Amphidinolide B: Total Synthesis, Structural Investigation, and Biological Evaluation

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Supporting Information

ABSTRACT: The total syntheses of amphidinolide B_1 and the proposed structure of amphidinolide B_2 have been accomplished. Key aspects of this work include the development of a practical, non-transition-metal-mediated method for the construction of the $C_{13}-C_{15}$ diene, the identification of α -chelation and dipole minimization models for diastereoselective methyl ketone aldol reactions, the discovery of a spontaneous Horner–Wadsworth–Emmons macrocyclization strategy, and the development of a novel late stage method for construction of an allylic epoxide moiety. The originally proposed structure for amphidinolide B_2 and diastereomers thereof display potent antitumor activities with IC₅₀ values ranging from 3.3 to 94.5 nM against human solid and blood



tumor cells. Of the different stereoisomers, the proposed structure of amphidinolide B_2 is over 12-fold more potent than the $C_{8,9}$ -epimer and C_{18} -epimer in human DU145 prostate cancer cells. These data suggest that the epoxide stereochemistry is a significant factor for anticancer activity.

■ INTRODUCTION

First reported in 1986, the amphidinolide family of natural products has long captured the attention of the scientific community.¹ To date, over 30 members of this family have been isolated.² Given their fascinating structures and diverse biological activity, these targets have attracted considerable attention in both the synthetic^{3–5} and biological communities.⁶

From this diverse collection of compounds, the amphidinolide B subfamily possesses some of the most intriguing structural features and biological activity. Kobayashi and coworkers reported the isolation of the 26-membered macrolide (amphidinolide B) from the dinoflagellate *Amphidinium* sp. in small amounts (Figure 1).^{2b} The planar original structure was proposed as compound 1. Subsequent reisolation by Shimizu and co-workers as well as structure determination through X-ray crystallographic analysis by Clardy and co-workers provided the relative stereochemistry of amphidinolide B (which was renamed amphidinolide B_1) as compound 2.^{2c} In addition, the location of the methyl moiety of the dienyl system was reassigned to the C15 position. Absolute configuration of 2 was later established via degradation.^{2d} Shimizu and co-workers also reported the isolation of two related members of this family, amphidinolide B₂ (3) and $B_3(4)$, and proposed their structures based on analogy to 2 and comparison of the NMR spectra. More recently, Kobayashi and co-workers reported the isolation of additional members of this subfamily, amphidinolide B_4 and B_5 (5 and 6)^{2e} as well as amphidinolides B_6 and B_7 (not shown).^{2f} Structurally related amphidinolides G and H [e.g., amphidinolide H_1 (7) and amphidinolide $G_1(8)$] have also been reported.^{2p-r} In particular, amphidinolide B_1 (2) has proven to be the most cytotoxic

member of the amphidinolide family, demonstrating impressive potency in early cancer screening [IC₅₀ levels: L1210 murine leukemia cell line (0.14 ng/mL),^{2b} human colon tumor HCT 116 cell line (0.12 μ g/mL),^{2c} and KB cancer cell line (4.2 ng/mL)].^{1f}

In addition to the intriguing biological activity, macrolide 2 has a compelling architecture with nine stereogenic centers embedded within a 26-membered macrocycle including a reactive allylic epoxide moiety at C_6-C_9 and a highly substituted diene moiety at $C_{13}-C_{15}$. Consequently, this target 2 has attracted considerable synthetic attention from numerous laboratories⁷ including our own.⁸⁻¹⁰ In 2008, we reported the first total syntheses of amphidinolide B_1 (2) as well as the proposed structure of amphidinolide B_2 (3) which we established to be incorrect.⁴ Subsequent to our efforts, Fürstner and co-workers published their synthesis of 2 in 2009⁵ and more recently Nishiyama and co-workers completed their total synthesis in 2012.¹¹ It should be noted that Fürstner reported the first syntheses of related amphidinolides G and H in 2007.^{3aa} Herein, we disclose a full account of our work and the biological evaluation of synthesized analogues of amphidinolide B.

RESULTS AND DISCUSSION

Our initial retrosynthetic strategy is shown in Scheme 1. We intended to install the fragile allylic epoxide moiety at a late stage from the corresponding C_7 – C_9 enone motif. Macrolactonization of the corresponding seco acid 9 under Mitsunobu conditions¹² could form the key 26-membered ring system. The C_8 – C_9 bond

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would be accessible from a HWE olefination with the keto phosphonate 11 and aldehyde 10. The C_{18} stereogenic center could be constructed by a chelation-controlled aldol condensation between aldehyde 12 and α -benzyloxy ketone 13. The aldehyde 12 would be derived from the styrenyl compound 14 through a regioselective oxidative cleavage. The diene could be accessed from the coupling of an allylic metallo species such as 15 and the methyl ketone 16.

Synthesis of the nucleophilic components 24 and 25 and styrenyl compound 16 are shown in Scheme 2. The methyl ketone 16 could be prepared in four steps from L-lactic acid (17). Using Seebach chemistry,¹³ stereochemistry originally contained in the hydroxyl acid 17 was transferred cleanly to the lactone 20 with high levels of diastereoselecitivity. Treatment of the lactone with MeLi yielded the α -hydroxy methyl ketone which was protected as its TES ether 16. For the nucleophilic component 15, we envisioned multiple options including phosphonate 25 and Peterson-type approaches (e.g., 24). Starting from the

Scheme 1. Initial Retrosynthetic Analysis of Amphidinolide B₁



Scheme 2. Synthesis of Diene Precursors



commercially available oxazolidinone **21** and known iodide **22**,¹⁴ Evans alkylation followed by reduction and protection generated

the allyl silane 24. Bromination at C_{14} using NBS followed by Arbuzov reaction generated the allyl phosphonate 25 in 55% yield over two steps.

With the coupling partners in hand, we envisioned multiple options for their combination (Scheme 3). One of the initial



approaches we explored was a phosphonate olefination strategy. Unfortunately, using a range of bases (e.g., *n*-BuLi, *t*-BuLi, *s*-BuLi, KHMDS) we were unable to facilitate the olefination. An alternative strategy involving a Peterson olefination could arrive at the same target **26**. Anions derived from allyl trimethylsilanes can attack a carbonyl electrophile through either α -position or γ -position.¹⁵ Using conditions developed by Magnus and coworkers,¹⁶ we observed exclusive formation of the vinylsilane product **27** derived from reactivity in the γ position. A modified procedure using MgBr₂¹⁷ failed to override this preference.

While the vinyl silane product 27 is in theory still a viable intermediate in the synthesis of the diene 26, we were unable to facilitate the subsequent conversion to the target diene.¹⁸ We also explored a Sakurai-type approach¹⁹ to C-C bond formation using allyl silane 24. Treatment of 24 and 16 under a range of Lewis acidic conditions (e.g., TMSOTf, BF3·Et2O, Me2AlCl, AlMe₃, SnCl₄) did not provide any of the desired coupled material 28. Fortunately, treatment with freshly distilled $TiCl_4$ did facilitate the C_{14} - C_{15} bond construction in up to 65% yield and 6:1 dr (stereochemistry at C_{15} undetermined). While effective, this reaction did prove to be scale dependent (typically providing 50-65% yield at 0.65 mmol scale, but 20-30% yield at 2.0 mmol scale). Despite considerable effort to ascertain the nature of the scale dependence, we were unable to fully address this shortcoming. Consequently, this transformation was often run in parallel batches to bring through sufficient amounts of material. Subsequent elimination of the tertiary alcohol 28 was best facilitated using SOCl₂ and pyridine in toluene at -78 °C to yield the desired diene in near-quantitative yield as a 2.2:1 ratio of diene regioisomers (rr) (26:29). The diene rr was not impacted by use of a single C_{15} stereoisomer of alcohol 28. Use of alternate conditions (e.g., MsCl, POCl₃, Tf₂O, Burgess reagent, Martin's sulfurane) led to no product formation or inferior levels of regioselectivity. While these diene regioisomers 26 and 29 were not readily separable by column chromatography, removal of the silyl protecting group provided the corresponding regioisomeric diols 30 and 31 that were separable.

The functionalization of the polyene is shown in Scheme 4. Mitsunobu-type conversion of the C10 alcohol into the corresponding nitrile followed by silyl protection yielded 14. Next, we required the selective functionalization of the $C_{18}-C_{19}$ alkene in the presence of the less sterically hindered 1,1disubstituted alkene at C_{13} and the more electron rich alkene at C₁₄-C₁₅. We had hoped that regioselective dihydroxylation at C_{18} – C_{19} could be possible by exploiting a π -stacking interaction between the phenyl ring and aromatic systems present in Sharpless ligands. While this sort of phenomenon has been used previously to explain enhanced enantioselectivity²⁰ and even diastereoselectivity²¹ in Sharpless asymmetric dihydroxylation, we were unaware of prior examples to exploit purely a regioselective directing effect. To our delight, the AD Mix β^* [the * denotion represents higher percentages of K₂OsO₄·2H₂O and (DHQD)₂PHAL as compared to the commercial amounts] provided excellent regioselectivity for the desired location. As hypothesized, use of standard OsO₄/NMO conditions provided a complex mixture of products. Interestingly, AD Mix α^* also proved to be a poor reagent of this transformation. While we can rationalize the poor outcome of the ligandless system, a rational for the reactivity differences between the pseudoenantiomeric ligand systems is less obvious. One possible explanation may be that the C₁₆ stereocenter exerts a pronounced influence on the conformation of the neighboring alkenes, resulting in a mismatched interaction between the (DHQ)₂PHAL ligand in AD mix α and the desired alkene. It is also clear that a conformational change occurs between the unconjugated system 33 and the conjugated system 14, dihydroxylation of polyene 33 results in a regioselective oxidation at $C_{18}-C_{19}$; however, the reaction does not show any diastereoselectivity. Periodate cleavage under standard conditions yielded the aldehydes 12 and 35.

Synthesis of the eastern subunit⁴ and southern fragment are shown in Scheme 5. Evans *syn* glycolate reaction between oxazolidinone 36^{22} and readily available aldehyde 37^8 provided the desired product in reasonable diastereoselectivity (7:1 dr)



Scheme 5. Synthesis of Eastern and Southern Subunits



and good yield. After TES silvlation, subsequent conversion to the thioester with lithium thiolate followed by cuprate addition yielded the methyl ketone 13. This strategy for synthesis of the $C_{19}-C_{26}$ subunit has been subsequently exploited by Fürstner

and co-workers.^{3aa,5} The carboxylic acid **11** was available in four straightforward steps from the commercially available ester **38**.

With the subunits now constructed, our efforts shifted toward their combination. We were particularly focused on addressing the diastereoselectivity in the methyl ketone aldol reaction between **12** and **13**. Prior to our entry into the field, Pattenden^{7e} and Kobayashi^{7k} had independently explored related coupling strategies with limited success (Scheme 6). In both cases, poor



diastereoselectivity was observed (3:2 dr) as well as low chemical yield (50% in Kobayashi's case, no chemical yield reported in Pattenden's example). Stereocontrolling models for β -silyloxy aldehydes such as 40 and 44 have been proposed;²³ however, it is unclear if the tertiary nature of the C₁₆ silyl ether is compatible with that analysis. In addition, models have been developed for exploiting the stereochemical controlling nature of α -chiral ketones (albeit primarily on ethyl ketone substrates). We had hypothesized the poor stereocontrol in these pioneering examples by Pattenden and Kobayashi could be attributed to the absence of a chelating protecting group at C_{21} , which rigidifies the facial selectivity of the enolate in the aldol transition state. Compelling evidence of the potential for this strategy can be found in Chakraborty's related work toward amphidinolides G and H (which lack the C₁₆ hydroxyl functionality).²⁴ Pioneering precedent for the stereocontrolling ability of α -oxy enolates had been reported by Paterson,²⁵ Heathcock,²⁶ and Masamune;²⁷ however, the majority of the examples were on ethyl ketones. Aldol reactions using methyl ketones have been reported to proceed through both chair and boat transition states, which could lead to an erosion in stereoinduction.²⁸ White,²⁹ Trost,³⁰ and Evans³¹ reported some of the earliest successful examples in this area. Other laboratories have subsequently explored this transformation with varying degrees of stereocontrol.³²

Our exploration into the diastereoselective aldol reaction is detailed in Scheme 7. We initially employed the unconjugated diene **35** as a model for the actual system. To our considerable disappointment, this substrate **35** performed poorly in the transformation, providing low diastereoselectivity (2:1 dr) in modest chemical yield (60%) (eq 1). Similar results were observed with the achiral aldehyde 3,3-dimethylbutanal (2–3:1 dr) (eq 2).



Scheme 7. Methyl Ketone Aldol Reaction

Undeterred by these discouraging model studies, we next studied the conjugated diene system 12 (eq 3). We were delighted to see that this transformation proceeded with high levels of stereocontrol; we only observed a single diastereomer in 69% yield. The configuration of the newly established stereocenter was confirmed by advanced Mosher ester method.³³ The subtle nature of the controlling influences of this aldol reaction warrants further comment. We are hesitant to put forth a model for the controlling influences of this transformation; however, it is important to note that both the trisubstituted alkene at $C_{14}-C_{15}$ and the chelating protecting group at C21 appear to work in concert to influence the stereochemical outcome of the aldol reaction. The C₁₆ silyl ether does not appear to exert considerable influence as this chelation strategy has subsequently proven useful in amphidinolide G/H series by both Fürstner^{3aa,5} and Zhao³⁴ which does not contain the C₁₆ alcohol moiety.

The construction of the macrocyclization precursor is shown in Scheme 8. Silylation of the C₁₈ alcohol proceeded smoothly with TBSOTf and a large excess of triethylamine.³⁵ Interestingly, use of near stoichiometric amounts of an amine base [e.g., 2,6-lutidine (1.2 equiv)] led to elimination (~20%) and alkene isomerization (~10%). Regioselective reduction of the C₉ nitrile was accomplished with DIBAL-H. Horner-Wadsworth-Emmons olefination of the resultant aldehyde using Ba(OH)₂³⁶ yielded the desired α,β -unsaturated ketone in good yield and excellent E/Z selectivity. Alternate conditions on related systems (DBU, LiCl or KHMDS, THF) were less effective for this olefination.³⁷ Subsequent selective cleavage of the C₂₅ TES ether produced the key Mitsunobu macrolactonization¹² precursor. Unfortunately, this substrate was resistant to all efforts to facilitate the macrocycle formation under these sterochemically invertive conditions. Scheme 8. Unsuccessful Macrocyclization Strategy



Based on our frustrations with facilitating a successful Mitsunobu macrolactonization, we decided a reorganization of our strategy for macrocyclic formation was needed. Specifically, we turned to inverting the C_{25} stereocenter prior to coupling, forming the C_1 ester bond in an intermolecular fashion and constructing the C_8-C_9 alkene through an intramolecular pathway. Toward this end, synthesis of a modified methyl ketone 54 was necessary (Scheme 9). Selective TES removal could be accomplished under aqueous acidic conditions in excellent yield. Martin's modified Mitsunobu conditions proceeded smoothly to give the inverted C_{25} ester. Finally, saponification of the PNB moiety with $Ba(OH)_2 \cdot 8H_2O$ and methanol followed by TMS protection revealed the coupling partner 54.

Synthesis of the modified aldehyde coupling partner is illustrated in Scheme 10. Conversion of the C₉ nitrile to its corresponding acetate was accomplished via DIBAL-H reduction to the aldehyde followed by NaBH₄ reduction to the alcohol and acylation. Regioselective functionalization at C₁₈ was again accomplished using Sharpless AD conditions followed by periodate cleavage to reveal the C₁₈ aldehyde.

While the original route to the aldehydes **12** and **57** was effective, it suffered from logistical impracticalities; notably, the titanium-mediated coupling of the allyl silane proved to be a scale-dependent reaction and the thionyl chloride elimination step gave a mixture of olefin products. Consequently, we sought to develop a more scalable approach to this subunit (Scheme 11).

Scheme 9. Second-Generation Synthesis of Eastern Subunit



Scheme 10. Modification of the Western Subunit



Starting from the commercially available Oppolzer sultam derivative 59, cuprate addition in accord with conditions developed by Paquette³⁸ cleanly generated the key C_{11} stereocenter in compound 60. After functional group manipulation to provide aldehyde 61, we were gratified to find that key sequential Wittig/Horner-Wadsworth-Emmons (HWE) reactions cleanly introduced the triene 63 as a single stereoisomer. After DIBAL-H reduction, the resulting trienyl alcohol proved remarkably reactive to Sharpless asymmetric epoxidation conditions (proceeding at -78 °C to -50 °C) to provide epoxide 64. In fact, if this reaction was performed at higher temperatures (e.g., -20 °C), another product was observed that appeared to be the result of a 1,2-alkyl shift to produce an aldehyde.³⁹ Subsequent Red-Al reduction provided the diol 65. Finally, protecting group exchange followed by 1° TES deprotection and oxidation provided the C_9-C_{18} subunit 57. The high efficiency of this route (11.5% overall yield) provided us with access to gram quantities of aldehyde 57.

With these modified coupling partners in hand, we revisited our aldol coupling strategy (Scheme 12). We were pleased to observe that similar levels of diastereoselectivity continued to be observed using the LDA conditions (62% yield, >20:1 dr). Silylation of the C₁₈ alcohol proved significantly more complicated than expected. Use of TBSOTf led to significant exchange of the C₂₅ TMS ether to the corresponding TBS ether. Use of TESOTf minimized this problem, providing a 95% yield of a 5:1 ratio of the C₂₅ TMS to C₂₅ TES. Both of these

50₂ 59 OPMB Mg, CuBr•DMS LiCl, TMSCl, 97% Ċ 58 60 **ÓPMB** DIBAL-H 90% TBSCI, imid. 98%; DDQ, pH 7, 93%; DMP, CH2Cl2, 71% CO₂Et .CHO 62 ⊕ PPh₂ PhMe, 94%; OTBS (EtO)₂P(O)CH₂CO₂Et NaH, DME, reflux OTBS 63 72% 61 DIBAL-H, 83%; (+)-DIPT Ti(Oi-Pr)₄, TBHP -78 to -50 °C, 80% OH Red-Al, THE 0°C, 70% OTBS OP. 64 64 CIC(O)CCl₃, pyr; CSA, CH₂Cl₂ / EtOH (1:1); Ac₂O, pyr, CH₂Cl₂ 75% (3 steps) 65 P₁ = H, P₂ = TBS **66** $P_1 = TCA, P_2 = Ac$ NH₄OH CH₂Cl₂ / EtOH (1:1); TESOTI, Et₃N, -78 °C 98% (2 steps) OTES **TESO** THF / HOAc / H₂O (8:8:1), 0°C; DMP, CH₂Cl₂ 74% (2 steps) OAc .OAc 57 67

Scheme 11. Second Generation Approach to the $C_0 - C_{10}$

Western Fragment of Amphidinolide B₁

compounds were productive intermediates and could be converted to the corresponding C_{25} alcohol using HOAc/ THF/H₂O conditions. Intermolecular Yamaguchi esterification provided the phosphonate 71. Next, removal of the C₉ acetate was cleanly accomplished using Ba(OH)₂•8H₂O. Subsequent oxidation using TPAP provided the aldehyde cyclization precursor 73. Interestingly, this compound proved to be surprisingly reactive—leading to spontaneous macrocyclization under the TPAP conditions. Ba(OH)₂·8H₂O was added to drive this transformation to completion with an 85% overall yield from the alcohol 72.

With the macrocycle in hand, the remaining challenges to be addressed were the successful incorporation of the C_6-C_9 allylic epoxide and global deprotection (Scheme 13). CBS reduction⁴⁰ proceeded smoothly to provide the desired allylic alcohol 75 in excellent diastereoselectivity. Subsequent Sharpless asymmetric epoxidation⁴¹ provided the desired epoxide 76 in 10:1 dr. Incorporation of the alkene could be best accomplished by inversion at C_7 to provide the selenide 78⁴² followed by TPAP

Scheme 12. Spontaneous Macrocyclization Strategy



mediated oxidation⁴³ to the selenoxide and syn elimination to yield the allylic epoxide as a single olefin geometry. Despite compelling precedent from Nicolaou and co-workers for a related deprotection in the synthesis of amphidinolide N,^{2v} we were unable to effect PMB removal using DDQ under buffered aqueous or anhydrous conditions. Based on this unfortunate result, it became clear that an alternate C₂₁ protecting group strategy was necessary to complete the total synthesis of amphidinolide B₁.

We initially considered a 3,4-dimethoxybenzyl (DMB) protecting group as a viable alternative to the C₂₁ PMB group as DMB groups are more readily oxidized by DDQ (Scheme 14). We synthesized the necessary $C_{19}-C_{26}$ subunit via an analogous route as described previously. Interestingly, aldol reaction with the C₁₈ aldehyde 82 proceeded smoothly but in significantly lower diastereoselectivity (4:1 dr). We are unsure as to the cause of this difference between the PMB and DMB series. Next, we explored the possibility of deprotecting the C₂₁ DMB group using DDQ. Interestingly, treatment of 83 with DDQ under buffer aqueous conditions led to formation of the acetal 86 in modest yield, but as a single diastereomer. This product is likely the result of initial oxidation to the oxonium ion followed by nucleophilic attack by the C22 OTES ether. Subsequent silyl migration to the C₁₈ position would produce the observed product 86. Despite this encouraging early result, we were unable to increase the chemical yield of this transformation through variation of oxidant or reaction conditions. Consequently, this approach was abandoned.

Given our setbacks with benzyl-derived protection at C_{21} , we revisited our central strategy for stereocontrol at C_{18} through the C_{21} chelation model described previously (Scheme 15). Traditionally, silyl protecting groups are not viewed as moieties that participate in chelation;⁴⁴ however, it is important to acknowledge work by Heathcock,⁴⁵ Eliel and Frye,⁴⁶ Willard⁴⁷ and Evans⁴⁸ which showed that silyl chelation is feasible under certain conditions. We were particularly drawn to Heathcock's work in the area in which TMEDA was a key additive in their chelation-controlled aldol reaction (eq 1). Alternatively, a dipole

Scheme 13. Incorporation of the Allylic Epoxide Moiety



minimization stereocontrol model produced the alternate facial attack on the aldehyde (eq 2). Silyl protection at C_{21} would



Scheme 14. C₂₁ Dimethoxybenzyl Protection Series

Scheme 15. Heathcock's Pioneering Stereoselective Aldol Work



simplify the endgame deprotection sequence and circumvent the problematic DDQ deprotection strategy.

The synthesis of the C_{21} silyl methyl ketone **98** is shown in Scheme 16. One attractive advantage to this strategy is that it greatly simplifies the synthesis of the methyl ketone as differential protection at C_{21} and C_{22} is no longer needed. Olefination of aldehyde **37** provided the $\alpha_{,\beta}$ -unsaturated enone **93**. Sharpless dihydroxylation under buffered conditions yielded the diol in good diastereoselectivity. Bis-silyl protection using TESOTf cleanly provided the tris-TES methyl ketone **95**. The C_{25} TES ether could be cleanly removed using HOAc/THF/H₂O conditions followed by inversion and protection as its C_{25} TMS ether.

Exploration of the key aldol reaction on the C_{21} silyl series generated intriguing results (Scheme 17 and Table 1). Initial studies with the C_{25} epimer **95** exhibited poor diastereoselectivity, but good reactivity under the Heathcock's TMEDA conditions (entry 1). It should be noted that the replacement of THF with



Scheme 17. Exploration into Aldol Stereoselectivity



Table 1. Exploration into Aldol Stereoselectivity

entry	substrates	conditions	yield (%) [dr]
1	95, 82	THF, −78 °C	64 [1.1:1 (99:102)]
2	95, 82	Et₂O, −78 °C	<10 [1.5:1 (99:102)]
3	98, 82	THF, −78 °C	67 [1:5 (100:103)]
4	98, 82	THF, −40 °C	68 [1.8:1 (100:103)]
5	98, 5 7	THF, −100 °C	65 [1:8 (101:104)]
6	98, 57	THF, −40 °C	66 [1.2:1 (101:104)]

 Et_2O led to a dramatic decrease in chemical yield (entry 2). Interestingly, use of the required C_{25} stereochemistry for amphidinolide B led to a more stereoselective aldol process (entries 3–6). If the reaction was conducted at -78 °C, the transformation appeared to proceed through a dipole minimization model to yield the 18*R* stereochemistry as the major product (5:1 **103:100**). If the otherwise identical transformation was conducted at -40 °C, the stereoselectivity for the transformation reversed to now favor the 18*S* stereochemistry. This divergent stereocontrol model could be attributed to reversible nature of aldol reactions.⁴⁹ A similar phenomenon was observed with the aldehyde **57** (entries 5 and 6). While unexpected, these results opened the door for the synthesis of both amphidinolide B₁ and B₂.

Synthesis of the macrocycle using the aldol products was accomplished based on close analogy to our C_{21} PMB series (Scheme 18). TES protection at C_{18} using TESCI/DMAP





conditions followed by C_{25} desilylation revealed the C_{25} alcohol. Yamaguchi esterification followed by saponification of the acetate protecting group produced the C_9 alcohol. As employed in our previous macrocyclization, TPAP oxidation produced the reactive aldehyde, which appeared to undergo spontaneous macrocyclization. The cyclization could be driven to completion by addition of Ba(OH)₂·8H₂O for 18S series and LiCl/Hunig's base for the 18*R* series for optimal yields. While the yields for these macrocyclizations were not as high as the C_{21} PMB series, the ability to successfully close the macrocycle was gratifying. Fortuitously, macrocycle **109** produced crystals suitable for X-ray crystallographic analysis, thereby confirming the stereochemical assignment (Figure 2).

With the macrocycle in hand and the stereochemistry firmly established, the remaining hurdles for completing the synthesis



Figure 2. ORTEP Representation of Macrocycle 109.

were incorporation of the allylic epoxide moiety and global deprotection. We first advanced the 18S stereoisomer, which would lead to the synthesis of amphidinolide B_2 (Scheme 19). CBS reduction produced the allylic alcohol in good diastereoselectivity. Although epoxidation under tartrate-mediated Sharpless conditions as used previously proved ineffective, titanium-mediated epoxidation (in the absence of tartrate ligand) produced the epoxide alcohols in modest diastereoselectivity (2:1 dr, 74% yield). Given the increased steric demands of the Ti-tartate complex, we attribute the reactivity difference between the C₂₀ OPMB series and the C₂₀ OTES series to a conformational change to the macrocycle, which increases the steric hindrance surrounding the allylic alcohol moiety. While we cannot rigorously assign the stereochemistry of the newly created stereocenters, they were assigned based on literature precedent.^{40,50} Incorporation of the selenide followed by TPAP-mediated oxidation and syn elimination provided the allyl epoxide 116. Unfortuantely, we were again unable to effect global desilylation under a range of conditions (e.g., TAS-F or HF·pyr). Fortunately, the ordering of the final events could be altered (global deprotection followed by syn elimination using TMSOOTMS for selenide oxidation) to reveal the proposed structure for amphidinolide B_2 (3). It should be noted that the use of TMSOOTMS has not been previously reported for selenide oxidation/elimination. H₂O₂ was ineffective in this transformation, likely due to unwanted Baeyer-Villigertype oxidation of the C₂₀ ketone. To our surprise, the spectral data did not match the literature values for amphidinolide B₂. We had presumed that the incorrect epoxide had been utilized. Consequently, we repeated the analogous endgame, but this product 114 also did not match the natural product. It became clear that the assignment of amphidinolide B₂ was incorrect. Comparison of the synthesized material with the literature data revealed a large chemical shift difference for H_{14} [isolation data: 5.93 (bs), synthetic 3: 6.06 (bs), synthetic 114: 6.08 (bs)] and H_{19a,b} [isolation data: 3.09 (dd, *J* = 2.3, 8.8 Hz, 1H) and 2.69 (dd, *J* = 8.6, 17.7 Hz, 1H), synthetic 3: 3.05 (m) and 2.48 (dd, *J* = 8.0, 17.0 Hz), synthetic 114: 2.90 (dd, J = 9.9, 17.1 Hz) and 2.45 (m)]. Careful inspection of the structure elucidation paper^{2c} revealed that while amphidinolide B1 was assigned by crystallographic analysis, amphidinolide B2 was assigned based on analogy and comparison of the NMR spectra. We hypothesize that the actual structure of amphidinolide B2 likely contains the opposite stereochemistry at C_{16} (e.g., 117), thereby maintaining a syn relationship between the two alcohols at C_{16} and C_{18} .

The successful completion of amphidinolide B_1 is shown in Scheme 20. Starting from the macrocycle **109**, CBS reduction provided the allylic alcohol **118** in 74% yield (3.5:1 dr). Titanium-mediated epoxidation of the alkene was high yielding, Scheme 19. Synthesis of Proposed Structure for Amphidinolide B₂



but unselective (1:1.5 dr). Incorporation of the aryl selenide using previously described conditions followed by global deprotection and selenide oxidation/*syn* elimination yielded amphidinolide B₁ (**2**). Comparison of the synthetic material with literature values showed excellent agreement (¹H NMR, ¹³C NMR, $[\alpha]_D$). For analogue studies, we also carried the epimeric epoxide series onto 8,9-*epi*-amphidinolide B₁ (**121**).

To assess the cytotoxic effects of the proposed structure of amphidinolide B₂ on human solid and blood cancer cells, cell viability assays were conducted using a nine cancer cell line panel.58 Compound 3 differentially reduced cell viabilities at 0.1 μ M in human DU145 prostate cancer, MDA-MB-435 breast cancer, OCI-LY3 lymphoma, K562 CML, MOLT-4 ALL, Reh ALL, U266 myeloma, KG1a AML and HL60 AML cells (Table 2). To further understand these antitumor activities of amphidinolide B_{22} we determined IC₅₀ values = 36.4 ± 2.9 nM, 94.5 ± 8.0 nM, 3.3 ± 0.9 nM and 7.4 ± 0.6 nM against DU145, MDA-MB-435, KG1a and HL60 AML cancer cells, respectively (Table 3). Next, compared the potency of compound 3 to $C_{8,9}$ epimer 114 and C₁₈ epimer 121 in DU145 prostate cancer cells, compound 3 exhibited over 12-fold increase in biological activity (Figure 3). These findings suggest that the stereochemistry of the epoxide plays an important role in eliciting potent anticancer effects.

Cell viability assays were carried out as described in the Experimental Section. Human DU145 prostate cancer, MDA-MB-435 breast cancer, OCI-LY3 lymphoma, K562 CML,

MOLT-4 ALL, Reh ALL, U266 myeloma, KG1a AML and HL60 AML cells were seeded in 96-well plates (5000/well for DU145 and MDA-MB-435, 10000/well for OCI-LY3, K562, MOLT-4, Reh, U266, KG1a and HL60), incubated overnight at 37 °C in 5% (v/v) CO₂ and exposed to 0.1 μ M of compound 3 for 72 h. Absorbance was measured at 490 nm using an automated ELISA plate reader. Each experiment was performed in quadruplicate. Data are mean ± SD.

IC₅₀ values of compound **3** were determined in human DU145 prostate cancer, MDA-MB-435 breast cancer, KG1a AML and HL60 AML cells. Cells were seeded in 96-well plates (5000/well for DU145 and MDA-MB-435, 10000/well for KG1a and HL60), incubated overnight at 37 °C in 5% (v/v) CO₂ and exposed to compound **3** in a dose-dependent manner for 72 h. Absorbance was measured at 490 nm using an automated ELISA plate reader. Each experiment was performed in quadruplicate. Data are mean \pm SD.

CONCLUSION

In summary, the first total synthesis of amphidinolide B_1 and the proposed structure for amphidinolide B_2 has been achieved. In our initial approach to this family, we utilized a chelationcontrolled aldol reaction to provide excellent stereoselectivity at C_{18} . Unfortunately, late-stage removal of the PMB group proved not feasible in our hands. Consequently, a revised approach utilizing a dipole-minimized aldol provided access to a stereoselective aldol product. The stereoselectivity in this key aldol Scheme 20. Synthesis of Amphidinolide B_1 and 8,9-epi-Amphidinolide B_1



process could be controlled by variation of the temperature. Other highlights of the synthetic route include two non-metalcatalyzed methods to construct the $C_{13}-C_{15}$ diene, a spontaneous HWE macrocyclization strategy, and a novel late-stage epoxidation/elimination strategy to incorporate the sensitive C_6-C_9 allylic epoxide moiety.

The proposed structure for amphidinolide B_2 displays potent and stereoselective antitumor activities against human solid and blood tumor cells at low nanomolar concentrations that include highly aggressive and metastatic prostate and breast cancer cells. In contrast, $C_{8,9}$ -epimer and C_{18} epimer exhibit an over 12-fold decrease in antitumor activity in comparison. The stereochemistry of the epoxide plays a key role in the anticancer effects.

EXPERIMENTAL SECTION

General Methods. Infrared spectra were recorded neat unless otherwise indicated and are reported in cm⁻¹. ¹H NMR spectra were recorded in deuterated solvents and are reported in ppm relative to tetramethylsilane and referenced internally to the residually protonated solvent. ¹³C NMR spectra were recorded in deuterated solvents and are reported in ppm relative to tetramethylsilane and referenced internally



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Figure 3. Comparison of Synthetic Analogues for Biological Activity.

to the residually protonated solvent. HRMS data was collected using a TOF mass spectrometer.

Routine monitoring of reactions was performed using EM Science DC-Alufolien silica gel, aluminum-backed TLC plates. Flash chromatography was performed with the indicated eluents on EM Science Gedurian 230–400 mesh silica gel.

Air- and/or moisture-sensitive reactions were performed under usual inert atmosphere conditions. Reactions requiring anhydrous conditions were performed under a blanket of argon, in glassware dried in an oven at 120 $^{\circ}$ C or by flame, then cooled under argon. Dry THF and DCM were obtained via a solvent purification system. All other solvents and commercially available reagents were either purified via literature procedures or used without further purification.



Lactone 20. To a solution of diisopropylamine (1.67 g, 2.3 mL, 16.5 mmol) in THF (7.6 mL) at -78 °C was added *n*-BuLi (6.6 mL,

Table 2. Effects of Com	pound 3 on Viabilitie	s of Human Cancer	: Cells ((% control)
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DU145	MDA-MB-435	OCI-LY3	K562	MOLT-4	Reh	U266	KG1a	HL60
31 ± 5	54 ± 10	0	67 ± 6	47 ± 4	82 ± 5	76 ± 5	10 ± 2	0

16.5 mmol, 2.5 M in hexanes) dropwise. After 10 min, the white slurry was warmed to -10 °C for 30 min. Then, the solution was cooled to -78 °C, and another 58 mL of THF was added slowly to the above solution. Next, a solution of Seebach lactone 18⁵¹ (2.37 g, 15.0 mmol) in THF (15 mL) was added dropwise to the above solution. After 20 min, a solution of cinnamyl bromide 19 (4.43 g, 3.33 mL, 22.5 mmol) in THF (10 mL) was added dropwise. After 30 min, the reaction was warmed slowly to -10 °C over 2 h. After 5 min, the reaction was quenched with satd aq NH₄Cl (30 mL) and warmed to rt. After 10 min, the organic layer was separated, and the aqueous layer was extracted with Et₂O $(3 \times 30 \text{ mL})$. The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2-10% EtOAc/ hexanes, to give 20 (3.74 g, 13.7 mmol, 91%) as a white solid: $[\alpha]^{23}_{D}$ +45.5 (c 1.21, CHCl₃); IR (thin film) 3027, 2963, 1796, 1484, 1173, 1138, 970, 692 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.28–7.42 (m, 5H), 6.58 (d, J = 15.6 Hz, 1H), 6.23 (dt, J = 15.6, 6.0 Hz, 1H), 5.24 (s, 1H), 2.72 (dd, J = 6.0, 10.8 Hz, 1H), 2.62 (dd, J = 6.0, 10.8 Hz, 1H), 1.51 (s, 3H), 0.98 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 175.5, 137.1, 135.3, 128.8, 127.9, 126.5, 122.4, 108.9, 80.2, 40.3, 34.8, 23.5, 23.1; HRMS (FAB+) calcd for $C_{17}H_{23}O_3$ (M + H) 275.1647, found 275.1650.



Methyl Ketone 16. To a solution of lactone acetal 20 (1.37 g, 5.0 mmol) in THF (100 mL) at -78 °C was added MeLi (4.64 mL, 5.6 mmol, 1.2 M in Et₂O) via syringe pump. After 15 min, the resulting solution was warmed to rt. After an additional 10 min, the solvent was concentrated in vacuo, diluted with Et₂O (50 mL), filtrated through a plug of Celite, and concentrated again in vacuo. The yellow oil was purified by chromatography over silica gel, eluting with 10-30% EtOAc/hexanes, to give a ketone intermediate (0.78 g, 3.85 mmol, 77%) as a colorless oil. To a solution of the above intermediate (340 mg, 1.66 mmol) in THF (8.3 mL) at -78 °C was added 2,6-lutidine (0.27 g, 0.29 mL, 2.49 mmol). After 5 min, TESOTf (0.53 g, 0.45 mL, 1.99 mmol) was added. After 30 min, the reaction was warmed to rt. After 5 min, the reaction was quenched with satd aq NH₄Cl (10 mL). After 10 min, the organic layer was separated, and the aqueous layer was extracted with Et₂O (3 \times 15 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2-6% EtOAc/hexanes, to give 16 (448 mg, 1.41 mmol, 88%) as a colorless oil: $[\alpha]^{23}_{D}$ +0.7 (*c* 0.70, CHCl₃); IR (thin film) 3027, 2955, 2916, 2876, 1718, 1457, 1415, 1350, 1239, 1166, 1131, 1016, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.20–7.33 (m, 5H), 6.38 (d, J = 15.9 Hz, 1H), 6.09-6.20 (m, 1H), 2.40-2.63 (m, 2H), 2.21 (s, 3H), 1.38 (s, 3H), 1.01 (t, J = 7.5 Hz, 9H), 0.68 (q, J = 7.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 214.2, 137.8, 133.6, 128.9, 127.6, 126.5, 125.1, 83.0, 45.2, 26.2, 25.4, 7.6, 7.1; HRMS (FAB+) calcd for C₁₉H₃₁O₂Si (M + H) 319.2093, found 319.2094.



AllyIsilane 23. To a solution of NaHMDS (5.6 mL, 5.6 mmol, 1.0 M in THF) at -78 °C was added a solution of imide 21 (1.09 g, 4.67 mmol) in THF (4.6 mL). After 20 min, a solution of allyl iodide 22^{52} (3.6 g, 14 mmol) in THF (3 mL) was added via cannula. After 10 min, the reaction was warmed slowly to 10 °C over 4 h and quenched with satd aq NH₄Cl (25 mL). After 10 min at rt, the organic layer was separated and the aqueous layer was extracted with Et₂O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 4–15% EtOAc/hexanes, to give 23 (1.46 g, 4.06 mmol, 87%) as a colorless oil: $[\alpha]^{23}_{D}$ +30.6 (*c* 0.66, CHCl₃); IR (thin film) 2954, 1780, 1699, 1632, 1454, 1385, 1349, 1246, 1207, 1101, 851, 701 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ

7.20–7.35 (m, 5H), 4.66–4.69 (m, 1H), 4.69 (s, 1H), 4.64 (s, 1H), 4.14–4.19 (m, 2H), 4.06 (q, J = 6.9 Hz, 1H), 3.26 (dd, J = 3.3, 13.2 Hz, 1H), 2.71 (dd, J = 3.9, 13.2 Hz, 1H), 2.52 (dd, J = 7.5, 14.4 Hz, 1H), 2.04 (dd, J = 7.5, 14.4 Hz, 1H), 1.59 (d, J = 3.6 Hz, 2H), 1.17 (d, J = 6.9 Hz), 0.04 (s. 9H); ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 153.5, 145.1, 135.8, 129.9, 129.3, 127.7, 109.7, 66.3, 55.7, 42.6, 38.4, 36.2, 26.8, 17.4, -1.0; HRMS (FAB+) calcd for C₂₀H₃₀NO₃Si (M + H) 360.1995, found 360.1989.



Allylsilane 24. To a solution of 23 (1.88 g, 5.23 mmol) in THF (35 mL) at 0 °C were added sequentially MeOH (0.20 g, 0.25 mL, 6.28 mmol) followed by LiBH₄ (3.14 mL, 6.28 mmol, 2.0 M in THF). After 2 h at 0 °C, the reaction was warmed to rt. After 2 h, the reaction was quenched with satd aq sodium tartrate (25 mL). After stirring for 20 min, the organic layer was separated, and the aqueous layer was extracted with Et_2O (3 × 25 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15-25% Et₂O/pentane, to give free alcohol SI-1 (0.96 g, 5.18 mmol, 99%) as a colorless oil: $[\alpha]_{D}^{23}$ (+) 9.4 (c 0.84, CHCl₃); IR (thin film) 3337 (br), 3072, 2954, 2916, 1631, 1455, 1419, 1248, 1036, 849 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5,60 (d, J = 15.6 Hz, 2H), 3.44-3.52 (m, 2H), 2.03-2.10 (m, 1H), 1.77-1.89 (m, 2H), 1.54 (s, 2H), 1.41 (t, OH), 0.91 (d, J = 6.3 Hz, 3H), 0.04 (s, 9H); ¹³C NMR (75 MHz, $CDCl_3$) δ 146.6, 109.2, 68.7, 43.1, 34.2, 26.7, 17.2, -0.9; HRMS (FAB+) calcd for $C_{10}H_{22}OSi$ (M) 186.1440, found 186.1447.



TIPS Ether 24. To a solution of alcohol intermediate **SI-1** (308 mg, 1.65 mmol) in CH₂Cl₂ at -78 °C were added sequentially Et₃N (0.25 g, 0.34 mL, 2.48 mmol) and TIPSOTf (607 mg, 0.53 mL, 1.98 mmol). After 20 min, the reaction was warmed to rt. After 5 min, the reaction was quenched with satd aq NH₄Cl (25 mL). After 5 min, the organic layer was separated, and the aqueous layer was extracted with Et₂O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 1% EtOAc/hexanes, to give **24** (560 mg, 1.64 mmol, 99%) as a colorless oil: $[α]^{23}_{D}$ –4.9 (*c* 0.98, CHCl₃); IR (thin film) 3072, 2944, 2866, 1631, 1463, 1387, 1248, 1103, 1068, 878, 849, 796, 681 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.57–4.58 (m, 1H), 4.54 (s, 1H), 3.47–3.51 (m, 2H), 2.18 (dd, *J* = 5.4, 13.8, 1H), 1.78–1.82 (m, 1H), 1.63–1.71 (m, 1H), 1.51 (s, 2H), 1.04–1.08 (m, 21H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.20 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 146.6, 108.8, 68.9, 42.7, 34.6, 26.7, 18.5, 17.0, 12.4, –0.9; HRMS (FAB+) calcd for C₁₉H₄₂OSi₂ (M⁺) 342.2774, found 342.2772.



Tertiary Alcohol 28. To a solution of allylsilane 24 (214 mg, 0.62 mmol) and ketone 16 (96 mg, 0.3 mmol) in CH₂Cl₂ (3.0 mL) at -78 °C was added freshly distilled TiCl₄ (117.6 mg, 70 μ L, 0.62 mmol) via microsyringe dropwise. After 15 min, the dark red solution was quenched with satd aq K₂CO₃ (5 mL) and warmed to rt immediately. After 20 min, the organic layer was separated and the aqueous layer was extracted with Et₂O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting carefully with 0.5–2% EtOAc/hexanes, to give 28 (382 mg,

0.19 mmol, 65%) as a colorless oil of a mixture of isomers (dr = 6:1). Major isomer: ¹H NMR (300 MHz, CDCl₃) δ 7.24–7.39 (m, 5H), 6.35–6.44 (m, 2H), 4.93 (s, 1H), 4.84 (s, 1H), 3.57–3.61 (m, 1H), 3.43–3.48 (m, 1H), 2.55–2.60 (m, 2H), 2.35–2.40 (m, 1H), 2.31 (dd, *J* = 5.7, 10.8, 1H), 2.16 (d, *J* = 9.9 Hz, 1H), 2.03 (dd, *J* = 5.7, 10.8 Hz, 1H), 1.85–1.88 (m, 1H), 1.33 (s, 3H), 1.20 (s, 3H), 1.08–1.10 (m, 21H), 0.99 (t, *J* = 6.0 Hz, 9H), 0.94 (d, *J* = 5.1 Hz, 3 H), 0.70 (q, *J* = 6.0 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 146.1, 138.0, 132.5, 128.9, 127.9, 127.4, 126.4, 115.4, 81.9, 77.7, 68.6, 43.3, 42.2, 42.1, 34.8, 23.1, 21.3, 18.5, 17.5, 12.4, 7.7, 7.4; HRMS (FAB+) calcd for C₃₅H₆₄O₃Si₂ (M⁺) 588.4394, found 588.4382.



Dienes 26 and 29. To a solution of 28 (118 mg, 0.20 mmol) in PhMe (5.0 mL) at -78 °C were added sequentially pyridine (79.0 mg, 0.08 mL, 1.0 mmol) and SOCl $_2$ (71.4 mg, 44 μL , 0.60 mmol) via microsyringe. After 30 min, the reaction was quenched with satd aq NaHCO₃ (10 mL) and warmed to rt. After 20 min, the organic layer was separated, and the aqueous layer was extracted with Et_2O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting carefully with 0.5-1% EtOAc/hexanes, to give a mixture of 26 and 29 (114 mg, 0.20 mmol, 99%, 2.2:1 ratio) separable by HPLC as a colorless oil. Conjugate diene **26**: $[\alpha]^{23}_{D}$ -43.2 (c 0.1, CHCl₃); IR (thin film) 2954, 2866, 1640, 1459, 1156, 1119, 1013, 882, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.20–7.34 (m, 5H), 6.35 (d, J = 12.0 Hz, 1H), 6.18 (dt, J = 12.0, 5.4 Hz, 1H), 5.89 (s, 1H), 5.01 (s, 1H), 4.82 (s, 1H), 3.44 (d, J = 4.2 Hz, 2H), 2.48–2.50 (m, 2H), 2.35 (dd, *J* = 3.6, 9.9 Hz, 1H), 1.84 (d, *J* = 0.9 Hz, 3H), 1.82 (dd, *J* = 3.6, 9.9 Hz, 1H), 1.66-1.72 (m, 1H), 1.46 (s, 3H), 1.07-1.09 (m, 21H), 1.00 (t, J = 5.7 Hz, 9H), 0.78 (d, J = 5.6 Hz, 3H), 0.64 (q, J = 5.7 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 145.7, 142.8, 138.3, 131.9, 128.8, 127.4, 127.2, 126.3, 125.5, 114.5, 78.7, 68.6, 46.3, 42.6, 34.9, 27.2, 18.5, 16.7, 15.0, 12.4, 7.6, 7.2; HRMS (FAB+) calcd for C35H62O2Si2 (M+) 570.4288, found 570.4277.

Unconjugated diene 29: $[\alpha]^{23}_{D} - 1.3$ (*c* 1.74, CHCl₃); IR (thin film) 3026, 2942, 2865, 1643, 1495, 1462, 1381, 1241, 1097, 1012, 882 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.22–7.36 (m, 5H), 6.38 (d, *J* = 12.0 Hz, 1H), 6.16–6.20 (m, 1H), 5.16 (s, 1H), 4.92 (s, 1H), 4.89 (s, 1H), 4.88 (s, 1H), 3.55–3.59 (m, 1H), 3.44–3.47 (m, 1H), 2.84 (s, 2H), 2.47–2.50 (m, 2H), 2.25 (dd, *J* = 3.6, 9.9 Hz, 1H), 1.74–1.85 (m, 2H), 1.43 (s, 3H), 1.08–1.10 (m, 21H), 1.00 (t, *J* = 6.0 Hz, 9H), 0.92 (d, *J* = 4.8 Hz, 3H), 0.64 (q, *J* = 6.0 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 152.0, 146.7, 138.3, 132.2, 128.9, 127.5, 127.2, 126.4, 113.9, 111.1, 78.3, 68.8, 46.4, 39.8, 38.8, 34.6, 27.5, 18.5, 17.2, 12.4, 7.6, 7.3.



Diols 30 and 31. To a mixture of **26** and **29** (75 mg, 0.13 mmol) in THF (0.1 mL) at 0 °C was added TBAF (0.6 mL, 1.0 M in THF). After 10 min, the reaction was warmed to rt. After 1 h, the reaction was quenched with satd aq NH₄Cl (3 mL). After 10 min, the mixture was extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel,

eluting with 20% EtOAc/hexanes, to give diol **30** (20 mg, 0.069 mmol, 61%) as a white foam: $[\alpha]^{23}{}_{\rm D}$ –4.9 (*c* 0.98, CHCl₃); IR (thin film) 3373 (br), 3026, 2955, 2925, 2870, 1722, 1627, 1448, 1373, 1032, 966, 897, 741, 692 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.38 (m, 5H), 6.46 (d, *J* = 15.6 Hz, 1H), 6.12–6.20 (m, 1H), 5.97 (s, 1H), 5.07 (s, 1H), 4.86 (s, 1H), 3.28–3.40 (m, 2H), 2.66 (dd, *J* = 7.2, 14.0 Hz, 1H), 2.46 (dd, *J* = 8.0, 14.0 Hz, 1H), 2.26 (dd, *J* = 5.6, 13.6 Hz, 1H), 1.89 (s, 3H), 1.84–1.88 (m, 1H), 1.61–1.66 (m, 1H), 1.42 (s, 3H), 0.85 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.0, 142.9, 137.7, 133.8, 129.0, 127.8, 126.5, 125.9, 125.1, 115.2, 76.0, 68.5, 44.4, 42.5, 34.9, 28.1, 16.6, 15.4; HRMS (ES+) calcd for C₂₀H₂₈O₂Na (M + Na) 323.1987, found 323.1996.

Diol 31: colorless oil; $[\alpha]^{23}_{D}$ +7.6 (*c* 1.21, CHCl₃); IR (thin film) 3373, 3081, 3026, 2957, 2926, 2870, 1724, 1644, 1496, 1449, 1373, 1112, 1034, 967, 900, 739, 692 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.39 (m, 5H), 6.49 (d, *J* = 15.6 Hz, 1H), 6.16–6.23 (m, 1H), 5.23 (s, 1H), 4.96 (s, 3H), 3.48–3.52 (m, 2H), 2.85–2.96 (m, 2H), 2.62–2.66 (m, 1H), 2.48–2.52 (m, 1H), 2.18–2.24 (m, 1H), 1.86–1.93 (m, 2H), 1.40 (s, 3H), 0.94 (d, *J* = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 151.9, 146.9, 137.7, 134.3, 128.9, 127.7, 126.6, 125.7, 114.3, 111.8, 75.7, 68.6, 45.0, 40.1, 39.2, 34.3, 28.2, 17.1; HRMS (ES+) calcd for C₂₀H₂₈O₂Na (M + Na) 323.1987, found 323.1989.



Nitrile 14. To a solution of **30** (98 mg, 0.33 mmol) in Et₂O (1.6 mL) at 0 $^{\circ}\text{C}$ were added sequentially PPh3 (342 mg, 1.3 mmol) and DEAD (0.24 mL, 1.3 mmol, 40% w/w in PhMe). After 15 min, acetone cyanohydrin (70.0 mg, 74.5 µL, 0.82 mmol) was added dropwise. After 5 min, the reaction was warmed to rt. After 6 h, the reaction was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give a corresponding nitrile intermediate (80 mg, 0.26 mmol, 81%) as a colorless oil: $[\alpha]^2$ ¹³_D +7.8 (c 0.99, CHCl₃); IR (neat) 3479 (br), 2962, 2927, 2246, 1733, 1496, 1456, 1420, 1381, 1105, 968, 902, 743, 694 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) δ 7.26–7.37 (m, 5H), 6.47 (d, J = 16.0 Hz, 1H), 6.12–6.18 (m, 1H), 5.95 (s, 1H), 5.12 (s, 1H), 4.92 (s, 1H), 2.66 (dd, *J* = 7.2, 14.0 Hz, 1H), 2.48 (dd, J = 7.2, 14.0 Hz, 1H), 2.05–2.20 (m, 4H), 1.88 (s, 3H), 1.76–1.88 (m, 1H), 1.75 (br, 1H), 1.43 (s, 3H), 0.97 (d, J = 6.6 Hz, 3H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl_3) δ 143.8, 143.5, 137.5, 134.0, 129.0, 127.9, 126.5, 125.5, 124.3, 119.1, 116.4, 76.0, 44.8, 44.5, 29.3, 28.1, 24.2, 19.5, 15.5; HRMS (FAB+) calcd for C₂₁H₂₈NO (M + 1) 310.2171, found 310.2144.



To a solution of the above intermediate (80 mg, 0.26 mmol) in CH₂Cl₂ (1.0 mL) at -78 °C was added 2,6-lutidine (111.0 mg, 0.12 mL, 1.04 mmol) followed by TESOTF (136.9 mg, 0.12 mL, 0.52 mmol). After 30 min, the reaction was warmed to rt and quenched with satd aq NH₄Cl (3 mL). After 5 min, the mixture was extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 1% EtOAc/hexanes, to give nitrile 14 (80 mg, 0.19 mmol, 72%) as a colorless oil: $[\alpha]^{23}_{D} - 37.6$ (*c* 0.98, CHCl₃); IR (neat) 2957, 2933, 2876, 2240, 1733, 1496, 1457, 1420, 1381, 1160, 1122, 1016, 967, 902, 742, 693 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.21–7.33 (m, 5H), 6.33 (d, *J* = 11.7 Hz, 1H), 6.15–6.19 (m, 1H), 5.87 (s, 1H), 5.07 (s, 1H), 4.87 (s, 1H), 2.50 (d, *J* = 5.4 Hz, 2H), 2.15 (dd, *J* = 5.4, 9.9 Hz, 1H), 2.00–2.06 (m, 3H), 1.81 (s, 3H), 1.72–1.79 (m, 1H), 1.47 (s, 3H), 1.02 (t, *J* = 5.7 Hz, 9H), 0.85 (d, *J* = 5.1 Hz, 3H), 0.67 (q, *J* = 5.1 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 143.9, 143.7, 138.1, 131.9, 129.0, 127.4, 127.3, 126.3, 124.6, 119.0, 116.0, 78.8, 46.0, 44.9, 29.3,



Aldehyde 12. To a solution of 14 (75 mg, 0.18 mmol) in *t*-BuOH/ H₂O (3 mL, 1:1) were added MeSO₂NH₂ (25.6 mg, 0.27 mmol) and AD mix β^{*53} (250 mg). After 16 h, the reaction was quenched with satd aq Na₂S₂O₃ (3 mL). After 20 min, the mixture was extracted with EtOAc (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 20% EtOAc/ hexanes, to give a mixture of diol isomers (73 mg, dr = 6:1, 0.16 mmol, 89%) as a colorless oil. The diol substrate was applied to the next step without further purification.

A solution of the mixture described above in THF/Et₂O (2.8 mL, 1:1) was added NaIO₄ (304 mg, 1.42 mmol) and H₂O (1.4 mL). After 2 h, the reaction was quenched with satd aq NaCl (3 mL), and the mixture was extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give aldehyde 12 (54 mg, 0.15 mmol, 96%) as a colorless oil: $[\alpha]^{23}_{D}$ -12.6 (c 1.22, CHCl₃); IR (neat) 3081, 2957, 2917, 2876, 2246, 1722, 1628, 1457, 1417, 1381, 1240, 1163, 1126, 1046, 1017, 904, 802, 743 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 9.66 (t, J = 3.2 Hz, 1H), 5.99 (s, 1H), 5.10 (s, 1H), 4.89 (s, 1H), 2.61 (dd, *J* = 3.2, 14.8 Hz, 1H), 2.47 (dd, *J* = 2.8, 14.8 Hz, 1H), 2.32 (dd, J = 5.2, 16.8 Hz, 1H), 2.15-2.25 (m, 2H), 2.08 (dd, J = 7.2, 13.6 Hz, 1H), 1.83-1.92 (m, 1H), 1.80 (s, 3H), 1.47 (s, 3H), 1.04 (d, J = 6.4 Hz, 3H), 0.96 (t, J = 8.0 Hz, 9H), 0.63 (q, J = 8.0 Hz, 6 H);¹³C NMR (100 MHz, CDCl₃) δ 203.2, 143.2, 142.7, 125.2, 118.9, 116.8, 54.5, 44.8, 30.1, 29.6, 28.3, 24.3, 19.8, 15.2, 7.5, 7.2; HRMS (FAB+) calcd for $C_{20}H_{36}O_2SiN (M + 1) 350.2515$, found 350.2508.



Nitrile 33. To a solution of **31** (30 mg, 0.10 mmol) in Et₂O (0.5 mL, 0.2 M) at 0 °C were added sequentially PPh₃ (104 mg, 0.4 mmol) and DEAD (72 μ L, 0.4 mmol, 40% solution in PhMe). After 15 min, acetone cyanohydrin (21.2 mg, 23.0 μ L, 0.25 mmol) was added dropwise. After 5 min, the reaction was warmed to rt. After 6 h, the reaction was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give a corresponding nitrile intermediate (25 mg, 0.08 mmol, 81%) as a colorless oil.

To a solution of the above nitrile (25 mg, 0.08 mmol) in CH₂Cl₂ (0.32 mL, 0.25 M) at -78 °C were added sequentially 2,6-lutidine (34.3 mg, 37 μ L, 0.32 mmol) and TESOTf (42.1 mg, 36 μ L, 0.16 mmol). After 30 min, the reaction was warmed to rt and quenched with satd aq NH₄Cl (3 mL). After 5 min, the mixture was extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 1% EtOAc/hexanes, to give **33** (26 mg, 62 μ mol, 77%) as a colorless oil: [α]²³_D -13.6 (*c* 0.28, CHCl₃); IR (neat) 2957, 2933, 2911, 2875, 2247, 1238, 1157, 1123, 1063, 1005, 967, 905, 742, 693 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.17–7.26 (m, SH), 6.35 (d, *J* = 15.9 Hz, 1H), 6.08–6.19 (m, 1H), 5.14 (s, 1H), 4.93 (s, 2H), 4.71 (s, 1H), 2.75–2.85 (m, 2H), 2.46

(dd, J = 0.6, 6.9 Hz, 2H), 2.24–2.32 (m, 1H), 1.97–2.17 (m, 4H), 1.38 (s, 3H), 0.95–1.04 (m, 12H), 0.58–0.68 (m, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 151.7, 144.9, 138.1, 132.3, 128.9, 127.4, 127.1, 126.4, 119.2, 115.8, 111.1, 78.2, 46.4, 42.3, 38.2, 28.9, 27.7, 24.3, 20.0, 7.6, 7.3.



Aldehyde 35. To a solution of 33 (26 mg, 62 μ mol) in *t*-BuOH/ H₂O (1 mL, 1:1) were added MeSO₂NH₂ (8.8 mg, 93 μ mol) and AD mix β^{*53} (87 mg). After 16 h, the reaction was quenched with satd aq Na₂S₂O₃ (3 mL). After 20 min, the mixture was extracted with EtOAc (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 20% EtOAc/ hexanes, to give a mixture of the diol isomers (25.5 mg, dr = 1.3:1, 56 μ mol, 90%) as a colorless oil. The diol substrate was applied to the next step without further purification.

A solution of the mixture described above in THF/Et₂O (1.24 mL, 1:1) was added NaIO₄ (120 mg, 0.56 mmol) and H₂O (0.62 mL). After 2 h, the reaction was quenched with satd aq NaCl (3 mL) and the mixture was extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give aldehyde **35** (18.5 mg, 0.053 mmol, 95%) as a colorless oil: $[\alpha]^{23}_{D} - 11.4$ (*c* 1.57, CHCl₃); IR (neat) 2958, 2915, 2876, 2737, 2246, 1722, 1645, 1457, 1417, 1375, 1239, 1165, 1123, 1045, 1005, 909, 743, 726 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.66 (t, *J* = 3.0 Hz, 1H), 5.28 (s, 1H), 4.95 (s, 1H), 4.94 (s, 1H), 4.90 (s, 1H), 2.76 (s, 2H), 2.20–2.62 (m, 5H), 2.02–2.06 (m, 2H), 1.46 (s, 3H), 1.08 (d, *J* = 6.3 Hz, 3H), 0.96 (t, *J* = 7.8 Hz, 9H), 0.63 (q, *J* = 7.8 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 203.3, 150.5, 144.3, 119.0, 116.2, 112.6, 54.5, 42.1, 38.4, 28.9, 28.5, 24.3, 24.0, 19.9, 7.5, 7.2.



Methyl Ketone 13. To a solution of imide 36^{22} (107 mg, 0.3 mmol) in PhMe (0.7 mL) at -50 °C were added sequentially Et₃N (34.4 mg, 48.2 µL, 0.34 mmol) and Bu₂BOTf (86.0 mg, 80.0 µL, 0.32 mmol). After 1.5 h, a solution of aldehyde 37^8 (46 mg, 0.20 mmol) in PhMe (0.25 mL) was transferred dropwise into the reaction via cannula. After 40 min, the reaction was warmed to -30 °C within 30 min. After 1 h, the reaction was quenched with aq phosphate buffer solution (0.5 mL, pH 7), MeOH (0.5 mL), and THF (0.5 mL). Next, the mixture was warmed up to rt. A solution of H_2O_2 (0.5 mL, 30% H_2O_2 in H_2O) in MeOH (0.5 mL) was added dropwise. After 1 h, the reaction mixture was extracted with EtOAc $(3 \times 15 \text{ mL})$ and the combined organic extract was diluted with an aqueous phosphate buffer solution (20 mL, pH 7). The mixture was concentrated in vacuo and extracted again with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15-20% EtOAc/hexanes, to give an aldol adduct SI-4 (83 mg, 0.14 mmol, 72%) as a colorless oil: $[\alpha]^{23}_{D}$ – 3.9 (*c* 0.83, CHCl₃); IR (thin film) 3490 (br), 2956, 2875, 1781, 1708, 1612, 1514, 1389, 1248, 1211, 1051, 1012, 744 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.25–7.39 (m, 7H), 6.92 (d, J = 8.8 Hz, 2H, 5.36 (d, J = 2.0 Hz, 1H), 4.64–4.72 (m, 1H), 4.64 (d, J =10.8 Hz, 1H), 4.49 (d, J = 10.8 Hz, 1H), 4.20–4.29 (m, 2H), 3.84–3.90 (m, 1H), 3.82 (s, 3H), 3.61-3.64 (m, 1H), 3.34 (dd, J = 2.8, 13.2 Hz, 1H), 2.78 (dd, J = 3.6, 13.5 Hz, 1 H), 2.31 (d, J = 10.0 Hz, 1H), 1.74-1.81 (m, 1H), 1.56–1.62 (m, 1H), 1.40–1.48 (m, 1H), 1.15 (d, J = 5.6 Hz, 3H), 1.03 (d, J = 6.8 Hz, 3H), 0.99 (t, J = 7.6 Hz, 9H), 0.62 (q, J = 7.6 Hz, 6H); 13 C NMR (75 MHz, CDCl₃) δ 171.6, 160.0, 153.6, 135.6, 130.7, 129.8, 129.6, 129.4, 127.8, 114.3, 78.4, 76.6, 73.0, 67.4, 67.3, 56.2,

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55.7, 43.1, 38.2, 34.6, 23.6, 16.2, 7.3, 5.3; HRMS (FAB+) calcd for $C_{32}H_{48}NO_7Si\ (M+1)\ 586.3200,$ found 586.3202.



TES Ether SI-5. To a solution of the aldol adduct SI-4 (78 mg, 0.13 mmol) in CH₂Cl₂ (0.3 mL) at 0 °C were added sequentially 2,6lutidine (27.8 mg, 29.8 µL, 0.26 mmol) and TESOTf (68.5 mg, 60.1 µL, 0.26 mmol). After 1 h, the reaction was warmed to rt and quenched with satd aq NH₄Cl (3 mL). After 5 min, the mixture was extracted with Et₂O (3 \times 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 4-6% EtOAc/hexanes, to give the corresponding protected alcohol SI-5 (84 mg, 0.12 mmol, 90%) as a colorless oil: $[\alpha]_{D}^{23}$ -33.8 (c 1.9, CHCl₃); IR (thin film) 2956, 2911, 2875, 1785. 1704, 1612, 1513, 1456, 1381, 1248, 1082, 1011, 739 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 7.34–7.26 (m, 7H), 6.84 (d, J = 8.4 Hz, 2H), 5.25 (d, J = 6.0 Hz, 1H), 4.64 (d, J = 11.7 Hz, 1H), 4.52 (d, J = 11.7 Hz, 1H), 4.46-4.49 (m, 1H), 4.03-4.12 (m, 3H), 3.73-3.98 (m, 1H), 3.79-3.81 (m, 1H), 3.74 (s, 3H), 3.11 (dd, J = 3.0, 13.2 Hz, 1H), 2.40 (dd, J = 3.0, 13.5 Hz, 1H), 1.47–1.55 (m, 2H), 1.09 (d, J = 6.0 Hz, 3H), 0.87–0.97 (m, 21H), 0.53–0.63 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 172.3, 159.9, 153.3, 135.8, 130.6, 130.3, 129.7, 129.4, 127.8, 114.0, 79.6, 77.8, 73.4, 67.6, 66.8, 56.4, 55.6, 44.7, 37.8, 33.9, 24.1, 15.2, 7.5, 7.4, 5.8, 5.4; HRMS (FAB+) calcd for C₃₈H₆₀NO₇Si₂ (M - 1) 698.3908, found 698.3890.



Methyl Ketone 13. To a solution of EtSH (75 mg, 90.2 μ L, 1.21 mmol) in THF (9.0 mL) at 0 °C was added *n*-BuLi (0.43 mL, 1.07 mmol, 2.5 M in hexane). After 20 min, a solution of the above intermediate (500 mg, 0.714 mmol) in THF (3.0 mL) was transferred via cannula dropwise. After 20 min, the reaction was quenched with satd aq NH₄Cl (10 mL). After 5 min, the mixture was extracted with Et₂O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by a plug of silica gel, eluting with 10% EtOAc/hexanes, to give the corresponding thioester intermediate (375 mg, 0.68 mmol, 95%) as a colorless oil.

To a suspension of CuI (796 mg, 4.18 mmol) in Et₂O (9.0 mL) at 0 °C was added MeLi (5.2 mL, 8.36 mmol, 1.6 M in Et₂O). After 15 min, the colorless solution was cooled to -50 °C, and a solution of the above thioester intermediate (375 mg, 0.68 mmol) in Et_2O (2.4 mL) was transferred into the reaction dropwise via cannula. After 2 h, the reaction was quenched with satd aq NH_4Cl (10 mL) at -50 °C, warmed to rt, and extracted with Et_2O (3 × 20 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2-4% EtOAc/hexanes, to give methyl ketone 13 (326 mg, 0.60 mmol, 89%) as a colorless oil: $[\alpha]_{D}^{23}$ +31.0 (c 0.80, CHCl₃); IR 2956, 2912, 2876, 1716, 1613, 1514, 1457, 1416, 1249, 1172, 1082, 1037, 1007, 820, 740 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.23 (d, J = 8.7 Hz, 2H), 6.86 (d, J = 8.7 Hz, 2H), 4.40 (d, J = 11.4 Hz, 1H), 4.38 (d, J = 11.4 Hz, 1H), 3.81 (s, 3H), 3.69-3.81 (m, 3H), 2.11 (s, 3H), 1.45-1.48 (m, 3H), 1.09 (d, J = 6.0 Hz, 3H), 0.85–0.97 (m, 21H), 0.53–0.61 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 211.5, 159.8, 130.1, 129.9, 114.1, 88.9, 73.1, 67.4, 55.7, 44.9, 33.5, 27.4, 23.9, 14.3, 7.4, 7.3, 5.7, 5.4; HRMS (FAB+) calcd for $C_{29}H_{55}O_5Si_2$ (M + 1) 539.3588, found 539.3559.



Carboxylic Acid 11. To a solution of 38 (320 mg, 0.35 mL, 2.50 mmol) in THF/H₂O (35 mL, 1:1) at rt were added K₂OsO₄·H₂O (14 mg, 15 μ mol) and NaIO₄ (2.4 g, 11.25 mmol). After 3 h, the reaction was quenched with satd aq Na₂S₂O₃ (5 mL). After 20 min, the mixture was extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give an aldehyde intermediate (247 mg, 1.90 mmol, 76%) as a colorless oil. The aldehyde intermediate SI-7 was used directly in the next step without further purification.

To a solution of the above aldehyde intermediate **SI**-7 (247 mg, 1.90 mmol) in CH₂Cl₂ (5.4 mL) at rt was added Wittig reagent Ph₃P= C(Me)CO₂t-Bu (725 mg, 2.0 mmol). After 30 min, the reaction was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give Wittig product **SI-8** (377 mg, 1.56 mmol, 82%) as a colorless oil: IR (neat) 2977, 1739, 1704, 1652, 1456, 1436, 1367, 1289, 1252, 1159, 1121, 1083, 852, 743 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.59 (dt, *J* = 1.2, 7.5 Hz, 1H), 3.65 (s, 3H), 2.32 (t, *J* = 7.5 Hz, 2H), 2.17 (q, *J* = 7.5 Hz, 2H), 1.71–1.83 (m, SH), 1.47 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 174.1, 167.8, 139.9, 130.5, 80.5, 51.9, 33.9, 28.5, 28.3, 24.2, 12.8; HRMS (FAB+) calcd for C₁₃H₂₃O₄ (M + 1) 243.1596, found 243.1598.



Phosphonate SI-9. To a solution of methyl diethylphosphonate (318 mg, 0.3 mL, 2.09 mmol) in THF (10 mL) at -78 °C was added n-BuLi (0.83 mL, 2.09 mmol, 2.5 M in hexane). After 20 min, this resulted solution was transferred dropwise into a solution of the above Wittig product SI-8 (202 mg, 0.83 mmol) in THF (4 mL) at -78 °C via cannula. After 30 min, the reaction was quenched with satd aq NH₄Cl (3 mL). The mixture was extracted with EtOAc (3×10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 80% EtOAc/hexanes, to give phosphonate intermediate SI-9 (254 mg, 0.65 mmol, 78%) as a colorless oil: IR (neat) 2978, 2932, 1701, 1652, 1392, 1366, 1254, 1162, 1024, 965, 794 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.58 (dt, J = 1.2, 7.5 Hz, 1H), 4.07–4.17 (m, 4H), 3.09 (s, 1H), 3.01 (s, 1H), 2.63 (t, J = 7.2 Hz, 2H), 2.13 (q, J = 7.5 Hz, 2H), 1.75 (s, 3H), 1.67–1.75 (m, 2H), 1.46 (s, 9H), 1.31 (t, J = 7.2 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 202.0, 167.8, 140.2, 130.4, 80.5, 63.0, 43.7, 42.0, 28.5, 28.1, 22.7, 16.7, 12.8; HRMS (FAB+) calcd for $C_{17}H_{32}O_6P$ (M + 1) 363.1937, found 363.1942.



Acid 11. To a solution of the above phosphonate SI-9 (102 mg, 0.28 mmol) in CH₂Cl₂ (1.5 mL) at 0 °C was added TFA (1.15 g, 0.75 mL, 9.7 mmol). After 5 min, the reaction was warmed to rt. After 2 h, the reaction was concentrated in vacuo and purified by chromatography over silica gel, eluting with 4% MeOH/CH₂Cl₂, to give 11 (84.8 mg, 0.28 mmol, 99%) as a colorless oil: IR (neat) 3418, 2986, 2934, 1772, 1715, 1395, 1209, 1163, 1023, 973, 799, 698 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.86 (t, *J* = 7.5 Hz, 1H), 6.07 (br, 1H), 4.12–4.23 (m, 4H), 3.18 (s, 1H), 3.10 (s, 1H), 2.64 (t, *J* = 7.2 Hz, 2H), 2.22 (q, *J* = 7.5 Hz, 2H), 1.82 (s, 3H), 1.71–1.80 (m, 2H), 1.30 (t, *J* = 7.5 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 202.2, 170.5, 143.5, 128.5, 63.2, 43.6, 41.9, 28.2, 22.5, 16.7, 12.5; HRMS (FAB+) calcd for C₁₃H₂₄O₆P (M + 1) 307.1311, found 307.1322.



Alcohol 48. To a solution of 13 (73 mg, 0.14 mmol) in Et_2O (0.7 mL) at -78 °C was added LDA⁵⁴ (0.15 mL, 1.0 M in THF). After 20 min, a precooled (-78 °C) solution of aldehyde 12 (31.5 mg,

0

0.09 mmol) in Et₂O (0.4 mL) was transferred in one portion via cannula at -78 °C. After 20 min, the reaction was guenched with satd ag NH₄Cl (3 mL), and the mixture was extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give 48 (55 mg, 0.06 mmol, 69%) as a colorless oil: $[\alpha]^{23}_{D}$ +21.3 (c 0.60, CHCl₃); IR (neat) 3496, 2955, 2911, 2876, 1715, 1614, 1515, 1457, 1418, 1379, 1249, 1086, 1036, 1008, 903, 807, 740 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.27 (d, J = 8.7 Hz, 2H), 6.85 (d, J = 8.7 Hz, 2H), 5.87 (s, 1H), 5.09 (s, 1H), 4.89 (s, 1H), 4.47 (d, J = 10.8 Hz, 1H), 4.33-4.36 (m, 2H), 3.80 (s, 3H), 3.76–3.80 (m, 3H), 3.61 (s, 1H), 2.95 (dd, J = 7.2, 18.0 Hz, 1H), 2.05-2.40 (m, 6H), 1.88-1.95 (m, 1H), 1.81 (s, 3H), 1.52 (s, 3H), 1.37 - 1.52 (m, 5H), 1.07 (d, J = 6.9 Hz, 3H), 1.05 (d, J = 6.0Hz, 3H), 0.83-0.98 (m, 30H), 0.51-0.67 (m, 18H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3) \delta 212.1, 159.7, 144.8, 143.2, 130.1, 124.3, 118.9,$ 116.7, 114.1, 88.5, 79.7, 72.9, 67.3, 65.1, 55.7, 48.4, 47.2, 45.1, 44.9, 33.5, 29.6, 26.7, 24.3, 23.6, 19.8, 15.2, 13.9, 7.5, 7.4, 7.3, 7.0, 5.7, 5.4; HRMS (FAB+) calcd for $C_{49}H_{90}O_7Si_3N$ (M + 1) 888.6025, found 888.6052.



Silyl Ether 49. To a solution of 48 (55 mg, 60 μ mol) in CH₂Cl₂ (0.15 mL) at rt was added Et₃N (0.15 mL). The solution was then cooled to -78 °C. After 5 min, TBSOTf (65.8 mg, 60.4 µL, 0.25 mmol) was added dropwise. After 5 min, the reaction was warmed to rt. After 20 min, the reaction was quenched with satd aq NH_4Cl (3 mL), and the mixture was extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 3% EtOAc/hexanes, to give 49 (53 mg, 53 µmol, 88%) as a colorless oil: $[\alpha]^{23}_{D}$ +35.1 (c 0.50, CHCl₃); IR (neat) 2954, 2928, 2876, 1714, 1615, 1515, 1458, 1381, 1250, 1167, 1125, 1065, 1006, 987, 835, 776, 740 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.27 (d, *J* = 8.4 Hz, 2H), 6.85 (d, *J* = 8.7 Hz, 2H), 5.80 (s, 1H), 5.07 (s, 1H), 4.89 (s, 1H), 4.51 (d, J = 10.5 Hz, 1H), 4.21-4.25 (m, 2H), 3.81 (s, 3H),3.64-3.81 (m, 3H), 3.17 (dd, J = 9.3, 18.0 Hz, 1H), 2.58 (dd, J = 2.7, 17.7 Hz, 1H), 2.38 (dd, J = 4.5, 16.8 Hz, 1H), 2.19 (dd, J = 6.9, 16.8 Hz, 1H), 2.05-2.10 (m, 2H), 1.86-1.94 (m, 2H), 1.79 (s, 3H), 1.66-1.79 (m, 1H), 1.58-1.62 (m, 1H), 1.40 (s, 3H), 1.28-1.35 (m, 2H), 1.07 (d, J = 6.3 Hz, 3H), 1.05 (d, J = 5.4 Hz, 3H), 0.86–0.97 (m, 30H), 0.78 (s, 9H), 0.50–0.65 (m, 18H), 0.05 (s, 3H), -0.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 212.2, 159.5, 143.6, 143.1, 130.3, 129.9, 124.9, 119.0, 116.6, 113.9, 89.1, 77.3, 72.3, 67.2, 66.1, 55.6, 49.7, 46.4, 45.5, 44.7, 33.7, 29.4, 29.0, 26.3, 24.3, 23.5, 19.7, 18.4, 15.4, 12.9, 7.7, 7.4, 7.3, 5.7, 5.3, -3.7, -4.2; HRMS (FAB+) calcd for C₅₅H₁₀₄O₇Si₄N (M + 1) 1002.6890, found 1002.6923.



Acid 9. To a solution of 49 (11 mg, 11 μ mol) in PhMe (0.25 mL) at -78 °C was added DIBAL-H (13.2 μ L, 13.2 μ mol, 1.0 M in PhMe) dropwise. After 10 min, another portion of DIBAL-H (6.0 μ L, 6.0 μ mol, 1.0 M in PhMe) was added. The reaction was quenched sequentially with MeOH (0.5 mL), satd aq tartaric acid (0.1 mL), and H₂O (2 mL). Next, the mixture was warmed to rt. After 1 h, the mixture was extracted with Et₂O (3 × 10 mL), and the dried (MgSO₄) extract was concentrated in vacuo to give the unstable aldehyde 10 as a colorless oil. The aldehyde was applied immediately to the next step without further purification.

To a solution of **11** (4.0 mg, 0.012 mmol) in THF/H₂O (0.2 mL, 40:1) at rt was added Ba(OH)₂·8H₂O (7.6 mg, 0.024 mmol). After 30 min, a solution of the above aldehyde intermediate **10** in THF (0.15 mL) was added dropwise via cannula. After 16 h, the reaction was quenched with satd aq NH₄Cl (2 mL) and the mixture was extracted with Et₂O (3×10 mL). The dried (MgSO₄) extract was concentrated in vacuo was applied immediately to the next step without further purification.

To a flask containing crude 50 at 0 °C was added a premixed solution of HOAc/THF/H₂O (4.0 mL, 8:4:1) dropwise. After 2 h, the solution was allowed to warm to 10 °C. After 6 h, the reaction was warmed to rt. After 6 h, the reaction was concentrated in vacuo. Benzene was added to further dry the product 9 (11.3 mg): ¹H NMR (300 MHz, CDCl₃) δ 7.30 (d, J = 8.8 Hz, 2H), 6.84–6.98 (m, 4H), 6.10 (d, J = 15.9 Hz, 2H), 5.83 (s, 1H), 4.99 (s, 1H), 4.83 (s, 1H), 4.52 (d, J = 10.5 Hz, 1H), 4.12-4.27 (m, 2H), 3.81 (s, 3H), 3.68-3.80 (m, 3H), 3.16 (m, 1H), 2.59-2.68 (m, 3H), 2.28–2.38 (m, 1H), 2.21–2.25 (m, 2H), 1.91–2.11 (m, 4H), 1.82 (s, 3H), 1.75 (s, 3H), 1.71–1.82 (m, 4H), 1.64–1.69 (m, 1H), 1.43 (s, 3H), 1.25–1.35 (m, 4H), 1.09 (d, J = 4.0 Hz, 3H), 0.88–1.01 (m, 31H), 0.81 (s, 9H), 0.55-0.68 (m, 18H), 0.09 (s, 3H), 0.01 (s, 3H); ^{13}C NMR (100 MHz, CDCl₃) δ 212.5, 201.2, 168.6, 159.6, 147.0, 145.0, 144.0, 142.1, 132.1, 130.1, 129.9, 128.9, 125.7, 115.5, 113.9, 88.5, 77.3, 72.2, 67.2, 66.1, 55.6, 49.6, 47.0, 46.0, 43.9, 40.3, 39.3, 33.3, 32.3, 31.5, 29.1, 28.6, 26.3, 23.3, 23.5, 23.0, 19.6, 18.3, 14.7, 14.5, 12.5, 7.7, 7.4, 7.3, 7.0, 6.2, 5.6, -3.8, -4.1.



Alcohol 52. To a flask containing 13 (120 mg, 0.22 mmol) at 0 °C was added a premixed solution of HOAc/THF/H₂O (4.0 mL, 8:8:1) dropwise. After 10 min, the solution was allowed to warm to 5 °C. After 2 h, the reaction was quenched with satd aq NaHCO3 (10 mL) dropwise. Next, the mixture was extracted with Et_2O (3 × 10 mL), and the dried $(MgSO_4)$ extract was concentrated in vacuo to give product 52 (90 mg, 0.21 mmol, 95%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 17.7 (c 0.34, CHCl₃); IR (thin film) 3413, 2958, 2910, 2875, 1712, 1613, 1514, 1457, 1354, 1302, 1249, 1172, 1076, 1055, 1009, 820, 740 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 7.23 \text{ (d, } J = 8.4 \text{ Hz}, 2\text{H}) 6.86 \text{ (d, } J = 8.4 \text{ Hz}, 2\text{H}),$ 4.43 (d, J = 11.4 Hz, 1H), 4.38 (d, J = 11.4 Hz, 1H), 3.81–3.91 (m, 2H), 3.80 (s, 3H), 3.74 (d, J = 6.3 Hz, 1H), 2.40 (br, 1H), 2.14 (s, 3H), 1.49-1.65 (m, 2H), 1.32–1.40 (m, 1H), 1.14 (d, J = 6.0 Hz, 3H), 0.85–0.99 (m, 12H), 0.58 (q, J = 7.8 Hz, 6H); ¹³C NMR (400 MHz, CDCl₃) δ 210.8, 159.8, 130.2, 129.7, 114.2, 88.4, 75.3, 72.9, 66.2, 55.7, 43.5, 33.5, 27.5, 24.2, 15.1, 7.4, 5.6; HRMS (FAB+) calcd for $C_{23}H_{41}O_5Si (M + 1)$ 425.2723, found 425.2730.



Ester 53. To a solution of **52** (237 mg, 0.56 mmol), *p*-NO₂-benzonic acid (374 mg, 2.24 mmmol), and PPh₃ (587 mg, 2.24 mmol) in THF (3.7 mL) at 0 °C was added DEAD (1.0 mL, 1.04 g, 2.24 mmol, 40% w/w in PhMe) dropwise. After 10 min, the reaction was warmed to rt. After 2 h, the reaction was concentrated in vacuo and purified by

chromatography over silica gel, eluting with 10–15% EtOAc/hexanes, to give **53** (46 mg, 0.084 mmol, 80%) as a colorless oil: $[\alpha]^{23}{}_{\rm D}$ (+) 38.1 (*c* 0.48, CHCl₃); IR (thin film) 2954, 2875, 1719, 1610, 1528, 1514, 1457, 1349, 1275, 1102, 1036, 1013, 823, 720 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.28 (d, *J* = 8.8 Hz, 2H), 8.19 (d, *J* = 8.8 Hz, 2H), 7.26 (d, *J* = 8.4 Hz, 2H), 6.89 (d, *J* = 8.4 Hz, 2H), 5.24–5.29 (m, 1H), 4.53 (d, *J* = 11.2 Hz, 1H), 4.38 (d, *J* = 11.2 Hz, 1H), 3.80–3.83 (m, 3H), 2.16 (s, 3H), 2.01–2.08 (m, 1H), 1.67–1.69 (m, 1H), 1.55–1.61 (m, 1H), 1.39 (d, *J* = 6.0 Hz, 3H), 0.93 (t, *J* = 8.0 Hz, 9H), 0.89 (d, *J* = 6.8 Hz, 3H), 0.59 (q, *J* = 8.0 Hz, 6H); ¹³C NMR (400 MHz, CDCl₃) δ 211.3, 164.8, 159.9, 150.8, 136.5, 131.0, 130.3, 129.6, 123.9, 114.2, 87.6, 77.4, 73.1, 71.3, 55.7, 39.9, 33.7, 28.0, 21.4, 14.7, 7.4, 5.5.



Silyl Ether 54. To a solution of 53 (125 mg, 0.23 mmol) in MeOH (4.7 mL) was added Ba(OH)₂.8H₂O (36.1 mg, 0.12 mmol). After 1 h, the reaction was quenched with satd aq $NH_4Cl(5 mL)$. The mixture was extracted with Et₂O (3 \times 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10-15% EtOAc/hexanes, to give an alcohol intermediate SI-10 (63 mg, 0.15 mmol, 65%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 38.4 (c 1.87, CHCl₃); IR (thin film) 3439, 2958, 2933, 2875, 1713, 1613, 1514, 1458, 1376, 1302, 1249, 1079, 1035, 820, 786, 739 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 7.24 (d, J = 8.7 \text{ Hz}, 2\text{H}), 6.87 (d, J = 8.7 \text{ Hz}, 2\text{H}), 4.44$ (d, J = 11.4 Hz, 1H), 4.38 (d, J= 11.4 Hz, 1H), 3.75-3.80 (m, 6H), 2.14 (s, 3H), 1.70 (br, 1H), 1.50–1.59 (m, 1H), 1.27–1.35 (m, 2H), 1.16 (d, J = 6.0 Hz, 3H), 0.92 (t, J = 7.8 Hz, 9H), 0.85 (d, J = 6.9 Hz, 3H), 0.58 (q, J = 6.9 Hz, 6H); ¹³C NMR (400 MHz, CDCl₃) δ 211.5, 159.8, 130.2, 129.7, 114.2, 88.3, 73.0, 66.4, 55.7, 43.9, 33.8, 27.7, 24.9, 14.7, 7.4, 5.6; HRMS (FAB+) calcd for C₂₃H₄₀O₅SiNa (M + Na) 447.2543, found 447.2561.

To a solution of the alcohol intermediate **SI-10** (77 mg, 0.18 mmol) in CH₂Cl₂ (0.9 mL) at -78 °C were added 2,6-lutidine (78 mg, 84.8 μ L, 0.73 mmol) and TMSOTf (80 mg, 65.2 µL, 0.36 mmol). After 20 min, the reaction was quenched with satd aq NH₄Cl (5 mL) and extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give product 54 (80 mg, 0.16 mmol, 90%) as a colorless oil: $[\alpha]_{D}^{23}$ (+) 31.8 (c 2.80, CHCl₃); IR (thin film) 2955, 2911, 2876, 1715, 1613, 1514, 1458, 1414, 1372, 1352, 1249, 1124, 1088, 1038, 1006, 820, 787, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.23(d, J = 8.4 Hz, 2H), 6.86 (d, J = 8.4 Hz, 2H), 4.42 (d, J = 11.4 Hz, 1H), 4.36 (d, J = 11.4 Hz, 1H), 3.74-3.85 (m, 5H), 3.68 (d, J = 6.9 Hz, 1H), 2.09 (s, 3H), 1.55-1.61 (m, 1H), 1.41-1.50 (m, 1H), 1.25–1.35 (m, 1H), 1.12 (d, J = 6.0 Hz, 3H), 0.92 (t, J = 7.8 Hz, 9H), 0.82 (d, J = 6.6 Hz, 3H), 0.58 (q, J = 7.8 Hz, 6H), 0.07 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 211.0, 159.7, 130.2, 129.7, 114.1, 89.3, 73.1, 66.2, 55.6, 45.2, 32.4, 26.9, 25.3, 12.9, 7.4, 5.7, 0.7; HRMS (ES+) calcd for C₂₆H₄₈O₅NaSi₂ (M + Na) 519.2938, found 519.2914.



Acetate 55. To a solution of 33 (45 mg, 0.11 mmol) in PhMe (0.53 mL) at $-78 \text{ }^{\circ}\text{C}$ was added DIBAL-H (0.15 mL, 0.15 mmol, 1.0 M in PhMe). After 10 min, the reaction was quenched sequentially

with MeOH (0.2 mL), satd aq tartaric acid (0.2 mL) and H_2O (2.0 mL). After stirring 12 h at rt, the mixture was extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give the corresponding crude aldehyde (0.11 mmol).

To a mixed solution of the above aldehyde (0.11 mmol) in CH₂Cl₂ (0.5 mL) and EtOH (0.5 mL) at 0 °C was added NaBH₄ (8.4 mg, 0.22 mmol). After 15 min, the reaction was quenched with a brine solution (5 mL) and extracted with Et₂O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 6% EtOAc/hexanes, to give the corresponding alcohol intermediate (38 mg, 0.09 mmol, 85%) as a colorless oil.

To a solution of the above alcohol intermediate (29 mg, 0.07 mmol) in CH₂Cl₂ at 0 °C were added sequentially pyridine (21.4 mg, 22.1 μ L, 0.27 mmol), DMAP (8.3 mg, 0.07 mmol), and Ac₂O (13.9 mg, 12.8 µL, 0.14 mmol). After 15 min, the reaction was quenched with satd aq NH₄Cl (5 mL) and extracted with Et₂O (3 \times 10 mL). The dried $(MgSO_4)$ extract was concentrated in vacuo to give product 55 (31 mg, 64 μ mol, 94%) as a colorless oil: $[\alpha]^{23}_{D}$ (-) 39.3 (c 0.40, CHCl₃); IR (thin film) 3026, 2955, 2913, 2875, 1741, 1496, 1457, 1367, 1236, 1158, 1121, 1053, 1016, 966, 898, 793, 741, 692 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 7.15–7.28 (m, 5H), 6.32 (d, J = 15.9 Hz, 1H), 6.07–6.17 (m, 1H), 5.85 (s, 1H), 4.96 (s, 1H), 4.79 (s, 1H), 3.98-4.07 (m, 2H), 2.41-2.52 (m, 2H), 2.10 (dd, J = 5.7, 13.5 Hz, 1H), 2.00 (s, 3H), 1.85 (dd, J = 7.8, 13.2 Hz, 1H), 1.79 (s, 3H), 1.58-1.67 (m, 2H), 1.43 (s, 3H), 1.28-1.36 (m, 1H), 0.97 (t, J = 7.5 Hz, 9H), 0.75(d, J = 6.6 Hz, 3H), 0.61 (q, J=7.5 Hz, 6 H); $^{13}\mathrm{C}$ NMR (75 MHz, $\mathrm{CDCl}_3)$ δ 171.6, 145.3, 143.0, 138.3, 132.0, 128.9, 127.4, 127.3, 126.3, 125.3, 114.9, 78.7, 63.3, 46.4, 46.2, 35.6, 29.0, 27.4, 21.4, 19.6, 15.0, 7.6, 7.2; HRMS (FAB+) calcd for $C_{29}H_{46}O_3SiNa (M + Na) 493.3114$, found 493.3130.



Aldehyde 57. To a solution of 55 (31 mg, 66 μ mol) in *t*-BuOH (0.55 mL) and H₂O (0.55 mL) at rt was added AD mix β^{*53} (92 mg) and CH₃SO₂NH₂ (9.5 mg, 0.10 mmol). After 16 h, the reaction was quenched with 10% aq Na₂S₂O₃ (2 mL). After 30 min, the mixture was extracted with EtOAc (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 20% EtOAc/hexanes, to give the corresponding diol (29 mg, 58 μ mol, 88%, 6:1 dr).

To a solution of the above diol (26 mg, 52 μ mol) in THF/Et₂O/H₂O (1.7 mL, 1:1:1) at rt was added NaIO₄ (110 mg, 0.52 mmol). After 2 h, the reaction was guenched with a brine solution (2 mL) and extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo to give the aldehyde 57 (20.4 mg, 52 μ mol, 99%) as a colorless oil. The aldehyde intermediate was applied to the next step without further purification: $[\alpha]_{D}^{23} = -11.6$ (*c* 1.83, CHCl₃); IR (neat) 2957, 2877, 1741, 1724, 1458, 1368, 1239, 1050, 1017, 743 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 9.66 (t, J = 3.0 \text{ Hz}, 1\text{H}), 6.02 (s, 1\text{H}), 5.03 (s, 1\text{H}),$ 4.83 (s, 1H), 4.14–4.04 (m, 2H), 2.65 (dd, J = 15.1, 2.9 Hz, 1H), 2.46 (dd, *J* = 15.1, 3.1 Hz, 1H), 2.14 (dd, *J* = 13.6, 5.8 Hz, 1H), 2.04 (s, 3H), 1.92 (dd, J = 13.5, 7.8 Hz, 1H), 1.82 (s, 3H), 1.70 - 1.60 (m, 2H), 1.48 (s, 3H), 1.45–1.39 (m, 1H), 1.00–0.94 (m, 9H), 0.87 (d, J = 6.5 Hz, 3H), 0.68-0.56 (m, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 203.0, 171.2, 144.3, 141.2, 125.7, 115.1, 62.8, 54.0, 45.8, 35.3, 28.7, 27.9, 20.9, 19.2, 14.7, 7.1, 6.7, 6.6, 5.8; HRMS (EI⁺) calcd for C₂₂H₄₀O₄Si (M⁺) 396.2696, found 396.2678.



Sultam 60. Following the similar procedure described by Paquette,^{38a} Mg (36.0 g, 1.5 mol) was stirred vigorously at rt in a dry flask under Ar. After 120 h, when a black coating formed inside the flask, THF (200 mL) and 1,2-dibromoethane (2.60 g, 1.2 mL, 13.9 mmol) were added sequentially. After 30 min, a solution of allyl chloride **58** (17.0 g, 75.0 mmol) in THF (80 mL) was added slowly to the Mg slurry over 5 h. The resulted mixture was stirred overnight at rt to give 300 mL of Grignard reagent (0.12 M, 47%) as a gray solution. The concentration of the Grignard reagent was determined by the titration using menthol in the presence of 1,10-phenanthroline.⁵⁵

Separately, CuBr·SMe2 (7.29 g, 35.5 mmol) and LiCl (1.61 g, 37.9 mmol) were dissolved in THF (80 mL) and added to the solution of Grinard reagent (263 mL, 31.5 mmol) at -78 °C via syringe. TMSCl (3.96 g, 4.5 mL, 36.5 mmol) was then added followed by a solution of known sultam 59⁵⁶ (6.9 g, 24.3 mmol) in THF (60 mL). After another 90 min, the reaction was quenched with NH₄Cl-NH₄OH (9:1, pH 9, 60 mL), warmed to rt. The aqueous layer was extracted with EtOAc ($3 \times$ 200 mL). The organic phase was washed with satd aq NaCl (100 mL). The dried extract $(MgSO_4)$ was concentrated in vacuo and purified by chromatography over silica gel, eluting with 8-15% EtOAc/hexanes, to give the product 60 (11.2 g, 34.4 mmol, 97%) as a colorless oil: $[\alpha]^{23}_{D}$ = -68.0 (c 0.51, CHCl₃); IR (neat) 2959, 2927, 2851, 1693, 1512, 1454, 1328, 1246, 1032 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.28 (d, J = 8.7 Hz, 2H), 6.88 (d, J = 8.7 Hz, 2H), 5.12 (s, 1H), 4.96 (s, 1H), 4.44 (t, J = 11.8 Hz, 2H), 3.94 (t, J = 11.8 Hz, 2H), 4.00 (dd, J = 14.0 Hz, 2H), 3.88 (t, *J* = 6.2 Hz, 1H), 3.82 (s, 3H), 3.46 (dd, *J* = 23.0, 13.8 Hz, 2H), 2.78 (dd, *J* = 16.1, 5.7 Hz, 1H), 2.51 (dd, J = 16.1, 7.6 Hz, 1H), 2.30–2.40 (m, 1H), 2.02-2.15 (m, 4H), 1.82-1.96 (m, 3H), 1.28-1.45 (m, 3H), 1.15 (s, 3H), 0.99 (d, J = 6.4 Hz, 3H), 0.98 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 159.1, 144.1, 130.6, 129.3, 113.7, 113.6, 72.4, 71.7, 65.2, 55.3, 53.0, 48.3, 47.7, 44.7, 42.5, 40.8, 38.6, 32.9, 28.0, 26.5, 20.8, 19.9; HRMS (ES⁺) calcd for C₂₆H₃₇NO₅SNa (M + Na) 498.2290, found 498.2271.



Aldehyde SI-13. To a stirred solution of sultam 60 (11.0 g, 23.1 mmol) in CH₂Cl₂ (115 mL) at -78 °C was added DIBAL-H (50.8 mL, 50.8 mmol, 1.0 M in CH₂Cl₂). After 2 h, the reaction was carefully quenched with methanol (2.0 mL) and poured into aq sodium potassium tartrate (250 mL, 10% aq) at rt. The reaction flask was rinsed with an additional portion of CH_2Cl_2 (150 mL). After 3 h, the aqueous layer was extracted with CH_2Cl_2 (3 × 150 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give the aldehyde SI-13 (5.9 g, 22.6 mmol, 98%) as a colorless oil. Further elution with 5% MeOH/EtOAc gave recovered auxiliary SI-14 (4.9 g, 22.4 mmol, 97%). **SI-13**: $[\alpha]_{D}^{23}$ = +5.93 (c 0.91, CHCl₃); IR (neat) 2956, 2929, 2837, 1723, 1612, 1513, 1247, 1077, 1034 cm $^{-1}$; ¹H NMR (300 MHz, CDCl₃) δ 9.75 (t, J = 1.5 Hz, 1H), 7.29 (d, J = 8.7 Hz, 2H), 6.90 (d, J = 8.7 Hz, 2H), 5.14 (d, J = 1.2 Hz, 1H), 4.95 (s, 1H), 4.44 (s, 2H), 3.93 (s, 2H), 3.83 (s, 3H), 2.47 (ddd, J = 14.0, 4.0, and 1.3 Hz, 1H), 2.17–2.34 (m, 2H), 2.01–2.11 (m, 2H), 0.98 (d, J = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 202.6, 159.2, 143.9, 130.3, 129.3, 114.1, 113.8, 72.4, 71.7, 55.3, 50.6, 41.0, 26.3, 20.1; HRMS (ES⁺) calcd for C₁₆H₂₂O₃Na (M + Na) 285.1467, found 285.1494.



Aldohol SI-15. To a stirred solution of aldehyde **SI-13** (5.6 g, 21.4 mmol) in CH₂Cl₂ (110 mL) at -78 °C was added DIBAL-H (28.3 mL, 28.3 mmol, 1.0 M in CH₂Cl₂). After 1 h, the reaction was carefully quenched with methanol (2.0 mL) and poured into aq sodium potassium tartrate (250 mL, 10% aq) at rt. The reaction flask was rinsed with an additional portion of CH₂Cl₂ (150 mL). After 3 h, the aqueous layer was extracted with CH₂Cl₂ (3 × 150 mL). The dried extract

(MgSO₄) was concentrated in vacuo to give the alcohol **SI-15** (5.6 g, 20.8 mmol, 97%) as a colorless oil: $[\alpha]^{23}{}_{\rm D} = -2.94$ (c 0.51, CHCl₃); IR (neat) 3407, 2926, 2868, 1612, 1513, 1461, 1248, 1059, 1036 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.29 (d, J = 8.7 Hz, 2H), 6.90 (d, J = 8.7 Hz, 2H), 5.10 (s, 1H), 4.94 (s, 1H), 4.44 (t, J = 12.2 Hz, 2H), 3.91 (t, J = 13.4 Hz, 2H), 3.82 (s, 3H), 3.61–3.78 (m, 2H), 2.15 (dd, J = 13.8, 6.0 Hz, 1H), 1.89–1.96 (m, 1H), 1.89–1.96 (m, 1H), 1.72–1.86 (m, 1H), 1.57–1.69 (m, 1H), 1.23–1.45 (m, 1H), 0.91 (d, J = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 159.2, 144.6, 130.4, 129.3, 113.8, 113.4, 72.6, 71.6, 61.0, 55.3, 41.3, 39.7, 27.5, 19.7, 18.8; HRMS (ES⁺) calcd for C₁₆H₂₄O₃Na (M + Na) 287.1623, found 287.1649.



TBS Ether SI-16. To a stirred solution of alcohol SI-15 (5.5 g, 20.8 mmol) in DMF (50 mL) at rt were sequentially added imidazole (3.4 g, 50.0 mmol) and TBSCl (3.8 g, 25.2 mmol). After 1 h, the reaction was quenched with satd aq NH₄Cl (50 mL) and extracted with Et₂O $(3 \times 150 \text{ mL})$. The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/ hexanes, to give TBS ether SI-16 (7.7 g, 20.3 mmol, 97%) as a colorless oil: $\left[\alpha\right]^{23}_{D} = -3.27$ (c 1.31, CHCl₃); IR (neat) 2954, 2928, 2856, 1513, 1249, 1094, 835 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.29 (d, J = 8.7 Hz, 2H), 6.90 (d, J = 8.7 Hz, 2H), 5.11 (s, 1H), 4.93 (s, 1H), 4.44 (s, 2H), 3.92 (s, 2H), 3.83 (s, 3H), 3.63-3.70 (m, 2H), 2.13 (dd, J = 13.8, 6.3 Hz, 1H), 1.88–1.96 (m, 1H), 1.76–1.86 (m, 1H), 1.57–1.69 (m, 1H), 1.28–1.35 (m, 1H), 0.91 (s, 9H), 0.89 (d, J = 6.6 Hz, 3H), 0.06 (s, 6H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl_3) δ 159.1, 144.7, 130.6, 129.3, 113.8, 112.8, 72.6, 71.6, 61.3, 55.3, 41.4, 39.8, 27.5, 26.0, 19.6, 18.3, -5.3; HRMS (ES⁺) calcd for C₂₂H₃₈O₃NaSi (M + Na) 401.2488, found 401.2489.



Alcohol SI-17. To a stirred solution of TBS ether SI-16 (3.85 g, 10.2 mmol) in CH₂Cl₂/pH 7 buffer (10: 1, 110 mL) was added DDQ (2.77 g, 12.2 mmol) at rt. After 1 h, the reaction was quenched with satd aq NaHCO₃ (50 mL) and extracted with Et₂O (3×100 mL). The dried $(MgSO_4)$ extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 8% EtOAc/hexanes, to give a mixture of product SI-17 and 4-methoxybenzaldehyde (3.90 g, 1:1 mol/mol, 9.9 mmol, 97%) as a colorless oil. An analytically pure sample was prepared by chromatography over silica gel, eluting with 3-5% EtOAc/hexanes, for characterization, but the product mixture was used in the subsequent step without complete removal of 4-methoxybenzaldehyde. SI-17: $[\alpha]^{23}_{D} = -6.09$ (*c* 1.21, CHCl₃); IR (neat) 3338, 2955, 2929, 2858, 1471, 1463, 1255, 1098, 835 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.09 (d, J = 1.5 Hz, 1H), 4.89 (s, 1H), 4.07 (d, *J* = 6.3 Hz, 2H), 3.61–3.75 (m, 2H), 2.13 (dd, *J* = 13.5, 6.0 Hz, 1H), 1.75– 1.95 (m, 2H), 1.55–1.66 (m, 1H), 1.41–1.50 (m, 1H), 1.27–1.38 (m, 1H), 0.91 (s, 9H), 0.89 (d, J = 6.6 Hz, 3H), 0.07 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 147.6, 110.7, 65.8, 61.2, 41.2, 39.6, 27.6, 26.0, 19.7, 18.3, -5.3; HRMS (EI⁺) calcd for $C_{14}H_{31}O_2Si (M + H) 259.2093$, found 259.2091.



Aldehyde 61. To a stirred solution of alcohol SI-17 and 4-methoxybenzaldehyde (7.8 g, 1:1 mol/mol, 19.7 mmol) in CH_2Cl_2 (200 mL) were sequentially added NaHCO₃ (3.0 g, 35.7 mmol) and DMP (10.0 g, 23.7 mmol) at rt. After 1 h, the reaction was quenched with satd aq NaHCO₃ (50 mL) and extracted with Et₂O (3 × 150 mL).

The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 3% EtOAc/hexanes, to give aldehyde **61** (4.6 g, 17.7 mmol, 90%) as a colorless oil: $[\alpha]^{23}_{D} = -8.20$ (*c* 1.21, CHCl₃); IR (neat) 2956, 2929, 2857, 1698, 1255, 1099, 835 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.56 (s, 1H), 6.28 (s, 1H), 6.07 (s, 1H), 3.60–3.73 (m, 2H), 2.28 (dd, *J* = 14.1, 6.9 Hz, 1H), 2.12 (dd, *J* = 13.8, 8.1 Hz, 1H),1.77–1.86 (m, 1H), 1.53–1.64 (m, 1H), 1.29–1.39 (m, 1H), 0.91 (s, 9H), 0.89 (d, *J* = 6.6 Hz, 3H), 0.07 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 194.7, 149.0, 135.2, 61.1, 39.5, 35.2, 28.4, 25.9, 25.5, 19.4, –5.4; HRMS (EI⁺) calcd for C₁₄H₂₈O₂Si (M) 256.1859, found 256.1861.



Methyl Ketone SI-18. A solution of aldehyde **61** (4.5 g, 17.5 mmol) and ylide **62**⁵⁷ (10.2 g, 30.7 mmol) in toluene (60 mL) was refluxed in a sealed tube (oil bath 112 °C). After 16 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes (1% Et₃N added), to give diene **SI-18** (5.0 g, 16.1 mmol, 92%) as a slightly yellow oil: $[\alpha]^{23}_{D} = -41.1$ (*c* 0.53, CHCl₃); IR (neat) 2955, 2928, 2857, 1671, 1255, 1100, 836, 775 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.90 (s, 1H), 5.29 (s, 1H), 5.14 (s, 1H), 3.62–3.73 (m, 2H), 2.37 (s, 3H), 2.30 (dd, *J* = 10.2, 4.8 Hz, 1H), 2.05 (dd, *J* = 10.5, 6.3 Hz, 1H), 1.97 (d, *J* = 10.2 Hz, 3H), 1.71–1.78 (m, 1H), 1.59–1.64 (m, 1H), 1.34–1.38 (m, 1H), 0.91 (s, 9H), 0.89 (d, *J* = 6.6 Hz, 3H), 0.07 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 200.3, 143.7, 140.9, 137.8, 118.9, 61.0, 45.1, 39.6, 28.4, 25.9, 25.7, 19.3, 18.3, 13.1, -5.3; HRMS (FAB⁺) calcd for C₁₈H₃₅O₂Si (M + H) 311.2406, found 311.2400.



Triene Ester 63. To a stirred slurry of NaH (1.29 g, 32.2 mmol) in DME (50 mL) was added ethyl 2-(diethoxyphosphoryl)acetate (6.48 g, 5.74 mL, 28.9 mmol) at rt. After 1 h, a solution of methyl ketone SI-18 (5.00 g, 16.1 mmol) in DME (25 mL) was added. The resulted solution was refluxed for 3 h and then quenched with $H_2O(15 \text{ mL})$ and extracted with Et₂O (3×150 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2% EtOAc/hexanes (1% Et₃N added), to give triene ester 63 (4.42 g, 11.6 mmol, 70%) as a colorless oil: $[\alpha]_{D}^{23} = -34.7$ (*c* 1.66, CHCl₃); IR (neat) 2955, 2928, 2857, 1716, 1610, 1255, 1163, 1098, 836, 775 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.24 (s, 1H), 5.92 (s, 1H), 5.13 (s, 1H), 4.93 (s, 1H), 4.19 (q, J = 7.1 Hz, 2H), 3.61–3.68 (m, 2H), 2.36 (s, 3H), 2.20 (dd, J = 13.3, 5.9 Hz, 1H), 1.93–2.00 (m, 4H), 1.53–1.71 (m, 2H), 1.31 (t, *J* = 7.2 Hz, 3H), 0.89 (s, 9H), 0.87 (d, *J* = 6.9 Hz, 3H), 0.05 (s, 6H); 13 C NMR (100 MHz, CDCl₃) δ 167.4, 156.4, 144.7, 137.8, 132.4, 116.4, 115.9, 61.2, 59.7, 45.7, 39.7, 28.3, 25.9, 19.5, 18.3, 15.8, 15.5, 14.4, -5.3; HRMS (EI⁺) calcd for $C_{22}H_{40}O_3Si(M + H)$ 380.2747, found 380.2732.



Allyl Alcohol SI-19. To a stirred solution of triene ester 63 (8.61 g, 22.6 mmol) in THF (200 mL) was added DIBAL-H (46 mL, 46.0 mmol, 1 M in toluene) at -78 °C. After 1.5 h, the reaction was quenched with MeOH (1.0 mL) and poured into aq sodium potassium tartrate (350 mL, 10% aq) at rt. The reaction flask was rinsed with an additional

portion of Et₂O (50 mL). After 3 h, the aqueous layer was extracted with Et₂O (3 × 200 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 8% EtOAc/ hexanes, to give allyl alcohol **SI-19** (6.20 g, 18.3 mmol, 81%) as a colorless oil: $[\alpha]^{23}_{D} = -36.6$ (*c* 1.66, CHCl₃); IR (neat) 3327, 2954, 2928, 2857, 1471, 1462, 1376, 1255, 1098, 1006, 835, 775 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.97 (s, 1H), 5.80 (t, *J* = 6.3 Hz, 1H), 5.08 (s, 1H), 4.87 (s, 1H), 4.34 (d, *J* = 6.4 Hz, 2H), 3.46-3.72 (m, 2H), 2.18 (dd, *J* = 13.6, 6.0 Hz, 1H), 1.99 (d, *J* = 0.8 Hz, 3H), 1.93 (dd, *J* = 13.5, 5.2 Hz, 1H), 1.86 (s, 3H), 1.54-1.73 (m, 2H), 1.28-1.37 (m, 1H), 0.90 (s, 9H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.05 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 145.2, 139.3, 137.6, 128.1, 125.9, 115.4, 61.3, 60.1, 46.0, 39.7, 28.3, 25.9, 19.5, 18.3, 15.6, 14.2, -5.3; HRMS (EI⁺) calcd for C₂₀H₃₈O₂Si (M) 338.2641, found 338.2612.



Epoxide 64. To a stirred solution of (+)-DIPT (41.5 mg, 0.18 mmol) and 4 Å molecular sieves (about 200 mg) in CH₂Cl₂ (4.0 mL) were sequentially added $Ti(O-i-Pr)_4$ (34 mg, 34.6 μ L, 0.12 mmol) and TBHP (236 µL, 1.30 mmol, 5.0–6.0 M in decane) at -20 °C. After 20 min, the reaction mixture was cooled to -78 °C, and a precooled solution (-78°C) of allyl alcohol SI-19 (200 mg, 0.59 mmol) in CH2Cl2 (4.0 mL) was added via cannula. The resulted solution was warmed to -50 °C. After another 60 min, the reaction was guenched with pH 7 phosphate buffer (0.5 mL), filtered over Celite, and extracted with Et_2O (3 × 20 mL). The dried organic layers (MgSO₄) were concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give epoxide **64** (167 mg, 0.47 mmol, 80%) as a colorless oil: $[\alpha]_{D}^{23} = -17.3$ (*c* 1.66, CHCl₃); IR (neat) 3430, 2954, 2927, 2856, 1471, 1463, 1378, 1255, 1097, 836, 775 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.89 (s, 1H), 5.03 (s, 1H), 4.86 (s, 1H), 3.86–3.92 (m, 1H), 3.73–3.81 (m, 1H), 3.62–3.71 (m, 2H), 3.02 (dd, J = 10.5, 6.3 Hz, 1H), 2.14 (dd, J = 13.5, 5.6 Hz, 1H), 1.91 (dd, J = 13.5, 8.4 Hz, 1H), 1.84 (d, J = 1.1 Hz, 3H), 1.54-1.66 (m, 2H), 1.45(s, 3H), 1.27–1.34 (m, 1H), 0.91 (s, 9H), 0.83 (d, J = 6.4 Hz, 3H), 0.06 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 144.0, 137.5, 126.2, 115.2, 63.7, 61.3, 45.6, 39.8, 28.2, 26.0, 19.3, 18.3, 16.7, 14.8, -5.3; HRMS (CI⁺) calcd for $C_{20}H_{39}O_3Si (M + H) 355.2669$, found 355.2666.



Diol 65. To a stirred solution of epoxide 64 (1.5 g, 4.23 mmol) in THF (30 mL) was added Red-Al (1.5 mL, 9.91 mmol, 65% w/v in toluene) at 0 °C. After 1 h, another portion of Red-Al (1.5 mL, 9.91 mmol, 65% w/v in toluene) was added. After another 1.5 h, the reaction was quenched with H₂O (0.10 mL), and extracted with CH₂Cl₂ (3 \times 150 mL). The dried organic layers (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 6-12% EtOAc/hexanes (1% Et₃N added), to give diol 65 (1.05 g, 2.96 mmol, 70%) as a colorless oil: $[\alpha]_{D}^{23} = -29.4$ (c 0.81, CHCl₃); IR (neat) 3389, 2955, 2928, 2858, 1471, 1462, 1382, 1255, 1097, 836, 775 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.10 (s, 1H), 5.04 (d, J = 0.9 Hz, 1H), 4.84 (s, 1H), 3.62–3.77 (m, 4H), 3.04 (s, br, 1H), 2.60 (s, br, 1H), 2.16 (dd, J = 13.2, 5.1 Hz, 1H), 1.88–1.96 (m, 3H), 1.79 (d, J = 0.9 Hz, 3H), 1.55–1.70 (m, 2H), 1.37 (s, 1H), 1.27–1.34 (m, 1H), 0.91 (s, 9H), 0.86 (d, J = 6.5 Hz, 3H), 0.07 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 145.1, 141.7, 125.1, 114.4, 77.1, 61.6, 60.4, 46.1, 40.1, 39.7, 28.6, 28.2, 26.0, 19.6, 18.4, 15.1, -5.2; HRMS (ES^+) calcd for $C_{20}H_{40}O_3SiNa$ (M + Na) 379.2644, found 379.2643.



Ester 66. To a stirred solution of diol **65** (2.10 g, 5.89 mmol) in CH_2Cl_2 (50 mL) were sequentially added pyridine (1.37 g, 1.40 mL, 17.7 mmol) and trichloroacetyl chloride (1.29 g, 0.79 mL, 7.09 mmol). After 3 h, the reaction was quenched with satd aq NH_4Cl (10 mL) and extracted with ether (3 × 30 mL). The dried extract (MgSO₄) was concentrated in vacuo to give crude ester (3.30 g) as a colorless oil, which was used in the next step without further purification.

To a stirred solution of crude ester (3.30 g) in $CH_2Cl_2/EtOH$ (1:1, 50 mL) was added CSA (2.31 g, 9.88 mmol) at 0 °C. After 1.5 h, the reaction was quenched with satd aq NaHCO₃ (10 mL) and extracted with ether (3 × 40 mL). The dried extract (MgSO₄) was concentrated in vacuo to give crude diol (2.02 g), which was used in the next step without further purification.

To a stirred solution of crude diol (2.02 g) in CH₂Cl₂ (50 mL) were added sequentially pyridine (1.53 g, 1.57 mL, 19.5 mmol) and Ac₂O (0.99 g, 0.92 mL, 9.73 mmol). After 1.5 h, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with ether (3 × 40 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give **66** (1.90 g, 4.42 mmol, 75% over 3 steps) as a colorless oil: $[\alpha]^{23}_{D} = -14.1 (c 1.16, CHCl_3)$; IR (neat) 3481, 2962, 2928, 1766, 1739, 1720, 1458, 1368, 1247, 828, 682 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.01 (s, 1H), 5.04 (s, 1H), 4.85 (s, 1H), 4.47–4.36 (m, 2H), 4.15–4.06 (m, 2H), 2.12–2.02 (m, 3H), 2.03 (s, 3H), 1.97–1.89 (m, 3H), 1.83 (s, 3H), 1.70–1.60 (m, 2H), 1.50–1.38 (m, 1H), 1.41 (s, 3H), 0.89 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 161.9, 144.1, 141.4, 124.9, 115.3, 74.6, 66.5, 62.3, 45.6, 37.8, 35.2, 28.7, 28.3, 21.0, 19.4, 14.9; HRMS (EI⁺) calcd for C₁₈H₂₇O₅Cl₃ (M⁺) 428.0924, found 428.0932.



TES Ether 67. To a stirred solution of alcohol **66** (1.90 g, 4.4 mmol) in CH₂Cl₂/EtOH (1:1, 50 mL) was added NH₃·H₂O (15 mL) at rt. After 1 h, the reaction was quenched with satd aq NH₄Cl (15 mL) and extracted with ether (3×40 mL). The dried extract (MgSO₄) was concentrated in vacuo to afford crude diol (1.55 g), which which was used in the next step without further purification.

To a stirred solution of crude diol (1.55 g) in CH₂Cl₂/Et₃N (1:1, 30 mL) was added freshly distilled TESOTf (3.52 g, 3.01 mL, 13.1 mmol) at -78 °C. After 20 min, the reaction was quenched with satd aq NaHCO₃ (10 mL) and extracted with ether (3 \times 40 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2% EtOAc/hexanes, to give TES ether 67 (2.12 g, 4.09 mmol, 93% over two steps) as a colorless oil: $[\alpha]^{23}_{D} = -18.4$ (c 1.11, CHCl₃); IR (neat) 29554, 2912, 2876, 1748, 1458, 1238, 1086, 1016, 741 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 5.89 (s, 1\text{H}), 5.00 (d, J = 1.2 \text{ Hz}, 1\text{H}), 4.81 (d, J = 1.2 \text{ Hz})$ Hz, 1H), 4.18-4.05 (m, 2H), 3.72-3.63 (m, 1H), 3.54-3.41 (m, 1H), 2.15 (dd, J = 13.5, 5.4 Hz, 1H), 2.05 (s, 3H), 1.95-1.89 (m, 2H), 1.87-1.78 (m, 1H), 1.77 (d, J = 1.2 Hz, 3H), 1.71–1.58 (m, 2H), 1.49–1.39 (m, 1H), 1.42 (s, 3H), 1.12–0.91 (m, 18H), 0.87 (d, J = 6.6 Hz, 3H), 0.74– 0.50 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 171.1, 144.8, 142.3, 124.3, 114.4, 77.2, 62.3, 59.5, 46.0, 44.4, 35.3, 28.8, 27.8, 21.0, 19.3, 14.6, 7.2, 6.9, 6.8, 6.4, 5.8, 4.4; HRMS (EI⁺) calcd for C₂₈H₅₆O₄Si₂Na (M + Na) 535.3615, found 535.3637.



Aldehyde 57. TES ether 67 (2.1 mg, 4.09 mmol) was dissolved in a stirred solution of HOAc/THF/H₂O (34 mL, 8:8:1) at 0 °C. After 1.5 h, the reaction was then quenched with solid NaHCO₃ and extracted with ether (4 \times 50 mL). The dried extract (MgSO₄) was concentrated in vacuo to afford crude alcohol (2.0 g), which was used in the next step without further purification.

To a stirred solution of crude alcohol (2.0 g) in CH_2Cl_2 (20 mL) were added sequentially solid NaHCO₃ (1.0 g, 11.9 mmol) and DMP (2.16 g, 5.09 mmol) at rt. After 1 h, the reaction was quenched with satd aq NaHCO₃ (15 mL) and extracted with ether (3 × 40 mL). The dried extract (MgSO₄) was concentrated in vacuo andpurified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give aldehyde 57 (1.36 g, 3.43 mmol, 74% over two steps) as a colorless oil.



Aldol Product SI-20. To a solution of methyl ketone 54 (38 mg, 76 μ mol) in Et₂O (0.4 mL) at -78 °C was added LDA⁵⁴ (85.0 μ L, 1.0 M in THF). After 20 min, a precooled (-78 °C) solution of aldehyde intermediate (20 mg, 50 μ mol) in Et₂O (0.25 mL) was transferred in one portion via cannula at -78 °C. After 20 min, the reaction was quenched with satd aq $NH_4Cl (3 mL)$ and extracted with $Et_2O (3 \times 10 mL)$. The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give the alcohol intermediate SI-20 (28 mg, 31 μ mol, 62%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 15.7 (*c* 1.55, CHCl₃); IR (neat) 3515, 2955, 2912, 2876, 1741, 1717, 1615, 1515, 1417, 1368, 1247, 1117, 1037, 1008, 899, 806, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.28 (d, J = 9.0 Hz, 2H), 6.84 (d, J = 8.7 Hz, 2H), 5.87 (s, 1H), 5.01 (s, 1H), 4.83 (s, 1H), 4.50 (d, J = 11.1 Hz, 1H), 4.34–4.51 (m, 1H), 4.33 (d, J = 11.1 Hz, 1H), 4.04–4.14 (m, 2H), 3.80 (s, 3H), 3.71–3.78 (m, 3H), 3.03 (dd, J = 7.5, 17.4 Hz, 1H), 2.29 (dd, J = 4.5, 17.4 Hz, 1H), 2.12 (dd, J = 5.7, 8.7 Hz, 1H), 2.03 (s, 3H), 1.83-1.96 (m, 2H), 1.82 (s, 3H), 1.59-1.73 (m, 3H), 1.52 (s, 3H), 1.42–1.49 (m, 4H), 1.11 (d, J = 6.0 Hz, 3H), 0.86–0.98 (m, 21H), 0.82 (d, J = 6.6 Hz, 3H), 0.51–0.67 (m, 12H), 0.06 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 211.7, 171.6, 159.6, 144.6, 143.8, 130.3, 130.1, 125.2, 115.6, 114.0, 88.4, 79.8, 77.6, 72.8, 66.2, 65.1, 63.3, 55.6, 48.5, 46.7, 46.2, 44.8, 35.7, 32.5, 29.1, 26.5, 25.2, 21.4, 19.7, 15.1, 13.3, 7.5, 7.4, 7.0, 5.7, 0.7; HRMS (ES+) calcd for C₄₈H₈₈O₉Si₃Na (M + Na) 915.5634, found 915.5564.



TES Ethers 68 and 69. To a solution of the above alcohol intermediate (40 mg, 45 μ mol) in CH₂Cl₂ (0.15 mL) and Et₃N (0.15 mL) at -78 °C was added TESOTF (32.0 mg, 28.3 μ L, 0.12 mmol). After

20 min, the reaction was quenched with satd aq NH₄Cl (3 mL). The mixture was extracted with Et₂O (3×10 mL), and the dried (MgSO₄) extract was concentrated in vacuo to give 69 (36 mg, 36 μ mol, 82%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 18.6 (c 1.58, CHCl₃); IR (neat) 2955, 2926, 2876, 2855, 1742, 1716, 1615, 1515, 1458, 1366, 1249, 1127, 1084, 1006, 899, 835, 776, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.4 Hz, 2H), 6.88 (d, J = 8.4 Hz, 2H), 5.84 (s, 1H), 5.04 (s, 1H), 4.89 (s, 1H), 4.59 (d, J = 10.8 Hz, 1H), 4.29–4.35 (m, 1H), 4.27 (d, J = 10.8 Hz, 1H), 4.10– 4.17 (m, 2H), 3.84 (s, 4H), 3.76–3.78 (m, 1H), 3.71 (d, J = 7.2 Hz, 1H), 3.19 (dd, J = 8.4, 17.6 Hz, 1H), 2.54 (dd, J = 3.3, 17.6 Hz, 1H), 2.11-2.15 (m, 1H), 2.06 (s, 3H), 1.86-1.98 (m, 2H), 1.85 (s, 3H), 1.63-1.77 (m, 4H), 1.52–1.56 (m, 1H), 1.45 (s, 3H), 1.30–1.38 (m, 2H), 1.13 (d, J = 6.4 Hz, 3H), 0.83–0.98 (m, 30H), 0.86 (d, J = 6.4 Hz, 3H), 0.52–0.64 (m, 18H), 0.15 (s, 9H); 13 C NMR (75 MHz, CDCl₃) δ 211.3, 171.2, 159.1, 144.4, 142.6, 130.1, 129.7, 125.1, 115.0, 113.5, 88.2, 77.4, 76.4, 72.0, 65.9, 63.0, 55.3, 50.0, 46.5, 46.0, 44.0, 35.3, 32.3, 29.7, 28.7, 28.0, 24.7, 21.0, 19.3, 14.9, 13.3, 7.3, 7.1, 7.0, 6.8, 6.6, 6.4, 5.3, 5.1, 0.4; HRMS (ES+) calcd for C₅₄H₁₀₂O₉Si₄Na (M + Na) 1029.6499, found 1029.6470.



Alcohol 70. To a flask with 69 (180 mg, 0.18 mmol) at 0 °C was added a freshly prepared stock solution of HOAc/THF/H2O (3.0 mL, 8:8:1) dropwise. After 40 min, the reaction was guenched with satd aq NaHCO₃ (20 mL) carefully and extracted with Et_2O (3 × 10 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give 70 (102 mg, 0.11 mmol, 61%) as a colorless oil: $[\alpha]^{23}{}_{D}$ (+) 23.7 (c 0.60, CHCl₃); IR (neat) 3546, 2955, 2926, 2876, 2855, 1742, 1716, 1615, 1515, 1458, 1366, 1249, 1127, 1084, 1006, 899, 835, 776, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.4 Hz, 2H), 6.89 (d, J = 8.4 Hz, 2H), 5.87 (s, 1H), 5.02 (s, 1H), 4.86 (s, 1H), 4.55 (d, J = 10.8 Hz, 1H), 4.23-4.29 (m, 1H), 4.23 (d, J = 10.8 Hz, 1H), 4.12-4.17 (m, 2H), 3.85 (s, 4H), 3.75-3.85 (m, 3H), 3.18 (dd, J = 9.2, 17.6 Hz, 1H), 2.54 (dd, J = 2.8, 17.6 Hz, 1H), 2.13 (dd, J = 5.6, 13.6 Hz, 1H), 2.06 (s, 3H), 1.86-1.94 (m, 2H), 1.83 (s, 3H), 1.63-1.80 (m, 4H), 1.52-1.56 (m, 1H), 1.45 (s, 3H), 1.38-1.45 (m, 2H), 1.19 (d, J = 6.0 Hz, 3H), 0.88-1.01(m, 33H), 0.55-0.68 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 211.3, 171.2, 159.1, 144.9, 141.7, 130.0, 129.8, 125.5, 114.9, 113.5, 88.5, 77.4, 77.2, 76.4, 71.8, 65.8, 63.2, 55.3, 49.5, 46.2, 45.8, 44.1, 35.4, 33.2, 29.7, 28.7, 28.5, 24.5, 21.0, 19.3, 14.9, 13.6, 7.3, 7.0, 6.95, 6.87, 5.2, 5.1; HRMS (ES+) calcd for $C_{51}H_{94}O_9Si_3Na$ (M + Na) 957.6103, found 957.6157.



Ketophosphonate 71. To a solution of phosphonate acid 11 (74 mg, 0.24 mmol) in PhMe (0.5 mL) at rt was added triethylamine (26.2 mg, 36.0 μ L, 0.24 mmol) and 2,4,6-trichlorobenzoyl chloride (56 mg, 36.0 μ L, 0,24 mmol). After 6 h, solvent of the reaction was removed in vacuo. A solution of the alcohol 70 (60 mg, 64 μ mol) in PhMe (0.5 mL) was transferred into the above flask via cannula and DMAP (29.6 mg, 0.24 mmol) were added sequentially. After 16 h, the reaction was purified by chromatography over silica gel, eluting with 70% EtOAc/hexanes, to give 71 (50 mg, 41 μ mol, 64%) as a colorless oil: [α]²³_D (+) 25.3 (*c* 1.50, CHCl₃); IR (neat) 2954, 2933, 2876, 1739, 1712, 1613, 1515, 1461, 1366, 1251, 1165, 1124, 1026, 968, 901, 835, 770, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.31 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 6.69 (t, *J* = 7.2 Hz, 1H), 5.85 (s, 1H), 5.03 (br, 2H), 4.88 (s, 1H), 4.62 (d, *J* = 10.8 Hz, 1H), 4.25–4.31 (m, 1H), 4.25 (d, *J* = 10.8 Hz, 1H), 4.05–4.21 (m, 6H), 3.84 (s, 3H), 3.81–3.83 (m, 1H),

3.72 (d, *J* = 6.0 Hz, 1H), 3.16 (dd, *J* = 8.8, 17.6 Hz, 1H), 3.12 (s, 1H), 3.07 (s, 1H), 2.67 (t, *J* = 7.2 Hz, 2H), 2.60 (dd, *J* = 2.8, 17.6 Hz, 1H), 2.16 (dd, *J* = 7.6, 14.8 Hz, 2H), 2.08–2.12 (m, 1H), 2.06 (s, 3H), 1.83–1.94 (m, 4H), 1.83 (s, 3H), 1.81 (s, 3H), 1.61–1.77 (m, 5H), 1.52–1.55 (m, 1H), 1.43 (s, 3H), 1.40–1.43 (m, 1H), 1.36 (t, *J* = 6.8 Hz, 6H), 1.22 (d, *J* = 6.0 Hz, 3H), 0.90–1.00 (m, 30H), 0.86 (d, *J* = 6.4 Hz, 3H), 0.54–0.66 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 211.7, 201.5, 201.4, 171.2, 167.7, 159.2, 144.5, 142.3, 140.6, 129.9, 129.7, 129.0, 125.2, 115.0, 113.6, 87.4, 76.0, 72.0, 68.9, 66.0, 63.1, 62.6, 62.5, 55.3, 49.9, 46.8, 45.9, 43.4, 43.1, 41.8, 39.3, 35.3, 33.0, 28.7, 28.2, 27.7, 22.3, 21.0, 20.8, 19.3, 16.4, 16.3, 14.9, 14.1, 12.5, 7.3, 7.0, 6.9, 6.8, 5.2, 5.1; HRMS (ES+) calcd for C₆₄H₁₁₅O₁₄PSi₃Na (M + Na) 1245.7230, found 1245.7181.



Alcohol 72. To a flask containing 71 (50.0 mg, 41 μ mol) at rt was added a premixed solution of Ba(OH)2.8H2O (1.5 mL, 0.1 M in MeOH). After 10 min, the mixture was purified by chromatography over silica gel, eluting with 50% EtOAc/hexanes, to give 72 (42 mg, 36 μ mol, 88%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 19.8 (c 0.78, CHCl₃); IR (neat) 3444, 2954, 2929, 2876, 1713, 1614, 1515, 1458, 1379, 1251, 1165, 1125, 1059, 1024, 970, 900, 835, 809, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₂) δ 7.33 (d, J = 8.8 Hz, 2H), 6.89 (d, J = 8.8 Hz, 2H), 6.69 (t, J = 7.6 Hz, 1H), 5.85 (s, 1H), 5.03–5.06 (m, 2H), 4.85 (s, 1H), 4.62 (d, J = 10.4 Hz, 1H), 4.12–4.27 (m, 7H), 3.84 (s, 3H), 3.81 (dd, J = 3.2, 6.8 Hz, 1H) 3.70-3.76 (m, 3H), 3.19 (dd, J = 9.2, 17.2 Hz, 1H), 3.13 (s, 1H), 3.07 (s, 1H), 2.68 (t, I = 7.2 Hz, 2H), 2.60 (dd, I = 2.8, 17.6 Hz, 1H), 2.18 (q, I =7.6 Hz, 2H), 2.10 (dd, J = 5.2, 18.4 Hz, 1H), 1.83–1.91 (m, 3H), 1.83 (s, 3H), 1.81 (s, 3H), 1.51-1.81 (m, 7H), 1.43 (s, 3H), 1.38-1.43 (m, 1H), 1.36 (t, J = 7.2 Hz, 6H), 1.23 (d, J = 6.0 Hz, 3H), 0.87-1.00 (m, 33H),0.54–0.66 (m, 18H); $^{13}\mathrm{C}$ NMR (100 MHz, CDCl₃) δ 212.3, 201.5, 201.4, 167.7, 159.2, 144.9, 141.7, 140.7, 130.0, 129.7, 129.0, 125.6, 114.9, 113.5, 87.6, 77.5, 75.9, 72.0, 68.9, 66.2, 62.6, 62.5, 61.0, 55.3, 49.5, 46.6, 46.2, 43.4, 43.1, 41.8, 40.1, 39.5, 32.9, 29.7, 28.4, 28.3, 27.7, 22.3, 20.8, 19.2, 16.4, 16.3, 15.1, 14.1, 13.9, 12.5, 7.3, 7.0, 6.9, 6.8, 5.2, 5.1; HRMS (ES+) calcd for $C_{62}H_{113}O_{13}PSi_3Na$ (M + Na) 1203.7124, found 1203.7054.



Macrolactone 51. To a solution of 72 (42 mg, 36 μ mol) in CH₂Cl₂ (1.2 mL) at rt. was added TPAP (15 mg, 43 $\mu mol).$ After 1 h, THF (3.3 mL), H₂O (82.8 μ L) and Ba(OH)₂·8H₂O (43 mg, 0.13 mmol) were sequentially added to the reaction. After 3 h, the reaction was directly purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give 51 (30.6 mg, 30 μ mol, 85%) as a colorless oil: $[\alpha]_{D}^{23}$ (+) 1.6 (c 0.45, CHCl₃); IR (neat) 2954, 2925, 2875, 2854, 1710, 1673, 1614, 1515, 1461, 1377, 1251, 1171, 1120, 1068, 1018, 985, 835, 808, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.6 Hz, 2H), 6.94–7.01 (m, 1H), 6.88 (d, J = 8.6 Hz, 2H), 6.71 (t, J = 7.6 Hz, 1H), 6.09 (d, J = 16.0 Hz, 1H), 5.77 (s, 1H), 5.05 (s, 1H), 4.96-5.04 (m, 1H), 4.87 (s, 1H), 4.53 (d, J = 10.8 Hz, 1H), 4.21–4.27 (m, 1H), 4.21 (d, J = 10.8 Hz, 1H), 3.84 (s, 3H), 3.67–3.74 (m, 2H), 3.15 (dd, J = 9.2, 17.6 Hz, 1H), 2.56-2.67 (m, 3H), 2.08-2.27 (m, 3H), 1.90 (s, 3H), 1.78 (s, 3H), 1.47–1.78 (m, 9H), 1.43 (s, 3H), 1.26–1.42 (m, 2H), 1.25 (d, J = 6.0 Hz, 3H), 0.90–1.00 (m, 30H), 0.83 (d, J = 6.4 Hz, 3H), 0.54–0.66 (m, 18H); 13 C NMR (100 MHz, CDCl₃) δ 209.8, 200.8, 167.6, 159.2, 147.0, 144.5, 142.7, 140.8, 132.3, 130.0, 129.5, 129.2, 125.3, 115.5, 113.6,

87.9, 71.7, 68.9, 65.4, 55.3, 49.2, 47.1, 45.8, 41.8, 40.6, 37.4, 32.0, 31.5, 31.4, 27.4, 22.9, 21.0, 19.5, 15.2, 14.2, 12.5, 7.3, 7.0, 6.9, 6.8, 5.2, 5.1; HRMS (ES+) calcd for $C_{58}H_{100}O_9Si_3Na$ (M + Na) 1047.6573, found 1047.6494.



Allylic Alcohol 75. To a solution of (S)-2-methyl-CBS-oxazaborolidine (10.1 μ L, 10 μ mol, 1.0 M in PhMe) in CH₂Cl₂ (0.3 mL) at rt was added catechol borane (0.20 mL, 0.20 mmol, 1.0 M CH₂Cl₂). After 10 min, the resulted solution was cooled to -20 °C. A solution of macrolactone 51 (26 mg, 25 μ mol) in CH₂Cl₂ (0.65 mL) was added into the above solution dropwise via cannula. After 2 h, the reaction was quenched with MeOH (2.0 mL) and concentrated in vacuo. The resultant mixture was purified by preparative TLC, eluting with 25% EtOAc/hexane, to give 75 (14 mg, 14 μ mol, 56%) as a colorless oil: $[\alpha]^{23}_{D}(+)$ 7.6 (c 0.88, CHCl₃); IR (neat) 3459, 2954, 2917, 2875, 2849, 1709, 1614, 1515, 1461, 1377, 1251, 1173, 1125, 1070, 1018, 835, 776, 743 cm⁻¹; ¹H NMR (400 MHz, CDCl₂) δ 7.33 (d, *J* = 8.6 Hz, 2H), 6.89 (d, J = 8.6 Hz, 2H), 6.72 (t, J = 7.6 Hz, 1H), 5.79 (s, 1H), 5.68-5.72 (m, 1H), 5.49 (dd, J = 6.8, 15.2 Hz, 1H), 5.05 (s, 1H), 4.97–5.04 (m, 1H), 4.87 (s, 1H), 4.57 (d, J = 10.8 Hz, 1H), 4.28-4.32 (m, 1H), 4.22 (d, J = 10.8 Hz, 1H), 4.13-4.17 (m, 1H), 3.85 (s, 3H), 3.65-3.73 (m, 2H), 3.16 (dd, J = 9.2, 18.0 Hz, 1H), 2.53 (dd, J = 3.2, 18.0 Hz, 1H), 2.21-2.24 (m, 4H), 1.98-2.04 (m, 4H), 1.88 (s, 3H), 1.82 (s, 3H), 1.75-1.84 (m, 2H), 1.53-1.70 (m, 7H), 1.44 (s, 3H), 1.30-1.36 (m, 1H), 1.24 (d, J = 5.6 Hz, 3H), 0.89–1.00 (m, 30H), 0.82 (d, J = 6.0 Hz, 3H), 0.54– 0.66 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 210.6, 167.8, 159.2, 145.1, 142.6, 141.7, 134.5, 130.4, 129.9, 129.8, 128.5, 125.6, 115.1, 113.5, 88.1, 77.2, 76.6, 72.4, 71.7, 68.6, 65.5, 55.3, 53.5, 49.5, 46.4, 44.9, 41.1, 39.4, 36.6, 32.2, 31.4, 28.6, 27.5, 23.7, 21.1, 19.8, 15.0, 12.6, 11.9, 7.3, 7.0, 6.9, 6.8, 5.2, 5.1; HRMS (ES+) calcd for $C_{58}H_{102}O_9Si_3Na$ (M + Na) 1049.6729, found 1049.6678.



Hydroxyl Epoxide 76. To a solution of (+)-DIPT (0.11 mL, 89 µmol, 0.8 M in CH₂Cl₂) were added CH₂Cl₂ (0.30 mL) and 4 Å MS (30 mg). The resulting mixture was cooled to -20 °C, and Ti(O-*i*-Pr)₄ (21.6 mg, 20.0 µL, 76 µmol) was added. After 30 min, TBHP (14.2 µL, 76 μ mol, 5.5 M in decane) was added dropwise. After another 30 min, a solution of alcohol 75 (13 mg, 13 μ mol) in CH₂Cl₂ (0.30 mL) was added dropwise via cannula, and the reaction was allowed to warm to 0 °C. After 16 h, the reaction was cooled back to -20 °C, and a brine solution of NaOH (0.5 mL, 1.0 M) was added. After 5 min, the reaction was diluted with brine (0.5 mL) and CH_2Cl_2 (0.5 mL) and warmed to rt. Next, the mixture was extracted with CH_2Cl_2 (3 × 15 mL) and the dried (MgSO₄) extract concentrated in vacuo. The crude mixture was purified by preparative TLC, eluting with 25% EtOAc/hexane, to give vinyl epoxide 76 (7.0 mg, 7.0 μ mol, 54%) as a colorless oil: $[\alpha]^{23}_{D}$ (+) 12.2 (c 0.36, CHCl₃); IR (neat) 3467, 3017, 2954, 2917, 2875, 2849, 2107, 1709, 1515, 1462, 1378, 1251, 1173, 1125, 1070, 1018, 835, 756, 667 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 8.8 Hz, 2H), 6.77 (t, J = 7.6 Hz, 1H), 5.80 (s, 1H), 5.06 (s, 1H), 4.95-5.00 (m, 1H), 4.87 (s, 1H), 4.60 (d, J = 10.8 Hz, 1H), 4.22-4.30 (m, 1H), 4.21 (d, J = 10.8 Hz, 1H), 3.85 (s, 3H), 3.75–3.84 (m, 1H), 3.67– 3.74 (m, 2H), 3.22 (dd, J = 9.2, 18.0 Hz, 1H), 3.05-3.10 (m, 1H), 2.78-2.81 (m, 1H), 2.54 (dd, J = 3.2, 18.0 Hz, 1H), 2.38 (br, 1H), 2.21-2.30 (m, 4H), 1.89–1.92 (m, 1H), 1.89 (s, 3H), 1.84 (s, 3H), 1.75–1.86 (m, 3H), 1.53–1.70 (m, 7H), 1.45 (s, 3H), 1.30–1.36 (m, 1H), 1.26 (d, J = 5.6 Hz, 3H), 0.91–1.01 (m, 30H), 0.56–0.67 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 210.8, 167.8, 159.2, 144.7, 142.4, 141.2, 129.9, 129.8, 128.9, 125.6, 115.4, 113.6, 88.1, 77.1, 71.8, 68.7, 68.2, 65.4, 61.3, 55.3, 54.2, 49.8, 46.5, 45.4, 41.8, 39.5, 33.3, 31.1, 30.8, 28.5, 27.7, 24.1, 21.1, 20.1, 15.1, 12.5, 11.8, 7.3, 7.0, 6.9, 5.3, 5.1; HRMS (ES+) calcd for C₅₈H₁₀₂O₁₀Si₃Na (M + Na) 1065.6679, found 1065.6774.



Selenide 78. To a solution of vinyl epoxide 76 (5.0 mg, 5 μ mol) in THF (0.15 mL) at rt were added 77 (33 mg, 0.15 mmol) and PBu₃ $(29 \text{ mg}, 36.2 \,\mu\text{L}, 0.15 \text{ mmol})$. After 1 h, the reaction mixture was directly purified by preparative TLC, eluting with 25% EtOAc/hexane, to give selenide 78 (3.0 mg, 2.5 μ mol, 50%) as a colorless oil: $[\alpha]^{23}_{D}$ (-) 9.9 (c 0.22, CHCl₃); IR (neat) 2956, 2932, 2874, 1701, 1515, 1457, 1250, 1074, 1007, 806, 730 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.22 (d, J = 6.9 Hz, 1H), 7.84 (d, J = 7.8 Hz, 1H), 7.48-7.54 (m, 1H), 7.48-7.55 (m, 3H), 6.87 (d, J = 8.4 Hz, 2H), 6.69 (t, J = 7.6 Hz, 1H), 5.84 (s, 1H), 5.05 (s, 1H), 4.96-5.00 (m, 1H), 4.87 (s, 1H), 4.56 (d, J = 10.5 Hz, 1H), 4.25-4.30 (m, 1H), 4.21 (d, J = 10.5 Hz, 1H), 3.84 (s, 3H), 3.70 (s, 2H), 3.26 (dd, J = 9.6, 17.4 Hz, 1H), 3.20 (d, J = 8.1, 14.4 Hz, 1H), 2.99–3.04 (m, 1H), 2.96 (dd, *J* = 8.1, 15.0 Hz, 1H), 2.57 (dd, *J* = 2.8, 17.4 Hz, 1H), 2.21-2.30 (m, 4H), 1.89-1.92 (m, 1H), 1.84 (s, 3H), 1.79 (s, 3H), 1.45-1.78 (m, 10H), 1.42 (s, 3H), 1.30-1.36 (m, 1H), 1.25 (d, J = 5.6 Hz, 3H), 0.91–1.01 (m, 30H), 0.56–0.67 (m, 18H); ¹³C NMR (75 MHz, CDCl₃) δ 210.9, 167.5, 159.1, 147.7, 144.5, 142.4, 140.3, 133.4, 131.3, 129.9, 129.7, 129.2, 126.1, 126.0, 125.5, 115.4, 113.5, 88.5, 77.2, 71.7, 68.9, 65.6, 62.1, 59.6, 55.3, 49.6, 45.9, 45.7, 44.8, 41.1, 39.6, 31.7, 31.6, 30.9, 28.3, 26.2, 21.0, 19.9, 15.2, 12.6, 12.1, 7.3, 7.0, 6.9, 5.3, 5.1; HRMS (ES+) calcd for $C_{64}H_{106}NO_{11}Si_3Se (M + 1)$ 1228.6239, found 1228.6216.



Allyl Epoxide 79. To a solution of selenide 78 (5.7 mg, 5.0 μ mol) in CH₂Cl₂ (0.25 mL) at rt were added sequentially triethylamine (19 mg, 26.0 µL, 0.18 mmol), TPAP (16 mg, 45 µmol), and NMO (35 mg, 0.3 mmol). After 30 min, the reaction mixture was purified directly by preparative TLC, eluting with 25% EtOAc/hexanes, to give vinyl epoxide 79 (2.5 mg, 2.4 μ mol, 48%) as a colorless oil: $[\alpha]^{23}_{D}$ -1.3 (c 0.15, CHCl₃); IR (neat) 3467, 3017, 2954, 2917, 2875, 2849, 2107, 1709, 1515, 1462, 1378, 1251, 1173, 1125, 1070, 1018, 835, 756, 667 cm^{-1} ; ¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, J = 8.4 Hz, 2H), 6.89 (d, J = 8.4 Hz, 2H), 6.73 (t, J = 7.6 Hz, 1H), 5.85-5.92 (m, 1H), 5.78 (s, 1H), 5.27 (dd, J = 8.0, 15.2 Hz, 1H), 5.01-5.04 (m, 2H), 4.85 (s, 1H), 4.57 (d, J = 10.4 Hz, 1H), 4.28–4.32 (m, 1H), 4.21 (d, J = 10.4 Hz, 1H), 3.85 (s, 3H), 3.70–3.74 (m, 2H), 3.25 (dd, J = 9.6, 17.2 Hz, 1H), 3.09 (dd, J = 2.0, 6.4 Hz, 1H), 2.90–2.94 (m, 1H), 2.54 (dd, J = 2.8, 17.2 Hz, 1H), 2.06–2.28 (m, 4H), 1.94 (d, J = 2.4, 10.8 Hz, 1H), 1.86 (s, 3H), 1.80–1.86 (m, 3H), 1.79 (s, 3H), 1.53–1.70 (m, 7H), 1.46 (s, 3H), 1.30–1.36 (m, 1H), 1.25 (d, J = 6.0 Hz, 3H), 0.90–1.01 (m, 27H), 0.86 $(d, J = 6.0 \text{ Hz}, 3\text{H}), 0.54 - 0.68 \text{ (m, 18H)}; {}^{13}\text{C NMR} (100 \text{ MHz}, \text{CDCl}_3)$ δ 211.0, 167.6, 159.1, 144.6, 142.4, 140.5, 134.3, 130.1, 129.8, 129.5, 128.7, 125.2, 115.3, 113.5, 88.4, 76.4, 71.7, 68.9, 65.7, 59.6, 58.5, 55.3, 49.4, 46.1, 45.6, 40.6, 40.2, 32.1, 30.9, 30.5, 29.7, 28.0, 27.8, 20.4, 19.8,

15.2, 13.0, 12.5, 7.3, 7.0, 6.9, 6.8, 5.2, 5.1; HRMS (ES+) calcd for $C_{58}H_{100}O_9Si_3Na~(M + Na)$ 1047.6573, found 1047.6664.



TES Ether SI-21. To a stirred solution of diol **65** (470 mg, 1.32 mmol) in DCM/TEA (6 mL, 1:1) was added freshly distilled TESOTf (1.05 g, 0.89 mL, 3.96 mmol) at -78 °C. After 30 min, the reaction was quenched with satd aq NaHCO₃ (1 mL) and extracted with ether (3 × 20 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2–5% EtOAc/hexanes, to give **SI-21** (732 mg, 1.25 mmol, 95%) as a colorless oil: $[\alpha]^{23}_{D} = -21.3^{\circ}$ (*c* 1.37, CHCl₃); IR (neat) 2954, 2929, 2876, 1460, 1254, 1093, 1007, 835, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.88 (s, 1H), 4.99 (s, 1H), 4.80 (s, 1H), 3.61–3.71 (m, 3H), 3.47 (dt, *J* = 15.6, 5.4 Hz, 1H), 2.15 (dd, *J* = 13.5, 5.4 Hz, 1H), 1.80–1.99 (m, 5H), 1.77 (d, *J* = 0.9 Hz, 3H), 1.54–1.66 (m, 1H), 0.88–1.00 (m, 27H), 0.84 (d, *J* = 6.3 Hz, 3H), 0.55–0.66 (m, 12H), 0.063 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 145.2, 142.0, 124.6, 114.1, 77.2, 61.7, 59.3, 46.2, 44.5, 40.0, 28.7, 26, 19.6, 18.4, 14.7, 7.2, 7.0, 6.9, 6.8, 4.4, -5.3; HRMS (EI⁺) calcd for C₃₂H₆₈O₃Si₃ (M⁺) 584.4476, found 584.4500.



Alcohol SI-22. TES ether SI-21 (300 mg, 0.51 mmol) was dissolved in a stirred solution of HOAc/THF/H₂O (8 mL, 8:8:1) at 0 °C. After 1.5 h, the reaction was then quenched with solid NaHCO₃ and extracted with ether $(3 \times 20 \text{ mL})$. The dried extract $(MgSO_4)$ was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give alcohol SI-22 (241 mg, 0.51 mmol, 99%) as a colorless oil: $[\alpha]_{D}^{23} = -15.5^{\circ}$ (c 0.64, CHCl₃); IR (neat) 3437, 2954, 2928, 2876, 1461, 1254, 1099, 1008, 835 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.02 (s, 1H), 5.04 (s, 1H), 4.85 (s, 1H), 3.63–3.73(m, 4H), 2.79 (t, J = 5.6 Hz, 1H), 2.17 (dd, J = 13.6, 5.6 Hz, 1H), 1.88-1.97 (m, 3H), 1.74–1.83 (m, 1H), 1.79 (d, J = 1.2 Hz, 3H), 1.59–1.68 (m, 1H), 1.47 (s, 3H), 1.28–1.40 (m, 1H), 0.99–1.03 (t, J = 7.6 Hz, 9H), 0.92 (s, 9H), 0.88 (d, J = 6.4 Hz, 3H), 0.69 (q, J = 7.2 Hz, 6H), 0.078 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 145.0, 141.4, 125.6, 114.6, 80.0, 61.6, 60.1, 46.1, 42.8, 39.9, 29.7, 28.6, 27.6, 26.0, 19.5, 18.4, 14.9, 7.2, 6.9, 6.8, -5.3; HRMS (EI⁺) calcd for $C_{26}H_{54}O_3Si_2$ (M⁺) 470.3611, found 470.3604.



Aldehyde 82. To a stirred solution of alcohol SI-22 (1.29 g, 2.73 mmol) in DCM (40 mL, 1:1) were added sequentially DMP (2.17 g, 5.12 mmol) and NaHCO3 (1.68 g, 20.0 mmol) at rt. After 30 min, the reaction was quenched with satd aq NaHCO₃ (10 mL) and extracted with ether $(3 \times 40 \text{ mL})$. The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give aldehyde 82 (1.17 g, 2.49 mmol, 91%) as a colorless oil: $[\alpha]^{23}_{D} = -12.5$ (*c* 0.56, CHCl₃); IR (neat) 2955, 2929, 2877, 1725, 1255, 1099, 1007, 836, 726 cm⁻¹ ¹: ¹H NMR (300 MHz, CDCl₃) δ 9.69 (t, J = 3.3 Hz, 1H), 6.03 (s, 1H), 5.03 (s, 1H), 4.83 (s, 1H), 3.61-3.68(m, 2H), 2.64 (dd, J = 15.0, 3.0 Hz, 1H), 2.45 (dd, J = 15.0, 3.0 Hz, 1H), 2.15 (dd, J = 13.5, 6.0 Hz, 1H), 1.85-1.92 (m, 1H), 1.83 (d, J = 1.2 Hz, 3H), 1.54-1.64 (m, 1H), 1.49 (s, 3H), 1.25–1.35 (m, 2H), 0.95–1.00 (t, J = 7.7 Hz, 9H), 0.91 $(s, 9H), 0.85 (d, J = 6.5 Hz, 3H), 0.65 (q, J = 7.8 Hz, 6H), 0.061 (s, 6H); {}^{13}C$ NMR (100 MHz, CDCl₃) δ 203.1, 144.7, 141.0, 126.0, 114.9, 76.9, 61.5, 54.2, 46.0, 39.9, 28.5, 27.8, 26.0, 19.5, 18.3, 14.7, 7.1, 6.7, -5.3; HRMS (EI⁺) calcd for C₂₆H₅₂O₃Si₂ (M⁺) 468.3455, found 468.3448.



Aldol Adduct SI-24. To a stirred solution of SI-23 (1.60 g, 0.15 mmol) in CH₂Cl₂ (11.2 mL) at -60 °C were added sequentially Et₃N (0.44 g, 0.60 mL, 4.33 mmol) and Bu₂BOTf (1.19 g, 1.08 mL, 4.33 mmol). After 3 h, the resulted solution was warmed to 0 °C for 30 min and then cooled back to -60 °C. A solution of aldehyde 37^8 (1.12 g, 4.86 mmol) in DCM (4.8 mL) was transferred to the reaction mixture via cannula. After 2 h, the reaction was allowed to warm to 0 °C. After another 20 min, the reaction was quenched by addition of pH 7 phosphate buffer (20 mL) followed by MeOH (15 mL) and 30% H₂O₂ (4 mL). After 1 h, the reaction mixture was extracted with EtOAc $(4 \times 35 \text{ mL})$. The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15-20% EtOAc/hexanes, to give SI-24 (1.58 g, 2.57 mmol, 62%) as a colorless oil: $[\alpha]^{23}_{D} = -9.2$ (c 0.77, CHCl₃); IR (neat) 3493, 2957, 2876, 1781, 1709, 1593, 1517, 1455, 1390, 1265, 1240, 1159, 1052, 1028, 746 cm⁻¹; $^{1}\mathrm{H}$ NMR (400 MHz) δ 7.23–7.38 (m, 5H), 6.83–7.03 (m, 3H), 5.36 (d, J = 2.0 Hz, 1H), 4.63-4.71 (m, 2H), 4.51 (d, J = 10.8 Hz, 1H),4.21-4.29 (m, 2H), 3.93 (s, 3H), 3.84-3.90 (m, 1H), 3.88 (s, 3H), 3.62–3.66 (m, 1H), 3.32 (dd, J = 13.6, 3.6 Hz, 1H), 2.77 (dd, J = 13.6, 10.0 Hz, 1H), 2.32 (d, J = 10.0 Hz, 1H), 1.78–1.80 (m, 1H), 1.58–1.64 (m, 1H), 1.15 (d, J = 6.0 Hz, 3H), 1.03 (d, J = 6.8 Hz, 3H), 0.98 (t, J = 8.0 Hz, 9H), 0.62 (q, J = 8.0 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 177.2, 153.2, 149.1, 149.0, 135.2, 129.5, 129.4, 129.0, 127.4, 121.4, 112.1, 110.9, 78.1, 76.1, 66.9, 55.9, 55.7, 42.8, 37.7, 34.1, 23.2, 15.7, 6.9, 4.9; HRMS (ES⁺) calcd for C₃₃H₄₉NO₈SiNa (M + Na) 638.3125, found 638.3155.



TES Ether SI-25. To a stirred solution of adol adduct SI-24 (500 mg, 0.81 mmol) in DCM (3.32 mL) at 0 °C were added sequentially 2,6-lutidine (184 mg, 0.20 mL, 1.72 mmol) and TESOTf (287 mg, 0.25 mL, 1.09 mmol). After 1 h, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with Et₂O (3 \times 30 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15% EtOAc/hexanes, to give SI-25 (570 mg, 0.78 mmol, 96%) as a colorless oil: $[\alpha]_{D}^{23} = -40.7$ (*c* 0.42, CHCl₃); IR (neat) 2955, 2911, 2876, 1784, 1702, 1517, 1456, 1239, 1084, 740 cm⁻¹; ¹H NMR (300 MHz) δ 7.17-7.32 (m,H), 6.80-7.00 (m, 3H), 5.28 (d, J = 6.0 Hz, 1H), 4.71 (d, J = 11.7 Hz, 1H), 4.54 (d, J = 11.7 Hz, 2H),4.47-4.49 (m, 1H), 4.00-4.12 (m, 3H), 3.90 (s, 3H), 3.84 (s, 3H), 3.78-3.82 (m, 1H), 3.14 (dd, J = 3.0, 13.2 Hz, 1H), 2.40 (dd, J = 10.5, 13.5 Hz, 1H), 1.42-1.56 (m, 3H), 1.11 (d, J = 5.7 Hz, 3H), 0.89-0.97 (m, 21H), 0.55–0.63 (m, 12H); $^{13}\mathrm{C}$ NMR (100 MHz, CDCl₃) δ 171.9, 152.9, 148.7, 148.8, 135.3, 130.3, 129.3, 129.0, 127.3, 121.1, 111.9, 110.7, 73.4, 67.2, 66.4, 56.0, 55.9, 55.8, 44.4, 37.4, 33.5, 23.7, 14.8, 7.1, 6.9, 5.4, 5.3, 5.2, 5.0; HRMS (ES^+) calcd for $C_{39}H_{63}NO_8Si_2Na$ (M + Na) 752.3990, found 752.3992.



Thiol Ester SI-26. To a stirred solution of EtSH (90 mg, 0.107 mL, 1.45 mmol) in THF (12.6 mL) at 0 °C was added n-BuLi (0.51 mL, 1.27 mmol, 2.5 M in hexanes). After 1 h, a solution of SI-25 (610 mg, 0.83 mmol) in THF (2.7 mL) was added dropwise via cannula. After another 1 h, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with Et₂O (3 \times 30 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5-8% EtOAc/hexanes, to give SI-26 (470 mg, 0.76 mmol, 92%) as a colorless oil: $[\alpha]^{23}_{D} = +42.0$ (c 0.39, CHCl₃); IR (neat) 2955, 2911, 2876, 1683, 1517, 1458, 1419, 1378, 1266, 1240, 1161, 1079, 1032, 811, 740 cm⁻¹; ¹H NMR (400 MHz) δ 7.04 (dd, J = 1.6 Hz, 1H), 6.94 (dd, J = 1.6, 8.0 Hz, 1H), 6.86 (d, J = 8.4 Hz, 1H), 4.63 (d, J = 10.8 Hz, 1H), 4.43 (d, J = 10.8 Hz, 1H), 3.94 (s, 3H), 3.92–3.93 (m, 4H), 3.81-3.86 (m, 2H), 2.91 (q, J = 7.6 Hz, 2H), 1.45-1.57 (m, 3H), 1.30 (t, J = 7.2 Hz, 3H), 1.14 (d, J = 6.0 Hz, 3H), 0.90-0.98 (m, 21H), 0.53-0.62 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 202.2, 148.8, 148.6, 129.9, 120.6, 110.6, 110.7, 88.2, 72.9, 66.9, 55.9, 55.8, 44.9, 32.7, 23.4, 22.5, 14.6, 13.6, 7.0, 6.9, 5.3, 5.2, 5.0; HRMS (ES⁺) calcd for $C_{31}H_{58}O_6Si_2SNa (M + Na) 637.3390$, found 637.3407.



Methyl Ketone 81. To a stirred slurry of CuI (859 mg, 4.51 mmol) in Et₂O (8.3 mL) at 0 °C was added MeLi (5.6 mL, 9.6 mmol, 1.6 M in Et₂O). After 15 min, the colorless solution was cooled to -50 °C, and a solution of SI-26 (450 mg, 0.75 mmol) in Et₂O (4.2 mL) was transferred into the reaction mixture dropwise via cannula. After 2 h, the reaction was quenched with satd aq NH₄Cl (10 mL) at -50 °C, warmed to rt, and extracted with Et₂O (3×30 mL). The dried (MgSO₄) extract was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2-4% EtOAc/ hexanes, to give 81 (296 mg, 0.52 mmol, 71%) as a colorless oil: $[\alpha]^{23}_{D}$ = +22.6 (c 0.23, CHCl₂); IR (neat) 2955, 2911, 2876, 1716, 1517, 1457, 1267, 1240, 1082, 1031, 1007, 741 cm⁻¹; ¹H NMR (400 MHz) δ 6.82–6.90 (m, 3H), 4.50 (dd, J = 11.4, 15.0 Hz, 2H), 3.90 (s, 6H), 3.80-3.85 (m, 2H), 3.76 (d, J = 6.3 Hz, 1H), 2.13 (s, 3H), 1.50–1.52 (m, 3H), 1.13 (d, J = 6.0 Hz, 3H), 0.88-1.00 (m, 21H), 0.57-0.64 (m, 12H); 13C NMR (100 MHz, CDCl₃) & 210.6, 148.8, 129.9, 120.7, 111.4, 110.8, 88.5, 72.9, 67.0, 55.9, 55.8, 44.5, 33.1, 27.0, 23.5, 14.0, 7.0, 6.9, 5.3, 5.0; HRMS (ES⁺) calcd for C₃₀H₅₆O₆Si₂Na (M + Na) 591.3513, found 591.3527.



Aldol Adduct 83. To a stirred solution of methyl ketone 81 (30 mg, 0.0527 mmol) in Et₂O (0.5 mL) at -78 °C was added LDA⁵⁴ (64 μ L, 0.064 mmol, 1 M in THF). After 15 min, a precooled (-78 °C) solution of aldehyde 82 (50 mg, 0.105 mmol) in THF (0.5 mL) was added via cannula in one portion. After another 0.5 h, the reaction was quenched with satd aq NH₄Cl (2 mL) at -78 °C, warmed to rt, and extracted with ether (3 × 10 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 6–8%

EtOAc/hexanes, to sequentially give aldol adduct **SI-27** (10 mg, 0.0096 mmol, 18%) and **83** (39 mg, 0.0375 mmol, 71%) and as colorless oils. **83**: $[\alpha]^{23}_{D} = +9.1$ (*c* 0.58, CHCl₃); IR (neat) 3503, 2955, 2934, 2876, 1715, 1517, 1463, 1265, 1240, 1095, 1007, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.83–6.97 (m, 3H), 5.90 (s, 1H), 5.05 (s, 1H), 4.85 (s, 1H), 4.53 (d, *J* = 11.2 Hz, 1H), 4.44–4.50 (m, 1H), 4.39(d, *J* = 10.8 Hz, 1H), 3.91 (s, 6H), 3.76–3.83 (m, 3H), 3.66–3.70 (m, 2H), 3.05 (dd, *J* = 17.6, 6.8 Hz, 1H), 2.38 (dd, *J* = 17.6, 5.6 Hz, 1H), 2.19 (dd, *J* = 13.5, 4.8 Hz, 1H), 1.80–1.91 (m, 1H), 1.84 (s, 3H), 1.48–1.68 (m, 4H), 1.57 (s, 3H), 1.20–1.42 (m, 4H), 1.10 (d, *J* = 6.0 Hz, 3H), 0.86–1.00 (m, 42H), 0.57–0.66 (m, 18H), 0.08 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 211.0, 148.8, 148.6, 144.6, 142.9, 130.1, 125.2, 120.7, 114.9, 111.5, 110.7, 88.3, 79.5, 72.6, 66.9, 64.8, 61.4, 55.9, 55.8, 48.0, 46.7, 46.0, 44.8, 39.9, 33.1, 28.4, 23.3, 19.5, 14.7, 13.5, 7.1, 7.0, 6.9, 6.5, 5.3, 5.0, -5.2; HRMS (ES⁺) calcd for C₅₆H₁₀₇O₉Si₄ (M + H) 1035.6992, found 1035.7047.



Enone 93. To a stirred slurry of NaH (36 mg, 0.90 mmol, 60% w/w in mineral oil) in DME (2 mL) was added phosphonate 92 (138 mg, 0.83 mmol) at rt. After 1 h, a solution of aldehyde 37 (160 mg, 0.69 mmol) in DME (2 mL) was added via cannula. After another 6 h, the reaction was quenched with satd aq NH₄Cl (2 mL) and extracted with Et_2O (4 × 10 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give enone 93 (142 mg, 0.52 mmol, 76%) as a colorless oil: $[\alpha]^{23}_{D} = -46.0^{\circ}$ (c 1.47, CHCl₃); IR (neat) 2958, 2877, 1700, 1678, 1627, 1458, 1360, 1252, 1139, 1055, 984, 745 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.76 (dd, J = 15.9, 7.8 Hz, 1H), 6.09 (d, J = 15.9 Hz, 1H), 3.86– 3.79 (m, 1H), 2.58–2.53 (m, 1H), 2.26 (s, 3H), 1.41–1.55 (m, 2H), 1.18 (d, J = 6.0 Hz, 3H), 1.09 (d, J = 6.9 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9H), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9Hz), 0.64 (q, J = 6.0 Hz, 3H), 0.97 (t, J = 7.8 Hz, 9Hz), 0.97 (t, J = 7.8 Hz, 9Hz), 0.64 (q, J = 6.0 Hz, 3Hz), 0.97 (t, J = 7.8 Hz, 9Hz), 0.64 (q, J = 6.0 Hz, 3Hz), 0.97 (t, J = 7.8 Hz, 9Hz), 0.97 (t, J = 7.8 Hz), 0.91 (t, J = 7.8 HzJ = 7.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 198.8, 153.5, 129.6, 66.4, 46.3, 33.4, 27.0, 24.3, 20.3, 6.9, 5.2, 5.0; HRMS (EI⁺) calcd for C₁₅H₃₀O₂Si (M⁺) 270.2015, found 270.2008.



Diol 94. To a stirred solution of enone 93 (142 mg, 0.52 mmol) in t-BuOH/H₂O (5 mL, 1:1) at 0 °C were added sequentially AD-mix- α (0.735 g), NaHCO₃ (132 mg, 1.57 mmol), MeSO₂NH₂ (50.6 mg, 0.53 mmol), and K₂OsO₂(OH)₄ (1.9 mg, 0.005 mmol). After 8 h, the reaction was quenched with satd aq Na_2SO_3 (8 mL) and extracted with EtOAc (4 × 10 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15-40% EtOAc/hexanes, to give diol 94 (122 mg, 0.40 mmol, 77%) as a colorless oil: $[\alpha]_{D}^{23} = -29.2$ (*c* 1.5, CHCl₃); IR (neat) 3456, 2957, 2877, 1717, 1380, 1238, 1132, 1048, 1011, 744 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 4.28 (d, J = 3.6 Hz, 1H), 4.04–3.99 (m, 1H), 3.72–3.78 (m, 1H) and OH), 2.41 (d, J = 10.4 Hz, 1H), 2.30 (s, 3H), 1.89-1.96 (m, 1H), 1.71–1.77 (m, 1H), 1.40–1.47 (m, 1H), 1.22 (d, J = 6.0 Hz, 3H), 1.10 (d, J = 6.8 Hz, 3H), 1.00 (t, J = 7.8 Hz, 9H), 0.68 (q, J = 7.9 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 208.9, 77.7, 75.2, 66.9, 42.8, 34.0, 25.3, 23.1, 16.4, 6.9, 4.9; HRMS (ES⁺) calcd for $C_{15}H_{32}O_4SiNa$ (M + Na) 327.1968, found 327.1950.



TES Ether 95. To a stirred solution of diol **94** (800 mg, 2.63 mmol) in CH₂Cl₂ (10 mL) at -78 °C were added sequentially 2,6-lutidine (1.41 g, 1.53 mL, 13.1 mmol) and TESOTf (1.74 g, 1.49 mL, 6.58 mmol). After 30 min, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with Et₂O (4 \times 25 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give TES ether 95 (1.29 g, 2.42 mmol, 92%) as a colorless oil: $[\alpha]_{D}^{23} = -0.83^{\circ}$ (c 1.2, CHCl₃); IR (neat) 2957, 2878, 1716, 1458, 1238, 1005 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 4.07 \text{ (d, } J = 6.0 \text{ Hz}, 1 \text{H}), 3.78 - 3.85 \text{ (m, 1H)}, 3.70$ (dd, J = 5.9, 2.6 Hz, 1H), 2.20 (s, 3H), 1.60-1.64 (m, 1H), 1.45-1.51(m, 2H), 1.13 (d, J = 6.0 Hz, 3H), 0.94–1.00 (m, 27H), 0.83 (d, J = 6.6 Hz, 3H), 0.55–0.70 (m, 18H); 13 C NMR (75 MHz, CDCl₃) δ 210.0, 81.5, 78.6, 67.1, 45.4, 32.2, 27.3, 23.1, 14.0, 7.0, 6.84, 6.79, 5.2, 4.9, 4.8; HRMS (ES⁺) calcd for $C_{27}H_{60}O_4Si_3Na$ (M + Na) 555.3697, found 555.3683.



Alcohol 96. TES ether 95 (5.60 g, 10.5 mmol) was dissolved in a stirred solution of HOAc/THF/H₂O (107 mL, 8:8:1) at 0 °C. After 12 h, the reaction was quenched with solid NaHCO₃, filtered over Celite, and extracted with ether (4 × 100 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give alcohol 96 (3.90 g, 9.31 mmol, 89%) as a colorless oil: $[\alpha]^{23}_{D} = -30.5$ (*c* 1.45, CHCl₃); IR (neat) 3446, 2958, 2878, 1716, 1458, 1239, 1005, 739 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.18 (d, *J* = 5.6 Hz, 1H), 3.86–3.88 (m, 2H), 2.24 (s, 3H), 1.80–1.90 (m, 1H), 1.41–1.55 (m, 2H), 1.20 (d, *J* = 6.0 Hz, 3H), 0.97–1.05 (m, 18H), 0.87 (d, *J* = 6.8 Hz, 3H), 0.61–0.73 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 209.3, 81.4, 76.4, 65.6, 43.7, 31.6, 27.8, 23.6, 15.4, 7.0, 6.8, 5.2, 4.8, 4.7; HRMS (ES⁺) calcd for C₂₁H₄₆O₄Si₂Na (M + Na) 441.2832, found 441.2836.



PNB Ester 97. To a stirred solution of alcohol 96 (600 mg, 1.43 mmol) in THF (15 mL) at 0 °C were added sequentially PPh₃ (1.50 g, 5.72 mmol), 4-nitrobenzoic acid (0.96 g, 5.74 mmol), and DEAD (0.99 g, 0.90 mL, 5.70 mmol). After 1 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 1-3% EtOAc/hexanes, to give ester 97 (670 mg, 1.18 mmol, 82%) as a colorless oil: $[\alpha]^{23}_{D} = +13.6$ (*c* 1.08, CHCl₃); IR (neat) 2956, 2878, 1723, 1530, 1319, 1275, 1014, 721 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 8.31 (d, J = 9.0 \text{ Hz}, 2\text{H}), 8.23 (d, J = 9.0 \text{ Hz}, 2\text{H}),$ 5.26 (m, 1H), 4.17 (d, J = 4.8 Hz, 1H), 3.69 (t, J = 4.8 Hz, 1H), 2.18 (s, 3H), 2.02–2.12 (m, 1H), 1.76 (m, 1H), 1.45–1.49 (m, 1H), 1.38 (d, J = 6.1 Hz, 3H), 0.93-1.02 (m, 18H), 0.90 (d, J = 6.6 Hz, 3H), 0.56-0.70 (m, 12H); 13 C NMR (100 MHz, CDCl₃) δ 209.3, 164.3, 150.4, 136.1, 130.7, 123.4, 81.5, 78.2, 70.8, 40.2, 32.4, 27.9, 20.9, 14.8, 7.0, 6.8, 5.1, 4.8; HRMS (ES⁺) calcd for $C_{28}H_{49}NO_7Si_2Na$ (M + Na) 590.2945, found 590.2926.



Alcohol SI-28. To a stirred solution of ester 97 (700 mg, 1.23 mmol) in MeOH (20 mL) at 0 $^{\circ}$ C was added Ba(OH)₂·8H₂O (390 mg,

1.24 mmol). After 4 h, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with EtOAc (4 × 20 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 15% EtOAc/hexanes, to give alcohol **SI-28** (369 mg, 0.88 mmol, 72%) as a colorless oil: $[\alpha]^{23}_{D} = -7.6$ (*c* 1.2, CHCl₃); IR (neat) 3434, 2957, 2878, 1716, 1459, 1415, 1239, 1005, 739 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.17 (d, *J* = 5.4 Hz, 1H), 3.81 (m, 1H), 3.69 (dd, *J* = 5.4, 3.9 Hz, 1H), 2.22 (s, 3H), 1.83 (m, 1H), 1.61–1.69 (m, 1H), 1.24–1.30 (m, 1H), 1.20 (d, *J* = 6.0 Hz, 3H), 0.94–1.04 (m, 18H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.58–0.71 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 210.1, 81.5, 78.1, 66.2, 43.8, 32.9, 27.6, 24.4, 15.2, 7.0, 6.8, 5.2, 4.8; HRMS (ES⁺) calcd for C₂₁H₄₇O₄Si₂ (M + H) 419.3013, found 419.2993.



TMS Ether 98. To a stirred solution of alcohol SI-28 (1.90 g, 4.54 mmol) in $CH_2Cl_2~(25~mL)$ at $-78~^\circ C$ were added sequentially 2,6-lutidine (1.45 g, 1.58 mL, 13.5 mmol) and TMSOTf (1.51 g, 1.23 mL, 6.81 mmol). After 30 min, the reaction was quenched with satd aq NH₄Cl (10 mL) and extracted with Et₂O (4 \times 20 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10% EtOAc/hexanes, to give TMS ether 98 (2.12 g, 4.32 mmol, 95%) as a colorless oil: $[\alpha]^2$ '_D = +14.4 (c 2.2, CHCl₃); IR (neat) 2957, 2878, 1716, 1459, 1415, 1124, 1006, 841, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.03 (d, J = 6.3 Hz, 1H), 3.79–3.85 (m, 1H), 3.71 (dd, *J* = 6.3, 2.4 Hz, 1H), 2.19 (s, 3H), 1.73–1.79 (m, 1H), 1.45–1.54 (m, 1H), 1.23–1.32 (m, 1H), 1.16 (d, J= 6.0 Hz, 3H), 0.95–1.03 (m, 18H), 0.83 (d, J = 6.6 Hz, 3H), 0.58–0.70 (m, 12H); 13 C NMR (75 MHz, CDCl₃) δ 210.2, 81.8, 78.6, 65.9, 45.3, 31.3, 26.8, 24.7, 13.2, 7.0, 6.8, 5.3, 4.8, 0.3; HRMS (ES⁺) calcd for C₂₄H₅₄O₄Si₃Na (M + Na) 513.3224, found 513.3204.



Aldol Adducts 100 and 103. To a stirred solution of methyl ketone 98 (312 mg, 0.64 mmol) in THF (5 mL) at -78 °C was added LDA² (0.765 mL, 1 M in THF). After 15 min, TMEDA (133 mg, 0.172 mL, 1.14 mmol) was added. After 5 min, the reaction was warmed up to -40 °C, followed by the addition of a precooled (-40 °C) solution of aldehyde 82 (200 mg, 0.43 mmol) in THF (5 mL) via cannula in one portion. After another 0.5 h, the reaction was quenched with satd aq NH₄Cl (10 mL) -78 °C, warmed to rt, and extracted with ether (4 × 20 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 1–1.5% EtOAc/ hexanes, to give aldol adduct 100 (142 mg, 0.15 mmol, 35%) and 103 (114 mg, 0.12 mmol, 28%) as colorless oil. 100: [α]²³_D = -12.0 (*c* 1.3, CHCl₃); IR (neat) 3511, 2955, 2929, 2877, 1715, 1460, 1413, 1250, 1092, 1006, 838, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.89 (s, 1H), 5.02 (s, 1H), 4.83 (s, 1H), 4.33 (m, 1H), 4.10 (d, *J* = 5.7 Hz, 1H), 3.82

(m, 1H), 3.74 (s, 1H), 3.64–3.72 (m, 3H), 2.96 (dd, J = 17.7, 6.2 Hz, 1H), 2.60 (dd, J = 18.1, 6.4 Hz, 1H), 2.18 (dd, J = 12.8, 4.0 Hz, 1H), 1.81–1.90 (m, 2H), 1.84 (s, 3H), 1.47–1.67 (m, 4H), 1.56 (s, 3H), 1.22–1.38 (m, 3H), 1.15 (d, J = 6.0 Hz, 3H), 0.92–1.03 (m, 36H), 0.87 (d, J = 6.4 Hz, 3H), 0.80 (d, J = 6.7 Hz, 3H), 0.58–0.70 (m, 18H), 0.10 (s, 9H), 0.06 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 210.7, 144.7, 125.1, 114.7, 81.1, 79.4, 78.4, 65.9, 65.1, 61.5, 48.2, 47.2, 46.0, 45.0, 39.9, 31.1, 28.5, 26.2, 26.0, 24.7, 19.5, 18.3, 14.7, 13.8, 7.2, 7.1, 6.9, 6.6, 5.3, 4.9, 0.4, -5.2; HRMS (ES⁺) calcd for C₅₀H₁₀₆O₇Si₅Na (M + Na) 981.6683, found 981.6646.

MTPA Esters. To a solution of **100** (5 mg, 0.005 mmol) in CH₂Cl₂ (0.5 mL) were added sequentially DMAP (6.4 mg, 0.052 mmol) and (*R*)- or (*S*)-(+)-α-methoxy-α-trifluoromethylphenylacetyl chloride (6.6 mg, 4.9 μL, 0.026 mmol). After 10 min, the solution was evaporated, and the residue was loaded directly onto silica gel and purified by chromatography, eluting with 2 - 10% EtOAc/hexanes, to give product (*S*)- or (*R*)- **MTPA esters** (52–61%) as colorless oils. ¹H NMR difference in ppm [(*S*)-Mosher ester – (*R*)-Mosher ester, CDCl₃, CDCl₃, 300 MHz NMR] H₁₉ = 2.847 – 2.834 = +0.013, H₂₁: 3.996 – 3.961 = +0.035, H₂₂: 3.686 – 3.678 = +0.008, H₂₅: 3.848 – 3.818 = +0.030, H₃₁: 0.881 – 0.876 = +0.005, H₂₉: 1.888 – 1.895 = -0.007, H₁₄: 5.723 – 5.824 = -0.101, H₂₈: 4.905 – 4.925 = -0.020, H₁₂: 2.319 – 2.334 = -0.015, H₂₇: 1.130 – 1.137 = -0.007.



Aldol Adducts 101 and 104. *Method A* ($-100 \degree C$ *Conditions*). To a stirred solution of methyl ketone 98 (574 mg, 1.17 mmol) in THF (6 mL) at $-78 \degree C$ was added LDA² (1.38 mL, 1 M in THF). After 15 min, TMEDA (400 mg, 0.310 mL, 3.44 mmol) was added. After 5 min, the reaction was cooled to $-100 \degree C$, followed by the addition of a precooled ($-100 \degree C$) solution of aldehyde 57 (310 mg, 0.78 mmol) in THF (6 mL) via cannula in one portion. After another 0.5 h, the reaction was quenched with 1 M AcOH in THF (1.5 mL) at $-100 \degree C$. The reaction mixture was then warmed up to rt, diluted with satd aq NH₄Cl (10 mL) and extracted with ether (4 × 25 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 50% CH₂Cl₂/hexanes -2% EtOAc/hexanes, to give aldol adduct 104 (405 mg, 0.45 mmol, 58%) and 101 (50 mg, 0.056 mmol, 7%) as colorless oils.

Method B (-40 °C Conditions). To a stirred solution of methyl ketone 98 (37.2 mg, 0.0758 mmol) in THF (0.4 mL) at -78 °C was added LDA² (90 μ L, 0.09 mmol, 1 M in THF). After 15 min, TMEDA (15.5 mg, 20 μ L, 0.133 mmol) was added. After 5 min, the reaction was warmed to -40 °C, followed by the addition of a precooled (-40 °C) solution of aldehyde 57 (20 mg, 0.0504 mmol) in THF (0.4 mL) via cannula in one portion. After another 0.5 h, the reaction was quenched with satd aq NH₄Cl (2 mL) and extracted with ether (4 × 5 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 50% CH₂Cl₂/hexanes -2% EtOAc/hexanes, to give aldol adduct 101 (16.1 mg, 0.0181 mmol, 36%) and 104 (13.4 mg, 0.0151 mmol, 30%) as colorless oils. 101: $[\alpha]^{23}_{D} = -9.42$ (*c* 1.21, CHCl₃); IR (neat) 3516, 2956, 2913, 2877, 1743, 1719, 1458, 1414, 1370, 1249, 1116, 1088, 1008, 841, 742 cm⁻¹;

¹H NMR (300 MHz, CDCl₃) δ 5.89 (s, 1H), 5.02 (s, 1H), 4.83 (s, 1H), 4.38-4.27 (m, 1H), 4.06-4.10 (m, 3H), 3.86-3.75 (m, 1H), 3.72-3.68 (m, 1H), 3.66 (s, 1H, OH), 2.96 (dd, J = 17.9, 6.2 Hz, 1H), 2.59 (dd, J = 18.0, 6.1 Hz, 1H), 2.15 (dd, J = 13.2, 5.7 Hz, 1H), 2.04 (s, 3H), 1.94-1.76 (m, 4H), 1.81 (s, 3H), 1.70-1.63 (m, 2H), 1.54 (s, 3H), 1.48-1.38 (m, 2H), 1.27-1.22 (m, 1H), 1.13 (d, J = 5.9 Hz, 3H), 1.02-0.93 (m, 2H), 1.02-0.93 (m,27H), 0.89 (d, J = 6.3 Hz, 3H), 0.79 (d, J = 6.6 Hz, 3H), 0.59-0.69 (m, 18H), 0.09 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 210.8, 171.1, 144.3, 143.3, 124.8, 115.0, 81.0, 79.3, 78.4, 65.9, 65.0, 62.9, 48.2, 47.2, 45.8, 45.0, 35.3, 31.0, 28.7, 26.3, 24.6, 21.0, 19.3, 14.7, 13.8, 7.1, 7.0, 6.8, 6.6, 5.2, 4.9, 0.3; HRMS (ES⁺) calcd for $C_{46}H_{94}O_8Si_4Na$ (M + Na) 909.5924, found 909.5895. **104**: $[\alpha]_{D}^{23} = +1.76$ (c 1.25, CHCl₃); IR (neat) 3511, 2956, 2913, 2877, 1743, 1718, 1458, 1369, 1249, 1119, 1088, 1011, 841, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.06 (s, 1H), 5.03 (s, 1H), 4.88 (s, 1H), 4.17-4.06 (m, 4H), 3.88 (s, 1H, OH), 3.88–3.81 (m, 1H), 3.73–3.69 (m, 1H), 2.66-2.78 (m, 2H), 2.17 (dd, J = 13.5, 6.0 Hz, 1H), 2.05 (s, 3H), 1.97-1.93 (m, 1H), 1.85-1.78 (m, 1H), 1.82(s, 3H), 1.74-1.70 (m, 1H), 1.62–1.14 (m, 4H), 1.44 (s, 3H), 1.28–1.20 (m, 2H), 1.14 (d, J = 5.8 Hz, 3H), 1.02–0.94 (m, 27H), 0.91 (d, J = 6.2 Hz, 3H), 0.78 (d, J = 6.6 Hz, 3H), 0.72–0.57 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 210.5, 171.1, 144.5, 140.8, 125.4, 114.8, 81.3, 80.7, 78.4, 66.0, 65.7, 63.0, 47.1, 45.9, 45.0, 35.2, 31.0, 29.7, 28.7, 28.2, 24.6, 21.0, 19.4, 14.9, 13.6, 7.1, 7.0, 6.9, 6.8, 6.6, 5.2, 4.8, 0.3; HRMS (ES⁺) calcd for C46H94O8Si4Na (M + Na) 909.5924, found 909.5948.



TES Ether SI-29. To a stirred solution of aldol adduct 101 (295 mg, 0.332 mmol) in CH₂Cl₂ (10 mL) at rt were added sequentially DMAP (608 mg, 4.98 mmol) and TESCI (375 mg, 0.418 mL, 2.49 mmol). After 3 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2% EtOAc/hexanes, to give TES ether SI-29 (290 mg, 0.289 mmol, 87%) as a colorless oil: $[\alpha]^{23}_{D} = -31.4$ (c 0.85, CHCl₃); IR (neat) 2955, 2913, 2877, 1745, 1718, 1459, 1368, 1249, 1127, 1086, 1007, 841, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.78 (s, 1H), 5.03 (s, 1H), 4.92 (s, 1H), 4.20-4.08 (m, 3H), 4.04 (d, J = 5.7 Hz, 1H), 3.84-3.81 (m, 1H), 3.72-3.68 (m, 1H), 2.86 (dd, I = 18.0, 5.8 Hz, 1H), 2.72 (dd, I = 17.4, 7.2 Hz, 1H), 2.17 (dd, J = 13.8, 6.0 Hz, 1H), 2.05 (s, 3H), 1.93–1.64 (m, 5H), 1.84 (s, 3H), 1.54–1.39 (m, 3H), 1.45 (s, 3H), 1.27–1.21 (m 1H), 1.14 (d, J = 6.0 Hz, 3H), 1.03–0.87 (m, 39H), 0.78 (d, J = 6.7 Hz, 3H), 0.71–0.55 (m, 24H), 0.11 (s, 9H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl₃) δ 208.6, 171.1, 144.3, 143.0, 125.0, 115.1, 81.0, 78.3, 77.7, 66.1, 65.7, 63.0, 49.8, 48.5, 46.0, 44.9, 35.4, 31.0, 28.6, 27.9, 24.6, 21.0, 19.2, 14.5, 14.2, 7.2, 7.0, 6.9, 6.8, 5.2, 4.9, 0.4; HRMS (ES⁺) calcd for C₅₂H₁₀₈O₈Si₅Na (M + Na) 1023.6788, found 1023.6785.



Alcohol 105. To a stirred solution of TMS ether **SI-29** (290 mg, 0.289 mmol) in THF/H₂O (6.52 mL, 8:1) at -20 °C was added HOAc (4 × 1.45 mL) in four portions every 60 min. After 5 h, the reaction was quenched with solid NaHCO₃, filtered over Celite, and extracted with ether (4 × 15 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5–10% EtOAc/hexanes, to give alcohol **105** (220 mg, 0.237 mmol, 82%) as a colorless oil: $[\alpha]^{23}{}_{\rm D} = -31.6$ (*c* 1.01, CHCl₃); IR (neat) 3510, 2956, 2912, 2877, 1744, 1720, 1458, 1414, 1368, 1239, 1062, 1006, 741

cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.78 (s, 1H), 5.02 (s, 1H), 4.90 (s, 1H), 4.22–4.08 (m, 4H), 3.81–3.77 (m, 1H), 3.69–3.66(m, 1H), 2.94 (dd, *J* = 18.3, 6.1 Hz, 1H), 2.78 (dd, *J* = 18.3, 5.7 Hz, 1H), 2.15 (dd, *J* = 13.1, 5.7 Hz, 1H), 2.04 (s, 3H), 1.93–1.60 (m, 6H), 1.83 (s, 3H), 1.45–1.34 (m, 2H), 1.44 (s, 3H), 1.30–1.20 (m, 1H), 1.17 (d, *J* = 6.0 Hz, 3H), 1.04–0.87 (m, 39H), 0.82 (d, *J* = 6.0 Hz, 3H), 0.72–0.56 (m, 24H); ¹³C NMR (75 MHz, CDCl₃) δ 208.8, 171.2, 144.4, 142.6, 125.2, 115.1, 81.4, 78.1, 77.7, 66.2, 65.4, 63.1, 49.7, 49.0, 45.9, 44.2, 35.3, 32.5, 28.6, 28.0, 24.3, 21.0, 19.2, 15.5, 14.6, 7.2, 7.0, 6.8, 5.3, 5.2, 4.8; HRMS (ES⁺) calcd for C₄₉H₁₀₀O₈Si₄Na (M + Na) 951.6393, found 951.6398.



Phosphonate SI-30. To a stirred solution of acid 11 (450 mg, 1.47 mmol) in PhMe (3.2 mL) at rt were added sequentially Et₃N (149 mg, 0.204 mL, 1.47 mmol) and 2,4,6-trichlorobenzoyl chloride (346 mg, 0.222 mL, 1.47 mmol). After 12 h, the resulted solution was concentrated in vacuo. DMAP (180 mg, 1.47 mmol) was added, followed by the addition of a solution of alcohol 105 (240 m mg, 0.258 mmol) in PhMe (5.7 mL). After another 19 h, the reaction was quenched with satd aq NH₄Cl (8 mL) and extracted with EtOAc (4×50 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 20–60% EtOAc/hexanes, to give phosponate SI-30 (235 mg, 0.193 mmol, 78%) as a colorless oil: $[\alpha]^{23}_{D} = -23.6$ (*c* 0.83, CHCl₃); IR (neat) 2955, 2912, 2877, 1734, 1716, 1458, 1369, 1241, 1056, 1019, 970, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.69 (t, I = 6.3 Hz, 1H), 5.79 (s, 1H), 5.10-5.00 (m, 1H), 5.03 (s, 1H), 4.92 (s, 1H), 4.22-4.08 (m, 8H), 3.72 (dd, J = 5.1, 3.6 Hz, 1H), 3.13 (d, J = 27.8 Hz, 2H), 2.90 (dd, J = 18.0, 5.9 Hz, 1H), 2.77 (dd, J = 17.8, 6.0 Hz, 1H), 2.68 (t, J = 7.2 Hz, 2H), 2.18-2.07 (m, 3H), 2.06 (s, 3H), 1.94-1.68 (m, 9H), 1.842 (s, 3H), 1.82 (s, 3H), 1.46 (s, 3H), 1.43–1.39 (m, 2H), 1.36 (t, J = 7.0 Hz, 6H), 1.25 (d, J = 6.0 Hz, 3H), 1.03–0.92 (m, 36H), 0.90 (d, J = 6.4 Hz, 3H), 0.80 (d, J = 6.7 Hz, 3H), 0.70–0.55 (m, 24H); ¹³C NMR (75 MHz, CDCl₃) δ 208.4, 201.3, 171.1, 167.6, 144.3, 142.7, 140.5, 127.9, 125.1, 115.1, 80.9, 77.8, 77.6, 68.8, 65.6, 63.0, 62.9, 62.8, 48.7, 49.0, 45.9, 43.4, 41.7, 40.6, 35.3, 31.4, 28.6, 28.0, 27.7, 22.3, 21.0, 20.7, 19.2, 16.3, 16.2, 14.7, 14.5, 12.4, 7.2, 7.0, 6.9, 6.8, 5.3, 5.2, 4.9; HRMS (ES⁺) calcd for $C_{62}H_{121}O_{13}Si_4PNa$ (M + Na) 1239.7520, found 1239.7458.



Alcohol 107. To a stirred solution of ester SI-30 (230 mg, 0.189 mmol) in MeOH (10 mL) at rt was added Ba(OH), 8H2O (66.4 mg, 0.189 mmol). After 1 h, the reaction mixture was purified directly by chromatography over silica gel, eluting with 20-60% EtOAc/hexanes, to give alcohol 107 (204 mg, 0.168 mmol, 89%) as a colorless oil: $[\alpha]_{D}^{23} = -27.0$ (c 0.80, CHCl₃); IR (neat) 3434, 2955, 2877, 1716, 1458, 1376, 1242, 1019, 742 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$: δ 6.68 (t, J = 6.8 Hz, 1H), 5.78 (s, 1H), 5.11–5.00 (m, 1H), 5.00 (s, 1H), 4.86 (s, 1H), 4.19–4.05 (m, 6H), 3.73–3.66 (m, 3H), 3.07 (d, J = 27.8 Hz, 2H), 2.93 (dd, J = 18.0, 6.1 Hz, 1H), 2.78 (dd, J = 18.3, 5.7 Hz, 1H), 2.65 (t, J = 7.1 Hz, 2H), 2.22–2.11 (m, 3H), 1.92–1.59 (m, 8H), 1.81 (s, 6H), 1.43 (s, 3H), 1.42–1.39 (m, 3H), 1.33 (t, J = 7.0 Hz, 6H), 1.23 (d, *J* = 6.3 Hz, 3H), 1.02–0.85 (m, 39H), 0.78 (d, *J* = 6.5 Hz, 3H), 0.69–0.52 (m, 24H); 13 C NMR (75 MHz, CDCl₃) δ 209.0, 201.4, 167.6, 144.8, 142.3, 140.5, 129.0, 125.5, 114.9, 80.8, 77.8, 77.7, 68.8, 65.6, 62.6, 62.5, 61.0, 49.5, 49.0, 46.2, 43.3, 41.6, 40.6, 40.0, 31.4, 28.3, 28.1, 27.7, 22.3, 20.7, 19.3, 16.3,

16.2, 14.7 (2C), 12.4, 7.2, 7.0, 6.9, 6.8, 5.2, 5.1, 4.9; HRMS (ES⁺) calcd for $\rm C_{60}H_{119}O_{12}Si_4PNa(M+Na)$ 1197.7414, found 1197.7422.



Macrocycle 109. To a stirred solution of alcohol 107 (125 mg, 0.106 mmol) in CH₂Cl₂ (4 mL) at rt was added TPAP (45 mg, 0.127 mmol). After 0.5 h, the reaction mixture was diluted with THF $(6.5 \text{ mL})/\text{H}_2\text{O}$ (16 μ L), and Ba(OH)₂·8H₂O (3 × 74 mg, 0.636 mmol) was added in three portions every 30 min. After another 2 h, the reaction mixture was purified directly by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give macrocycle 109 (54 mg, 0.053 mmol, 50% over two steps) as colorless crystals: $[\alpha]^{23}_{D} = -27.0$ (c 0.40, CHCl₃); IR (neat) 2955, 2925, 2876, 1727, 1708, 1675, 1458, 1417, 1260, 1127, 1064, 1009, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.15–7.03 (m, 1H), 6.73 (t, J = 6.9 Hz, 1H), 6.19 (d, J = 16.3 Hz, 1H), 5.78 (s, 1H), 5.00 (s, 1H), 5.05-4.97 (m, 1H), 4.82 (s, 1H), 4.18–4.09 (m, 1H), 4.11 (d, J = 5.1 Hz, 1H), 3.62– 3.58 (m, 1H), 3.00-2.93 (m, 2H), 2.85-2.70 (m, 1H), 2.62-2.50 (m, 1H), 2.37-2.21 (m, 3H), 2.15-2.00 (m, 2H), 1.91-1.63 (m, 7H), 1.80 (s, 6H), 1.45–1.35 (m, 2H), 1.41 (s, 3H), 1.25 (d, J = 6.1 Hz, 3H), 1.08–0.83 (m, 39H), 0.75–0.47 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 206.2, 201.0, 167.6, 147.2, 144.7, 141.8, 140.6, 132.4, 129.2, 125.8, 115.4, 80.5, 79.6, 77.9, 68.3, 64.9, 49.7, 48.6, 46.7, 43.2, 41.1, 37.5, 31.0, 29.2, 28.9, 27.8, 22.6, 21.0, 18.8, 15.6, 13.3, 12.5, 7.3, 7.1, 7.0, 5.2, 4.8; HRMS (ES⁺) calcd for C₅₆H₁₀₆O₈Si₄Na (M + Na) 1041.6863, found 1041.6812.



TES Ether SI-31. To a stirred solution of aldol adduct 104 (440 mg, 0.496 mmol) in CH_2Cl_2 (20 mL) at rt were added sequentially DMAP (910 mg, 7.44 mmol) and TESCl (557 mg, 0.620 mL, 3.72 mmol). After 3 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 2% EtOAc/hexanes, to give TES ether SI-31 (445 mg, 0.444 mmol, 90%) as a colorless oil: $[\alpha]^{23}_{D} = -20.0 \ (c \ 0.24, \ CHCl_{3}); \ IR \ (neat) \ 2955, \ 2912, \ 2877, \ 1744,$ 1717, 1458, 1249, 1127, 1069, 1008, 840, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.81 (s, 1H), 5.00 (s, 1H), 4.85 (s, 1H), 4.43-4.30 (m, 1H), 4.16-4.03 (m, 3H), 3.88-3.78 (m, 1H), 3.69-3.72 (m, 1H), 2.88 (dd, J = 17.4, 4.8 Hz, 1H), 2.73 (dd, J = 17.4, 7.2 Hz, 1H), 2.13 (dd, J = 13.8, 6.0 Hz, 1H), 2.04 (s, 3H), 1.82-1.94 (m, 3H), 1.79 (s, 3H), 1.64–1.75 (m, 3H), 1.46 (s, 3H), 1.39–1.51 (m, 2H), 1.20– 1.28 (m, 1H), 1.14 (d, J = 6.0 Hz, 3H), 0.84–1.03 (m, 39H), 0.77 (d, J = 6.6 Hz, 3H), 0.56–0.71 (m, 24H), 0.11 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 208.4, 171.1, 144.3, 143.8, 124.3, 114.8, 81.1, 78.4, 77.3, 66.1 (2C), 62.9, 50.3, 48.7, 45.9, 45.2, 35.2, 30.7, 28.7, 26.9, 24.7, 21.0, 19.3, 14.8, 13.8, 7.2, 7.0, 6.9, 6.7, 5.4, 5.3, 5.0, 0.3; HRMS (ES⁺) calcd for $C_{52}H_{108}O_8Si_5Na$ (M + Na) 1023.6788, found 1023.6737.



Alcohol 106. To a stirred solution of TMS ether SI-31 (432 mg, 0.43 mmol) in THF/H₂O (9 mL, 8:1) at -20 °C was added HOAc

(8 mL). After 5 h, the reaction was quenched with solid NaHCO₃, filtered over Celite and extracted with ether $(4 \times 20 \text{ mL})$. The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 5-10-20% EtOAc/ hexanes, to give alcohol 106 (338 mg, 0.36 mmol, 85%) as a colorless oil: $[\alpha]^{23}_{D} = -20.8$ (c 1.01, CHCl₃); IR (neat) 3503, 2955, 2912, 2877, 1744, 1720, 1458, 1414, 1367, 1239, 1007, 741 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.85 (s, 1H), 5.02 (s, 1H), 4.86 (s, 1H), 4.50-4.40 (m, 1H), 4.18-4.03 (m, 3H), 3.85-3.77 (m, 1H), 3.67 (t, J = 5.0 Hz, 1H), 2.74–2.86 (m, 2H), 2.19 (br, OH), 2.14 (dd, J = 13.6, 6.3 Hz, 1H), 2.06 (s, 3H), 1.89–1.96 (m, 2H), 1.80 (s, 3H), 1.66– 1.85 (m, 5H), 1.45 (s, 3H), 1.39–1.42 (m, 1H), 1.19 (d, J = 6.0 Hz, 3H), 1.10-1.22 (m, 1H), 0.95-1.05 (m, 36H), 0.90 (d, J = 6.4 Hz, 3H), 0.85 (d, J = 6.4 Hz, 3H), 0.59–0.72 (m, 24H); ¹³C NMR (100 MHz, CDCl₃) δ 208.4, 171.2, 144.3, 143.5, 124.4, 115.0, 82.1, 78.5, 77.3, 66.0, 65.9, 63.0, 50.0, 48.0, 45.9, 44.4, 35.2, 32.3, 28.7, 27.3, 24.2, 21.0, 19.3, 15.4, 15.0, 7.3, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 4.9; HRMS (ES⁺) calcd for $C_{49}H_{100}O_8Si_4Na$ (M + Na) 951.6393, found 951.6418.



Phosphonate SI-32. To a stirred solution of acid 11 (837 mg, 2.73 mmol) in PhMe (6 mL) at rt were added sequentially Et₃N (276 mg, 0.379 mL, 2.73 mmol) and 2,4,6-trichlorobenzoyl chloride (641 mg, 0.411 mL, 2.73 mmol). After 12 h, the resulted solution was concentrated in vacuo. DMAP (333 mg, 2.73 mmol) was added, followed by the addition of a solution of alcohol 106 (445 mg, 0.479 mmol) in PhMe (10.5 mL). After another 19 h, the reaction was quenched with satd aq NH₄Cl (15 mL) and extracted with EtOAc (4 \times 50 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 20-60% EtOAc/hexanes, to give phosphonate SI-32 (450 mg, 0.100 mmol, 77%) as a colorless oil: $[\alpha]^{23}_{D} = -20.3$ (c 1.23, CHCl₃); IR (neat) 2955, 2913, 2877, 1740, 1716, 1458, 1368, 1243, 1056, 1019, 968, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.69 (t, J = 6.4 Hz, 1H), 5.82 (s, 1H), 5.10-5.02 (m, 1H), 5.01 (s, 1H), 4.84 (s, 1H), 4.42-4.32 (m, 1H), 4.21-4.02 (m, 7H), 3.76–3.68 (m, 1H), 3.12 (d, J = 22.8 Hz, 2H), 2.85 (dd, J = 17.4, 4.1 Hz, 1H), 2.65-2.76 (m, 3H), 2.12-2.22 (m, 2H), 2.10-2.04 (m, 1H), 2.05 (s, 3H), 1.58-1.93 (m, 10H), 1.82 (s, 3H), 1.79 (s, 3H), 1.45 (s, 3H), 1.43–1.39 (m, 1H), 1.35 (t, J = 7.1 Hz, 6H), 1.24 (d, J = 5.9 Hz, 3H), 0.87–1.03 (m, 39H), 0.79 (d, J = 6.6 Hz, 3H), 0.55–0.68 (m, 24H); ¹³C NMR (75 MHz, CDCl₃) δ 208.3, 201.4, 171.1, 167.6, 144.3, 143.7, 140.5, 129.0, 124.4, 114.9, 81.1, 77.9, 77.34, 68.8, 66.2, 62.9, 62.6, 62.5, 50.3, 49.0, 45.9, 43.3, 41.6, 40.8, 35.2, 31.1, 28.7, 27.7, 26.9, 22.3, 21.0, 20.7, 19.3, 16.3, 16.2, 14.8, 14.3, 12.4, 7.2, 7.1, 7.0, 6.9, 6.7, 5.3, 5.2, 5.0; HRMS (ES⁺) calcd for C₆₂H₁₂₁O₁₃Si₄PNa (M + Na) 1239.7520, found 1239.7563.



Alcohol 108. To a stirred solution of ester SI-32 (170 mg, 0.140 mmol) in MeOH (0.5 mL) at rt was added a saturated solution of Ba(OH)₂·8H₂O in MeOH (6.0 mL). After 20 min, the reaction mixture was purified directly by chromatography over silica gel, eluting with 20–60% EtOAc/hexanes, to give alcohol **108** (150 mg, 0.127 mmol, 91%) as a colorless oil: $[\alpha]_{23}^{23} = -20.3$ (*c* 0.60, CHCl₃); IR (neat) 3440,

2955, 2877, 1716, 1458, 1242, 1019, 969, 742 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 6.69 (t, J = 6.2 Hz, 1H), 5.82 (s, 1H), 5.10–5.03 (m, 1H), 5.00 (s, 1H), 4.83 (s, 1H), 4.42–4.31 (m, 1H), 4.20–4.08 (m, SH), 3.63–3.74 (m, 3H), 3.12 (d, J = 22.8 Hz, 2H), 2.78–2.72 (m, 2H), 2.66 (t, J = 7.1 Hz, 2H), 2.04–2.19 (m, 3H), 1.89–1.61 (m, 10H), 1.81 (s, 3H), 1.78 (s, 3H), 1.44 (s, 3H), 1.43–1.38 (m, 1H), 1.35 (t, J = 7.1 Hz, 6H), 1.24 (d, J = 5.9 Hz, 3H), 1.03–0.92 (m, 36H), 0.87 (d, J = 6.4 Hz, 3H), 0.79 (d, J = 6.6 Hz, 3H), 0.55–0.71 (m, 24H); ¹³C NMR (100 MHz, CDCl₃) δ 208.5, 201.5, 167.7, 144.6, 143.5, 140.6, 129.0, 124.7, 114.7, 81.2, 78.0, 68.8, 66.1, 62.6, 62.5, 61.1, 50.3, 49.1, 46.1, 43.5, 43.4, 41.8, 40.9, 39.8, 31.1, 28.5, 27.8, 27.1, 22.3, 20.8, 19.5, 16.4, 16.3, 14.9, 14.2, 12.4, 7.3, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 5.0; HRMS (ES⁺) calcd for C₆₀H₁₁₉O₁₂Si₄PNa (M + Na) 1197.7414, found 1197.7423.



Macrocycle 110. To a stirred solution of alcohol 108 (170 mg, 0.144 mmol) in CH₂Cl₂ (4 mL) at rt was added TPAP (61 mg, 0.173 mmol). After 0.5 h, the reaction mixture was diluted with CH_2Cl_2 $(3 \text{ mL})/\text{CH}_3\text{CN}(7 \text{ mL})$ and Hunig's base (297 mg, 0.4 mL, 2.29 mmol) was added, followed by the addition of LiCl (20 mg, 0.476 mmol). After 24 h, the reaction mixture was purified directly by chromatography over silica gel, eluting with 5% EtOAc/hexanes, to give macrocycle 110 (75 mg, 0.073 mmol, 51% over two steps) as a colorless oil: $[\alpha]^{23}_{D} =$ -10.0 (c 0.62, CHCl₃); IR (neat) 2955, 2913, 2876, 1708, 1674, 1457, 1417, 1375, 1240, 1124, 1072, 1008, 742 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) δ 6.88 (dt, J = 16.2, 7.4 Hz, 1H), 6.72 (t, J = 8.0 Hz, 1H), 6.10 (d, J = 16.0 Hz, 1H), 5.82 (s, 1H), 5.08-4.99 (m, 1H), 5.02 (s, 1H), 4.87(s, 1H), 4.30–4.22 (m, 1H), 4.01 (d, J = 6.4 Hz, 1H), 3.58 (dd, J = 6.4, 2.7 Hz, 1H), 2.86 (dd, J = 17.4, 8.6 Hz, 1H), 2.77 (dd, J = 15.4, 6.0 Hz, 1H), 2.62–2.54 (m, 2H), 2.36–2.19 (m, 4H), 2.14 (dd, J = 17.4, 8.6 Hz, 1H), 2.05–1.76 (m, 7H), 1.80 (s, 3H), 1.79 (s, 3H), 1.70–1.60 (m, 1H), 1.57–1.51 (m, 1H), 1.41 (s, 3H), 1.25 (d, J = 6.2 Hz, 3H), 1.05– 0.93 (m, 36H), 0.84 (d, J = 6.6 Hz, 3H), 0.76 (d, J = 6.6 Hz, 3H), 0.73-0.58 (m, 24H); ¹³C NMR (100 MHz, CDCl₃) δ 208.1. 201.0, 167.6, 147.3, 147.1, 144.0, 142.8, 140.8, 132.4, 129.3, 124.7, 115.1, 80.6, 77.4 (2C), 68.5, 65.1, 50.6, 49.0, 46.1, 41.7, 40.2, 37.2, 31.3, 30.8, 27.8, 27.4, 23.1, 21.0, 19.7, 15.3, 12.8, 12.5, 7.3, 7.1, 7.0, 6.9, 5.3, 5.2, 4.9; HRMS (ES⁺) calcd for C₅₆H₁₀₆O₈Si₄Na (M + Na) 1041.6863, found 1041.6824.



Allylic Alcohol 111. To a stirred solution of macrocycle 110 (107 mg, 0.105 mmol) in CH₂Cl₂ (5.4 mL) at -20 °C were added sequentially (*S*)-CBS (0.42 mL, 0.42 mmol, 1 M in PhMe) and BH₃·DMS (0.84 mL, 0.84 mmol, 1 M in THF). After 45 min, the reaction was quenched with MeOH (0.3 mL), diluted with aq NaHCO₃ (5 mL), and extracted with Et₂O (3 × 8 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 3–6% EtOAc/hexanes, to give allylic alcohol 111 (72 mg, 0.0704 mmol, 67%) as a colorless oil: $[\alpha]^{23}_{D} = -16.3$ (*c* 0.30, CHCl₃); IR (neat) 3431, 2954, 2913, 2876, 1708, 1674, 1458, 1414, 1376, 1241, 1128, 1073, 1009, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.80 (t, *J* = 8.0 Hz, 1H), 5.89 (s, 1H), 5.63–5.52 (m, 1H), 5.45 (dd, *J* = 15.4, 7.5 Hz, 1H), 5.10–5.00

(m, 1H), 4.98 (s, 1H), 4.82 (s, 1H), 4.32–4.20 (m, 1H), 4.19–4.09 (m, 1H), 4.04 (d, J = 6.2 Hz, 1H), 3.57 (dd, J = 6.2, 2.5 Hz, 1H), 2.87 (dd, J = 17.0, 9.0 Hz, 1H), 2.49 (d, J = 16.8 Hz, 1H), 2.28–2.20 (m, 2H), 2.13–1.98 (m, 3H), 1.90–1.61 (m, 9H), 1.86 (s, 3H), 1.75 (s, 3H), 1.55–1.46 (m, 2H), 1.39 (s, 3H), 1.26 (d, J = 6.0 Hz, 3H), 1.06–0.94 (m, 36H), 0.82 (d, J = 6.4 Hz, 3H), 0.73–0.60 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 208.5, 167.8, 144.6, 142.0, 141.3, 134.6, 130.8, 128.2, 125.4, 114.4, 80.6, 78.0, 72.9, 68.4, 65.2, 50.3, 49.2, 45.4, 41.8, 39.7, 36.8, 31.3, 30.4, 28.8, 28.2, 24.0, 20.9, 19.6, 15.4, 12.5, 12.4, 7.3, 7.1, 7.0, 5.3, 5.2, 4.9; HRMS (ES⁺) calcd for C₅₆H₁₀₈O₈Si₄Na (M + Na) 1043.7019, found 1043.7052.



Epoxide 113 and 112. To a stirred solution of allylic alcohol 111 (70 mg, 0.0685 mmol) in CH_2Cl_2 (6 mL) at -20 °C were added sequentially 4 Å MS (50 mg), TBHP (37 µL, 0.206 mmol, 5.5 M in decane), and Ti(O-i-Pr)₄ (23.3 mg, 24 μ L, 0.082 mmol). After 5 h, the reaction was quenched with aq NaHCO₃ (3 mL) and extracted with Et₂O (3 \times 7 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 6-10% EtOAc/hexanes, to give epoxide 113 (35 mg, 0.0342 mmol, 50%) and epoxide 112 (17 mg, 0.0166 mmol, 24%) as colorless oils. 113: $\left[\alpha\right]_{D}^{23} = -29.0 (c \ 0.42, CHCl_{3}); IR (neat) 3431, 2954, 2923, 2876, 1708,$ 1647, 1458, 1414, 1377, 1242, 1128, 1073, 1009, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.78 (t, J = 8.0 Hz, 1H), 5.88 (s, 1H), 5.05–4.98 (m, 1H), 5.01 (s, 1H), 4.86 (s, 1H), 4.30–4.20 (m, 1H), 4.05 (d, J = 6.1 Hz, 1H), 3.59 (dd, J = 6.1, 2.0 Hz, 1H), 3.52-3.43 (m, 1H), 3.22-3.15 (m, 1H), 2.91 (dd, J = 16.3, 9.1 Hz, 1H), 2.70 (dd, J = 6.0, 2.0 Hz, 1H), 2.44-2.25 (m, 3H), 2.18 (dd, J = 13.0, 5.6 Hz, 1H), 2.10-1.63 (m, 12H), 1.87 (s, 3H), 1.78 (s, 3H), 1.41 (s, 3H), 1.26 (d, J = 6.0 Hz, 3H), 1.15-1.09 (m, 1H), 1.07-0.89 (m, 36H), 0.80 (d, J = 6.4 Hz, 3H), 0.76–0.59 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 208.4, 167.7, 144.0, 142.1, 141.4, 128.6, 125.0, 114.9, 80.8, 78.2, 77.5, 71.8, 68.3, 65.6, 62.8, 56.4, 50.6, 49.1, 46.6, 42.1, 38.9, 33.1, 30.1, 29.4, 28.8, 28.3, 23.7, 21.0, 19.8, 15.5, 12.5, 12.4, 7.3, 7.1, 7.0, 5.3, 5.2, 5.0; HRMS (ES⁺) calcd for C₅₆H₁₀₈O₉Si₄Na (M + Na) 1059.6968, found 1059.7009. 112: $[\alpha]_{\rm D} = -26.2$ (c 0.60, CHCl₃); IR (neat) 3482, 2954, 2923, 2876, 1708, 1458, 1414, 1377, 1240, 1128, 1008, 742 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) $\delta 6.79$ (t, J = 7.7 Hz, 1H), 5.87 (s, 1H), 5.05–4.98 (m, 1H), 4.99 (s, 1H), 4.84 (s, 1H), 4.30–4.20 (m, 1H), 4.09 (d, J = 5.9 Hz, 1H), 3.75–3.69 (m, 1H), 3.57 (dd, J = 5.6, 3.2 Hz, 1H), 2.91–2.78 (m, 3H), 2.68–2.60 (m, 2H), 2.38-2.20 (m, 3H), 2.07 (dd, J = 12.9, 6.2 Hz, 1H), 1.93-1.62 (m, 10H), 1.85 (s, 3H), 1.76 (s, 3H), 1.50-1.44 (m, 2H), 1.40 (s, 3H), 1.25 (d, J = 6.0 Hz, 3H), 1.05–0.93 (m, 39H), 0.74–0.59 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 208.6, 167.8, 144.1, 142.5, 141.5, 128.4, 124.9, 114.9, 81.0, 78.2, 77.4, 69.0, 68.4, 65.5, 60.7, 54.9, 50.3, 49.2, 46.3, 41.7, 38.4, 33.3, 30.7, 30.3, 29.0, 27.9, 23.7, 21.0, 20.0, 15.3, 13.2, 12.4, 7.3, 7.1, 7.0, 6.9, 5.3, 5.2, 4.9; HRMS (ES⁺) calcd for C₅₆H₁₀₈O₉Si₄Na (M + Na) 1059.6968, found 1059.7009.



Selenide 115. To a stirred solution of epoxide **113** (42 mg, 0.0405 mmol) in THF (2 mL) at rt were added sequentially o-NO₂C₆H₄SeCN (184 mg, 0.809 mmol) and PBu₃ (164 mg, 202 μ L, 0.809 mmol). After 5 h, the reaction mixture was concentrated in vacuo purified by chromatography over silica gel, eluting with hexanes then with 4% EtOAc/hexanes, to give crude selenide **115** (30 mg) as a yellow oils which was used directly in next step without further purification.

Polyol SI-33. To a stirred solution of selenide 115 (30 mg) in THF/ DMF/H2O (10:1:0.02, 1.8 mL/180 µL/3.6 µL) at 0 °C was added TAS-F (33.7 mg, 0.123 mmol). The reaction mixture was then warmed to rt. After 2 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10-65% EtOAc/hexanes, to give polyol SI-33 (15 mg, 0.0196 mmol, 48% over two steps) as a yellow solid: $[\alpha]_{D}^{23} = -20.1$ (c 0.12, CHCl₃); IR (neat) 3447, 2925, 2854, 1701, 1520, 1456, 1334, 1273, 759, 732 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.28 (d, J = 8.3 Hz, 1H), 7.66 (d, J = 7.8 Hz, 1H), 7.56 (t, J = 8.1 Hz, 1H), 7.38 (t, J = 8.1 Hz, 1H), 6.81 (t, J = 6.4 Hz, 1H), 6.05 (s, 1H), 5.10–5.00 (m, 1H), 5.02 (s, 1H), 4.80 (s, 1H), 4.38 (d, J = 3.6 Hz, 1H), 4.18 (s, 1H), 4.17–4.11 (m, 1H), 4.07 (t, J = 9.6 Hz, 1H), 3.58 (t, J = 8.6 Hz, 1H), 3.18–3.10 (m, 1H), 2.97 (d, J = 8.1 Hz, 1H), 2.79 (dd, J = 12.8, 10.1 Hz, 1H), 2.37–2.28 (m, 1H), 2.40-2.30 (m, 1H), 2.22-2.04 (m, 3H), 1.94-1.76 (m, 7H), 1.83 (s, 3H), 1.79 (s, 3H), 1.72–1.62 (m, 3H), 1.38 (s, 3H), 1.31 (d, J = 6.1 Hz, 3H), 1.25–1.19 (m, 2H), 1.07 (d, J = 6.4 Hz, 3H), 0.83 (d, J = 6.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.1, 167.6, 149.6, 144.4, 141.0 (2C), 133.1, 132.5, 128.9, 128.3, 127.0, 126.0, 125.1, 115.0, 78.3, 77.5, 75.3, 68.7, 68.3, 62.3, 59.3, 46.7, 45.7, 45.5, 43.8, 40.4, 40.0, 32.8, 31.6, 29.1, 29.0, 28.1, 26.1, 21.2, 17.7, 16.2, 15.3, 12.5; HRMS (ES⁺) calcd for $C_{38}H_{55}NO_{10}NaSe (M + Na)$ 788.2889, found 788.2859.



Proposed Structure of Amphidinolide B₂ (3). To a stirred solution of selenide SI-33 (6.0 mg, 0.00784 mmol) in CH₂Cl₂ (1.2 mL) at rt were added sequentially NaHCO₃ (60 mg, 0.714 mmol) and TMSOOTMS (41.7 mg, 50 μL, 0.233 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc/hexanes, to give allylic epoxide 3 (3.0 mg, 0.00533 mmol, 68%): $[α]^{23}_{D} = -52.3 (c 0.21, CHCl_3); IR (neat) 3446, 2923, 2853, 1701, 1457, 1273, 1120 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.74 (t,$ *J*= 6.4 Hz, 1H), 6.06 (s, 1H), 5.92 (ddd,*J*= 15.0, 8.9, 4.4 Hz, 1H), 5.20 (dd,*J*= 15.5, 8.8 Hz, 1H), 5.11–5.07 (m, 1H), 5.07 (s, 1H), 4.86 (s, 1H), 4.28 (d,*J*= 5.4, 1H), 4.14 (s, OH), 4.14–4.09 (m, 1H), 3.73 (d,*J*= 5.6 Hz, OH), 3.69 (t,*J*= 9.5 Hz, 1H), 3.50 (d,*J*= 6.8 Hz, 1H), 3.23 (dd,*J*= 8.7, 2.2 Hz, 1H), 3.08–3.03 (m, 2H), 2.95

(d, *J* = 9.2 Hz, OH), 2.53–2.45 (m, 1H), 2.45–2.36 (m, 1H), 2.28 (dd, *J* = 13.2, 1.8 Hz, 1H), 2.17–2.12 (m, 3H), 1.97–1.93 (m, 4H), 1.85 (s, 3H), 1.82–1.79 (m, 1H), 1.80 (s, 3H), 1.78–1.75 (m, 1H), 1.64–1.60 (m, 1H), 1.34 (s, 3H), 1.31 (d, *J* = 6.1 Hz, 3H), 1.17–1.12 (m, 1H), 1.03 (d, *J* = 6.7 Hz, 3H), 0.98 (d, *J* = 5.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 212.5, 167.7, 144.7, 141.6, 139.5, 136.3, 128.4, 128.3, 124.9, 114.6, 78.1, 75.6, 69.28, 68.23, 61.45, 59.5, 47.1, 46.4, 44.1, 40.0, 39.4, 33.3, 31.0, 29.3, 28.3, 26.7, 21.2, 17.5, 15.9, 15.2, 12.6; HRMS (ES⁺) calcd for C₃₂H₅₀O₈Na (M + Na) 585.3403, found 585.3390.



Selenide SI-34. To a stirred solution of epoxide **112** (12 mg, 0.0116 mmol) in THF (0.7 mL) at rt were added sequentially o-NO₂C₆H₄SeCN (53 mg, 0.232 mmol) and PBu₃ (47 mg, 58 μ L, 0.232 mmol). After 0.5 h, the reaction mixture was concentrated in vacuo purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc/hexanes, to give crude selenide SI-34 (10.6 mg) as a yellow oil which was used directly in next step without further purification.

Polyol SI-35. To a stirred solution of selenide SI-34 (10.6 mg) in THF/DMF/H₂O (10:1:0.02, 1.0 mL/100 µL/2.0 µL) at 0 °C was added TAS-F (12 mg, 0.0434 mmol). The reaction mixture was then warmed to rt. After 2 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10-65% EtOAc/hexanes, to give polyol SI-35 (6.2 mg, 0.00811 mmol, 70% over two steps) as a yellow oil: $[\alpha]_{\rm D} =$ -47.0 (c 0.30, CHCl₃); IR (neat) 3446, 2925, 2854, 1701, 1515, 1456, 1332, 1271, 757, 731 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, J = 8.1 Hz, 1H), 7.70 (d, J = 8.0 Hz, 1H), 7.55 (t, J = 7.4 Hz, 1H),7.42 (t, J = 7.8 Hz, 1H), 6.85 (t, J = 6.3 Hz, 1H), 6.02 (s, 1H), 5.11-5.05 (m, 1H), 5.04 (s, 1H), 4.81 (s, 1H), 4.38 (d, J = 3.6 Hz, 1H), 4.32-4.25 (m, 1H), 4.17-4.12 (m, 1H), 3.89 (d, J = 4.6 Hz, 1H), 3.72-3.65 (m, 1H), 3.62-3.55 (m, 1H), 3.10-3.05 (m, 2H), 2.84 (dd, J = 14.8, 9.2 Hz, 1H), 2.52 (dd, J = 14.8, 2.5 Hz, 1H), 2.38 (d, J = 9.5 Hz, 1H), 2.40-2.20 (m, 3h), 1.97-1.74 (m, 8H), 1.83 (s, 3H), 1.76 (s, 3H), 1.70–1.60 (m, 3H), 1.36 (s, 3H), 1.33 (d, J = 6.1 Hz, 3H), 1.06 (d, J = 6.7 Hz, 3H), 0.87 (d, J = 6.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 212.0, 167.7, 147.7, 144.6, 141.3, 140.9, 133.7, 132.3, 130.1, 128.6, 126.5, 126.1, 125.6, 115.1, 78.0, 77.0, 75.5, 69.1, 67.7, 60.6, 58.4, 45.7, 45.6, 43.7, 43.1, 40.4, 39.2, 33.8, 30.3, 29.5, 28.9, 28.0, 27.0, 21.2, 19.7, 16.3, 15.2, 12.5; HRMS (ES^+) calcd for $C_{38}H_{55}NO_{10}NaSe$ (M + Na) 788.2889, found 788.2897.



Allylic Epoxide 114. To a stirred solution of selenide SI-35 (2.5 mg, 0.00327 mmol) in CH_2Cl_2 (0.5 mL) at rt were added sequentially NaHCO₃ (20 mg, 0.238 mmol) and TMSO-OTMS (19.1 mg, 23 μ L, 0.107 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc/hexanes, to give allylic epoxide 114 (1.2 mg, 0.00213 mmol, 65%): $[\alpha]_{\rm D} = -27.5$ (c 0.12, CHCl₃); IR (neat) 3443, 2924, 2852, 1703, 1457, 1379, 1272, 1118 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 6.71 - 6.63 \text{ (m, 1H)}, 6.08 \text{ (s, 1H)}, 5.88 - 5.78$ (m, 1H), 5.26 (dd, J = 15.1, 8.4 Hz, 1H), 5.12-5.05 (m, 1H), 5.05 (s, 1H), 4.84 (s, 1H), 4.29 (dd, J = 5.7, 1.5 Hz, 1H), 4.30-4.20 (m, 100)1H), 3.71 (t, J = 7.9 Hz, 1H), 3.66 (d, J = 5.7 Hz, 1H), 3.19 (d, J = 9.6Hz, 1H), 3.12 (dd, J = 8.6, 2.2 Hz, 1H), 3.01-2.87 (m, 2H), 2.46 (dd, J = 14.4, 2.1 Hz, 1H), 2.36-2.20 (m, 5H), 2.00-1.74 (m, 6H),1.83 (s, 3H), 1.77 (s, 3H), 1.60-1.55 (m, 1H), 1.35 (s, 3H), 1.33-1.28 (m, 1H), 1.30 (d, J = 6.1 Hz, 3H), 1.18–1.11 (m, 1H), 1.03 (d, I = 6.6 Hz, 3H), 0.88 (d, I = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 212.9, 167.5, 144.9, 141.6, 140.3, 136.4, 128.7, 128.6, 125.7, 115.0, 78.2, 76.6, 75.9, 68.5, 68.2, 60.2, 60.0, 45.8, 45.1, 43.8, 39.4, 39.3, 33.6, 31.0, 30.3, 28.5, 27.0, 21.2, 20.0, 15.8, 15.2, 12.7; HRMS (ES⁺) calcd for $C_{32}H_{50}O_8Na$ (M + Na) 585.3403, found 585.3394.



Allylic Alcohols 118 and 119. To a stirred solution of macrocycle 109 (50 mg, 0.049 mmol) in CH₂Cl₂ (2.3 mL) at -30 °C were added sequentially (S)-CBS (0.196 mL, 0.196 mmol, 1 M in PhMe) and BH₃·DMS (0.3934 mL, 0.393 mmol, 1 M in THF). After 45 min, the reaction was quenched with MeOH (0.3 mL), diluted with aq NaHCO₃ (3 mL), and extracted with Et_2O (4 × 6 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 3-6% EtOAc/hexanes, to give allylic alcohol 118 (29 mg, 0.028 mmol, 58%) and SI-36 (8 mg, 0.0078 mmol, 16%) as colorless oils. 118: $[\alpha]^{23}_{D} = -23.6$ (c 0.25, CHCl₃); IR (neat) 3503, 2954, 2911, 2876, 1707, 1458, 1376, 1240, 1128, 1007, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.81 (t, J = 6.9 Hz, 1H), 5.76 (s, 1H), 5.70–5.62 (m, 1H), 5.53 (dd, J = 13.2, 6.3 Hz, 1H), 5.00-4.95 (m, 1H), 4.98 (s, 1H), 4.84 (s, 1H), 4.15-4.04 (m, 3H), 3.58 (dd, *J* = 5.7, 2.4 Hz, 1H), 2.93 (dd, *J* = 18.4, 6.2 Hz, 1H), 2.78 (dd, J = 18.4, 6.0 Hz, 1H), 2.28–2.22 (m, 2H), 2.18–2.10 (m, 2H), 2.08-1.99 (m, 1H), 1.89-1.57 (m, 9H), 1.84 (s, 3H), 1.81 (s, 3H), 1.48-1.40 (m, 2H), 1.45 (s, 3H), 1.26 (d, J = 6.1 Hz, 3H), 1.07-0.91 (m, 36H), 0.88 (d, J = 6.6 Hz, 3H), 0.74–0.53 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 207.8, 167.8, 145.5, 142.5 (2C), 134.9, 129.7, 127.9, 125.7, 114.9, 80.6, 79.2, 77.9, 71.9, 68.2, 65.3, 49.6, 49.2, 45.6, 42.2, 39.4, 37.0, 31.8, 29.8, 29.0, 28.1, 23.6, 21.0, 19.9, 14.7, 12.7, 12.4, 7.2, 7.1, 7.0, 6.9, 6.8, 5.3, 5.2, 4.7; HRMS (ES⁺) calcd for $C_{56}H_{108}O_8Si_4Na$ (M + Na) 1043.7019, found 1043.7072. SI-36: $[\alpha]_{\rm D} = -29.1$ (c 0.80, CHCl₃); IR (neat) 3481, 2954, 2876, 1707, 1458, 1241, 1130, 1008, 742 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.74 (t, J = 7.4 Hz, 1H), 5.77 (s, 1H), 5.75–5.70 (m, 1H), 5.55 (dd, J = 15.5, 7.0 Hz, 1H), 5.00-4.96 (m, 1H), 4.99 (s, 1H), 4.83 (s, 1H), 4.22-4.15 (m, 1H), 4.11-4.06 (m, 1H), 4.07 (d, J = 5.8 Hz, 1H),

3.57 (dd, J = 5.6, 2.3 Hz, 1H), 2.96 (dd, J = 18.4, 7.4 Hz, 1H), 2.78 (dd, J = 18.3, 4.2 Hz, 1H), 2.23–2.08 (m, 4H), 1.95–1.52 (m, 11H), 1.834 (s, 3H), 1.80 (s, 3H), 1.48–1.40 (m, 1H), 1.43 (s, 3H), 1.25 (d, J = 6.1 Hz, 3H), 1.06–0.87 (m, 39H), 0.73–0.55 (m, 27H); ¹³C NMR (100 MHz, CDCl₃) δ 207.4, 167.9, 145.3, 142.2, 141.7, 134.2, 130.9, 128.3, 125.9, 114.9, 80.9, 79.0, 77.7, 73.2, 68.4, 65.1, 49.2, 48.8, 45.5, 42.1, 39.8, 36.8, 31.7, 30.1, 28.5, 28.3, 24.5, 21.0, 19.4, 15.0, 13.1, 12.4, 7.3, 7.1, 7.0, 6.9, 6.8, 5.2, 5.1, 4.8; HRMS (ES⁺) calcd for C₅₆H₁₀₈O₈Si₄Na (M + Na) 1043.7019, found 1043.6984.



Epoxides 119 and 120. To a stirred solution of allylic alcohol 118 (29 mg, 0.0284 mmol) in CH₂Cl₂ (2.2 mL) at -40 °C were added sequentially 4 Å MS (20 mg), TBHP (15.5 µL, 0.0852 mmol, 5.5 M in decane), and Ti(O-i-Pr)4 (16.1 mg, 16.6 µL, 0.0567 mmol). After 5 h, the reaction was quenched with aq NaHCO₃ (3 mL) and extracted with Et_2O (4 × 4 mL). The dried extract (MgSO₄) was concentrated in vacuo and purified by chromatography over silica gel, eluting with 6-10% EtOAc/hexanes, to give epoxide 120 (13.6 mg, 0.0131 mmol, 46%) and epoxide 119 (9.1 mg, 0.00867 mmol, 31%) as colorless oils. 120: $[\alpha]^{23}_{D} = -32.3$ (c 0.73, CHCl₃); IR (neat) 3482, 2954, 2911, 2876, 1706, 1458, 1380, 1239, 1131, 1073, 1009, 742 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 6.83 (t, J = 7.0 \text{ Hz}, 1\text{H}), 5.76 (s, 1\text{H}), 5.00-4.92$ (m, 1H),4.98 (s, 1H), 4.83 (s, 1H), 4.20-4.10 (m, 1H), 4.10(d, J = 5.5 Hz, 1H), 3.58 (dd, J = 5.4, 1.7 Hz, 1H), 3.60-3.50 (m, 1H), 3.12-3.05 (m, 1H), 2.98-2.92 (m, 2H), 2.90-2.82 (m, 1H), 2.77 (dd, J = 5.0, 2.0 Hz, 1H), 2.40–2.30 (m, 2H), 2.17 (dd, J = 12.7, 4.3 Hz, 1H), 2.12-2.00 (m, 1H), 1.91-1.61 (m, 8H), 1.82 (s, 3H), 1.79 (s, 3H), 1.50-1.40 (m, 3H), 1.42 (s, 3H), 1.30-1.20 (m, 1H), 1.26 (d, J = 6.0 Hz, 3H), 1.08–0.87 (m, 39H), 0.75–0.50 (m, 27H); ¹³C NMR (75 MHz, CDCl₃) δ 208.1, 167.8, 144.7, 141.8, 141.7, 128.2, 125.9, 115.0, 80.5, 79.4, 77.9, 70.4, 68.2, 65.1, 60.8, 55.8, 49.8, 48.9, 45.9, 42.9, 39.3, 33.6, 30.2, 29.2, 28.6, 28.5 24.0, 21.1, 19.5, 15.1, 12.9, 12.4, 7.3, 7.1, 6.9, 5.2, 4.7; HRMS (ES⁺) calcd for $C_{56}H_{108}O_9Si_4Na$ (M + Na) 1059.6968, found 1059.7001. **119**: $[\alpha]^{23}_{D} = -25.7$ (*c* 0.42, CHCl₃); IR (neat) 3482, 2954, 2876, 1708, 1458, 1378, 1240, 1130, 1008, 742 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 6.76 (t, J = 6.1 Hz, 1H), 5.83 (s, 1H), 5.05-4.90 (m, 1H), 5.01 (s, 1H), 4.86 (s, 1H), 4.15-4.10 (m, 1H), 4.08 (d, J = 5.8 Hz, 1H), 3.58 (dd, J = 5.5, 2.6 Hz, 1H), 3.59-3.49 (m, 1H), 3.11-3.03 (m, 1H), 2.95-2.91 (m, 2H), 2.77 (dd, J = 5.5, 2.0 Hz, 1H), 2.35-2.20 (m, 3H), 2.02-1.59 (m, 10H), 1.83 (s, 3H), 1.81 (s, 3H), 1.44 (s, 3H), 1.50–1.40 (m, 3H), 1.25 (d, J = 6.0 Hz, 3H), 1.06–0.89 (m, 39H), 0.74–0.52 (m, 27H); ¹³C NMR (75 MHz, CDCl₃) δ 207.7, 167.8, 144.8, 142.3, 141.5, 128.4, 125.4, 115.3, 80.8, 78.9, 77.8, 71.5, 68.4, 65.3, 61.8, 55.5, 49.2 (2C), 45.4, 41.7, 38.8, 33.3, 30.4, 29.6, 28.5, 28.3, 23.9, 21.0, 20.0, 15.0, 13.2, 12.5, 7.2, 7.1, 7.0, 6.9, 5.2, 4.8; HRMS (ES⁺) calcd for C₅₆H₁₀₈O₉Si₄Na (M + Na) 1059.6968, found 1059.6982.



Selenide SI-37. To a stirred solution of epoxide **119** (8.5 mg, 0.00819 mmol) in THF (0.5 mL) at rt were added sequentially o-NO₂C₆H₄SeCN (37 mg, 0.164 mmol) and PBu₃ (33.2 mg, 41 μ L, 0.164 mmol). After 5 h, the reaction mixture was concentrated in vacuo purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc/hexanes, to give crude selenide **SI-37** (4.5 mg) as a yellow oil which was used directly in next step without further purification.

Polyol SI-38. To a stirred solution of selenide SI-37 (4.5 mg) in THF/DMF/H₂O (10:1:0.02, 0.50 mL/50 µL/1 µL) at 0 °C was added TAS-F (5 mg, 0.0180 mmol). The reaction mixture was then warmed to rt. After 2 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10-65% EtOAc/ hexanes, to give polyol SI-38 (2.0 mg, 0.00261 mmol, 32% over two steps) as a yellow oil: $[\alpha]^{23}_{D} = -41.7$ (c 0.12, CHCl₃); IR (neat) 3446, 2924, 2854, 1701, 1519, 1457, 1378, 1334, 1121, 759, 732 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, J = 8.0 Hz, 1H), 7.70 (d, J = 8.0 Hz, 1H), 7.55 (t, J = 7.4 Hz, 1H), 7.42 (t, J = 7.5 Hz, 1H), 6.88 (t, J = 6.3 Hz, 1H), 5.87 (s, 1H), 5.15-5.08 (m, 1H), 5.03 (s, 1H), 4.82 (s, 1H), 4.50 (s, 1H), 4.20-4.10 (m, 2H), 3.78-3.69 (m, 1H), 3.20-3.10 (m, 1H), 2.94-2.77 (m, 3H), 2.39-2.33 (m, 1H), 2.30-2.13 (m, 2H), 2.07-1.50 (m, 11H), 1.84 (s, 6H), 1.41 (s, 3H), 1.40–1.28 (m, 5H), 1.07 (d, J = 6.5 Hz, 3H), 0.77 (t, J = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.4, 167.7, 149.7, 144.0, 143.0, 141.1, 133.1, 132.6, 128.8, 128.4, 127.1, 125.9, 123.9, 115.2, 77.6, 76.0, 75.3, 68.3, 66.0, 62.1, 59.0, 46.8, 46.1, 45.3, 44.7, 40.3, 40.2, 32.7, 31.9, 29.2, 28.9, 28.0, 26.3, 21.2, 17.8, 15.9, 15.4, 12.5; HRMS (ES⁺) calcd for C₃₈H₅₅NO₁₀NaSe (M + Na) 788.2889, found 788.2891.



Amphidinolide B₁ (2). To a stirred solution of selenide SI-38 (2.0 mg, 0.00261 mmol) in CH₂Cl₂ (0.4 mL) at rt were added sequentially NaHCO₃ (20 mg, 0.238 mmol) and TMSO–OTMS (16.6 mg, 20 μ L, 0.0929 mmol). After 1.5 h, the yellow color vanished, and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc/ hexanes, to give amphidinilide B_1 (2) (1.0 mg, 0.00178 mmol, 68%): $[\alpha]^{23}_{D} = -63.7 \ (c \ 0.08, \text{CHCl}_3) \ [\text{lit.}^{2c} \ [\alpha]^{23}_{D} = -62.5 \ (c \ 0.39, \text{CHCl}_3) \];$ ¹H NMR (400 MHz, CDCl₃) δ 6.78 (t, J = 7.8 Hz, 1H), 5.99 (s, 1H), 5.93 (ddd, J = 15.2, 8.5, 4.8 Hz, 1H), 5.18 (dd, J = 15.8, 8.6 Hz, 1H), 5.06 (m, 1H), 5.05(s, 1H), 4.84 (s, 1H), 4.34 (dd, J = 4.8, 1.4 Hz, 1H), 4.20 (m, 1H), 3.92 (d, J = 3.3 Hz, 1H), 3.88 (d, J = 5.0 Hz, 1H), 3.73 (ddd, J = 10.3, 8.8, 1.5 Hz, 1H), 3.19 (d, J = 10.0 Hz, 1H), 3.16 (dd, J = 8.3, 2.0 Hz, 1H), 2.94 (ddd, J = 8.9, 2.6, 2.2 Hz, 1H), 2.86 (d, J = 7.3 Hz, 1H), 2.80 (dd, J = 15.9, 3.2 Hz, 1H), 2.43 (m, 1H), 2.38 (m, 1H), 2.25 (m, 1H), 2.20 (m, 1H), 2.17 (m, 1H), 2.16 (s, 1H), 1.98–1.91 (m, 4H), 1.80 (s, 6H), 1.76 (dd, J = 14.5, 5.2 Hz, 1H), 1.64 (m, 1H), 1.49 (ddd, J = 13.6, 10.9, 3.0 Hz, 1H), 1.44 (s, 3H), 1.31 (m, 1H), 1.30 (d, J = 6.1 Hz, 3H), 1.26 (m, 1H),

1.03 (d, *J* = 6.6 Hz, 3H), 0.90 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 212.5, 167.8, 144.5, 143.2, 140.0, 135.5, 128.6, 128.5, 124.4, 114.9, 77.9, 76.1, 75.7, 68.4, 66.7, 60.2, 47.0, 46.0, 45.4, 39.5, 39.4, 33.3, 31.0, 29.4, 28.4, 26.9, 21.1, 18.3, 15.7, 15.2, 12.5; HRMS (ES⁺) calcd for C₃₂H₅₀O₈Na (M + Na) 585.3403, found 585.3411.



Selenide SI-39. To a stirred solution of epoxide **120** (14.5 mg, 0.0139 mmol) in THF (0.8 mL) at rt were added sequentially o-NO₂C₆H₄SeCN (63 mg, 0.279 mmol) and PBu₃ (56.7 mg, 70 μ L, 0.279 mmol). After 1 h, the reaction mixture was concentrated in vacuo purified by chromatography over silica gel, eluting with Hexanes then with 4% EtOAc/hexanes, to give crude selenide **SI-39** (15.2 mg) as a yellow oil which was used directly in next step without further purification.

Polyol SI-40. To a stirred solution of selenide SI-39 (15.2 mg, 0.0122 mmol) in THF/DMF/H2O (10:1:0.02, 1.6 mL/0.16 mL/ 3.2 $\mu L)$ at 0 $^{\circ}C$ was added TAS-F (16.8 mg, 0.0610 mmol). The reaction mixture was then warmed up to rt. After 2 h, the reaction mixture was concentrated in vacuo and purified by chromatography over silica gel, eluting with 10-65% EtOAc/hexanes, to give polyol SI-40 (9.1 mg, 0.0119 mmol, 86% over two steps) as yellow oils: $[\alpha]^{23}_{D} = -51.8$ (c 0.44, CHCl₂); IR (neat) 3447, 2926, 2855, 1701, 1514, 1456, 1332, 1271, 1037, 902, 756, 731 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.26 (d, J = 8.3 Hz, 1H), 7.73 (d, J = 8.0 Hz, 1H), 7.55 (t, J = 7.3 Hz, 1H), 7.36 (t, J = 7.9 Hz, 1H), 6.81 (t, J = 7.0 Hz, 1H), 5.97 (s, 1H), 5.10-5.03 (m, 1H), 5.03 (s, 1H), 4.83 (s, 1H), 4.47 (s, OH), 4.40-4.32 (m, 1H), 4.14 (dd, J = 14.2, 7.2 Hz, 1H), 3.88-3.78 (m, 1H), 3.42-3.35 (m, 1H), 3.05-2.97 (m, 1H and OH), 2.85 (d, J = 4.8 Hz, 2H), 2.50-2.38 (br, 1H), 2.30-2.20 (m, 2H), 2.07 (s, OH), 2.11-2.02 (m, 1H), 1.96-1.65 (m, 11H), 1.83 (s, 6H), 1.44 (s, 3H), 1.40–1.32 (m, 2H), 1.33 (d, J = 5.8 Hz, 3H), 1.08 (d, J = 6.3 Hz, 3H), 0.90 (d, J = 4.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.6, 167.9, 147.6, 144.0, 143.5, 141.2, 133.6, 132.3, 130.6, 128.7, 126.4, 126.0, 124.0, 115.5, 78.0, 76.1, 75.0, 69.5, 66.3, 60.7, 58.7, 46.1, 45.8, 44.8, 43.8, 40.5, 39.5, 33.5, 30.4 (2C), 28.2, 27.7, 27.1, 20.8, 19.6, 16.5, 15.2, 12.5; HRMS (ES⁺) calcd for C₃₈H₅₅NO₁₀NaSe (M + Na) 788.2889, found 788.2934.



Allylic Epoxide 121. To a stirred solution of selenide **SI-40** (2.7 mg, 0.00353 mmol) in CH₂Cl₂ (0.5 mL) at rt were added sequentially NaHCO₃ (30 mg, 0.357 mmol) and TMSO–OTMS (22.5 mg, 27 μ L, 0.126 mmol). After 1.5 h, the yellow color vanished and the reaction mixture was purified by preparative TLC, eluting with 60% EtOAc/ hexanes, to give allylic epoxide **121** (1.3 mg, 0.00231 mmol, 65%): $[\alpha]^{23}_{D} = +10.0 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 2923, 2853, 1701, 1457, 1200 (c 0.13, CHCl_3); IR (neat) 3422, 1200 (c 0.13, CHCl_3); IR (neat) 3420 (c 0.13$

1377, 1261, 1103; ¹H NMR (400 MHz, CDCl₃) δ 6.72 (t, *J* = 4.9 Hz, 1H), 6.04 (s, 1H), 5.88–5.80 (m, 1H), 5.28 (dd, *J* = 15.4, 8.4 Hz, 1H), 5.10–5.05 (m, 1H), 5.06 (s, 1H), 4.84 (s, 1H), 4.35 (dd, *J* = 4.2, 1.5 Hz, 1H), 4.30–4.20 (m, 1H), 4.13 (s, 1H), 3.79 (t, *J* = 9.8 Hz, 1H), 3.77 (d, *J* = 4.6 Hz, 1H), 3.40 (d, *J* = 10.2 Hz, 1H), 3.09 (dd, *J* = 8.5, 2.1 Hz, 1H), 2.98–2.93 (m, 2H), 2.76 (dd, *J* = 15.4, 2.4 Hz, 1H), 2.38–2.23 (m, SH), 2.35 (s, 1H), 1.98 (m, 1H), 1.95 (m, 1H), 1.88 (m, 1H), 1.85 (m, 1H), 1.84 (s, 6H), 1.78 (dd, *J* = 14.6, 4.7 Hz, 1H), 1.70 (m, 1H), 1.64 (m, 1H), 1.32 (m, 1H), 1.30 (d, *J* = 6.1 Hz, 3H), 1.13 (m, 1H), 1.03 (d, *J* = 6.6 Hz, 3H), 0.83 (d, *J* = 6.4 Hz, 3H) ¹³C NMR (100 MHz, CDCl₃) δ 212.6, 167.5, 144.6, 143.2, 140.8, 135.9, 128.9, 128.7, 124.3, 114.9, 78.3, 76.1, 75.5, 68.3, 66.8, 60.3, 59.2, 46.5, 46.1, 45.6, 39.6, 33.3, 31.2, 30.5, 29.7, 28.6, 27.1, 21.0, 19.9, 15.7, 15.4, 12.7; HRMS (ES⁺) calcd for C₃₂H₅₀O₈Na (M + Na) 585.3403, found 585.3409.

Cell Viability Assays. MTS assays were conducted for cell viability as described by the supplier (Promega).⁵⁸ Human DU145 prostate cancer, OCI-LY3 lymphoma, K562 CML, MOLT-4 ALL, Reh ALL, U266 myeloma, KG1a AML, HL60 AML, and MDA-MB-435 breast cancer cells were seeded in 96-well plates, incubated overnight at 37 °C in 5% (v/v) CO₂, and exposed to **3**, **114**, or **121** at 0.1 μ M or in a dosedependent manner for 72 h. DMSO was used as the vehicle control. Cell viability was determined by tetrazolium conversion to its formazan dye and absorbance was measured at 490 nm using an automated ELISA plate reader.

ASSOCIATED CONTENT

Supporting Information

Copies of ¹H and ¹³C spectra for all new compounds. X-ray crystallographic data (CIF) for compound **109**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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REFERENCES

 Reviews of the amphidinolides: (a) Ishibashi, M.; Kobayashi, J. Heterocycles 1997, 44, 543–572. (b) Chakraborty, T. K.; Das, S. Curr. Med. Chem.: Anti-Cancer Agents 2001, 1, 131–49. (c) Kobayashi, J.; Shimbo, K.; Kubota, T.; Tsuda, M. Pur. App. Chem. 2003, 75, 337–342. (d) Kobayashi, J.; Tsuda, M. Nat. Prod. Rep. 2004, 21, 77–93. (e) Colby, E. A.; Jamison, T. F. Org. Biomol. Chem. 2005, 3, 2675–2684. (f) Kobayashi, J.; Kubota, T. J. Nat. Prod. 2007, 70, 451–460. (g) Hiersemann, N.; Kobayashi, J. J. Antibiot. 2008, 61, 271–284. (h) Fürstner, A. Israel J. Chem. 2011, 51, 329–345.

(2) Amphidinolide A: (a) Kobayashi, J.; Ishibashi, M.; Nakamura, H.; Ohizumi, Y.; Yamasu, T.; Sasaki, T.; Hirata, Y. *Tetrahedron Lett.* **1986**, 27, 5755–5758. Amphidinolide B: (b) Ishibashi, M.; Ohizumi, Y.; Hamashima, M.; Nakamura, H.; Hirata, Y.; Sasaki, T.; Kobayashi, J. *J. Chem. Soc., Chem. Commun.* **1987**, 1127–29. (c) Bauer, I.; Maranda, L.; Shimizu, Y.; Peterson, R. W.; Cornell, L.; Steiner, J. R.; Clardy, J. *J. Am. Chem. Soc.* **1994**, *116*, 2657–58. (d) Ishibashi, M.; Ishiyama, H.; Kobayashi, J. *Tetrahedron Lett.* **1994**, *35*, 8241–42. (e) Tsuda, M.;

Kariya, Y.; Iwamoto, R.; Fukushi, E.; Kawabata, J.; Kobayashi, J. Mar. Drugs 2005, 3, 1-8. (f) Oguchi, K.; Tsuda, M.; Iwamoto, R.; Okamoto, Y.; Endo, T.; Kobayashi, J.; Ozawa, T.; Masuda, A. J. Nat. Prod. 2007, 70, 1676-1679. Amphidnolide C/F: (g) Kobayashi, J.; Ishibashi, M.; Wälchli, N. R.; Nakamura, H.; Yamasu, T.; Hirata, Y.; Sasaki, T.; Ohizumi, Y. J. Am. Chem. Soc. 1988, 110, 490-94. (h) Kubota, T.; Tsuda, M.; Kobayashi, J. Org. Lett. 2001, 3, 1363-66. (i) Kubota, T.; Sakuma, Y.; Tsuda, M.; Kobayashi, J. Mar. Drugs 2004, 2, 83-87. (j) Tsuda, M.; Kariya, Y.; Iwamoto, R.; Fukushi, E.; Kawabata, J.; Kobayashi, J. Mar. Drugs 2005, 3, 1-8. (k) Kubota, T.; Suzuki, A.; Yamada, M.; Baba, S.; Kobayashi, J. Heterocycles 2010, 82, 333-338. Amphidinolide D: (1) Kobayashi, J.; Ishibashi, M.; Nakamura, H.; Ohizumi, Y.; Yamasu, T.; Hirata, Y.; Sasaki, T.; Ohta, T.; Nozoe, S. J. Nat. Prod. 1989, 52, 1036-1041. Amphidinolide E: (m) Kobayashi, J.; Ishibashi, M.; Murayama, T.; Takamatsu, M.; Iwamura, M.; Ohizumi, Y.; Sasaki, T. J. Org. Chem. 1990, 55, 3421-3423. (n) Kubota, T.; Tsuda, M.; Kobayashi, J. J. Org. Chem. 2002, 67, 1651-1656. Amphidinolide F: (o) Kobayashi, J.; Tsuda, M.; Ishibashi, M.; Shigemori, H.; Hirota, H.; Sasaki, T. J. Antibiot. 1991, 44, 1259-1261. Amphidinolides G/H: (p) Kobayashi, J.; Shigemori, H.; Ishibashi, M.; Yamasu, T.; Hirota, H.; Sasaki, T. J. Org. Chem. 1991, 56, 5221-5224. (q) Kobayashi, J.; Shimbo, K.; Sato, M.; Shiro, M.; Tsuda, M. Org. Lett. 2000, 2, 2805-2807. (r) Kobayashi, J.; Shimbo, K.; Sato, M.; Tsuda, M. J. Org. Chem. 2002, 67, 6565-6592. Amphidinolide J: (s) Kobayashi, J.; Sato, M.; Ishibashi, M. J. Org. Chem. 1993, 58, 2645-2646. Amphidinolide K: (t) Ishibashi, M.; Sato, M.; Kobayashi, J. J. Org. Chem. 1993, 58, 6928-6929. Amphidinolide L: (u) Tsuda, M.; Sasaki, T.; Kobayashi, J. J. Org. Chem. 1994, 59, 3734-3737. Amphidinolide M: (v) Kobayashi, J.; Yamaguchi, N.; Ishibashi, M. J. Org. Chem. 1994, 59, 3698-4700. Amphidinolide N: (w) Ishibashi, M.; Yamaguchi, N.; Sasaki, T.; Kobayashi, J. J. Chem. Soc., Chem. Commun. 1994, 1455-1456. Amphidinolide O/P: (x) Ishibashi, M.; Takahashi, M.; Kobayashi, J. J. Org. Chem. 1995, 60, 6062-6066. Amphidinolide Q: (y) Kobayashi, J.; Takahashi, M.; Ishibashi, M. Tetrahedron Lett. 1996, 37, 1449-1450. (z) Takahashi, Y.; Kubota, T.; Fukushi, E.; Kawabata, J.; Kobayashi, J. Org. Lett. 2008, 10, 3709-3711. Amphidinolides R and S: (aa) Ishibashi, M.; Takahashi, M.; Kobayashi, J. Tetrahedron 1997, 53, 7827-7832. Amphidinolide T: (bb) Kobayashi, J.; Kubota, T.; Endo, T.; Tsuda, M. J. Org. Chem. 2001, 66, 134-142. (cc) Kobayashi, J.; Kubota, T.; Endo, T.; Tsuda, M. Tetrahedron 2001, 57, 6175-6179. Amphidinolide U: (dd) Tsuda, M.; Endo, T.; Kobayashi, J. Tetrahedron 1999, 55, 1465-14570. Amphidnolide W: (ee) Shimbo, K.; Tsuda, M.; Izui, N.; Kobayashi, J. J. Org. Chem. 2002, 67, 1020-1023. Amphidinolide X: (ff) Tsuda, M.; Izui, N.; Shimbo, K.; Sato, M.; Fukushi, E.; Kawabata, J.; Katsumata, K.; Horiguchi, T.; Kobayashi, J. J. Org. Chem. 2003, 68, 5339-4345. Amphidinolide Y: (gg) Tsuda, M.; Izui, N.; Shimbo, K.; Sato, M.; Fukushi, E.; Kawabata, J.; Kobayashi, J. J. Org. Chem. 2003, 68, 9109-9112.

(3) Syntheses of amphidinolide natural products (excluding amphidinolide B): (a) Williams, D.; Kissel, W. S. J. Am. Chem. Soc. 1988, 120, 11198-11199. (b) Williams, D. R.; Myers, B. J.; Mi, L. Org. Lett. 2000, 2, 945-948. (c) Williams, D. R.; Meyer, K. G. J. Am. Chem. Soc. 2001, 123, 765-766. (d) Lam, H. W.; Pattenden, G. Angew. Chem., Int. Ed. 2002, 41, 508-511. (e) Maleczka, R. E., Jr.; Terrell, L. R.; Geng, F.; Ward, J. S., III. Org. Lett. 2002, 4, 2841-2844. (f) Fürstner, A.; Aissa, C.; Riveiros, R.; Ragot, J. Angew. Chem., Int. Ed. 2002, 41, 4763-4766. (g) Trost, B. M.; Chisholm, J. D.; Wrobleski, S. J.; Jung, M. J. Am. Chem. Soc. 2002, 124, 12420-12421. (h) Aiessa, C.; Riveiros, R.; Ragot, J.; Fürstner, A. J. Am. Chem. Soc. 2003, 125, 15512-15520. (i) Ghosh, A. K.; Liu, C. J. Am. Chem. Soc. 2003, 125, 2374-2375. (j) Lepage, O.; Kattnig, E.; Fürstner, A. J. Am. Chem. Soc. 2004, 126, 15970-15971. (k) Ghosh, A. K.; Gong, G. J. Am. Chem. Soc. 2004, 126, 3704-3705. (1) Trost, B. M.; Harrington, P. E. J. Am. Chem. Soc. 2004, 126, 5028-5029. (m) Trost, B. M.; Papillion, J. P. N. J. Am. Chem. Soc. 2004, 126, 13618-13619. (n) Trost, B. M.; Wrobleski, S. T.; Chisholm, J. D.; Harrington, P. E.; Jung, M. J. Am. Chem. Soc. 2005, 127, 13589-13597. (o) Harrington, P. E.; Chisholm, J. D.; Wrobleski, S. T. J. Am. Chem. Soc. 2005, 127, 13598-13610. (p) Colby, E. A.; O'Brien, K. C.; Jamison, T. F. J. Am. Chem. Soc. 2005, 127, 4297-4307. (q) Trost, B. M.; Papillon, J.

P. N.; Nussbaumer, T. J. Am. Chem. Soc. 2005, 127, 17921-17937. (r) Va, P.; Roush, W. R. J. Am. Chem. Soc. 2006, 128, 15960-15961. (s) Kim, C. H.; An, H. J.; Shin, W. K.; Yu, W.; Woo, S. K.; Jung, S. K; Lee, E. Angew. Chem., Int. Ed. 2006, 45, 8019-8021. (t) Ghosh, A. K.; Gong, G. J. Org. Chem. 2006, 71, 1085-1093. (u) Fürstner, A.; Kattnig, E.; Lepage, O. J. Am. Chem. Soc. 2006, 128, 9194-9204. (v) Nicoloau, K. C.; Brenzovich, W. E.; Bulger, P. G.; Francis, T. M. Org. Biomol. Chem. 2006, 4, 2119-2157. (w) Nicolaou, K. C.; Bulger, P. G.; Brenzovich, W. E. Org. Biomol. Chem. 2006, 4, 2158-2183. (x) Deng, L.-S.; Huang, X.-P.; Zhao, G. J. Org. Chem. 2006, 71, 4625-4635. (y) Jin, J.; Chen, Y.; Li, Y.; Wu, J.; Dai, W.-M. Org. Lett. 2007, 9, 2585-2588. (z) Va, P.; Roush, W. R. Tetrahedron 2007, 63, 5768-5796. (aa) Fürstner, A.; Bouchez, L. C.; Funel, J.-A.; Liepins, V.; Poree, F.-H.; Gilmour, R.; Beaufils, F.; Laurich, D.; Tamiya, M. Angew. Chem., Int. Ed. 2007, 46, 9265-9270. (bb) Dai, W.-M.; Chen, Y.; Jin, J.; Wu, J.; Lou, J.; He, Q. Synlett 2008, 1737-1741. (cc) Barbazanges, M.; Meyer, C.; Cossy, J. Org. Lett. 2008, 10, 4489-4492. (dd) Kim, C. H.; An, H. J.; Shin, W. K.; Yu, W.; Woo, S. K.; Jung, S. K.; Lee, E. Chem. Asian J. 2008, 3, 1523-1534. (ee) Rodriquez-Escrich, C.; Urpi, F.; Vilarrasa, J. Org. Lett. 2008, 10, 5191-5194. (ff) Hangyou, M.; Ishiyama, H.; Takahashi, Y.; Kobayashi, J. Org. Lett. 2009, 11, 5046-5049. (gg) Ko, H. M.; Lee, C. W.; Kwon, H. K.; Chung, H. S.; Choi, S. Y.; Chung, Y. K.; Lee, E. Angew. Chem., Int. Ed. 2009, 47, 2364-2366. (hh) Fürstner, A.; Flügge, S.; Larionov, O.; Takahashi, Y.; Kubota, T.; Kobayashi, J. Chem.-Eur. J. 2009, 15, 4011-4029. (ii) Yadav, J. S.; Reddy, C. S. Org. Lett. 2009, 11, 1705–1708. (jj) Li, H.; Wu, J.; Luo, J.; Dai, W.-M. Chem.-Eur. J. 2010, 16, 11530-11534. (kk) Wu, D.; Li, H.; Jin, J.; Wu, J.; Dai, W.-M. Synlett 2011, 895-898. (ll) Sun, L.; Wu, D.; Wu, J.; Dai, W.-M. Synlett 2011, 3036-3040. (mm) Mahapatra, S.; Carter, R. G. Angew. Chem., Int. Ed. 2012, 51, 7948-7851.

(4) (a) Lu, L.; Zhang, W.; Carter, R. G. J. Am. Chem. Soc. 2008, 130, 7253–7255. (b) Lu, L.; Zhang, W.; Carter, R. G. J. Am. Chem. Soc. 2008, 130, 11834.

(5) Fürstner, A.; Bouchez, L. C.; Morency, L.; Funel, J.-A.; Liepins, V.; Poree, F.-H.; Gilmour, R.; Laurich, D.; Beaufils, F.; Tamiya, M. *Chem.*—*Eur. J.* **2009**, *15*, 3983–4010.

(6) Biology and Biosynthesis of the Amphidinolides: (a) Matsunaga, K.; Nakatani, K.; Ishibashi, J.; Kobayashi, J.; Ohizumi, Y. *Biochem. Biophys. Acta* **1999**, *1427*, 23–32. (b) Sato, M.; Shimbo, K.; Tsuday, M.; Kobayashi, J. *Tetrahedron Lett.* **2000**, *41*, 503–506. (c) Kubota, T.; Tsuda, M.; Kobayashi, J. *Tetrahedron* **2001**, *57*, 5975–5977. (d) Tsuda, M.; Kubota, T.; Sakuma, Y.; Kobayashi, J. *Chem. Pharm. Bull.* **2001**, *49*, 1366–1367. (e) Tsuda, M.; Izui, N.; Sato, M.; Kobayashi, J. *Chem. Pharm. Bull.* **2002**, *50*, 976–977. (f) Saito, S.-Y.; Feng, J.; Kira, A.; Kobayashi, J.; Ohizumi, Y. *Biocehm. Biophys. Res. Commun.* **2004**, *320*, 961–965. (g) Kubota, T.; Iinuma, Y.; Kobayashi, J. *Biolog. Pharm. Bull.* **2006**, *29*, 1314–1318. (h) Fürstner, A.; Kattnig, E.; Kelter, G.; Fiebig, H.-H. *Chem.—Eur. J.* **2009**, *15*, 4030–4043. (i) Trigili, C.; Pera, B.; Barbazanges, M.; Cossy, J.; Meyer, C.; Pineda, O.; Rodriguez-Escrich, C.; Urpi, F.; Vilarrasa, J.; Diaz, J. F.; Barasoain, I. *ChemBioChem* **2011**, *12*, 1027–1030.

(7) (a) Chakraborty, T. K.; Thippeswamy, D.; Suresh, V. R.; Jayaprakash, S. Chem. Lett. 1997, 563-64. (b) Chakraborty, T. K.; Suresh, V. R. Chem. Lett. 1997, 565-66. (c) Lee, D. H.; Lee, S.-W. Tetrahedron Lett. 1997, 38, 7909-10. (d) Ohi, K.; Shima, K.; Hamada, K.; Saito, Y.; Yamada, N.; Ohba, S.; Nishiyama, S. Bull. Chem. Soc. Jpn. 1998, 71, 2433-40. (e) Cid, M. B.; Pattenden, G. Synlett 1998, 540-542. (f) Ohi, K.; Nishiyama, S. Synlett 1999, 571-72. (g) Ohi, K.; Nishiyama, S. Synlett 1999, 573-75. (h) Eng, H. M.; Myles, D. C. Tetrahedron Lett. 1999, 40, 2275-78. (i) Eng, H. M.; Myles, D. C. Tetrahedron Lett. 1999, 40, 2279-82. (j) Chakraborty, T. K.; Thippeswamy, D. Synlett 1999, 150-152. (k) Ishiyama, H.; Takemura, T.; Tsuda, M.; Kobayashi, J. J. Chem. Soc., Perkin Trans. 1 1999, 1163-1166. (l) Lee, D.-H.; Rho, M. D. Tetrahedron Lett. 2000, 41, 2573-76. (m) Ndubaku, C. O.; Jamison, T. F. 227th ACS National Meeting, Anaheim, Mar 28-Apr 2, 2004; American Chemical Society: Washington, DC, 2004; ORGN-392. (n) Mandal, A. K.; Schneekloth, J. S., Jr.; Crews, C. M. Org. Lett. 2005, 7, 3645-3648. (o) Mandal, A. K.; Schneekloth, J. S., Jr.; Crews, C. M. Org. Lett. 2005, 7, 5347. (p) Gopalaratham, A.; Nelson, S. G. Org. Lett. 2006, 8, 7-10.

(q) Mandal, A. K.; Schneekloth, J. S., Jr.; Kuramochi, K.; Crews, C. M. *Org. Lett.* **2006**, *8*, 427–430. (r) Sidera, M.; Costa, A. M.; Vilarrasa, J. *Org. Lett.* **2011**, *13*, 4934–4937.

(8) Zhang, W.; Carter, R. G.; Yokochi, A. F. T. J. Org. Chem. 2004, 69, 2569–72.

(9) (a) Zhang, W.; Carter, R. G. Org. Lett. 2005, 7, 4209–12.
(b) Zhang, W.; Carter. R. G. 227th ACS National Meeting, Anaheim, Mar 28–Apr 2, 2004; American Chemical Society: Washington, DC, 2004; ORG-398.

(10) Zhang, W. Ph.D. Dissertation, Oregon State University, 2006.

(11) Hara, A.; Morimoto, R.; Iwasaki, Y.; Saitoh, T.; Ishikawa, Y.; Nishiyama, S. *Angew. Chem., Int. Ed.* **2012**, *51*, 9877–9880.

(12) (a) Mitsunobu, O. Synthesis **1981**, 1981, 1–28. (b) Swamy, K. C. K.; Kumar, N. N. B.; Balaraman, E.; Kumar, K. V. P. P. Chem. Rev. **2009**, 109, 2551–2651.

(13) Seebach, D.; Naef, R.; Calderari, G. *Tetrahedron* **1984**, *40*, 1313–1324.

(14) Kang, K.-T.; U, J. S.; Park, D. K.; Kim, J. G.; Kim, W. J. Bull. Korean Chem. Soc. **1995**, *16*, 464–466.

(15) (a) Ager, D. J. Synthesis **1984**, 384–398. (b) Van Staden, L. F.; Gravestock, D.; Ager, D. J. Chem. Soc. Rev. **2002**, 31, 195–200.

(16) Ehlinger, Ed.; Magnus, P. J. Am. Chem. Soc. 1980, 102, 5004-5011.

(17) Lau, P. W. K.; Chan, T. H. Tetrahedron Lett. 1978, 19, 2383–2386.

(18) Evans, D. A.; Golob, A.-M. J. Am. Chem. Soc. 1975, 97, 4765–4766.

(19) Hosomi, A.; Sakurai, H. *Tetrahedron Lett.* **1976**, 17, 1295–1298.

(20) (a) Kolb, H. C.; Andersson, P. G.; Sharpless, K. B. J. Am. Chem. Soc. **1994**, 116, 1278–91. (b) Corey, E. J.; Guzman-Perez, A.; Noe, M. C. J. Am. Chem. Soc. **1995**, 117, 10805–16.

(21) (a) Mander, L. N.; Morris, J. C. J. Org. Chem. 1997, 62, 7497–99.
(b) Allen, P. A.; Brimble, M. A.; Prabaharan, H. Synlett 1999, 295–298.

(c) Carter, R. G.; Weldon, D. J. Org. Lett. 2000, 2, 3913-16.

(22) Evans, D. A.; Gage, J. R.; Leighton, J. L.; Kim, A. S. J. Org. Chem. **1992**, *57*, 1961–1963.

(23) (a) Evans, D. A.; Dart, M. J.; Duffy, J. L.; Yang, M. G. J. Am. Chem. Soc. 1996, 118, 4322–4343. (b) Evans, D. A.; Coleman, P. J.; Coté, B. J.

Org. Chem. **1997**, *62*, 788–789. (c) Evans, D. A.; Coleman, P. J.; Cole, B. J.

J.; Connell, B. T. J. Am. Chem. Soc. 2003, 125, 10893–10898.

(24) Chakraborty, T. K.; Suresh, V. R. Tetrahedron Lett. 1998, 39, 7775–7778.

(25) Paterson, I.; Lister, M. A.; Norcross, R. D. *Tetrahedron Lett.* **1992**, 33, 1767–1770.

(26) (a) Heathcock, C. H.; White, C. T. J. Am. Chem. Soc. **1979**, 101, 7076–7077. (b) Heathcock, C. H.; White, C. T.; Morrison, J. J.; VanDerveer, D. J. Org. Chem. **1981**, 46, 1296–1309.

(27) Masamune, S.; Ali, S. A.; Snitman, D. L.; Garvey, D. S. Angew. Chem., Int. Ed. **1980**, 19, 557–558.

(28) (a) Braun, M. Angew. Chem., Int. Ed. Engl. 1987, 26, 24–37. (b) Li,
Y.; Padden-Row, M. N.; Houk, K. N. J. Org. Chem. 1990, 55, 484–493.
(c) Bernardi, A.; Capelli, A. M.; Gcnnari, C.; Goodman, J. M.; Paterson,
I. J. Org. Chem. 1990, 55, 3576–3581. (d) Paterson, I.; Goodman, J. M.;
Lister, M. A.; Schumann, R. C.; McClure, C. K.; Norcross, R. D.

Tetrahedron **1990**, *46*, *4663–4684*. (e) Gustin, D. J.; VanNieuwenhze, M. S.; Roush, W. R. *Tetrahedron Lett.* **1995**, *36*, 3443–3446.

(29) White, J. D.; Bolton, G. L. J. Am. Chem. Soc. 1990, 112, 1626–1628.

(30) (a) Trost, B. M.; Urabe, H. J. Org. Chem. 1990, 55, 3982–3984.
(b) Trost, B. M.; Rodriguez, M. S. Tetrahedron Lett. 1992, 33, 4675–4678.

(31) Evans, D. A.; Carter, P. H.; Carreira, E. M.; Prunet, J. A.; Charette, A. B.; Lautens, M. Angew. Chem., Int. Ed. **1998**, 37, 2354–2359.

(32) (a) Wallace, G. A.; Scott, R. W.; Heathcock, C. H. *J. Org. Chem.* 2000, 65, 4145–4152. (b) Denmark, S. E.; Stavenger, R. A. *J. Am. Chem. Soc.* 2000, *122*, 8837–8847. (c) Hosaka, M.; Hayakawa, H.; Miyashita, M. *J. Chem. Soc., Perkin Trans.* 1 2000, 4227–4230. (d) Crimmins, M. T.; Katz, J. D.; McAtee, L. C.; Tabet, E. A.; Kirincich, S. J. *Org. Lett.* 2001, *3*, 949–952. (e) Nakamura, S.; Inagaki, J.; Sugimoto, T.; Kudo, M.; Nakajima, M.; Hashimoto, S. Org. Lett. 2001, 3, 4075-4078. (f) Crimmins, M. T.; Katz, J. D.; Washburn, D. G.; Allwein, S. P.; McAtee, L. F. J. Am. Chem. Soc. 2002, 124, 5661-5663. (g) Nakamura, S.; Inagaki, J.; Kudo, M.; Sugimoto, T.; Obara, K.; Nakajima, M.; Hashimoto, S. Tetrahedron 2002, 58, 10353-10374. (h) Heathcock, C. H.; McLaughlin, M.; Medina, J.; Hubbs, J. L.; Wallace, G. A.; Scott, R.; Claffey, M. M.; Hayes, C. J.; Ott, G. R. J. Am. Chem. Soc. 2003, 125, 12844-12849. (i) Smith, A. B., III; Fox, R. J.; Vanecko, J. A. Org. Lett. 2005, 7, 3099-3102. (j) Paterson, I.; Coster, M. J.; Chen, D. Y.-K.; Acena, J. L.; Bach, J.; Keown, L. E.; Trieselmann, T. Org. Biomol. Chem. 2005, 3, 2420-2430. (k) Kawahara, S.; Gaunt, M. J.; Scolaro, A.; Yamanoi, S.; Ley, S. V. Synlett 2005, 2031-2034. (1) Pellicena, M.; Solsona, J. G.; Romea, P.; Urpí, F. Tetrahedron Lett. 2008, 49, 5265-5267. (m) Lorenz, M.; Kalesse, M. Org. Lett. 2008, 10, 4271-4374. (n) Paterson, I.; Mühlthau, F. A.; Cordier, C. J.; Housden, M. P.; Burton, P. M.; Loiseleur, O. Org. Lett. 2009, 11, 353-356. (o) Paterson, I.; Findlay, A. D.; Noti, C. Chem. Asian. J. 2009, 4, 594-611. (p) Lorenz, M.; Bluhm, N.; Kalesse, M. Synthesis 2009, 3061-3066. (q) Lorente, A.; Pellicena, M.; Romea, P.; Urpí, F. Tetrahedron Lett. 2010, 51, 942-945. (r) Guérinot, A.; Lepesqueux, G.; Sablé, S.; Reymond, S.; Cossy, J. J. Org. Chem. 2010, 75, 5151–5163. (s) Smith, A. B., III; Dong, S.; Fox, R. J.; Brenneman, J. B.; Vanecko, J. A.; Maegawa, T. Tetrahedron 2011, 67, 9809-9828.

(33) (a) Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. **1991**, 113, 4092–96. (b) Dale, J. A.; Mosher, H. S. J. Am. Chem. Soc. **1973**, 95, 512–19. (c) Sullivan, G. R.; Dale, J. A.; Mosher, H. S. J. Org. Chem. **1973**, 38, 2143–47.

(34) Deng, L.; Ma, Z.; Zhang, Y.; Zhao, G. Synlett 2007, 87-80.

(35) Magnus, P.; Carter, R.; Davies, M.; Elliott, J.; Pitterna, T. Tetrahedron **1996**, *52*, 6283–6306.

(36) Paterson, I.; Yeung, K. Tetrahedron Lett. 1993, 34, 5347-5350.

(37) Blanchette, M. A.; Choy, W.; Davis, T.; Essenfeld, A. P.; Masamune, S.; Roush, W. R.; Sakai, T. *Tetrahedron Lett.* **1984**, 25, 2183–2186.

(38) (a) Boulet, S. L.; Paquette, L. A. Synthesis 2002, 895–900.
(b) Lipshultz, B. H.; Hackmann, C. J. J. Org. Chem. 1994, 59, 7437–7444. (c) Zhou, X. T.; Liang, L.; Furkert, D. K.; Wells, C. E.; Carter, R. G. Angew. Chem., Int. Ed. 2006, 45, 7622–7626.

(39) (a) Maruoka, K.; Ooi, T.; Yamamoto, H. J. Am. Chem. Soc. **1989**, 111, 6431–6432. (b) Maruoka, K.; Ooi, T.; Nagahara, S.; Yamamoto, H. *Tetrahedron* **1991**, 47, 6983–6998. (c) Suda, K.; Kikkawa, T.; Nakajima, S.-I.; Takanami, T. J. Am. Chem. Soc. **2004**, 126, 9554–9555.

(40) (a) Hirao, A.; Itsuno, S.; Nakahama, S.; Yamazaki, N. J. Chem. Soc., Chem. Commun. **1981**, 315–317. (b) Corey, E. J.; Bakshi, R. K.; Shibata, S. J. Am. Chem. Soc. **1987**, 109, 5551–5553. (c) Corey, E. J.; Bakshi, K.; Shibata, S.; Chen, C.; Singh, V. K. J. Am. Chem. Soc. **1987**, 109, 7925– 7926. (d) Mathre, D. J.; Jones, T. K.; Xavier, L. C.; Blacklock, T. J.; Reamer, R. A.; Mohan, J. J.; Jones, E. T. T.; Hoogsteen, K.; Baum, M. W; Grabowski, E. J. J. J. Org. Chem. **1991**, 56, 751–762.

(41) (a) Katsuki, T.; Sharpless, K. B. J. Am. Chem. Soc. **1980**, *102*, 5974–5976. (b) Gao, Y.; Klunder, J. M.; Hanson, R. M.; Masamune, H.; Ko, S. Y.; Sharpless, K. B. J. Am. Chem. Soc. **1987**, *109*, 5765–5780. (c) Ahmed, A.; Hoegenar, E. K.; Enev, S. V.; Hanbeur, M.; Kaehlig, H.; Ohler, E.; Mulzer, J. J. Org. Chem. **2003**, *68*, 3026–3042.

(42) Grieco, P. A.; Gilman, S.; Nishizawa, M. J. Org. Chem. **1976**, 41, 1485–1486.

(43) Zhou, X.-T.; Carter, R. G. Angew. Chem., Int. Ed. 2006, 45, 1787–1790.

(44) (a) Frye, S. V.; Eliel, E. Tetrahedron Lett. 1986, 27, 3223–3226.
(b) Kahn, S. D.; Keck, G. E.; Hehre, W. J. Tetrahedron Lett. 1987, 28, 279–280.
(c) Keck, G. E.; Castellino, S. Tetrahedron Lett. 1987, 28, 281–284.
(d) Shambiayati, S.; Blake, J. F.; Wierschke, S. G.; Jorgensen, W. J.; Schreiber, S. L. J. Am. Chem. Soc. 1990, 112, 697–703.

(45) Van Draanen, N. A.; Arseniyadis, S.; Crimmins, M. T.; Heathcock, C. H. J. Org. Chem. **1991**, *56*, 2499–2506.

(46) Chen, X.-N.; Hortelano, E. R.; Eliel, E. L.; Frye, V. S. J. Am. Chem. Soc. **1992**, 114, 1778–1784.

(47) Willard, P. G.; Hintze, M. J. J. Am. Chem. Soc. 1987, 109, 5539–5541.

(48) (a) Evans, D. A.; Allison, B. D.; Yang, M. G. *Tetrahedron Lett.* **1999**, 40, 4457–4460. (b) Evans, D. A.; Halstead, D. P.; Allison, B. D. *Tetrahedron Lett.* **1999**, 40, 4461–4464.

(49) Heathcock, C. H.; Lampe, J. J. Org. Chem. **1983**, 48, 4330–4337. (50) Lurain, A. E.; Carroll, P. J.; Walsh, P. J. J. Am. Chem. Soc. **2005**, 70, 1262–1268.

(51) Seebach, D.; Naef, R.; Calderari, G. *Tetrahedron* **1984**, *40*, 1313–1324.

(52) The iodide was prepared from commercially available 2-(chloromethyl)allyltrimethylsiliane [NaI (2.5 equiv), acetone (0.5 M), 16 h, flask covered in aluminum foil, rt, 99%]. ¹H NMR (300 MHz, CDCl₃) δ 5.16 (s, 1H), 4.70 (s, 1H), 3.88 (s, 2H), 1.72 (s, 2H), 0.04 (s, 9H).

(53) AD Mix $\beta^* = (DHQD)_2PHAL (15.2 mg), K_2OsO_4 \cdot 2H_2O (2.55 mg), K_2CO_3 (293.6 mg), K_3Fe(CN)_6 (699.6 mg). Commercially available AD mix <math>\beta$ proved to be slow and inefficient.

(54) Preparation of LDA solution: To a solution of diisopropylamine (101.9 mg, 0.14 mL, 1.0 mmol) in THF (0.46 mL) at -78 °C was added *n*-BuLi (0.4 mL, 1.0 mmol, 2.5 M in THF). After 5 min, the white slurry was warmed to -10 °C and stirred for an additional 15 min.

(55) Titration of Grignard reagent: To a stirred solution of menthol (15.6 mg, 0.1 mmol) and 1,10-phenanthroline (2 mg) in THF (0.5 mL) was added prepared Grignard reagent solution until a burgundy color persisted. The concentration was calculated using the formula: [RMgX] = 0.1 mmol/volume of added RMgX in mL. For the references, see: (a) Lin, H, -S; Paquette, L. A. Synth. Commun. **1994**, *24*, 2503. (b) Watson, S. C.; Eastham, J. F. J. Organomet. Chem. **1967**, *9*, 165.

(56) Vanderwalle, M.; Van der Eychen, J.; Oppolzer, W.; Vullioud, C. *Tetrahedron* **1986**, *42*, 4035.

(57) Aitken, R. A.; Atherton, J. I. J. Chem. Soc., Perkin Trans. 1 1994, 1281.

(58) Nam, S.; Williams, A.; Vultur, A.; List, A.; Bhalla, K.; Smith, D.; Lee, F. Y.; Jove, R. *Mol. Cancer Ther.* **2007**, *6*, 1400–1405.