 7.72 (dt, J = 6.9, 1.2 Hz, 1 H, 5-H).

 ^{13}C NMR (CDCl₃, 125.8 MHz): δ = 10.0, 21.3, 25.3 (3 q, CH₃), 34.3 (s, CMe₂), 46.6 (t, CH₂), 48.8 (d, CH), 112.5 (d, C-6), 118.1 (d, C-8), 121.3 (d, C-5), 121.5 (d, C-7), 121.7 (s, C-3), 126.6, 127.6, 128.8, 136.7 (3 d, s, C_6H_5), 139.0 (s, C-2), 141.2 (s, C-9), 173.9 (s, C=O).

MS (EI): m/z (%) = 319 (53, [M]⁺), 258 (17), 229 (17), 228 (100, [M - Bn]⁺), 121 (30), 108 (22), 106 (17), 94 (25), 91 (38, [Bn]⁺), 86 (37), 84 (64), 78 (33), 70 (37), 69 (15), 67 (17), 55 (23), 47 (16), 43 (21), 41 (24).

HRMS: *m*/*z* calcd for C₂₀H₂₁N₃O: 319.1685; found: 319.1666.

Methyl *N*-(3-{3-[(*tert*-Butoxycarbonyl)amino]imidazo[1,2-*a*]py-ridin-2-yl}-3-methylbutanoyl)-l-alaninate (11)

Acid **8** (0.074 g, 0.222 mmol), H-Ala-OMe·HCl (0.036 g, 0.26 mmol) and BOP (0.115 g, 0.26 mmol) were dissolved in anhyd CH₂Cl₂ (2 mL), and then DIEA (0.1 mL, 0.66 mmol) was added. The reaction was stirred for 10 d at r.t. EtOAc and H₂O were added and the layers were separated. The organic layer was successively washed with sat. aq NaHCO₃ solution, brine and H₂O, dried (Na₂SO₄) and the solvent was removed under reduced pressure. Purification by column chromatography [silica gel; hexane–EtOAc (1:2), then MeOH–CH₂Cl₂ (1:1), and finally HPLC 50% *i*-PrOH–hexane, 64 mL/min, 124 bar] yielded 0.089 g (97%) of **11** as a colorless solid; mp 70–72 °C; $[\alpha]_D – 27.4$ (c = 0.95, MeOH).

IR (KBr): 3290 (N–H), 3055–2880 (=C–H, C–H), 1725 (C=O), 1655 (C=N) cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): $\delta = 1.06$ (d, J = 7.2 Hz, 3 H, CH₃), 1.54, 1.55 (2 s, 9 H, 6 H, t-C₄H₉, CH₃), 2.60–2.82 (m, 2 H, CH₂), 3.60 (s, 3 H, OCH₃), 4.35 (q, J = 7.2 Hz, 1 H, CH), 6.64, 6.74 (2 br s, 1 H, 1 H, NH), 6.82 (t, J = 6.2 Hz, 1 H, 6-H), 7.11–7.24 (m, 1 H, 7-H), 7.52 (d, J = 9.0 Hz, 1 H, 8-H), 7.86 (d, J = 6.2 Hz, 1 H, 5-H). ¹³C NMR (CDCl₃, 125.8 MHz): δ = 17.7, 28.2 [2 q, CH₃, C(CH₃)₃], 36.0 (s, *CM*e₂), 47.7 (d, CH), 50.6 (t, CH₂), 52.1 (q, OCH₃), 81.3 (s, *CM*e₃), 112.0 (d, C-6), 117.0 (d, C-8), 122.9 (d, C-5), 124.7 (d, C-7), 128.0 (s, C-3), 142.5 (s, C-2), 148.7 (s, C-9), 154.1, 171.4, 171.5 (3 s, C=O).

MS (EI): *m/z* (%) = 418 (6, [M]⁺), 345 (15), 344 (48), 318 (39), 242 (20), 223 (14), 216 (15), 215 (71), 214 (45), 200 (85), 188 (57), 175 (13), 174 (100), 173 (14), 158 (13), 147 (11), 121 (12), 79 (12), 78 (30), 57 (32).

HRMS: *m/z* calcd for C₂₁H₃₀N₄O₅: 418.2216; found: 418.2235.

Methyl 3-(3-{[*N*-(*tert*-Butoxycarbonyl)-l-alanyl]amino}imidazo[1,2-*a*]pyridin-2-yl)propanoate (12)

Amino ester **5** (0.026 g, 0.12 mmol), Boc-Ala-OH (0.022 g, 0.12 mmol) and DIEA (0.06 mL, 0.36 mmol) were dissolved in anhyd CH₂Cl₂ (3 mL), cooled in an ice bath, and then TFFH (0.042 g, 0.15 mmol) was added. The temperature was allowed to rise to r.t. After 4 d, the mixture was successively washed with aq 1 M HCl, sat. aq NaHCO₃ solution and brine, dried (Na₂SO₄) and the solvent was removed under reduced pressure. Purification by column chromatography [silica gel; hexane–EtOAc (4:1), then MeOH–CH₂Cl₂ (4:1), and finally HPLC 50% *i*-PrOH–hexane, 64 mL/min, 124 bar] gave 0.012 g (26%) of **12** as pale yellow oil; $[\alpha]_D$ –25.6 (*c* = 0.09, MeOH).

IR (KBr): 3295 (N–H), 3055–2855 (=C–H, C–H), 1715 (C=O), 1635 (C=N) cm⁻¹.

¹H NMR (CDCl₃, 500 MHz): $\delta = 1.46$ (s, 9 H, *t*-C₄H₉), 1.55 (d, J = 7.2 Hz, 3 H, CH₃), 2.75–2.82 (m, 2 H, CH₂), 2.91–2.97 (m, 2 H, CH₂), 3.60 (s, 3 H, OCH₃), 4.43 (q, J = 7.2 Hz, 1 H, CH), 5.15 (br s, 1 H, NH), 6.75 (td, J = 6.8, 1.1 Hz, 1 H, 6-H), 7.13 (ddd, J = 9, 6.8, 1.1 Hz, 1 H, 7-H), 7.46 (dt, J = 9, 1.1 Hz, 1 H, 8-H), 7.73 (d, J = 6.8 Hz, 1 H, 5-H), 8.75 (br s, 1 H, NH).

¹³C NMR (CDCl₃, 125.8 MHz): δ = 18.2 (q, CH₃), 21.9 (t, CH₂), 28.3 [q, C(CH₃)₃], 33.0 (t, CH₂), 38.6 (s, CMe₂), 50.6 (d, CH), 51.7 (q, OCH₃), 80.3 (s, CMe₃), 111.8 (d, C-6), 115.5 (s, C-3), 117.1 (d, C-8), 123.6 (d, C-5), 124.2 (d, C-7), 138.6 (s, C-2), 142.8 (s, C-9), 155.8, 172.7, 174.6 (3 s, C=O).

MS (EI): *m*/*z* (%) = 390 (15, [M]⁺), 334 (12), 247 (13), 246 (86), 219 (56), 218 (18), 214 (16), 186 (20), 160 (38), 159 (14), 146 (15), 131 (15), 79 (42), 73 (19), 59 (29), 57 (100), 55 (16), 44 (59), 43 (42).

HRMS: *m*/*z* calcd for C₁₉H₂₆N₄O₅: 390.1903; found: 390.1914.

Acknowledgment

Support by the Deutsche Forschungsgemeinschaft (Graduiertenkolleg 'Hydrogen Bridges and Hydrogen Transfer'), the Fonds der Chemischen Industrie and the Schering AG is most gratefully acknowledged. We also thank Simon Braun for his experimental help.

References

- (1) Sewald, N.; Jakubke, H.-D. *Peptides: Chemistry and Biology*; Wiley-VCH: Weinheim, **2002**, Chap. 7.
- (2) For excellent up to date reviews, see: (a) Zhu, J.; Bienaymé, H. Multicomponent Reactions; Wiley-VCH: Weinheim,
 2005. (b) Dömling, A. Chem. Rev. 2006, 106, 17. (c) Zhu, J. Eur. J. Org. Chem. 2003, 1133. (d) Dömling, A.; Ugi, I. Angew. Chem. Int. Ed. 2000, 39, 3168; Angew. Chem. 2000, 118, 3300. (e) Bienaymé, H.; Hulme, C.; Oddon, G.; Schmitt, P. Chem. Eur. J. 2000, 6, 3321. (f) Ugi, I.; Lohberger, S.; Karl, R. In Comprehensive Organic

Synthesis 2006, No. 16, 2677-2684 © Thieme Stuttgart · New York

Synthesis, Vol. 2; Trost, B. M.; Fleming, I., Eds.; Pergamon: Oxford, **1991**, Chap. 4.6. (g) Ugi, I. *J. Prakt. Chem.* **1997**, 339, 499.

- (3) (a) Reissig, H.-U.; Zimmer, R. Chem. Rev. 2003, 103, 1151.
 (b) Reissig, H.-U. Top. Curr. Chem. 1988, 144, 73.
- (4) Zimmer, R.; Ziemer, A.; Gruner, M.; Brüdgam, I.; Hartl, H.; Reissig, H.-U. Synthesis 2001, 1649.
- (5) Veljkovic, I.; Al-Ajmi, H.; Özbek, H.; Reissig, H.-U., manuscript in preparation.
- (6) (a) Groebke, K.; Weber, L.; Mehlin, F. Synlett 1998, 661.
 (b) Mandair, G. S.; Light, M.; Russell, A.; Hursthouse, M.; Bradley, M. Tetrahedron Lett. 2002, 43, 4267. (c) Lyon, M. A.; Kercher, T. S. Org. Lett. 2004, 6, 4989. (d) Blackburn, C.; Guan, B.; Fleming, P.; Shiosaki, K.; Tsai, S. Tetrahedron Lett. 1998, 39, 3635. Alternative methods for the preparation of imidazo[1,2-a]pyridines: (e) Krasovsky, A. L.; Nenajdenko, V. G.; Balenkova, E. S. Synthesis 2002, 1379.
- (7) (a) Katritzky, A. R.; Xu, Y.-J.; Tu, H. *J. Org. Chem.* 2003, 68, 4935. (b) Jaramillo, C.; de Diego, E.; Hamdouchi, C. *Synlett* 2002, 1544; and references cited therein.
- (8) (a) Kappe, C. O. Angew. Chem. Int. Ed. 2004, 43, 6250; Angew. Chem. 2004, 116, 6408. (b) Loupy, A. Microwaves in Organic Synthesis; Wiley-VCH: Weinheim, 2002. For microwave-assisted synthesis of fused 3-aminoimidazolines, see: (c) Ireland, S. M.; Tye, K.; Whittaker, M. Tetrahedron Lett. 2003, 44, 4369. (d) Varma, R. S.; Kumar, D. Tetrahedron Lett. 1999, 40, 7665.

- (9) Blackburn, C.; Guan, B. Tetrahedron Lett. 2000, 41, 1495.
- (10) Högberg, T.; Ström, P.; Ebner, M.; Rämsby, S. J. Org. *Chem.* **1987**, *52*, 2033.
- (11) Hünig, S.; Schaller, R. Angew. Chem., Int. Ed. Engl. 1982, 21, 36; Angew. Chem. 1982, 94, 1.
- (12) (a) Campagne, J.-M.; Coste, J.; Jouin, P. J. Org. Chem. 1995, 60, 5214. (b) Castro, B.; Dormoy, J. R.; Evin, G.; Selve, C. Tetrahedron Lett. 1975, 1219.
- (13) Carpino, L. A.; El-Faham, A. J. Am. Chem. Soc. 1995, 117, 5401.
- (14) Kunkel, E.; Reichelt, I.; Reissig, H.-U. *Liebigs Ann. Chem.* 1984, 512.
- (15) Casanova, J.; Newton, D. W.; Schuster, R. E. J. Am. Chem. Soc. 1966, 88, 3473.
- (16) Sheldrick, G. M. SHELX97 (Includes SHELXS97, SHELXL97, CIFTAB) Programs for Crystal Structure Analysis (Release 97-2); Universität Göttingen: Germany, 1998.
- (17) Atomic coordinates and further crystallographic details have been deposited at the Cambridge Crystallographic Data Centre, deposition number CCDC 601760, and copies of this data can be obtained in application to CCDC, University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, UK (fax: +44 1223 336033; e-mail: deposit@ccdc.cam.ac.uk).