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CO₂-expanded liquids as solvents to enhance activity of *Pseudozyma antarctica* lipase B towards *ortho*-substituted 1-phenylethanols

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Introduction

Biocatalysts, especially lipases, have been studied extensively [1,2]. Among them, *Pseudozyma* (Candida) antarctica lipase B (CAL-B, Novozym 435[®]) is one of the most widely used and outstanding biocatalysts; CAL-B catalyzes transesterification of a wide range of secondary alcohols in organic solvents smoothly with excellent enantioselectivity [3–7]. However, CAL-B-catalyzed transesterification of ortho-substituted 1-phenylethanol analogs suffers much lower conversions than those of meta and para- substituted analogs (Supplementary information Section 2 and Table S1). Due to the limitation of the substrate scope of CAL-B, the dynamic kinetic resolution of ortho-substituted 1-phenylethanols also resulted in considerable retardations in the reactivity [7]. For example, reaction of 1-(2'-bromophenyl)ethanol took 7 days to complete whereas that of 1-phenylethanol took 3 h. The lipase-catalyzed resolution reaction became rate-limiting in the overall reactions. It was also reported that reactivities of Burkholderia (Pseudomonas) cepacia lipase for transesterification of ortho-substituted 1-phenylethanol analogs were extremely lower than those of meta- and para-substituted analogs [8].

The limitation of biocatalysts has been overcome by mutations and chemical modifications of enzymes and solvent engineering [1,9-17]. For solvent engineering, sustainable solvents should be

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ABSTRACT

Pseudozyma (*Candida*) *antarctica* lipase B (CAL-B, Novozym 435[®]) is one of the most widely used and outstanding biocatalysts. However, CAL-B-catalyzed transesterification of *ortho*-substituted 1-phenylethanol analogs suffers low conversion. In this research, the reactions were accelerated by using CO₂-expanded liquids, liquids expanded by dissolving pressurized CO₂, such as CO₂-expanded hexane or CO₂-expanded MeTHF.

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should be chosen since they have been generating a massive amount of waste and burden both economically and environmentally [18,19]. Therefore, methods for minimizing the usage of solvents and replacing traditional organic solvents bv environmentally friendly alternatives have been studied extensively [20–22]. As sustainable solvents, CO₂ related solvents such as supercritical CO₂ (scCO₂), liquid CO₂ and CO₂-expanded liquids have attracted great attentions [23-27]. Among them, CO₂expanded liquids, liquids expanded by dissolving CO₂, have an advantage over scCO₂ and liquid CO₂; CO₂-expanded liquids can be used under lower pressures with wider temperature ranges than $scCO_2$ and liquid CO_2 [23]. As a liquid to be expanded by CO₂, it is desirable to use bio-based solvents such as 2-methyltetrahydrofuran (MeTHF), which can be derived from lignocellulosic biomass and has recently gained increasing interests as a promising solvent for various synthesis applications [28–30], including biocatalysis [31].

Previously, we have reported the utilization of CO_2 -expanded bio-based liquids as effective reaction media for transesterification of alcohols catalyzed by CAL-B [27,32,33]. As shown in Fig. 1, the reaction is accelerated especially for 1-(2'-bromophenyl)ethanol using CO_2 -expanded MeTHF. However, *ortho*-substituted 1-phenylethanol analogs, except 1-(2'-bromophenyl)ethanol, have not been used as substrates although these compounds are important as chiral intermediates for pharmaceuticals [34–38]. To assess the effectiveness of CO_2 -expanded liquids for biocatalysis further, in this study, we investigated CAL-B-catalyzed transesterification of









Fig. 1. CAL-B-catalyzed transesterification of 1-phenylethanol and 1-(2'-bromophenyl)ethanol in CO₂-expanded MeTHF [27,32,33]. ^aReaction conditions: substrate (0.40 mmol), vinyl acetate (0.53 mmol), Novozym 435[®] (10 mg), MeTHF (10 mL) or CO₂-expanded MeTHF (10 mL, MeTHF concentration 10% v/v, 6.0 MPa), 20 °C, 1 h [32]. ^bReaction conditions: substrate (0.10 mmol), vinyl acetate (0.53 mmol), Novozym 435[®] (10 mg), MeTHF (10 mg), MeTHF (10 mL) or CO₂-expanded MeTHF (10 mL) or CO₂-expanded MeTHF (10 mL, MeTHF concentration 10% v/v, 6.0 MPa), 20 °C, 5 h [33].

various *ortho*-substituted 1-phenylethanol analogs **1a-7a** in CO₂-expanded MeTHF and CO₂-expanded hexane (Fig. 2). **1a-5a** were used to examine the *ortho* substituent effect, while **6a** and **7a** were selected to examine the potential of this reaction to synthesize chiral **6b** and **7b**, key intermediates of NEK2 kinase inhibitor [34,36] and c-Met/ALK inhibitor (Crizotinib), [37,38] respectively. The activities of CAL-B were found to be significantly higher in both CO₂-expanded MeTHF and CO₂-expanded hexane than in the corresponding neat liquids without expansion with CO₂.

Results and discussion

CAL-B-catalyzed transesterification of various *ortho*-substituted 1-phenylethanol analogs **1a-7a** in MeTHF and in CO₂-expanded MeTHF was examined at 50 °C. As shown in Table 1, the reaction in CO₂-expanded MeTHF showed higher conversion than those in neat MeTHF for the reaction of **1a-5a**. However, the reactions of **6a** and **7a**, to afford the pharmaceutical intermediates, hardly proceeded both in MeTHF and in CO₂-expanded MeTHF. For the reaction of **1a-5a** in CO₂-expanded MeTHF, the smaller the size of the substituents are, the better the conversions were. As listed in **Table S2**, the similar effect of size of *ortho* substituents can be seen for other reactions such as the dynamic kinetic resolution of *ortho*substituted 1-phenylethanol analogs by CAL-B and dicarbonylchlorido (pentabenzylcyclopentadienyl)ruthenium in toluene [7] and

hydrolysis of ortho-substituted 1-phenylethyl acetate analogs by Bacillus subtilis esterase (BsE) [39]. It can be also seen for other biocatalytic reactions such as oxidation of ortho-substituted benzaldehyde analogs by Geotrichum candidum aldehyde dehydrogenase [40] and bovine lens aldehyde dehydrogenase [41], and asymmetric reduction of ortho-substituted acetophenone analogs by a mutant W288A of G. candidum acetophenone reductase (GcAPRD) [15]. On the other hand, the enantioselectivities were excellent for all reactions proceeded in CO2-expanded MeTHF; ee of transesterification products of **1a-5a** was >99% (*R*) regardless of the kind of substituents. The excellent enantioselectivity of CAL-B was also reported for transesterification in CO₂-expanded MeTHF using other types of substrates such as 1-adamantylethanol, 1-(1-naphthyl)ethanol, 2-octanol, etc. [27,32,33] and for transesterification in supercritical carbon dioxide using meta- and para-substituted 1-phenylethanol analogs such as 1-(3'-trifluoromethylphenyl) ethanol and 1-(4'-bromophenyl) ethanol [42]. Therefore, the excellent enantioselectivity of CAL-B was not affected when the conversion was improved by expanding the solvent by CO₂.

Then, the kind of solvents to be expanded by CO_2 was investigated for the CAL-B-catalyzed transesterification of **3a**. Since CO_2 expanded MeTHF was reported to be best among the biobased solvents tested for the transesterification of 1-adamantylethanol by CAL-B (γ -valerolactone, diethyl carbonate, MeTHF, 2-methylfuran, *p*-cymene, (+)-limonene and (-)-limonene) [32], conventional



Fig. 2. CAL-B-catalyzed transesterification of ortho-substituted 1-phenylethanol analogs (1a-7a) in CO2-expanded liquids [34,36-38].

Table 1

Substrate			Solvent	Product	Conv. ^a (%)	<i>ee</i> _p (%)
R OH	R = F	1a	MeTHF	1b	2.0	N.d.
L ŝ			CO ₂ -expanded MeTHF		24	>99
	Cl	2a	MeTHF	2b	<1	N.d.
Į į			CO ₂ -expanded MeTHF		11	>99
\sim	Br	3a	MeTHF	3b	<1 ^b	N.d.
			CO ₂ -expanded MeTHF		5.6 ^b	>99
	CH_3	4a	MeTHF	4b	<1	N.d.
			CO ₂ -expanded MeTHF		9.1	>99
	OCH ₃	5a	MeTHF	5b	1.2	N.d.
			CO ₂ -expanded MeTHF		18	>99
	CF ₃	6a	MeTHF	6b	<1	N.d.
			CO ₂ -expanded MeTHF		<1	N.d.
CI	ОН	7a	MeTHF	7b	<1	N.d.
F	<u> </u>		CO ₂ -expanded MeTHF		<1	N.d.
	`CI					

Comparison of (CAL_R_catalyze	d transectorification of	ortho_substituted 1	 al analogs in MeTH	E and in COsper	manded MeTHE
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Reaction conditions: **1a-7a** (0.10 mmol), vinyl acetate (0.20 mmol), Novozym 435[®] (10 mg), MeTHF (10 mL) or CO₂-expanded MeTHF (10 mL, MeTHF concentration 5% v/v, 6.0 MPa), 50 °C, 0.50 h. N.d.: not determined due to low conversions observed. *ee*_p: Enantiomeric excess of the product.

^a The reactions were stopped at low conversion to show differences between the two media.

^b Similar results shown in Fig. 1 were reported using different conditions [33].

solvents were also investigated in this study. Generally, hydrophobic solvents have been reported to be suitable for lipase-catalyzed transesterification [43,44]. For example, hexane was reported to be best among the solvents tested for the transesterification of 1-phenylethanol by CAL-B (vinyl acetate, THF, diisopropyl ether, chloroform, toluene, hexane, and isooctane) [45]. Based on these reports, p-cymene, hexane, THF, and vinyl acetate were chosen for the investigation for CAL-B-catalyzed transesterification of 3a at 50 °C. As shown in Table S3, CO₂ expanded hexane gave the best result, followed by CO₂ expanded *p*-cymene. When the reactions in solvents without expansion by CO₂ were compared, hexane also gave the best result, followed by p-cymene. The preliminary experiment using MeTHF, hexane, p-cymene, diethyl carbonate, and cyclopentyl methyl ether (CPME) at 20 °C also resulted in hexane being the best. The hydrophobicity of hexane, as well as *p*-cymene, is considered to be suitable for the reaction, and CO₂ may exert the additional positive effect.

Then, a conventional excellent solvent for CAL-B-catalyzed reaction, hexane, was used instead of MeTHF for the transesterification of **1a-7a** at 50 °C. The results are shown in Table 2. The reactions in CO₂-expanded hexane gave better conversions than those in neat hexane for all substrates tested. The conversions in neat hexane and CO₂-expanded hexane were also higher than those in neat MeTHF and CO₂-expanded MeTHF (Table 1), respectively. It is probably due to the high hydrophilicity of MeTHF as reported in other lipase-catalyzed reactions [44]. For **1a-3a**, as the steric hindrance of substituents are larger, the acceleration effects were more profound. Importantly, the enantioselectivities of reactions were excellent for all of the reactions proceeded except for **6a**.

The effect of temperature on CAL-B-catalyzed transesterification of **3a** in CO₂-expanded hexane was investigated. As shown in Fig. 3, the conversion increased from 20 °C to 50 °C, and the highest conversion was observed at 50 °C (23%), which is in agreement with the reported optimum temperature of CAL-B-catalyzed

Table 2

Comparison of CAL-B-catalyzed transesterification of ortho-substituted 1-phenylethanol analogs in hexane and in CO2-expanded hexane.

Substrate			Solvent	Product	Conv. ^a (%)	<i>ee</i> _p (%)	
	R = F	1a	Hexane	1b	30	>99	
			CO ₂ -expanded hexane		46	>99	
	Cl	2a	Hexane	2b	16	>99	
			CO ₂ -expanded hexane		30	>99	
	Br	3a	Hexane	3b	9.0	>99	
			CO ₂ -expanded hexane		23	>99	
	CH ₃	4a	Hexane	4b	11	>99	
			CO ₂ -expanded hexane		27	98	
\checkmark	OCH ₃	5a	Hexane	5b	29	>99	
			CO ₂ -expanded hexane		42	>99	
	CF ₃	6a	Hexane	6b	<1	N.d.	
			CO ₂ -expanded hexane		1.2	N.d.	
	CF ₃	6a	Hexane	6b	13 ^b	58	
			CO ₂ -expanded hexane		41 ^b	58	
CI OH		7a	Hexane	7b	<1	N.d.	
			CO ₂ -expanded hexane		<1	N.d.	
		7a	Hexane	7b	<1 ^b	N.d.	
CI			CO ₂ -expanded hexane		2.0 ^b	N.d.	

Reaction conditions: **1a-7a** (0.10 mmol), vinyl acetate (0.20 mmol), Novozym 435[®] (10 mg), hexane (10 mL) or CO₂-expanded hexane (10 mL, hexane concentration 5% v/v, 6.0 MPa), 50 °C, 0.50 h. *ee*_p: Enantiomeric excess of the product.

^a The reactions were stopped at low conversion to show differences between the two media.

^b 24 h.



Fig. 3. Effect of temperature on CAL-B-catalyzed transesterification of 1-(2'bromophenyl)ethanol **3a** in CO₂-expanded hexane. Reaction conditions: **3a** (0.10 mmol), vinyl acetate (0.20 mmol), Novozym 435[®] (10 mg), CO₂-expanded hexane (10 mL, hexane concentration 5% v/v, 6.0 MPa), 0.50 h. Enantiomeric excess of the product ee_p > 99% (R) under all temperatures examined.

transesterification [46–48]. The enantioselectivities of reactions were excellent ($ee_p > 99\%$ (*R*)), regardless of the reaction temperature.

CAL-B-catalyzed transesterification of **1a-7a** was also investigated using liquid CO₂ as a solvent at 20 °C. The reaction temperature using liquid CO₂ can be set only below the critical temperature of CO₂ (31.1 °C) [24]. As shown in Table S2, the reactions in liquid CO₂ at 20 °C gave lower conversions than those in CO₂-expanded hexane at 50 °C due to the difference in reaction temperature. However, the reactions of **2a-5a** in liquid CO₂ at 20 °C gave higher conversions than those in hexane at 20 °C. Therefore, the presence of a large concentration of CO₂ in the solvent is important for the CAL-B-catalyzed transesterification.

The CAL-B activity toward *ortho*-substituted 1-phenylethanol analogs, which was low in neat organic solvents, was enhanced in both CO_2 -expanded liquids tested (MeTHF and hexane). The mechanism of the CO_2 induced acceleration can be hypothesized that CO_2 induces enhanced transport properties [25–27] and improved flexibility of enzymes [49–52]. Particularly, the solvent-exposed residues α 5, the pseudo-lid covering the entrance of the active site, has high fluctuations in CO_2 and thus allows the lipase more tolerant to sterically hindered substrates with *ortho*-substitutions [49–52]. Additionally, a plausible reason for the difference in the degree of acceleration between the two solvents, of which more drastic are found for MeTHF than for hexane, is that CO_2 increases the hydrophobicity of MeTHF, whereas neat hexane is hydrophobic, so that the hydrophobicity of hexane does not change significantly by the expansion with CO_2 .

Conclusion

CO₂-expanded liquids were proven to be promising alternative solvents for CAL-B-catalyzed transesterification of sterically hindered *ortho*-substituted 1-phenylethanol analogs. This research will lead to the further development of biocatalysis in CO₂expanded liquids using different kinds of substrates and/or enzymes in the future, especially to diminish the detrimental effect of the *ortho* substituents.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.tetlet.2020.152424.

References

- R.A. Sheldon, D. Brady, Broadening the scope of biocatalysis in sustainable organic synthesis, ChemSusChem 12 (13) (2019) 2859–2881.
- [2] J.P. Adams, M.J.B. Brown, A. Diaz-Rodriguez, R.C. Lloyd, G.D. Roiban, Biocatalysis: a pharma perspective, Adv. Synth. Catal. 361 (11) (2019) 2421– 2432.
- [3] N.L.D. Nyari, G.L. Zabot, R. Zamadei, A.R. Paluzzi, M.V. Tres, J. Zeni, L.D. Venquiaruto, R.M. Dallago, Activation of Candida Antarctica Lipase B in pressurized fluids for the synthesis of esters, J. Chem. Technol. Biotechnol. 93 (3) (2018) 897–908.
- [4] A. Park, S. Kim, J. Park, S. Joe, B. Min, J. Oh, J. Song, S. Park, S. Park, H. Lee, Structural and experimental evidence for the enantiomeric recognition toward a bulky sec-alcohol by Candida Antarctica Lipase B, ACS Catal. 6 (11) (2016) 7458–7465.
- [5] Y. L. de los Santos, Y.L. Chew-Fajardo, G. Brault, N. Doucet, Dissecting the evolvability landscape of the CalB active site toward aromatic substrates, Sci. Rep. 9 (1) (2019) 1–14.
- [6] H. Chen, X. Meng, X. Xu, W. Liu, S. Li, The molecular basis for lipase stereoselectivity, Appl. Microbiol. Biotechnol. 102 (8) (2018) 3487–3495.
- [7] M. Päiviö, D. Mavrynsky, R. Leino, L.T. Kanerva, Dynamic kinetic resolution of a wide range of secondary alcohols: cooperation of dicarbonylchlorido (pentabenzylcyclopentadienyl)ruthenium and CAL-B, Eur. J. Org. Chem. 8 (2011) 1452–1457.
- [8] K. Nakamura, M. Kawasaki, A. Ohno, Lipase-catalyzed transesterification of aryl-substituted alkanols in an organic solvent, Bull. Chem. Soc. Jpn. 69 (4) (1996) 1079–1085.
- [9] K. Yasukawa, F. Motojima, A. Ono, Y. Asano, Expansion of the substrate specificity of porcine kidney D-amino acid oxidase for S-stereoselective oxidation of 4-Cl-benzhydrylamine, ChemCatChem 10 (16) (2018) 3500–3505.
- [10] H. Yamaguchi, M. Tatsumi, K. Takahashi, U. Tagami, M. Sugiki, T. Kashiwagi, M. Kameya, S. Okazaki, T. Mizukoshi, Y. Asano, Protein engineering for improving the thermostability of tryptophan oxidase and insights from structural analysis, J. Biochem. 164 (5) (2018) 359–367.
- [11] T. Ema, H. Inoue, Chemical modification of lipase for rational enhancement of enantioselectivity, Chem. Lett. 44 (10) (2015) 1374–1376.
- [12] B.J. Jones, H.Y. Lim, J. Huang, R.J. Kazlauskas, Comparison of five protein engineering strategies for stabilizing an α/β-hydrolase, Biochemistry 56 (50) (2017) 6521–6532.
- [13] A.A. Koesoema, Y. Sugiyama, K.T. Sriwong, Z. Xu, S. Verina, D.M. Standley, M. Senda, T. Senda, T. Matsuda, Reversible control of enantioselectivity by the length of ketone substituent in biocatalytic reduction, Appl. Microbiol. Biotechnol. 103 (23–24) (2019) 9529–9541.
- [14] A.A. Koesoema, D.M. Standley, T.K. Sriwong, M. Tamura, T. Matsuda, Access to both enantiomers of substituted 2-tetralol analogs by a highly enantioselective reductase, Tetrahedron Lett. 61 (13) (2020) 151682.
- [15] A.A. Koesoema, D.M. Standley, S. Ohshima, M. Tamura, T. Matsuda, Control of enantioselectivity in the enzymatic reduction of halogenated acetophenone analogs by substituent positions and sizes, Tetrahedron Lett. 61 (18) (2020) 151820.
- [16] C. Cao, T. Matsuda, Biocatalysis in organic solvents, supercritical fluids and ionic liquids, in: Organic Synthesis Using Biocatalysis, Elsevier, 2016, pp. 67– 97.
- [17] A. Hinzmann, N. Adebar, T. Betke, M. Leppin, H. Gröger, Biotransformations in pure organic medium: organic solvent-labile enzymes in the batch and flow synthesis of nitriles, Eur. J. Org. Chem. 2019 (41) (2019) 6911–6916.
- [18] R.K. Henderson, C. Jiménez-González, D.J.C. Constable, S.R. Alston, G.G.A. Inglis, G. Fisher, J. Sherwood, S.P. Binks, A.D. Curzons, Expanding GSK's solvent selection guide – embedding sustainability into solvent selection starting at medicinal chemistry, Green Chem. 13 (4) (2011) 854.
- [19] P.T. Anastas, M.M. Kirchhoff, Origins, current status, and future challenges of green chemistry, Acc. Chem. Res. 35 (9) (2002) 686–694.
- [20] P. Tundo, P. Anastas, D.S. Black, J. Breen, T.J. Collins, S. Memoli, J. Miyamoto, M. Polyakoff, W. Tumas, Synthetic pathways and processes in green chemistry. Introductory overview, Pure Appl. Chem. 72 (7) (2000) 1207–1228.
- [21] J.M. DeSimone, Practical approaches to green solvents, Science 297 (5582) (2002) 799–803.
- [22] D. Prat, A. Wells, J. Hayler, H. Sneddon, C.R. McElroy, S. Abou-Shehada, P.J. Dunn, CHEM21 selection guide of classical- and less classical-solvents, Green Chem. 18 (1) (2016) 288–296.

M. Otsu et al.

- [23] X. Han, M. Poliakoff, Continuous reactions in supercritical carbon dioxide: problems, solutions and possible ways forward, Chem. Soc. Rev. (2012) 1428– 1436.
- [24] H.N. Hoang, T. Matsuda, Biotransformation using liquid and supercritical CO2, in: Tomoko Matsuda (Ed.), Future Directions in Biocatalysis, Elsevier, 2017, pp. 3–25.
- [25] G.R. Akien, M. Poliakoff, A critical look at reactions in class I and II gasexpanded liquids using CO2 and other gases, Green Chem. 11 (8) (2009) 1083– 1100.
- [26] P.G. Jessop, B. Subramaniam, Gas-expanded liquids, Chem. Rev. 107 (6) (2007) 2666–2694.
- [27] H.N. Hoang, K.R.A. Are, T. Matsuda, CHAPTER 7. Biocatalysis in supercritical and liquid carbon dioxide and carbon dioxide-expanded liquids, in: A.J. Hunt, T.M. Attard (Eds.), Supercritical and Other High-pressure Solvent Systems: For Extraction, Reaction and Material Processing, The Royal Society of Chemistry, London, 2018, pp. 191–220, Green Chemistry Series.
- [28] V. Pace, P. Hoyos, L. Castoldi, P. Domínguez de María, A.R. Alcántara, 2-Methyltetrahydrofuran (2-MeTHF): a biomass-derived solvent with broad application in organic chemistry, ChemSusChem 5 (8) (2012) 1369–1379.
- [29] J.H. Clark, T.J. Farmer, A.J. Hunt, J. Sherwood, Opportunities for bio-based solvents created as petrochemical and fuel products transition towards renewable resources, Int. J. Mol. Sci (2015) 17101–17159.
- [30] Y. Gu, F. Jérôme, Bio-based solvents: an emerging generation of fluids for the design of eco-efficient processes in catalysis and organic chemistry, Chem. Soc. Rev. (2013) 9550–9570.
- [31] Z.-G. Chen, D.-N. Zhang, L. Cao, Y.-B. Han, Highly efficient and regioselective acylation of pharmacologically interesting cordycepin catalyzed by lipase in the eco-friendly solvent 2-methyltetrahydrofuran, Bioresour. Technol. 133 (2013) 82–86.
- [32] H.N. Hoang, Y. Nagashima, S. Mori, H. Kagechika, T. Matsuda, CO2-expanded bio-based liquids as novel solvents for enantioselective biocatalysis, Tetrahedron 73 (20) (2017) 2984–2989.
- [33] H.N. Hoang, E. Granero-Fernandez, S. Yamada, S. Mori, H. Kagechika, Y. Medina-Gonzalez, T. Matsuda, Modulating biocatalytic activity toward sterically bulky substrates in CO2-Expanded biobased liquids by tuning the physicochemical properties, ACS Sustain. Chem. Eng. 5 (11) (2017) 11051–11059.
- [34] B.H. Hoff, E. Sundby, Preparation of pharmaceutical important fluorinated 1arylethanols using isolated enzymes, Bioorg. Chem. 51 (2013) 31–47.
- [35] M.L. Contente, I. Serra, L. Palazzolo, C. Parravicini, E. Gianazza, I. Eberini, A. Pinto, B. Guidi, F. Molinari, D. Romano, Enzymatic reduction of acetophenone derivatives with a benzil reductase from Pichia Glucozyma (KRED1-Pglu): electronic and steric effects on activity and enantioselectivity, Org. Biomol. Chem. 14 (13) (2016) 3404–3408.
- [36] S. Solanki, P. Innocenti, C. Mas-Droux, K. Boxall, C. Barillari, R.L.M. Van Montfort, G.W. Aherne, R. Bayliss, S. Hoelder, Benzimidazole inhibitors induce a DFG-out conformation of never in mitosis gene a-related Kinase 2 (Nek2) without binding to the back pocket and reveal a nonlinear structure-activity relationship, J. Med. Chem. 54 (6) (2011) 1626–1639.
- [37] H. Nishii, T. Chiba, K. Morikami, T.A. Fukami, H. Sakamoto, K. Ko, H. Koyano, Discovery of 6-benzyloxyquinolines as c-Met selective kinase inhibitors, Bioorg. Med. Chem. Lett. 20 (4) (2010) 1405–1409.

- [38] P.D. de Koning, D. McAndrew, R. Moore, I.B. Moses, D.C. Boyles, K. Kissick, C.L. Stanchina, T. Cuthbertson, A. Kamatani, L. Rahman, et al., Fit-for-purpose development of the enabling route to Crizotinib (PF-02341066), Org. Process Res. Dev. 15 (5) (2011) 1018–1026.
- [39] G. Zheng, X. Liu, Z. Zhang, P. Tian, G. Lin, J. Xu, Separation of enantiopure Msubstituted 1-phenylethanols in high space-time yield using bacillus subtilis esterase, RSC Adv. 3 (43) (2013) 20446.
- [40] T. Hoshino, E. Yamabe, M.A. Hawari, M. Tamura, S. Kanamaru, K. Yoshida, A.A. Koesoema, T. Matsuda, Oxidation of aromatic and aliphatic aldehydes to carboxylic acids by *Geotrichum candidum* aldehyde dehydrogenase, Tetrahedron 76 (33) (2020) 131387.
- [41] T. Knaus, V. Tseliou, L.D. Humphreys, N.S. Scrutton, F.G. Mutti, A biocatalytic method for the chemoselective aerobic oxidation of aldehydes to carboxylic acids, Green Chem. 20 (17) (2018) 3931–3943.
- [42] T. Matsuda, K. Tsuji, T. Kamitanaka, T. Harada, K. Nakamura, T. Ikariya, Rate enhancement of lipase-catalyzed reaction in supercritical carbon dioxide, Chem. Lett. 34 (8) (2005) 1102–1103.
- [43] L.F. García-Alles, V. Gotor, Lipase-catalyzed transesterification in organic media: solvent effects on equilibrium and individual rate constants, Biotechnol. Bioeng. 59 (6) (1998) 684–694.
- [44] A. lemhoff, J. Sherwood, C.R. McElroy, A.J. Hunt, Towards sustainable kinetic resolution, a combination of bio-catalysis, flow chemistry and bio-based solvents, Green Chem. 20 (1) (2018) 136–140.
- [45] H.N. Hoang, T. Matsuda, Liquid carbon dioxide as an effective solvent for immobilized Candida Antarctica Lipase B catalyzed transesterification, Tetrahedron Lett. 56 (4) (2015) 639–641.
- [46] N.M. Hadzir, M. Basri, M.B.A. Rahman, C.N.A. Razak, R.N.Z.A. Rahman, A.B. Salleh, Enzymatic alcoholysis of triolein to produce wax ester, J. Chem. Technol. Biotechnol. 76 (5) (2001) 511–515.
- [47] G.D. Yadav, P.S. Lathi, Kinetics and mechanism of synthesis of butyl isobutyrate over immobilised lipases, Biochem. Eng. J. 16 (3) (2003) 245– 252.
- [48] B.M. Lue, S. Karboune, F.K. Yeboah, S. Kermasha, Lipase-catalyzed esterification of cinnamic acid and oleyl alcohol in organic solvent media, J. Chem. Technol. Biotechnol. 80 (4) (2005) 462–468.
- [49] M.J. Hernáiz, A.R. Alcántara, J.I. García, J.V. Sinisterra, Applied biotransformations in green solvents, Chem. - A Eur. J. 16 (31) (2010) 9422– 9437.
- [50] H. Monhemi, M.R. Housaindokht, How enzymes can remain active and stable in a compressed gas? New insights into the conformational stability of Candida Antarctica Lipase B in near-critical propane, J. Supercrit. Fluids 72 (2012) 161–167.
- [51] H. Monhemi, M.R. Housaindokht, Chemical modification of biocatalyst for function in supercritical CO2. In silico redesign of stable lipase, J. Supercrit. Fluids 117 (2016) 147–163.
- [52] H. Monhemi, M.R. Housaindokht, The molecular mechanism of protein denaturation in supercritical CO₂: the role of exposed lysine residues is explored, J. Supercrit. Fluids 147 (2019) 222–230.