

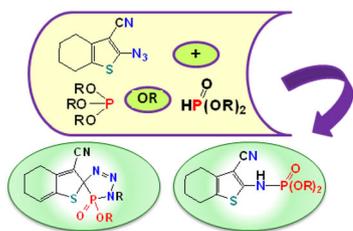
Spiro- and substituted tetrahydrobenzo[*b*]thiophene-triazaphospholes and phosphoramidates as potent antineoplastic agents: synthesis, biological evaluation, and SAR studies

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Abstract New and efficient conjugate addition reaction of trimethyl, triethyl, and triisopropyl phosphites with 2-azido-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile was developed and furnished spiro-triazaphosphole-oxide derivatives in $\approx 70\%$ yield. Contrary to these results, linear substituted phosphoramidates were obtained from the reaction of the azide with dimethyl, diethyl, and diisopropyl phosphites. *N*-Alkylamino- or 2-aminotetrahydrobenzo[*b*]thiophene-3-carbonitrile was also isolated in $\sim 10\%$ yield from the previous two reactions. In the context of this work, hexaalkylphosphorus triamides were caused to react with the same substrate afforded the corresponding phosphoric triamides only in the presence of a protonating agent (dil. alcohol). The three reactions proceeded smoothly, cleanly, and were completed within ~ 6 h at r.t. Pharmacological evaluation results of antibreast, anticolon, and antiprostata carcinoma cell lines properties for the products were discussed in terms of structure–activity relationship to define a chromophore for lead compounds.

Graphical abstract



Keywords Trialkyl phosphites · Dialkyl phosphites · Azides · Triazaphospholes · Phosphoramidates · Anticancer agents

Introduction

The search for biologically promising new chemical entities against the deadly cancer disease remains an important theme in drug discovery [1]. The preliminary cancer treatment options are usually a combination of surgery, radio-, and chemotherapy. The latter one is considered one of the effective approaches in suppressing tumor growth and eradication of tumors. However, many patients, who undergoing chemotherapy, suffer from associated serious side effects emphasizing the need to new anticancer agents with improved efficacy and reduced side effects are still needed. Generally, cancers of the breast, lung, colon, and prostate are the most frequent types of cancer worldwide [1–3]. This paper along with our recent efforts focuses on developing, identifying, and optimizing new entities of anticancer properties [4–10].

Organophosphonates, over the last three decades, have become notably recognized for their pharmacological antiinflammatory [11, 12] and anticancer activity [13, 14], especially when associated with various heterocycles [15]. One of the most important of these bioactive heterocycles is the triazoles, which have not been widely reported in the literature [16, 17]. Also, phosphoramidates in particular constitute an important class of compounds that attracts medicinal chemists because of their wide use in drug development [18–20].

Based on these considerations and with the aim of synthesizing new phosphorus esters containing one, two or

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three amino functions directly linked to the phosphorus atom, we describe herein the reactions between 2-azido-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**1**) and the phosphorus reagents trialkyl phosphites (TAPs), dialkyl phosphites (DAPs), and hexaalkylphosphorus triamides. Pharmacological evaluation results of antibreast, anticolon, and antiprostata carcinoma cell lines properties for the products were also discussed in terms of structure–activity relationship (SAR) in an attempt to define a chromophore for lead compounds.

Results and discussion

Chemistry

The azide **1** readily reacted with trimethyl, triethyl, and triisopropyl phosphites (**2a–2c**) in absolute tetrahydrofuran (THF) under ambient conditions in 4–6 h to give spiro-triazaphosphole-oxides **4a–4c** in good yields. Alkylated 2-aminotetrahydrobenzo[*b*]thiophene-3-carbonitriles **5a–5c** were also isolated ($\approx 10\%$ yield) from the reactions.

Compounds **4a–4c** could be formed in a one-step mechanism [1 + 4] cycloaddition to give initially the intermediates **3** [21]. Further extrusion of the appropriate alcohol moiety, and an alkyl group shift led to the formation of the triazaphosphole-oxide products **4a–4c** [21, 22]. The latter step was provoked by the formation of the highly

stable P=O bond [23]. Similar triazaphosphole ring was previously obtained from the reaction of trialkyl phosphites with 3- ω -azidoacetyl coumarin [24]. Furthermore, 4*H*-isooxazol-4-ylphosphonate was formed via [1 + 4] cycloaddition mechanism from the reaction of triketoin-dan-2-oxime with phosphorus reagents [25]. In support of the formation of the intermediates **3**, the ^{31}P NMR spectrum was recorded in an early stage of the reaction for the crude mixture of trimethyl phosphite and azide **1**. The spectrum (after 2 h) revealed two big peaks at $\delta = 141.3$ ppm $\text{P}(\text{OMe})_3$, and at 11.7 ppm that corresponds to the phosphazole product **4a** whereas only a minor peak was displayed at -9.6 ppm assigned to the intermediate **3**, which indicated the short life of this structure. Unfortunately, all trials to obtain any of **4a–4c** in proper crystallized form for running single crystal crystallography failed. On the other hand, the amines **5a–5c** arose from slight collapse of the intermediates **3** (due to unfortuitous moisture), resulting in its denitrogenation and the loss of phosphorus moiety (Scheme 1). ^{31}P NMR spectra of the crude mixtures of the reaction of $(\text{MeO})_3\text{P}$ and azide **1** after 2h (a), and at the end of the reaction (b) was displayed in Fig. 1.

Compounds **4a–4c** showed ^{31}P NMR chemical shifts around 12 ppm confirming the presence of N–P–C linkage in an azaphosphole nucleus [23, 24]. In their EI mass spectra, **4a–4c** showed the molecular ion peaks in addition to the expected fragmentations of the corresponding

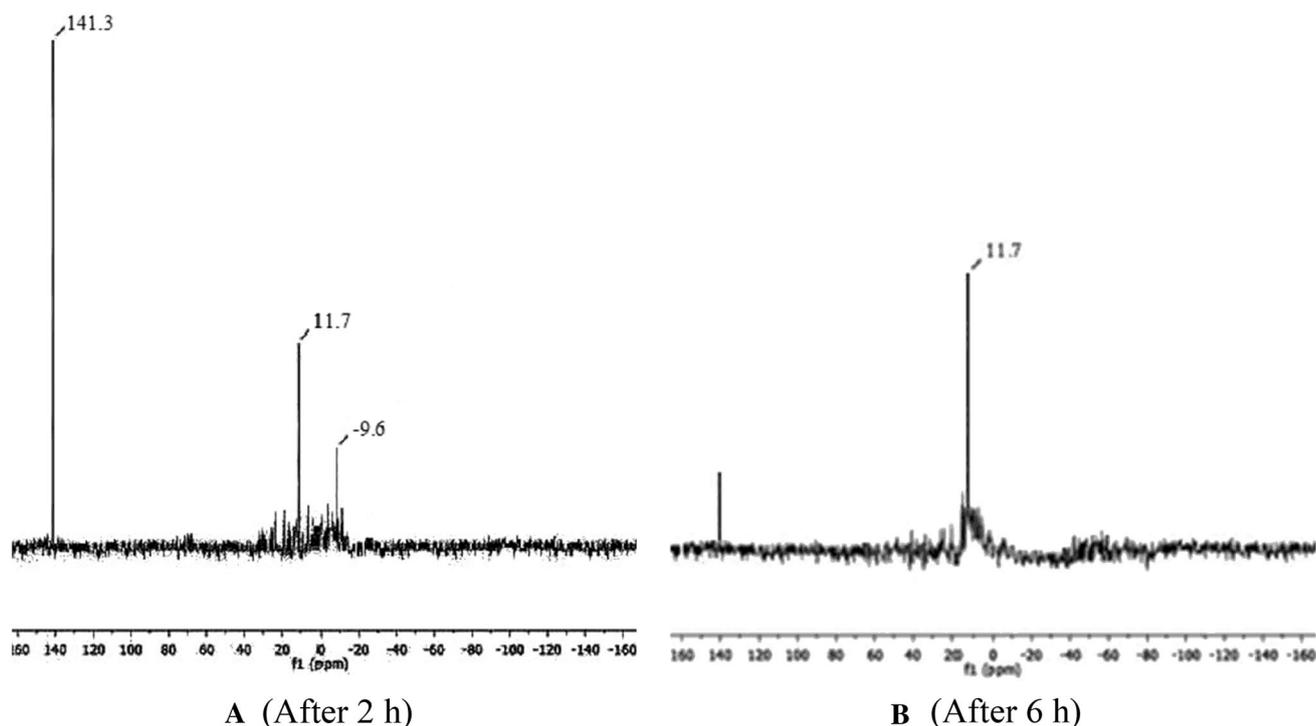
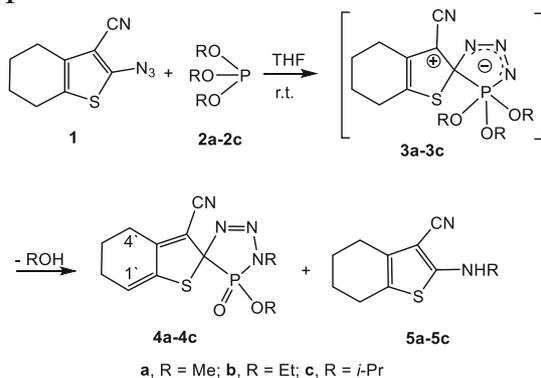


Fig. 1 ^{31}P NMR spectra of the crude mixtures of the reaction of $(\text{MeO})_3\text{P}$ and azide **1** after 2h (a), and at the end of the reaction (b)

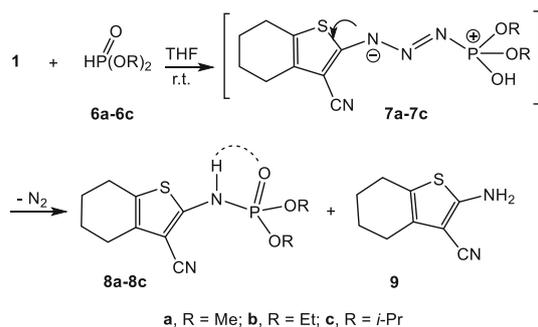
Scheme 1



compound. The IR spectrum of **4b** showed the presence of absorption bands at 2211 (CN), 1243 (P=O, free), and at 1029 (P–O–C) cm^{-1} . The strong presence of azide frequency in the IR spectrum of **1** at 2090 cm^{-1} was absent in the IR spectrum of **4a-4c**. In the ¹H NMR spectrum of **4b**, the two ethyl–methyl groups were observed as a multiplet within the range 1.16–1.25 ppm (6H, Me.C–N and MeC.OP), while the 1-H proton of the trihydrobenzene nucleus was displayed as a triplet (³*J*_{HH} = 6.6 Hz) at 6.93 ppm. A doublet of quartet (2H, ³*J*_{HH} = 6.7 Hz, ³*J*_{PH} = 8.3 Hz) was revealed at 3.79 ppm due to CH₂ protons linked to 3-N in the triazaphosphole ring. A quintet (2 H, ³*J*_{HH} = 5.6, ³*J*_{PH} = 8.9 Hz) at 4.02 ppm due to OCH₂ protons attached to the phosphorus atom. Moreover, the spiro-C was located as a doublet (¹*J*_{PC} = 133.6 Hz) at δ = 110.3 ppm in the ¹³C NMR spectrum of **4b**.

The reaction of dialkylphosphonates, dimethyl, diethyl, and diisopropyl phosphites (**6a-6c**) with the azide **1**, however, did not proceed in the same way, but gave the respective phosphoramidates **8a-8c** (~72 % yield) along with the known [26] 2-amino-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**9**, 10 % yield; Scheme 2). The reaction sequence firstly included the reaction of the azide with phosphites to give phosphorimidates **7a-7c**, which are analogous to the iminophosphoranes formed during the

Scheme 2

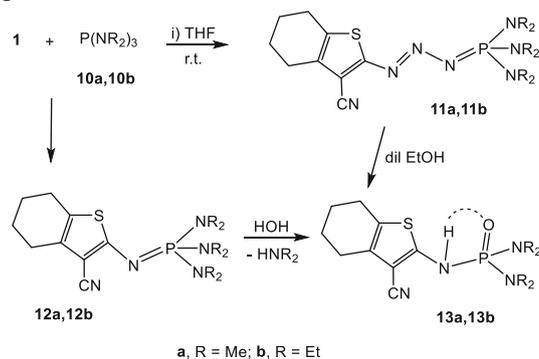


Staudinger reaction of azides with phosphines [27, 28]. Phosphorimidates **7** underwent rearrangement to their corresponding phosphoramidates **8** via a 1,2-proton shift by Lewis acid activation, whereas the amine **9** was formed through the Staudinger reduction [29, 30] (Scheme 2).

Structural reasons for **8** are: (a) elementary analyses and molecular weight determination (MS) confirmed the assigned structure; (b) the IR spectra (KBr) of **8a-8c** revealed the presence of NH and CN absorption bands around 3322 and 2198 cm^{-1} ; (c) The ¹H NMR spectrum of **8a** (CDCl₃) showed a signal at 11.35 ppm (NH) and a doublet (³*J*_{PH} = 11.5 Hz) at 3.79 ppm due to the two methoxyl groups attached to the phosphorus atom; and (d) ³¹P NMR signals of **8a-8c** were found at \approx 18 ppm.

Next, the azide **1** was allowed to react also with hexaalkyl-phosphorus triamides **10a** and **10b** in THF at 20 °C. The resulting crystalline 1:1 adducts (\approx 90 % yield) were formulated as phosphoranylidene-triazenes **11a**, **11b** [31, 32]. The ³¹P NMR signals of **11a**, **11b** were observed at \approx 40 ppm. Correct elemental analyses and expected ¹H, ¹³C NMR data also confirmed the assigned structure **11** (Scheme 3). Compounds **11a** and **11b** were water sensitive, quite stable only for few days in a desiccator. When a protonating agent (2 cm^3 dil. EtOH) was present in the reaction medium, the reaction markedly accelerated leading to the formation of the phosphoric triamides **13a** and **13b** (\approx 82 % yield). Furthermore, when compounds **11** were warmed in dil. alcohol, **13a** and **13b** were again obtained. According to the Staudinger mechanism, the first step is the nucleophile attack of the trivalent phosphorus reagent **10** on the terminal nitrogen of the azide **1** that results in the formation of the phosphazide **11** [29, 30]. Compound **11** is stable under drastic conditions due to the presence of the strong donor, hexaalkylphosphorane triamides moiety [33]. Quenching **11** with a molecule of H₂O afforded **13** via the intermediate **12** with concomitant loss of dialkylamine molecule (Scheme 3). The ³¹P NMR spectrum of these compounds showed signals around δ = 34 ppm. The ¹H NMR spectrum of **13a** revealed a

Scheme 3



doublet centered at 2.45 ppm ($^3J_{\text{PH}} = 9.4$ Hz) due to 12 H of the two magnetically equivalent dimethylamino groups.

Pharmacology

Evaluation of anticancer activity on the synthesized stable phosphorus compounds **4a–4c**, **8a–8c**, **13a**, and **13b** was performed at the Cancer Research Institute (CRI). First, all these compounds were evaluated in primary anticancer assay at 10^{-4} M concentration against MCV7 (breast), COLO205 (colon), and PC-3 (prostate cancer) cell lines (Table 1).

For CRI criteria, compounds that reduce the growth of any one of the cell lines to approximately 32 % or less are passed on for evaluation in a full panel of cell lines over a 3-long dose range. Triazaphospholes **4a**, **4b** and phosphoramidates **8a**, **8b** that met these criteria were evaluated for their anticancer activity following the known in vitro disease-oriented antitumor screening program, which is based on the use of a multiple panels of 12 human tumor cell lines [34]. Each compound is tested as a minimum of three concentrations at 10-fold dilution against every cell line in the panel. A 48 h continuous drug exposure protocol is used and sulforhodamine B (SRB) protein assay is used to estimate cell viability or growth [35, 36]. The response parameters GI_{50} , TGI, and LC_{50} (Table 2) refer to the drug concentration that produce 50 % inhibition, total growth inhibition, and 50 % cytotoxicity, and are expressed in 10^{-5} molar concentration. In the table, we report only the activity of those compounds, which exhibited GI_{50} , TGI, and LC_{50} less than 10×10^{-5} M. The GI_{50} reported data in Table 2 indicated that nominated compounds **4a**, **4b**, **8a**, and **8b** showed a good level of cytostatic activity at 10^{-5} M and in some cases at 10^{-6} M concentration against all subpanel cell lines. Compound **4a** displayed a significant growth inhibitory activity on the 12 cell lines showing GI_{50} values between 6.35 and 28.56×10^{-6} M, and total growth inhibitory activity at 2.789 – 8.888×10^{-5} M. **4a** also showed cytotoxic activity against all cell lines (4.411 – 8.564×10^{-6} M concentrations) with particular selectivity against breast and prostate cancer cell lines. Compound **4b** indicated more or less close activity to **4a**

against all tested cell lines. On the other hand, phosphoramidates **8a** and **8b** showed excellent response to the three parameters GI_{50} , TGI, and LC_{50} of the 5 colon cell lines. As such, **8b** produced GI_{50} between 6.84 and 21.36×10^{-6} M concentration, TGI between 3.542 and 5.942×10^{-5} M, and LC_{50} value between 5.084 and 6.306×10^{-5} M. For comparison, the standard drug, 5-fluorouracil showed GI_{50} between 11.36 and 41.63×10^{-6} M concentration, TGI between 3.368 and 6.651×10^{-5} M, and LC_{50} value between 3.894 and 8.894×10^{-5} M concentrations.

Considering the structure–activity relationship (SAR), as shown in Tables 1 and 2, the highest protection was observed for **4a**, followed by **4b** while **8a** and **8b** displayed less activity. On the other hand, compounds **4c**, **8c**, **13a**, and **13b** revealed only a weak antitumor activity toward all tested cell lines. It seems that the phosphonate group is essential for the biological activity. As far as cyclic substituted pyrazol-based phosphonate is concerned, it is obvious that the moieties attached to phosphorus determine the potency. As such, cyclic triazaphospholes (**4a**, **4b**) are more favorable for antitumor activity compared to open-chain phosphoramidates (**8a** and **8b**). Furthermore, the introduction of a long chain (e.g., isopropyl) or replacing the alkoxy group (Me or Et) with the amino group at the phosphor atom was totally unfavorable and dramatically decreased the antitumor potencies of compounds (**4c**, **8c**; **13a**, **13b**).

Conclusion

In conclusion, we have developed a benign convenient and rapid procedure for the synthesis of two types of phosphor families: spiro-triazaphosphole-oxides and phosphoramidate derivatives in significant yields. The products were evaluated for their in vitro antitumor activities against 12 carcinoma cell lines represent (breast, colon, and prostate cancer diseases). Four compounds out of 8 phosphor ester derivatives exhibited micromolar remarkable inhibition against several cancer cell lines in comparison to 5-fluorouracil.

Experimental section

Melting points were determined with open capillary tube on an Electrothermal (variable heater) melting point apparatus and were corrected. IR spectra were recorded on a JASCO FT-IR 6100 using KBr disc (JASCO, Japan). NMR spectra were measured with a JEOL E.C.A-500 MHz (^{13}C : 125.4 MHz, ^1H : 500.7 MHz, ^{31}P : 200.7 MHz) spectrometer (JEOL, Japan). ^{31}P NMR spectra were recorded with H_3PO_4 (85 %) as external reference. ^1H and ^{13}C NMR spectra were recorded with trimethylsilane as internal standard in CDCl_3 . Chemical

Table 1 Antiproliferative activity of **4a–4c**, **8a–8c**, **13a**, and **13b** at 10^{-4} M concentration expressed in growth percentage

Compounds	MCF7	COLO205	PC-3
4a	10	18	4
4b	35	42	24
4c	78	89	84
8a	22	33	14
8b	56	64	32
8c	87	101	73
13a	72	62	67
13b	73	101	77

Table 2 GI₅₀, TGI, and LC₅₀ values of compounds **4a**, **4b** and **8a**, **8b**

Panel/cell lines Type of test	4a			4b			8a			8b			5-Fluorouracil			
	GI ₅₀	TGI	LC ₅₀													
Breast cancer																
MCF7	0.635	4.662	5.084	0.503	4.732	3.190	0.947	8.852	6.143	0.764	6.864	–	2.284	4.831	7.521	
MDA-MB-231/ATCC	1.445	3.025	5.284	1.457	3.186	4.780	2.205	4.946	6.985	1.843	7.893	6.874	2.635	6.572	3.894	
MDA-MB-435	0.828	2.789	8.076	0.954	3.433	8.063	1.766	5.780	8.864	0.366	7.075	–	3.022	4.073	5.776	
NCI/ADR-RES	1.409	4.062	6.053	1.273	4.689	6.423	1.684	6.250	8.974	2.065	8.443	8.531	4.163	6.651	8.894	
HS578T	1.924	5.864	4.762	1.746	6.786	4.628	1.336	6.876	6.740	0.541	–	8.468	2.065	4.101	6.184	
Colon cancer																
COLO205	1.843	8.748	7.962	1.780	8.042	8.053	0.865	6.947	5.234	0.945	3.542	6.306	1.168	3.479	6.032	
HCC-2998	1.278	8.432	6.895	2.942	8.353	8.756	0.761	4.037	5.846	0.684	4.314	6.143	1.434	4.673	5.074	
HCT-15	2.633	8.888	8.564	3.653	6.406	8.684	1.776	3.621	6.346	2.136	5.942	5.084	1.776	4.055	8.063	
HCT-116	2.062	5.562	7.053	2.064	4.985	–	0.764	6.875	5.647	0.857	4.634	5.173	2.148	3.435	4.079	
SW-620	2.856	5.883	6.941	2.868	8.043	–	0.982	4.043	7.023	0.906	4.959	5.430	2.982	3.368	7.653	
Prostate cancer																
PC-3	1.243	3.356	4.411	0.997	4.636	4.650	1.053	6.620	5.878	0.788	2.865	–	1.439	4.658	6.008	
DU-145	1.053	4.325	4.664	2.843	3.513	5.844	1.448	5.974	5.036	0.816	3.906	–	1.136	4.502	6.016	

(–) means values $>10 \times 10^{-5}$

shifts (δ) are given in ppm. The mass spectra were performed at 70 eV on an MS-50 spectrometer provided with a data system spectrometer (Kratos, UK). Elemental analyses (C/H/N/S) were carried out at the Microanalysis Laboratory, Cairo University, Cairo, Egypt, using elementary Analysen-systeme GmbH-Vario EL III Element Analyzer, Germany. The values obtained are favorably agreed with the calculated ones (± 1). The appropriate precautions in handling moisture-sensitive compounds were observed and solvents were dried by standard techniques. TLC: Merck 0.2 mm silica gel 60 F154 anal aluminum plates. Column chromatography (CC): silica gel (Kieselgel 60 mesh, particle size 0.2–0.5 mm; E. Merck, Darmstadt). The substrate 2-azido-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**1**) was prepared according to the reported method [37].

Preparation of compounds **4a–4c** and **5a–5c** by reaction of azide **1** with trialkyl phosphites **2a–2c**

A mixture of 0.8 g of the azide **1** (3.9 mmol) and 4.1 mmol trimethyl, triethyl, or triisopropyl phosphite (**2a–2c**) in 15 cm³ THF was stirred at r.t. for ≈ 4 –6 h (TLC). Excess of the volatile materials was removed under vacuum, and the resulting residue was chromatographed on silica gel. Elution with *n*-hexane/CHCl₃ (9:1 v/v) produced alkylated amines **5a–5c**. Elution with *n*-hexane/CHCl₃ (7:3 v/v) afforded triazaphospholes **4a–4c**.

2-(Methylamino)-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**5a**)

Obtained as orange substance; yield: 766 mg (10.4 %). Compound **5a** is commercially available and described in

[38], but no physical data have been published. m.p.: 73 °C (from pentane); IR (KBr): $\bar{\nu} = 3334$ (NH), 2222 (CN) cm⁻¹; ¹H NMR (500.7 MHz, CDCl₃): $\delta = 2.68$ (s, 3H, MeN), 1.82, 2.79 (2 m, 2 \times 4H, 4 H₂C-hexyl), 6.98 (br, 1H, HN) ppm; ¹³C NMR (125.4 MHz, CDCl₃): $\delta = 163.2$ (C–NH), 130.4, 128.3 (C=C-hexyl), 118.7 (CN), 87.3 (C–CN), 34.7 (MeN), 26.4, 26.2, 23.5, 21.3 (CH₂-hexyl) ppm; MS (EI, 70 eV): m/z (%) = 192 (56) [M⁺], 191 (100) [M⁺-1].

2-(Ethylamino)-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**5b**)

Obtained as orange material; yield: 735 mg (9.6 %). Compound **5b** is commercially available, but no physical data have been published. m.p.: 49 °C (from pentane); IR (KBr): $\bar{\nu} = 3333$ (NH), 2217 (CN) cm⁻¹; ¹H NMR (500.7 MHz, CDCl₃): $\delta = 1.24$ (t, ³J_{HH} = 7.1 Hz, 3H, MeH₂C), 1.82, 2.95 (2 m, 2 \times 4H, 4 H₂C-hexyl), 3.54 (q, ³J_{HH} = 6.8 Hz, 2H, H₂CMe), 6.88 (br, 1H, HN) ppm; ¹³C NMR (125.4 MHz, CDCl₃): $\delta = 164.3$ (C–NH), 130.5, 128.6 (C=C-hexyl), 118.8 (CN), 87.7 (C–CN), 43.6 (CH₂Me), 26.4, 26.2, 23.5, 21.3 (CH₂-hexyl), 15.3 (HC₂Me) ppm; MS (EI, 70 eV): m/z (%) = 206 (64) [M⁺], 205 (100) [M⁺-1].

2-(Isopropylamino)-4,5,6,7-tetrahydrobenzo[*b*]thiophene-3-carbonitrile (**5c**, C₁₂H₁₆N₂S)

Yellow material; yield: 850 mg (10.3 %); m.p.: 62 °C (from CH₂Cl₂/diethylether 1:1); IR (KBr): $\bar{\nu} = 3325$ (NH), 2218 (CN) cm⁻¹; ¹H NMR (500.7 MHz, CDCl₃): $\delta = 1.24$ (d, ³J_{HH} = 7.1 Hz, 2 \times 3H, 2 Me.C), 1.82, 2.93 (2 m, 2 \times 4H, 4 H₂C-hexyl), 3.01 (m, 1H, HC.(Me)₂), 7.81 (br,

1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 162.2$ (C–NH), 130.1, 128.6 (C=C-hexyl), 118.6 (CN), 86.7 (C–CN), 45.5 (CH_2Me_2), 26.4, 26.2, 23.5, 21.3 (CH_2 -hexyl), 23.7 (HCMe_2) ppm; MS (EI, 70 eV): m/z (%) = 219 (47) [M^+ -1].

4'-Methoxy-3'-methyl-3',4',5,6-tetrahydro-4H-spiro[1-benzothiophene-2,5'-[1,2,3,4]triazaphosphole]-3-carbonitrile 4'-oxide (4a, C₁₁H₁₃N₄O₂PS)

Strew yellow needles; yield: 0.77 g (66.4 %); m.p.: 146 °C (CH_2Cl_2); IR (KBr): $\bar{\nu} = 2214$ (CN), 1237 (P=O, free), 1034 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.21$, 2.15 (2 m, 2 \times 2H, 2 H_2C -hexyl), 2.78 (t, $^3J_{\text{HH}} = 6.8$ Hz, 2H, H_2C -hexyl), 3.51 (d, $^3J_{\text{PH}} = 7.1$ Hz, 3H, Me.N), 3.78 (d, $^3J_{\text{PH}} = 6.7$ Hz, 3H, Me.O), 6.87 (t, $^3J_{\text{HH}} = 6.8$ Hz, 1H, HC-hexyl) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 158.5$, 149.7, 115.8, 27.1, 26.2, 21.9 (C-hexyl), 123.1 (CN), 110.6 (d, $^1J_{\text{PC}} = 136.4$ Hz, C–P), 95.1 (C–CN), 55.3 (d, $^2J_{\text{PC}} = 8.7$ Hz, MeO), 28.6 (d, $^2J_{\text{PC}} = 8.4$ Hz, MeN) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 11.7$ ppm; MS (EI, 70 eV): m/z (%) = 296 (25) [M^+], 268 (37) [M^+ -28 (N_2)], 253 (25) [M^+ -54 ($\text{N}_2 + \text{CN}$)], 227 (23) [M^+ -69 ($\text{N}_2 + \text{CN} + \text{Me}$)], 149 (100) [M^+ -147 ($\text{N}_2 + \text{CN} + \text{Me} + \text{P}(\text{O})(\text{OMe})$)].

4'-Ethoxy-3'-ethyl-3',4',5,6-tetrahydro-4H-spiro[1-benzothiophene-2,5'-[1,2,3,4]triazaphosphole]-3-carbonitrile (4b, C₁₃H₁₇N₄O₂PS)

Strew yellow needles; yield: 0.94 g (74.2 %); m.p.: 132 °C (cyclohexane); IR (KBr): $\bar{\nu} = 2211$ (CN), 1243 (P=O, free), 1029 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.16$ – 1.25 (m, 6H, Me.C–N, MeC.OP), 1.11, 2.45 (2 m, 2 \times 2H, 2 H_2C -hexyl), 2.78 (t, $^3J_{\text{HH}} = 6.8$ Hz, 2H, H_2C -hexyl), 3.79 (dq, $^3J_{\text{HH}} = 6.7$, $^3J_{\text{PH}} = 8.3$ Hz, 2H, H_2CN), 4.02 (qt, $^3J_{\text{HH}} = 5.6$, $^3J_{\text{PH}} = 8.9$ Hz, 2H, H_2CO), 6.93 (t, $^3J_{\text{HH}} = 6.6$ Hz, 1H, HC-hexyl) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 159.7$, 149.4, 118.6, 26.7, 25.3, 22.4 (C-hexyl), 122.5 (CN), 110.3 (d, $^1J_{\text{PC}} = 133.6$ Hz, C–P), 94.7 (C–CN), 63.6 (d, $^2J_{\text{PC}} = 12.7$ Hz, CH_2O), 43.7 (d, $^2J_{\text{PC}} = 11.9$ Hz, CH_2N), 19.7 (d, $^3J_{\text{PC}} = 8.3$ Hz, MeCH_2O), 14.6 (d, $^3J_{\text{PC}} = 7.9$ Hz, MeCH_2N) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 12.4$ ppm; MS (EI, 70 eV): m/z (%) = 324 (43) [M^+], 296 (37) [M^+ -28 (N_2)], 270 (29) [M^+ -54 ($\text{N}_2 + \text{CN}$)], 241 (56) [M^+ -83 ($\text{N}_2 + \text{CN} + \text{Et}$)], 149 (100) [M^+ -175 ($\text{N}_2 + \text{CN} + \text{Et} + \text{P}(\text{O})(\text{OEt})$)].

4'-Isopropoxy-3'-isopropyl-3',4',5,6-tetrahydro-4H-spiro[1-benzothiophene-2,5'-[1,2,3,4]triazaphosphole]-3-carbonitrile 4'-oxide (4c, C₁₅H₂₁N₄O₂PS)

Strew yellow crystals; yield: 0.98 g (71.2 %); m.p.: 152 °C (MeCN); IR (KBr): $\bar{\nu} = 2213$ (CN), 1240 (P=O, free), 1031 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 0.96$, 2.12 (2 m, 2 \times 2H, 2 H_2C -hexyl), 1.26 (dd, $^3J_{\text{HH}} = 4.6$, $^4J_{\text{PH}} = 3.8$ Hz, 2 \times 3H, Me_2CHN), 1.33 (dd, $^3J_{\text{HH}} = 6.7$,

$^4J_{\text{PH}} = 4.1$ Hz, 2 \times 3H, Me_2CHO), 2.69 (t, $^3J_{\text{HH}} = 6.5$ Hz, 2H, H_2C -hexyl), 3.68 (dsept, $^3J_{\text{PH}} = 7.9$ Hz, 1H, HC.N), 4.17 (dsept, $^3J_{\text{PH}} = 8.2$ Hz, 1H, HC.O), 6.49 (t, $^3J_{\text{HH}} = 6.4$ Hz, 1H, HC-hexyl) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 161.2$, 149.6, 120.6, 26.6, 25.6, 22.6 (C-hexyl), 122.8 (CN), 109.8 (d, $^1J_{\text{PC}} = 139.5$ Hz, C–P), 93.8 (C–CN), 75.5 (d, $^2J_{\text{PC}} = 12.9$ Hz, CH.O), 49.6 (d, $^2J_{\text{PC}} = 11.9$ Hz, HC.N), 24.1 (d, $^3J_{\text{PC}} = 7.9$ Hz, Me_2CHO), 21.3 (d, $^3J_{\text{PC}} = 8.5$ Hz, Me_2CHN) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 11.9$ ppm; MS (EI, 70 eV): m/z (%) = 352 (47) [M^+], 324 (36) [M^+ -28 (N_2)], 298 (25) [M^+ -54 ($\text{N}_2 + \text{CN}$)], 255 (46) [M^+ -97 ($\text{N}_2 + \text{CN} + \text{C}_3\text{H}_7$)], 149 (100) [M^+ -203 ($\text{N}_2 + \text{CN} + \text{C}_3\text{H}_7 + \text{P}(\text{O})(\text{OC}_3\text{H}_7)$)].

Preparation of phosphoramidates 8a–8c by reaction of azide 1 with dialkyl phosphites 6a–6c

A mixture of 0.8 g azide **1** (3.9 mmol) and 4.1 mmol of dimethyl, diethyl, or diisopropyl phosphite (**6a–6c**) in 15 cm^3 THF was stirred at r.t. for 6–10 h (TLC) and volatile materials were removed under vacuum. The resulting residue was chromatographed on silica gel with *n*-hexane/ CHCl_3 (8:2, v/v) to give 2-amino-4,5,6,7-tetrahydrobenzo-*[b]*thiophene-3-carbonitrile (**9**) as buff substance in 10.3 % yield; m.p. 147 °C (Ref. [26] 145 °C). Elution with *n*-hexane/ CHCl_3 (1:1, v/v) afforded **8a–8c**.

*Dimethyl 3-cyano-4,5,6,7-tetrahydrobenzo[*b*]thiophen-2-ylphosphoramidate (8a, C₁₁H₁₅N₂O₃PS)*

Colorless crystals; yield: 0.76 g (68.2 %); m.p.: 174 °C (EtOH); IR (KBr): $\bar{\nu} = 3322$ (NH), 2198 (CN), 1234 (P=O, bonded), 1029 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.91$, 2.45 (2 m, 4 \times 2H, 4 H_2C -hexyl), 3.79 (d, $^3J_{\text{PH}} = 11.5$ Hz, 2 \times 3H, 2 Me.OP), 11.35 (br, 1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 151.6$ (d, $^2J_{\text{PC}} = 31.5$ Hz, C–NH), 134.4, 130.2 (C = C-hexyl), 113.8 (CN), 74.3 (C–CN), 52.4 (d, $^2J_{\text{PC}} = 12.9$ Hz, 2 MeOP), 26.4, 26.1, 23.6, 22.3 (CH_2 -hexyl) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 17.6$ ppm; MS (EI, 70 eV): m/z (%) = 285 (53) [M^+ -1], 259 (34) [M^+ -27 (H + CN)], 150 (100) [M^+ -136 (H + CN + P(O)(OMe)₂)].

*Diethyl 3-cyano-4,5,6,7-tetrahydrobenzo[*b*]thiophen-2-ylphosphoramidate (8b, C₁₃H₁₉N₂O₃PS)*

Colorless crystals; yield: 0.86 g (70.6 %); m.p.: 162 °C (EtOH); IR (KBr): $\bar{\nu} = 3325$ (NH), 2194 (CN), 1229 (P=O, bonded), 1032 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.32$ (dt, $^3J_{\text{HH}} = 6.7$, $^4J_{\text{PH}} = 4.4$ Hz, 2 \times 3H, 2 MeCOP), 1.93, 2.46 (2 m, 4 \times 2H, 4 H_2C -hexyl), 4.17 (dq, $^3J_{\text{HH}} = 6.7$, $^3J_{\text{PH}} = 10.6$ Hz, 2 \times 2H, 2 H_2COP), 11.29 (br, 1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 151.3$ (d, $^2J_{\text{PC}} = 29.6$ Hz, C–NH), 134.2, 129.7 (C=C-hexyl), 113.4 (CN), 74.7 (C–CN), 62.4 (d, $^2J_{\text{PC}} = 13.5$ Hz, 2 CH_2OP), 26.3, 26.1, 23.4, 21.9 (CH_2 -

hexyl), 16.3 (d, $^3J_{PC} = 8.2$ Hz, 2*Me*.COP) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 19.3$ ppm; MS (EI, 70 eV): m/z (%) = 313 (48) [$\text{M}^+ - 1$], 287 (34) [$\text{M}^+ - 27$ (H + CN)], 150 (100) [$\text{M}^+ - 164$ (H + CN + P(O)(OEt) $_2$)].

*Diisopropyl 3-cyano-4,5,6,7-tetrahydrobenzo[*b*]thiophen-2-ylphosphoramidate (8c, C₁₅H₂₃N₂O₃PS)*

Colorless crystals; yield: 1.02 g (76.4 %); m.p.: 170 °C (EtOH); IR (KBr): $\bar{\nu} = 3329$ (NH), 2196 (CN), 1231 (P=O, bonded), 1031 (P–O–C) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.34, 1.56$ (2 dd, $^3J_{\text{HH}} = 6.7, ^4J_{\text{PH}} = 4.7$ Hz, 4 × 3H, (Me₂CO) $_2$ P), 1.93, 2.51 (2 m, 4 × 2H, 4 H₂C-hexyl), 4.64–4.81 (2 m, 2H, (HCO) $_2$ P), 10.91 (br, 1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 151.5$ (d, $^2J_{PC} = 31.2$ Hz, C–NH), 133.9, 129.9 (C = C-hexyl), 113.5 (CN), 75.6 (C–CN), 71.2 (d, $^2J_{PC} = 12.7$ Hz, 2 CH.OP), 26.4, 26.2, 23.5, 21.6 (CH₂-hexyl), 24.2 (d, $^3J_{PC} = 8.5$ Hz, 4 Me.COP) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 18.4$ ppm; MS (EI, 70 eV): m/z (%) = 341 (56) [$\text{M}^+ - 1$], 315 (43) [$\text{M}^+ - 27$ (H + CN)], 150 (100) [$\text{M}^+ - 192$ (H + CN + P(O)(OC₃H₇) $_2$)].

Preparation of compounds 11a and 11b by reaction of azide 1 with 10a, 10b

Hexamethyl- or hexaethylphosphorus triamide (10a, 10b, 5.2 mmol) in 5 cm³ THF was added dropwise to 0.8 g of azide 1 (3.9 mmol) in 10 cm³ THF. The reaction mixture was stirred at r.t. for ≈ 4 h (TLC). The precipitate was collected and washed several times with light petroleum (40–60 °C) to give 11a and 11b. Compounds 11a and 11b are pure enough for carrying out the spectroscopic analyses, and stable for a week at –10 °C under argon.

2-[3-[Tris(dimethylamino)phosphoranylidene]triaz-1-en-1-yl]-4,5,6,7-tetrahydro-1-benzothiophene-3-carbonitrile (11a, C₁₅H₂₆N₇PS)

Yellow material; yield: 1.26 g (90 %); m.p.: 193 °C (CHCl₃); IR (KBr): $\bar{\nu} = 2206$ (CN), 1333 (P=N), 1301, 923 (P(NMe₂) $_3$) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.81, 2.61$ (2 m, 2 × 4H, 4 H₂C-hexyl), 2.75, 2.81 (2 d, $^3J_{\text{PH}} = 8.9$ Hz, 18H, (Me₂N) $_3$ P) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 160.6$ (C–N=N), 131.1, 128.7 (C=C-hexyl), 117.5 (CN), 69.4 (C–CN), 37.8 (d, $^2J_{PC} = 13.5$ Hz, (Me₂N) $_3$ P), 26.3, 26.1, 23.6, 21.4 (CH₂-hexyl) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 40.2$ ppm; MS (EI, 70 eV): m/z (%) = 367 (35) [M^+], 339 (24) [$\text{M}^+ - 28$ (N₂)], 313 (29) [$\text{M}^+ - 54$ (N₂ + CN)], 150 (100) [$\text{M}^+ - 217$ (N₂ + CN + C₆H₁₈N₃P)].

2-[3-[Tris(diethylamino)phosphoranylidene]triaz-1-en-1-yl]-4,5,6,7-tetrahydro-1-benzothiophene-3-carbonitrile (11b, C₂₁H₃₈N₇PS)

Yellow material; yield: 1.49 g (92 %); m.p.: 184 °C (EtOH); IR (KBr): $\bar{\nu} = 2210$ (CN), 1330 (P=N), 1317,

898 (P(NEt₂) $_3$) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 0.95, 1.39$ (2 dt, $^3J_{\text{HH}} = 8.5, ^4J_{\text{PH}} = 6.4$ Hz, 18H, {(Me.C) $_2$ N} $_3$ P), 1.84, 2.63 (2 m, 4 × 2H, 4 H₂C-hexyl), 2.91, 3.26 (2 dq, $^3J_{\text{HH}} = 8.5, ^3J_{\text{PH}} = 9.1$ Hz, 12H, [(H₂C) $_2$ N] $_3$ P) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 160.4$ (C–N=N), 131.3, 128.6 (C=C-hexyl), 117.4 (CN), 69.3 (C–CN), 39.5 (d, $^2J_{PC} = 13.7$ Hz, [(CH₂) $_2$ -N] $_3$ P), 26.5, 26.2, 23.6, 21.2 (CH₂-hexyl), 14.7 (d, $^3J_{PC} = 8.4$ Hz, [(Me.C) $_2$ N] $_3$ P) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 40.4$ ppm; MS (EI, 70 eV): m/z (%) = 451 (34) [M^+], 423 (26) [$\text{M}^+ - 28$ (N₂)], 397 (19) [$\text{M}^+ - 54$ (N₂ + CN)], 150 (100) [$\text{M}^+ - 301$ (N₂ + CN + C₁₂H₃₀N₃P)].

Reaction of 1 with hexaalkylphosphorus triamides 10a and 10b in the presence of a protonating agent. Preparation of compounds 13a and 13b

Phosphorus triamides 10a, 10b (5.2 mmol) in 5 cm³ of dry THF were added in one portion to a mixture of 0.8 g azide 1 (3.9 mmol) and 2 cm³ dil. EtOH (1:1) in 10 cm³ THF. The reaction mixture was stirred at r.t. for ≈ 6 h (TLC). The solvent was evaporated to dryness; the residue was washed several times with light petroleum (40–60 °C), and crystallized from the proper solvent to give 13a and 13b, respectively.

N''-(3-Cyano-4,5,6,7-tetrahydro-1-benzothien-2-yl)-N,N,N',N'-tetramethylphosphoric triamide (13a, C₁₃H₂₁N₄OPS)

Pale yellow crystals; yield: 0.97 g (79.6 %); m.p.: 165 °C (MeCN); IR (KBr): $\bar{\nu} = 3325$ (NH), 2205 (CN), 1232 (P=O, bonded) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.91, 2.32$ (2 m, 4 × 2H, 4 H₂C-hexyl), 2.45 (d, $^3J_{\text{PH}} = 9.4$ Hz, 2 × 6H, (Me₂N) $_2$ P), 11.21 (br, 1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 150.6$ (d, $^2J_{PC} = 28.3$ Hz, C–NH), 136.1, 130.2 (C=C-hexyl), 113.7 (CN), 87.3 (C–CN), 35.5 (d, $^2J_{PC} = 13.5$ Hz, (Me₂N) $_2$ P), 26.4, 26.2, 23.5, 21.8 (CH₂-hexyl) ppm; ^{31}P NMR (200.7 MHz, CDCl_3): $\delta = 34.7$ ppm; MS (EI, 70 eV): m/z (%) = 311 (45) [$\text{M}^+ - 1$], 285 (34) [$\text{M}^+ - 27$ (H + CN)], 150 (100) [$\text{M}^+ - 162$ (H + CN + C₄H₁₂N₂OP)].

N''-(3-Cyano-4,5,6,7-tetrahydro-1-benzothien-2-yl)-N,N,N',N'-tetraethylphosphoric triamide (13b, C₁₇H₂₉N₄OPS)

Pale yellow crystals; yield: 1.1 g (82.4 %); m.p.: 153 °C (CHCl₃); IR (KBr): $\bar{\nu} = 3327$ (NH), 2205 (CN), 1228 (P=O, bonded) cm^{-1} ; ^1H NMR (500.7 MHz, CDCl_3): $\delta = 1.32$ (dt, $^3J_{\text{HH}} = 7.7, ^4J_{\text{PH}} = 5.4$ Hz, 2 × 6H, [MeC.C) $_2$ N] $_2$ P), 1.92, 2.45 (2 m, 4 × 2H, 4 H₂C-hexyl), 3.26 (dq, $^3J_{\text{HH}} = 7.7, ^3J_{\text{PH}} = 8.6$ Hz, 2 × 4H, [(H₂C) $_2$ N] $_2$ P), 11.25 (br, 1H, HN) ppm; ^{13}C NMR (125.4 MHz, CDCl_3): $\delta = 150.4$ (d, $^2J_{PC} = 28.6$ Hz, C–NH), 135.8, 130.1 (C=C-hexyl), 113.6 (CN), 88.2 (C–CN), 40.8 (d,

$^2J_{PC} = 13.5$ Hz, $[(MeCH_2)_2N]_2P$, 26.3, 25.9, 23.5, 21.2 (CH₂-hexyl), 16.3 (d, $^3J_{PC} = 8.2$ Hz, $[(MeCH_2)_2N]_2P$) ppm; ^{31}P NMR (200.7 MHz, CDCl₃): $\delta = 33.3$ ppm; MS (EI, 70 eV): m/z (%) = 367 (42) [M⁺-1], 341 (31) [M⁺-27 (H + CN)], 150 (100) [M⁺-218 (H + CN + C₈H₂₀N₂OP)].

Antitumor activity screening

Determination of GI₅₀, TGI, and LC₅₀ values: A total of 12 human tumor cell lines, derived from three cancer types (breast, colon, and prostate), formed the basis of this test [35, 36]. The tumor cells were cultured in RPMI1640 medium supplemented with 5 % fetal calf serum and 2 mM L-glutamine. The tumor cells are inoculated over a series of standard 48-well microtiter plates in 100 cm³ of medium. Density of inoculums depends on the type of tumor cell and on its growth characteristics. These cells are then preincubated on the microtiter plate for 24 h before adding the compounds. They were tested in DMSO solution at three different concentrations (10⁻⁴, 10⁻⁵, and 10⁻⁶ M). After an incubation of the chemical agent for 48 h with the tumor cell lines, a sulforhodamine B (SRB) protein assay was used to estimate cell viability or growth. The cytotoxic effects are evaluated, and the assay was used to estimate cell viability or growth, and the assay results and dose–response parameters were calculated as previously described [34].

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