



Subscriber access provided by University of Glasgow Library

# Biomimetic Oxidative Deamination Catalysis via ortho-Naphthoquinone-Catalyzed Aerobic Oxidation Strategy

Gangadhararao Golime, Ganganna Bogonda, Hun Young Kim, and Kyungsoo Oh ACS Catal., Just Accepted Manuscript • DOI: 10.1021/acscatal.8b00992 • Publication Date (Web): 27 Apr 2018 Downloaded from http://pubs.acs.org on April 27, 2018

# **Just Accepted**

Letter

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

# Biomimetic Oxidative Deamination Catalysis via *ortho*-Naphthoquinone-Catalyzed Aerobic Oxidation Strategy

Gangadhararao Golime, Ganganna Bogonda, Hun Young Kim\* and Kyungsoo Oh\*

Center for Metareceptome Research, College of Pharmacy, Chung-Ang University, 84 Heukseok-ro, Dongjak, Seoul 06974, Republic of Korea

KEYWORDS: aerobic oxidation, deamination, organocatalysis, ortho-naphthoquinone, prototropic rearrangement

**ABSTRACT:** An *ortho*-naphthoquinone-catalyzed *oxidative* deamination reaction has been developed where the molecular oxygen and water serve as the sole oxidant and nucleophile. The current *aerobic* deamination reaction proceeds via the ketimine formation between *ortho*-naphthoquinones and amines followed by the prototropic rearrangement and hydrolysis by water, representing a biomimetic *oxidative* deamination of amine species in the human body by liver and kidneys. The compatibility of *ortho*-naphthoquinone organocatalysts with molecular oxygen and water opens up a new biomimetic catalyst system that can function as versatile deaminases for a variety of amine-containing molecules such as amino acids and DNA nuclear bases.

Amine dehydrogenases are periplasmic quinoproteins involving the bacterial growth on primary amines as a source of carbon and nitrogen.<sup>1</sup> Aromatic amine dehydrogenase (AADH) is specific for primary amines while methylamine dehydrogenase (MADH) displays its specificity for smaller amines such as methylamine and ethylamine.<sup>2</sup> These enzymes are tryptophan tryptophyquinone (TTQ)-dependant and catalyze the deamination of primary amines to aldehydes and ammonia (Scheme 1A). Another class of enzymes that perform the oxidation of primary amines to aldehydes is periplasmic as well as human copper amine oxidases (CuAOs) that utilize a redox cofactor, topaquinone (TPQ), and molecular oxygen in a variety of biological processes such as the oxidation of neurotransmitters in animals, thus leading to the formation of ammonia and hydrogen peroxide as byproducts.<sup>3</sup> Recently, tremendous research efforts have been made to the development of CuAOs-like biomimetic catalysts that could bring the significant substrate scope that natural enzymes can not offer (Scheme 1B).<sup>4</sup> Thus, the synthetic *ortho*-quinone catalysts enabled the aerobic oxidation of  $\alpha$ -branched primary benzylic amines<sup>5</sup> and secondary amines.<sup>6</sup> However, the current CuAOslike biomimetic catalyst systems convert primary and secondary amines to imines and ketimines, respectively. A subsequent hydrolysis of imines and ketimines will be required to obtain the true enzymatic oxidation products, aldehydes or ketones, and also lead to the recovery of amine starting materials up to 50% yields. This means that the CuAOs-like biomimetic catalysts scavenge the 50% of amine substrates, only leading to the maximum yield of 50% of aldehyde/ketone products. Previously, the Corey group demonstrated the transamination reaction protocol that combined the amines (R<sub>2</sub>CHNH<sub>2</sub>) and mesitylglyoxals (Ar-CH<sub>2</sub>CHO) to form Schiff bases which underwent prototropic rearrangement to isomeric Schiff bases. The subsequent acid hydrolysis of the newly formed Schiff bases led to the desired ketones.<sup>7</sup> Recently, the group of Srogl reported the aerobic oxidation of ascorbic acid to dehydroascorbic acid that condensed with amines to give

aldehydes and ketones upon acid hydrolysis.<sup>8</sup> At the present time there exist no practical catalytic systems that perform the direct aerobic oxidation of amines to aldehydes and ketones without using the separate hydrolysis step and the stoichiometric amount of oxidants.<sup>9</sup> As a consequence, the development of one-pot catalytic aerobic oxidation of primary amines to aldehydes and ketones, *the true enzyme-mimetic catalyst system*, remains significant unmet need in aerobic oxidation protocols.<sup>10</sup> Herein, we report the development of an unprecedented promiscuous enzyme-mimetic organocatalyst system that skillfully promotes the ADH-like oxidation of primary amines, the CuAOs-like oxidation of  $\alpha$ -branched primary amines, and the deaminase-like reactivity for DNA nuclear bases.

SCHEMES 1. (A) Amine Dehydrogenases and Cupper Amine Oxidases. (B) Proposed Enzyme-mimetic Catalytic System

60



With an aim of developing an enzyme-mimetic catalyst system that can utilize water (like ADHs) and molecular oxygen (like CuAOs), we envisioned the ortho-naphthoquinonecatalyzed direct aerobic oxidation of amines to aldehydes and ketones. Previously, we had shown that the aerobic oxidations of amines were possible via the modular catalyst systems of ortho-naphthoquinone with Cu(OAc)<sub>2</sub> for homo-coupled imines, with TFA for cross-coupled imines, and with Ag<sub>2</sub>CO<sub>3</sub> for the oxidation of secondary amines.<sup>11</sup> Encouraged by the fact that the ortho-naphthoquinone catalyst displays robust stability with acidic and basic conditions, we investigated the compatibility of the catalyst system with water (Table 1) as the initial condensation reaction between catalysts and amines produce water molecules during the catalytic cycle. Thus, benzylamine 1a was treated with a catalytic amount of o-NQ1 (15 mol%) in CH<sub>3</sub>CN/H<sub>2</sub>O at ambient temperature. The <sup>1</sup>H NMR analysis of the reaction mixture revealed the formation of imine intermediate 3a in > 90 % yields within 48 h. The imine intermediate was stable under the reaction conditions with intact o-NQ1. However, upon increasing the reaction temperature to 80 °C, the imine 3a was hydrolyzed by the residual water in CH<sub>3</sub>CN to give benzaldehyde 2a and benzylamine 1a where the catalyst o-NQ1 quickly converted benzylamine 1a to benzaldehyde 2a (entry 1). While the crude reaction mixture showed the *exclusive* formation of 2a, the isolated yield was 67 % due to the volatile nature of benzaldehyde 2a. The use of other benzylamines with higher molecular masses led to the improved isolated yields of aldehydes 2b-2c in 79-84 % yields (entry 2-3). After gaining the valuable reaction profile, we turned attention to *sec*-primary amine, our 1phenylethylamine, as substrate for the direct oxidation of amines. At ambient temperature, the use of 5 mol % o-NQ1 only led to the formation of ketimine 3d in 32 % yield alongside 4 % yield of ketone 2d (entry 4). Increasing the catalyst loading to 10 % mol improved the yields of ketone 2d and ketimine 3d to 25 % and 46 %, respectively (entry 5). Screening of other ortho-naphthoquinone catalysts (o-NQ) was performed (entry 6-14), but only o-NQ3 possessed the similar catalytic activity as o-NQ1 (entry 7). The solvent screening also confirmed that CH<sub>3</sub>CN was the optimal reaction solvent (entry 15-18). To further optimize the catalytic activity of o**NQ1**, the reaction temperature was investigated (entry 19-20), where the optimal reaction temperature of 80 °C provided the ketone **2d** and the ketimine **3d** in 72 % yield and 7 % yield, respectively. The reaction conversion was clearly catalyst loading-dependant, thus the optimal loading of *o*-**NQ1** was set as 15 mol % (entry 21). The control experiments confirmed that the molecular oxygen in air was enough to induce the direct oxidation of amine without an oxygen balloon (entry 22) and the reaction required molecular oxygen to render the catalyst turnover (entry 23). Interestingly, the Stahl's catalyst system,<sup>12</sup> *tert*-butyl-2-yydoxybenzoquinone (TBHBQ), under our optimized conditions provided the ketone **2d** and the ketimine **3d** in 56 % yield and 14 % yield, respectively.

 Table 1. Optimization of the Direct Aerobic Oxidation of

 Amines to Aldehydes/Ketones<sup>a</sup>

R I NH2	air			
<b>1a</b> , R, R <sup>1</sup> = H		2a-d	3a-d	
1b, R = 4-OMe, R <sup>1</sup> =	= H			
1c, R =3,4-Dioxolan	yl, R <sup>1</sup> = H			
1d, R = H, R <sup>1</sup> = Me				
	<i>o</i> -NQ1, R <sup>1</sup> , R <sup>3</sup> =	H, $R^2 = Ph$	<i>o</i> -NQ2, R <sup>1</sup> , R <sup>3</sup> = H, R <sup>2</sup> = 4-OMePr	1

R <sup>1</sup> − NQ3, R <sup>1</sup> , R <sup>3</sup> = H, R <sup>2</sup> = 4-FPh         − NQ4, R <sup>1</sup> , R <sup>2</sup> , R <sup>1</sup> = H           − NQ5, R <sup>1</sup> = H, R <sup>2</sup> = Ph, R <sup>3</sup> = OMe         − NQ6, R <sup>1</sup> = Me, R <sup>2</sup> = Ph, R <sup>3</sup> = H           − NQ7, R <sup>1</sup> , R <sup>2</sup> = Ph, R <sup>3</sup> = H         − NQ8, R <sup>1</sup> , R <sup>2</sup> = pPr, R <sup>3</sup> = H						
	$R^2$	<i>o</i> -NQ9, R <sup>1</sup> , R <sup>3</sup>	= H, R <sup>2</sup> = C <sub>5</sub> H <sub>1</sub>	o-NQ1	<b>0</b> , R <sup>1</sup> , R <sup>3</sup> = H, F	<sup>2</sup> = OMe
Entry	1	Cat (mol %)	Solvent	T (°C)	Yield <b>2</b> (%) <sup>b</sup>	Yield <b>3</b> (%) <sup>b</sup>
1	1a	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2a</b> , 67 <sup>c</sup>	<b>3a</b> , 0
2	1b	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2b</b> , 79 <sup>°</sup>	<b>3b</b> , 0
3	1c	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2c</b> , 84 <sup>c</sup>	<b>3c</b> , 0
4	1d	<b>o-NQ1</b> (5)	CH₃CN	23	<b>2d</b> , 4	<b>3d</b> , 32
5	1d	<b>o-NQ1</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 25	<b>3d</b> , 46
6	1d	<b>o-NQ2</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 6	<b>3d</b> , 36
7	1d	<b>o-NQ3</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 20	<b>3d</b> , 44
8	1d	<b>o-NQ4</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 2	<b>3d</b> , 11
9	1d	<b>o-NQ5</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 1	<b>3d</b> , 40
10	1d	<b>o-NQ6</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 1	<b>3d</b> , 21
11	1d	<b>o-NQ7</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 2	<b>3d</b> , 8
12	1d	<b>o-NQ8</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 2	<b>3d</b> , 17
13	1d	<b>o-NQ9</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 5	<b>3d</b> , 37
14	1d	<b>o-NQ10</b> (10)	CH <sub>3</sub> CN	23	<b>2d</b> , 1	<b>3d</b> , 21
15	1d	<b>o-NQ1</b> (10)	THF	23	<b>2d</b> , 6	<b>3d</b> , 44
16	1d	<b>o-NQ1</b> (10)	PhCH₃	23	<b>2d</b> , 12	<b>3d</b> , 54
17	1d	<b>o-NQ1</b> (10)	$CH_2CI_2$	23	<b>2d</b> , 14	<b>3d</b> , 45
18	1d	<b>o-NQ1</b> (10)	EtOH	23	<b>2d</b> , 5	<b>3d</b> , 36
19	1d	<b>o-NQ1</b> (10)	CH₃CN	60	<b>2d</b> , 52	<b>3d</b> , 21
20	1d	<b>o-NQ1</b> (10)	CH₃CN	80	<b>2d</b> , 72	<b>3d</b> , 7
21	1d	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2d</b> , 100	<b>3d</b> , 0
22 <sup><i>a</i></sup>	1d	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2d</b> , 98	<b>3d</b> , 2
23 <sup>e</sup>	1d	<b>o-NQ1</b> (15)	CH₃CN	80	<b>2d</b> , 18	<b>3d</b> , 4
24	1d	<b>TBHBQ</b> (15)	CH₃CN	80	<b>2d</b> , 56	<b>3d</b> , 14

<sup>a</sup>Reaction conditions: **1** (0.4 mmol), **o-NQ** catalyst (mol %) in solvent (0.4 M) for 36-48 h. <sup>b</sup>Unless stated otherwise, yield was determined by <sup>1</sup>H NMR. <sup>c</sup>Isolated yields using a mixture of CH<sub>3</sub>CN:H<sub>2</sub>O (3:1). <sup>d</sup>Reaction using oxygen balloon. <sup>e</sup>Reaction under argon. TBHBQ = *tert*-Butyl-2-hydroxybenzoquinone.

With the optimized reaction conditions in place, the substrate scope of the *o*-NQ1-catalyzed direct oxidation of amines to ketones was studied using a variety of *sec*-primary amines with different electronic and steric characters (Scheme 2). 1-

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15 16 17

18

19 20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

1

2

3

4

5

6

7

8

9

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57 58 59

60

Phenylethyl derivatives with different electronic character were all suitable substrates, providing excellent isolated yields of ketones (2d-2o) in 79-97 % yield. The sole formation of products was observed, and the analytically pure products were obtained after filtering through a short pad of silica to remove the catalyst o-NQ1. Upon scaling up the reaction, the recovery of catalyst o-NQ1 was attempted. However, the recovered *o*-NQ1 remained < 5 % yields due to the formation of several polar by-products including diacid derivatives.<sup>13</sup> Also, the work-up procedure required phase separation, leading to somewhat diminished isolated yields of ketone products. The 10 slight loss of isolated products was mainly due to the volatile 11 nature of ketones. 1-(Naphthalen-1-yl)ethylamine 1p required 20 mol % catalyst loading to provide the corresponding ketone 12 **2p** in 74 % yield. The use of 1-(thiophen-2-yl)ethylamine **1q** 13 also provide an excellent yield of ketone 2q in 94 %. Another 14 sec-primary amine, diphenylmethanamine 1r, also underwent 15 the desired oxidation to give ketone 2r in 65 % yield. The 16 current catalyst system failed to show the catalytic activity for 17 1-phenylpropan-1-amine 1s, where the ketone 2s was isolated 18 in 20 % yield. Interestingly, a cyclic sec-primary amine 1t was 19 directly oxidized under the reaction conditions to give ketone 20 2t in 60 % yield. Based on the structural feature of 1s and 1t, it 21 could be concluded that the flexible conformation of the ethyl 22 group in 1s rendered the steric repulsion that prevented the catalyst turn-over activity. The aryl group in sec-primary 23 amines turned out to be not required since 1-(adamantan-1-24 yl)ethylamine 1u smoothly underwent the oxidation to give 25 the ketone 2u in 56 % yield. 26

SCHEMES 2. Scope of sec-Primary Amines in the o-NQ1-**Catalyzed Direct Oxidation of Amines to Ketones** 



<sup>a</sup>20 mol % catalyst.

The fact that the catalyst *o*-NO1 enables the direct oxidation of sec-primary amines to ketones through the formation of ketimine intermediate led us to re-optimize the reaction conditions to selectively obtain ketimines 3 (see Supporting Information for detail). Scheme 3 lists the scope of o-NQ1catalyzed aerobic oxidation of sec-primary amines to ketimines. Thus, with the cooperative action of *o*-NQ1 and acetic acid the oxidation reaction was interrupted at the ketimine stage thanks to the dehydrating effect of 3 Å molecular sieves.<sup>14</sup> The optimized reaction condition was widely applicable to sec-primary amines, leading to excellent isolated yields of ketimines (3d-3r). However, the corresponding ketimines of 1-(naphthalen-1-yl)ethylamine 1p and 1-(adamantan-1-yl)ethylamine 1u could not be identified, where the ketones 2p and 2u were the only observed products. This result may signify the steric requirement for the ketimine formation. 1-Phenylpropan-1-amine 1s and cyclic sec-primary amine 1t were not oxidized, instead the catalyst o-NQ1 was captured by the substrates as benzooxazine derivatives (3s-**3t**).<sup>15</sup> Nevertheless, the results in Scheme 3 represent the prac-

tical aerobic oxidation protocol for ketimine synthesis from the sec-primary amines.

SCHEMES 3. Scope of o-NQ1-Catalyzed Aerobic Oxidation of Amines to Ketimines



<sup>a</sup>NMR yield of 96 %. <sup>b</sup>15 mol% catalyst loading at 80 °C. <sup>c</sup>Reaction at 60 °C. d100 mol % catalyst loading.

The o-NQ1-catalyzed direct oxidations of amines to ketones mimic amine dehydrogenases as well as amine oxidases that use water molecule and molecular oxygen, respectively, for their catalytic activities. While the employment of alkenyl and alkynyl amines, 4a and 4b, was partially successful in the direct deamination protocol, due to the nucleophilic character of water and by-product, H<sub>2</sub>O<sub>2</sub> at elevated reaction temperature the formation of ketimines 7a and 7b instead of the corresponding ketone products was observed in 79-88% yields (Scheme 4). Also, the reaction under an oxygen balloon helped the catalyst turn-over at 23 °C, possibly facilitating the redox process between ortho-naphthoquinone and 2aminonaphthalen-1-ol.<sup>16</sup> In addition, the decarboxylative deamination of amino acids revealed the importance of a neighboring aryl group for the catalyst turn-over number, where phenylglycine 8 was converted to benzaldehyde in 92% yield.<sup>17</sup> To explore the possibility of *o*-NQ1 as deaminasemimetics<sup>18</sup> that removes amino groups from DNA nuclear bases, we investigated the conversion of cytosines 9 to uracils 10.<sup>19</sup> After some experimentation, we found that a stoichiometric amount of o-NQ1 could exert the formation of uracils 10 in 63-71% yields.

SCHEMES 4. o-NQ1-Mediated Direct Deamination of Various Amines (Yields using internal standard)



A proposed reaction mechanism for the direct aerobic deamination is depicted in Scheme 5 based on the HRMS-ESI analysis of the reaction mixture. The ortho-naphthoquinone catalyst, o-NQ1, condenses with amine 1 to give water and o-NQketimine. While there are two ketone moieties that amine 1

can condense with, the crystallographic evidence of benzooxazine derivative **3t** clearly demonstrates the site of attack being the 2-carbon position of o-NQ1. The prototropic rearrangement then occurs to give Naphthol-ketimine. Our mass spectra analysis revealed the peak corresponding to either o-NQketimine or Naphthol-ketimine as [M+H]<sup>+</sup>. Naphtholaminal should be derived from the reaction of the Naphthol**ketimine** with another molecule of amine **1**. Alternatively, the Naphthol-ketimine can isomerize to Naphthol-enamine in such cases the catalyst arrest happens to give benzooxazine derivatives such as 3s and 3t. The Naphthol-aminal undergoes the equilibrium-driven dissociation of ketimine 3 to give Naphthol-amine that on exposure to air spontaneously oxidizes to regenerate catalytically active species to re-enter the catalytic cycle.<sup>11</sup> The formations of Naphthol-amine as well as the reduced form of o-NQ1, Catechol, were also established from the MS analysis of the reaction mixture. Ketimine 3 is hydrolyzed to give ketone 2, and amine 1 re-enters the catalytic cycle. From the MS analysis of the reaction mixture we also identified the formation of unsubstituted Ketimine, possibly from the reaction between the ketone 2 and NH<sub>3</sub> byproduct. Also, we observed the formation of Catecholproduct complex, representing the resting state of Catechol intermediate (see Supporting Information for HRMS-ESI data).

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58 59

60

SCHEMES 5. A Proposed Catalytic Cycle for the Aerobic Deamination Based on HRMS-ESI Analysis



In summary, we have developed a novel *aerobic* deamination reaction to directly access to ketones and ketimines from  $\alpha$ -branched primary amines. The direct formation of ketones from amines has been, for the first time, achieved in good to excellent yields thanks to the mild organocatalyst, *ortho*naphthoquinone, with a distinctive amine activation mode. Another key feature of the deamination catalyst is its applicability to a diverse array of amine-containing molecules, where the unprecedented deamination activity for amino acids and DNA nuclear bases has been demonstrated with the specific substrate dependency. Further extension of the aerobic deamination reactions is currently underway, and our results will be reported in due course.

# ASSOCIATED CONTENT

## **Supporting Information**

The Supporting Information is available free of charge on the ACS Publications website.

Experimental procedures and characterization data for all new compounds (PDF)

# **AUTHOR INFORMATION**

#### **Corresponding Authors**

\*E-mail: <u>kyungsoooh@cau.ac.kr</u>. \*E-mail: hunykim@cau.ac.kr

### Notes

The authors declare no competing financial interest.

### ACKNOWLEDGMENT

This research was supported by the National Research Foundation of Korea (NRF) grants funded by the Korean government (MSIP) (NRF-2015R1A5A1008958 and NRF-2015R1C1A2A01053504).

### REFERENCES

(1) For reviews, see: (a) *Principles and Application of Quinoproteins*; Davidson. V. L., Ed.; Marcel Dekker, Inc., New York, 1993. (b) Davidson, V. L. Electron Transfer in Quinoproteins. *Arch. Biochem. Biophys.* **2004**, *428*, 32-40.

(2) (a) Eady, R. R.; Large, P. J. Microbial Oxidation of Amines. Spectral and Kinetic Properties of the Primary Amine Dehydrogenase of *Pseudomonas* AM 1. *Biochem. J.* **1971**, *123*, 757-768. (b) de Beer, R.; Duine, J. A.; Frank Jzn, J.; Large, P. J. The Prosthetic Group of Methylamine Dehydrogenase from *Pseudomonas* AM1: Evidence for a Quinone Structure. *Biochim. Biophys. Acta* **1980**, *622*, 370-374. (c) Hyun, Y. L.; Davidson, V. L. Mechanistic Studies of Aromatic Amine Dehydrogenase, a Tryptophan Tryptophylquinone Enzyme. *Biochemistry* **1995**, *34*, 816-823. (d) Roujeinikova, A.; Hothi, P.; Masgrau, L.; Sutcliffe, M. J.; Scrutton, N. S.; Leys, D. New Insights into the Reductive Half-reaction Mechanism of Aromatic Amine Dehydrogenase Revealed by Reaction with Carbinolamine Substrates. *J. Biol. Chem.* **2007**, *282*, 23766-23777.

(3) For reviews, see: (a) Mure, M.; Mills, S. A.; Klinman, J. P. Catalytic Mechanism of the Topa Quinone Containing Copper Amine Oxidases. *Biochemistry* 2002, *41*, 9269-9278. (b) Mure, M. Tyrosine-Derived Quinone Cofactors. *Acc. Chem. Res.* 2004, *37*, 131-139. (c) Klinman, J. P.; Bonnot, F. Intrigues and Intricacies of the Biosynthetic Pathways for the Enzymatic Quinocofactors: PQQ, TTQ, CTQ, TPQ, and LTQ.*Chem. Rev.* 2014, *114*, 4343-4365.

(4) For recent reviews, see: (a) Chen, B.; Wang, L.; Gao, S. Recent Advances in Aerobic Oxidation of Alcohols and Amines to Imines. *ACS Catal.* **2015**, *5*, 5851-5876. (b) Wendlandt, A. E.; Stahl, S. S. Quinone-Catalyzed Selective Oxidation of Organic Molecules. *Angew. Chem. Int. Ed.* **2015**, *54*, 14638-14658. (c) Largeron M. Aerobic Catalytic Systems Inspired by Copper Amine Oxidases: Recent Developments and Synthetic Applications. *Org. Biomol. Chem.* **2017**, *15*, 4722-4730.

(5) Qin, Y.; Zhang, L.; Lv, J.; Luo, S.; Cheng J.-P. Bioinspired Organocatalytic Aerobic C–H Oxidation of Amines with an *ortho*-Quinone Catalyst. *Org. Lett.* **2015**, *17*, 1469-1472.

(6) For examples, see: (a) Yuan, H.; Yoo, W.-J.; Miyamura, H.; Kobayashi, S. Discovery of a Metalloenzyme-like Cooperative Catalytic System of Metal Nanoclusters and Catechol Derivatives for the Aerobic Oxidation of Amines. J. Am. Chem. Soc. 2012, 134, 13970-13973. (b) Jawale, D. V.; Gravel, E.; Shah, N.; Dauvois, V.; Li, H.; Namboothiri, I. N. N.; Doris, E. Cooperative Dehydrogenation of N-Heterocycles Using a Carbon Nanotube–Rhodium Nanohybrid. Chem. – Eur. J. 2015, 21, 7039-7042. (c) Wendlandt, A. E.; Stahl, S. S. Bioinspired Aerobic Oxidation of Secondary Amines and Nitrogen Heterocycles with a Bifunctional Quinone Catalyst. J. Am. Chem.

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

Soc. 2014, 136, 506-512. (d) Wendlandt, A. E.; Stahl, S. S. Modular o-Quinone Catalyst System for Dehydrogenation of Tetrahydroquinolines under Ambient Conditions. J. Am. Chem. Soc. 2014, 136, 11910-11913.

(7) (a) Corey, E. J.; Achiwa, K. Oxidation of Primary Amines to Ketones. J. Am. Chem. Soc. **1969**, *91*, 1429-1432. (b) Buckley, T. F.; Rapoport, H. Mild and Simple Biomimetic Conversion of Amines to Carbonyl Compounds. J. Am. Chem. Soc. **1982**, *104*, 4446-4450.

(8) Srogl, J.; Voltrova, S. Copper/Ascorbic Acid Dyad as a Catalytic System for Selective Aerobic Oxidation of Amines. *Org. Lett.* **2009**, *11*, 843-845.

(9) (a) Matsuo, J.; Kawana, A.; Fukuda, Y.; Mukaiyama, T. Oxidative Deamination of Various Primary Amines to the Corresponding Carbonyl Compounds by Using *N-tert*-Butylphenylsulfinimidoyl Chloride. *Chem. Lett.* 2001, 712-713. (b) Nicolaou, K. C.; Mathison, C. J. N.; Montagnon, T. o-Iodoxybenzoic Acid (IBX) as a Viable Reagent in the Manipulation of Nitrogen- and Sulfur-Containing Substrates: Scope, Generality, and Mechanism of IBX-Mediated Amine Oxidations and Dithiane Deprotections. J. Am. Chem. Soc. 2004, 126, 5192-5201. (c) Hamamoto, H.; Suzuki, Y.; Takahashi, H.; Ikegami, S. Direct Transformation of Benzilic Amines to Carbonyls Using Polyacrylamide-bound Tungstate under Phase-Transfer Catalysis Conditions. *Tetrahedron Lett.* 2007, 48, 4239-4242. (d) Galletti, P.; Funiciello, F.; Soldati, R.; Giacomini, D. Selective Oxidation of Amine to Aldehydes or Imines using Laccase-Mediated Bio-Oxidation. *Adv. Synth. Catal.* 2015, 357, 1840-1848.

(10) For the earlier attempts with low yields of 10-40 %, see: (a) Yoneda, F.; Sakuma, Y.; Kadokawa, Y. Koshiro, A. Oxidation of Amines to Carbonyl Compounds by Pyrimido[4,5-b]quinoline-2,4-(3H,10H)-dione (5-Deazaflavin). Chem. Lett. 1979, 8, 1467-1468. (b) Yoneda, F.; Kakagawa, K. Autorecylcling in the Oxidation of Alcohols and Amines by 1,6-Dimethylpyrimido-[4,5-c]pyridazine-5,7(1H,6H)-dione (4-Deazatoxoflavins). J. Chem. Soc., Chem. Commun. 1980, 878-879. (c) Yoneda, F.; Nakagawa, K.; Noguchi, M.; Higuchi, M. Syntheses of 1, 6-Dimethylpyrimido [4,5-c] pyridazine-5, 7(1H,6H)-diones (4-Deazatoxoflavins) and Their Use in the Autorecycling Oxidation of Alcohols and Amines. Chem. Pharm. Bull. 1981, 29, 379-385. (d) Nagamatsu, T.; Hashiguchi, Y.; Sakuma, Y.; Yoneda, F. Autorecycling Oxidation of Amines to Carbonyl Compounds Catalyzed by 3,4-Disubstituted 4-Deazatoxoflavin Derivatives. Chem. Lett. 1982, 11, 1309-1312. (d) Ohshiro, Y.; Itoh, S.; Kurokawa, K.; Kato, J.; Hirao, T.; Agawa, T. Micelle Enhanced Oxidation of Amines by Coenzyme PQQ. Tetrahedron Lett. 1983, 24, 3465-3468

(11) Goriya, Y.; Kim, H. Y.; Oh, K. *o*-Naphthoquinone-Catalyzed Aerobic Oxidation of Amines to (Ket)imines: A Modular Catalyst Approach. *Org. Lett.* **2016**, *18*, 5174-5177.

(12) A single example of *sec*-primary amine oxidation by TBHBQ to imine in 69% yield was described, see: Wendlandt, A. E.; Stahl, S. S. Chemoselective Organocatalytic Aerobic Oxidation of Primary Amines to Secondary Imines. *Org. Lett.* **2012**, *14*, 2850-2853.

(13) After the reaction, the formation of diacid derivatives, possibly formed by the H<sub>2</sub>O<sub>2</sub>-promoted cleavage of **o**-NQ1, was confirmed from the MS analysis. For the formation of diacids from the oxidation of *ortho*-quinones, see: (a) Speier, G.; Tyeklar, Z. Kinetics and Mechanism of the Oxidation of 3,5-Di-*t*-butyl-*o*-benzoquinone with Hydrogen Peroxide in Aqueous Methanol Solution. *J. Chem. Soc., Perkin Trans 2*, **1981**, 1176-1179. (b) Sawaki, Y.; Foote, C. S. Mecha-

nism of C-C Cleavage of Cyclic 1,2-Diketones with Alkaline Hydrogen Peroxide. The Acyclic Mechanism and Its Application to the Basic Autooxidation of Pyrogallol. J. Am. Chem. Soc. **1983**, 105, 5035-5040.

(14) For selected examples, see: (a) Wanner, M. J.; van der Haas, R. N. S.; de Cuba, K. R.; van Maarseveen, J. H.; Hiemstra, H. Catalytic Asymmetric Pictet-Spengler Reactions via Sulfenyliminium Ions. Angew. Chem. Int. Ed. 2007, 46, 7485-7487. (b) Lou, S.; Moquist, P. N.; Schaus, S. E. Asymmetric Allylboration of Acyl Imines Catalyzed by Chiral Diols. J. Am. Chem. Soc. 2007, 129, 15398-15404. (c) Itoh, J.; Fuchibe, K.; Akiyama, T. Chiral Phosphoric Acid Catalyzed Enantioselective Friedel-Crafts Alkylation of Indoles with Nitroalkenes: Cooperative Effect of 3 Å Molecular Sieves. Angew. Chem. Int. Ed. 2008, 47, 4016-4018. (d) Li, N.; Song, J.; Tu, X.-F.; Liu, B.; Chen, X.-H.; Gong, L.-Z. Organocatalytic Asymmetric Intramolecular [3+2] Cycloaddition: A Straightforward Approach to Access Multiply Substituted Hexahydrochromeno[4,3-b]pyrrolidine Derivatives in High Optical Purity. Org. Biom. Chem. 2010, 8, 2016-2019. (e) Takizawa, S.; Kiriyama, K.; Ikeki, K.; Sasai, H. A Bifunctional Spiro-type Organocatalyst with High Enantiocontrol: Application to the Aza-Morita-Baylis-Hillman Reactions. Chem. Commun. 2011, 47, 9227-9229.

(15) CCDC 1826024 contains the supplementary crystallographic data for **3t**. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data request/cif.

(16) For the aerobic oxidation of naphthalene-1,2-diols, see: Kim, H. Y.; Takizawa, S.; Oh, K. Copper-catalyzed Divergent Oxidative Pathways of 2-Naphthol Derivatives: *ortho*-Naphthoquinone versus 2-BINOLs. *Org. Biomol. Chem.* **2016**, *14*, 7191-7196.

(17) The use of **o-NQ1** for phenylalanine and leucine did not show the catalytic activity, only leading to the corresponding aldehydes in 10-20% yields. For the pioneering deamination attempts of carboxylic acids, see: a) Itoh, S.; Kato, N.; Ohshiro, Y.; Agawa, T. Oxidative Decarboxylation of  $\alpha$ -Amino Acids with Coenzyme PQQ. *Tetrahedron Lett.* **1984**, *25*, 4753-4756. (b) Mure, M.; Suzuki, A.; Itoh, S.; Ohshiro, Y. Oxidative C–C Fission (Dealdolation) of  $\beta$ -Hydroxy Amino Acids by Coenzyme PQQ. *J. Chem. Soc., Chem. Commun.* **1990**, 1608-1612.

(18) For the cytosine deaminase activity, see: (a) Kream, J.; Chargaff, E. On the Cytosine Deaminase of Yeast. J. Am. Chem. Soc. **1952**, 74, 5157-5160. (b) Cohen, S. S.; Barner, H. D. The Conversion of 5-Methyldeoxycytidine to Thymidine *in vitro* and *in vivo*. J. Biol. Chem. **1957**, 226, 631-642. (c) Ipata, P. L.; Marmocchi, F.; Magni, G.; Felicioli, R.; Polidoro, G. Baker's Yeast Cytosine Deaminase. Enzymic Properties and Allosteric Inhibition by Nucleosides and Nucleotides. Biochemistry, **1971**, 10, 4270-4276. (d) Hitchcock, D. S.; Fedorov, A. A.; Fedrov, E. V.; Almo, S. C.; Raushel, F. M. Discovery of a Bacterial 5-Methylcytosine Deaminase. Biochemistry, **2014**, 53, 7426-7435.

(19) The chemical conversion of cytosine to uracil requires nitrous acid followed by strong basic conditions, see: Swigor, J. E.; Pittman, K. A. Synthesis of 1-(2-Deoxy-2-fluoro-β-D-arabinofuranosyl)-5-iodo [2-14C]uracil. *J. Label. Compd. Radiopharm*, **1985**, *22*, 931-937.

59 60

