ARTICLE IN PRESS

Bioorganic & Medicinal Chemistry Letters xxx (2017) xxx-xxx



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters



journal homepage: www.elsevier.com/locate/bmcl

Exploration of phenylpropanoic acids as agonists of the free fatty acid receptor 4 (FFA4): Identification of an orally efficacious FFA4 agonist

Steven M. Sparks^{*}, Christopher Aquino, Pierette Banker, Jon L. Collins, David Cowan, Caroline Diaz, Steven T. Dock, Donald L. Hertzog, Xi Liang, Erin D. Swiger, Josephine Yuen, Grace Chen, Channa Jayawickreme, David Moncol, Christopher Nystrom, Vincent Rash, Thomas Rimele, Shane Roller, Sean Ross

GlaxoSmithKline, Enteroendocrine Discovery Performance Unit and Platform Technology and Science, 5 Moore Drive, Research Triangle Park, NC 27709, United States

ARTICLE INFO

Article history: Received 15 December 2016 Revised 10 January 2017 Accepted 11 January 2017 Available online xxxx

Keywords: Free fatty acid receptor 4 GPR120 Free fatty acid receptor 1 Phenylpropanoic acid Type 2 diabetes

ABSTRACT

The long chain free fatty acid receptor 4 (FFA4/GPR120) has recently been recognized as lipid sensor playing important roles in nutrient sensing and inflammation and thus holds potential as a therapeutic target for type 2 diabetes and metabolic syndrome. To explore the effects of stimulating this receptor in animal models of metabolic disease, we initiated work to identify agonists with appropriate pharmacokinetic properties to support progression into in vivo studies. Extensive SAR studies of a series of phenylpropanoic acids led to the identification of compound **29**, a FFA4 agonist which lowers plasma glucose in two preclinical models of type 2 diabetes.

© 2017 Elsevier Ltd. All rights reserved.

Free fatty acids play important roles in biological processes contributing to both health and disease. As essential nutritional components, free fatty acids are utilized as building blocks for energy storage and cell membrane integrity and as substrates for critical cellular pathways (i.e. Randle cycle¹). In addition to these roles, free fatty acids have been recognized as important signaling molecules involved in a variety of physiological processes including inflammation and insulin resistance and hence contribute to the pathologies of type 2 diabetes and obesity.

Recently, numerous G-protein coupled receptors (GPCRs) have been identified that are activated by free fatty acids.^{2–6} The GPCRs can be grouped according to the carbon chain length (short chain: <6 carbons; medium chain: 6–12 carbons; long chain: >12 carbons) and level of unsaturation of the fatty acids which activate the receptors, with free fatty acid receptor 2 (FFA2/GPR43) and free fatty acid receptor 3 (FFA3/GPR41) activated by short chain fatty acids, G-protein coupled receptor 84 (GPR84) activated by medium chain fatty acids, free fatty acid receptor 1 (FFA1/GPR40) activated by saturated and unsaturated long-chain fatty acids, and free fatty

* Corresponding author at present address: Viamet Pharmaceuticals, 4505 Emperor Blvd., Suite 300, Durham, NC 27703, United States.

E-mail address: ssparks@viamet.com (S.M. Sparks).

acid receptor 4 (FFA4/GPR120) activated by unsaturated long-chain fatty acids including omega-3 fatty acids (ω -3 FAs).^{7,8}

Our interest in understanding the physiological processes governed by the long-chain fatty acid receptors led us to further consider FFA1 and FFA4 as candidates for small molecule drug discovery efforts. Previous reports have highlighted our group's efforts leading to the identification of the FFA1 biased agonist GW9508 (~100-fold selective for FFA1 over FFA4).^{9,10} These studies provided the first evidence that small molecule mediated FFA1 agonism could be utilized for the pharmacological regulation of insulin secretion.¹¹ FFA1 is expressed highest in the pancreas, with insulin secreting β-cells abundantly expressing the receptor, and has also been shown to also be present in intestine and brain.^{12,13} FFA1 amplified glucose-stimulated insulin secretion has been extensively studied and provides the basis for the current interest in FFA1 selective small molecule agonists for the treatment of type 2 diabetes.^{14–18}

FFA4 has been less extensively studied but numerous recent reports highlight the receptor in several pathologies associated with type 2 diabetes and the components of metabolic syndrome.^{8,19–21} FFA4 is abundantly expressed in several tissues including the intestine,²² adipose,²³ and lung²⁴ and is present in a variety of cell types including proinflammatory macrophages,⁸ intestinal L- and K-cells,^{22,25} the mouse enteroendocrine STC-1 cell

Please cite this article in press as: Sparks S.M., et al. Bioorg. Med. Chem. Lett. (2017), http://dx.doi.org/10.1016/j.bmcl.2017.01.034

http://dx.doi.org/10.1016/j.bmcl.2017.01.034 0960-894X/© 2017 Elsevier Ltd. All rights reserved.

ARTICLE IN PRESS

line,²⁶ and in the taste buds of the tongue.²⁷ This pattern of expression is consistent with the nutrient sensing and anti-inflammatory roles that have been reported for FFA4.^{21,28–30} Indeed, seminal studies aimed at identifying the receptors involved in nutrient stimulated glucagon-like peptide-1 (GLP-1) secretion have identified FFA4 as a candidate GPCR involved in long-chain fatty acid stimulated GLP-1 secretion.²² In addition the anti-inflammatory and insulin sensitizing effects of the ω -3 fatty acids DHA and EPA have been reported to occur through FFA4.^{8,30–32} Further understanding of the therapeutic potential of FFA4 activation would be greatly aided by the identification of small molecule agonists with good pharmacokinetic properties.^{33–40} With this goal in mind, efforts directed toward the identification of small molecule FFA4 agonists culminating in identification of phenylpropanoic acid agonists with pharmacokinetic properties appropriate for in vivo studies are described herein.

Compound **1** was identified as starting point for optimization following a cross-screening effort of our FFA1 program compounds (Fig. 1). The potency and efficacy of compound **1** was determined using a ten-point dose response curve in mammalian U2OS cells transiently expressing either human FFA1 or FFA4 using a FLIPR readout. The data revealed that compound **1** was 17-fold selective for FFA1 over FFA4. Thus, a primary goal of this exercise was to explore the SAR against both receptors while identifying compounds with selectivity for the FFA4 receptor.

In addition, several other considerations were made for the optimization exercise. While the structural features of the endogenous ligands for long-chain fatty acid receptors suggest obtaining orthosteric agonists with drug-like properties may be challenging, the optimization efforts were committed to maintaining molecular weight in the range of compound **1** (MW < 400) while hoping to realize high plasma drug concentrations, as plasma protein binding was expected to limit the free fraction of the drug. With these goals in mind, our efforts directed toward the identification of improved small molecule FFA4 agonists commenced.

The general synthesis of the phenylpropanoic acid agonists is shown in Scheme 1. Alkylation of substituted 4-hydroxyphenylpropanoic esters with substituted benzyl halides or benzyl alcohols followed by ester hydrolysis directly yielded the desired phenylpropanic acids. The ease of synthesis of the desired compounds coupled with the commercial availability of the starting materials allowed for rapid SAR generation.

Phenylpropanoic acid headgroups were prepared as shown in Scheme 2. In general, ketone or benzaldehyde derivatives (I) were olefinated and reduced to provide the corresponding propanoates (III), which in some cases required demethylation (BBr₃) to provide the desired phenols. Hydroxyl derivative **4** was obtained via the aldol reaction of the corresponding benzaldehyde derivative (I) with bis-lithiated species of acetic acid. The phenylpropanoic acid derivative for alcohol **13** was obtained from the corresponding





Scheme 1. General syntheses of the phenylpropanoic acids. Reagents and conditions: (a) K_2CO_3 , DMF (X = Cl or Br) or DIAD, PPh₃ (X = OH); (b) NaOH or LiOH, THF/ MeOH/H₂O.



Scheme 2. General syntheses of the phenylpropanoic phenols. Reagents and conditions: (a) DIBAL-H, toluene; (b) NaH or *t*-BuOK, (EtO)₂POCH₂CO₂R, DMF; (c) *n*-BuLi, AcOH, THF; (d) 10% Pd/C, H₂, THF/MeOH or NiCl₂, NaBH₄, THF; (e) BBr₃, DCM (f) TiCl₄, MeOCHCl₂, DCM; (g) 2-methyl-2-propen-1-ol, DIAD, PPh₃, toluene, 250 °C.

aldehyde (**IV**) which in turn was prepared by a titanium-mediated formylation/deprotection sequence from **III**. Isobutyl derivative **14** was prepared by the Mitsunobu-initiated Claisen-rearrangement of the corresponding phenol (**III**, R' = H) with 2-methyl-2-propenol followed by alkene reduction to afford intermediate **V**.

Benzyl alcohols and their activated derivatives were prepared as outlined in Scheme 3. Benzoic acids with the desired 2,5-disubstitution pattern were reduced directly to the desired benzyl alcohol derivatives (VI). Variation of ether group at the 2-position holding the 5-methyl substituent constant (VII) was accomplished starting from 5-methyl salicylic acid by simple alkylation followed by reduction. Formylation of 4-trifluoromethyl phenol followed by reduction and conversion to the mesylate derivative X allowed for incorporation of a trifluoromethyl group at the 5-position of the benzyl alcohol derivatives. 5-Methyl-3-alkoxy-disubstituted benzyl alcohols were prepared starting from 3-methoxy-5-bromobenzene utilizing a sequence consisting of deprotection followed by alkylation to furnish bromide XI, which underwent subsequent metal-halogen exchange and trapping to provide the intermediate aldehyde which was reduced to afford the desired benzyl alcohol derivatives (XII).



Scheme 3. General synthetic routes to benzyl alcohol derivatives. Reagents and conditions: (a) LiAlH₄ or BH₃ \bullet THF, THF; (b) RI, K₂CO₃, DMF; (c) 3,4-dihydro-2H-pyran, PPTS, DCM; (d) *n*-BuLi; DMF; (e) NaBH₄, MeOH; (f) MsCl, Et₃N, DCM; (g) BBr₃, DCM.



Scheme 4. Synthesis of compound **34**. Reagents and conditions: (a) NBS, CCl₄ (b) Ethyl 3-(4-hydroxy-3-methylphenyl)propanoate, K₂CO₃, DMF (c) Phenol, Pd(OAc)₂, di-*t*-BuXPhos, K₃PO₄, toluene (d) LiOH, 1,4-dioxane/H₂O.

Table 1

Cmpd

2

3

4

5

6

7

8

9

10

11

12

13

14

In vitro FFA4 agonism: Phenylpropanoic acid SAR.

R2, R3, R5

H, H, H

ннн

H, H, H

Н, Н, Н

Me, H, H

H. Me. H

H. Me. Me

Me, Me, H

-(CH)4-, H

H. OMe. H

H, CH₂OH, H

H, CH₂CH(CH₃)₂, H

H. Cl. H



hFFA4 EC50^a (% Max Resp.)

681 (105)

9977 (85)

11,380 (97)

>30,000

722 (90)

201 (108)

660 (102)

304 (77)

108 (80)

2292 (105)

1992 (110)

9660 (161)

5911 (53)

Select compounds were derivatized following construction of the benzyl ether bond (Scheme 4), for instance, compounds containing 5-methyl-3-alkoxy substitution on the benzyl tail were prepared starting from 3,5-dimethylbromobenzene. Radical halogenation followed by benzyl ether formation provided the arylbromide intermediate **XIII** which underwent palladium catalyzed etherification⁴¹ followed by saponification to provide the desired diphenylether **31**.

Our initial studies explored substitution about the phenylpropanoic acid headgroup while holding a 2-thiotrifluoromethylbenzyl tail constant (Table 1). Parent compound 2 provided a nearly balanced FFA1/FFA4 (2-fold selective for FFA4) starting point and it was hoped that further improvements in potency for FFA4 could be realized utilizing the benzyl tail of compound 2. Addition of polarity (3 & 4) along the propanoic acid chain or simple substitution (5) eroded activity at both receptors. Incorporation of a methyl substituent at R2 (6) or dimethyl substitution at R3 and R5 (8) provided no improvement in potency, while methyl substitution at R3 (7) increased activity at FFA4 while diminishing FFA1 activity. Dimethyl substitution (9) provided a selective FFA4 agonist while growing the substitution in the form of a naphthalene ring (10) led to diminished activity at FFA4. Small nonpolar substituents at R3, such as the chloro group of **11**, provided a further improvement in FFA4 potency and afforded excellent selectivity over FFA1, which stands in contrast to the incorporation of more polar (12 & 13) or larger (14) R3 substituents which led to a reduction in activity at both receptors.

With the identification of small nonpolar substituents at R3 of the phenylpropanoic acid ring providing improved activity at FFA4, exploration of the benzyl SAR was undertaken (Table 2). Small substituents were well tolerated at both the *ortho* and *meta* positions (15–23) with the 2-bromo- (19), 2-isopropoxy- (18), and 3-methyl-substituents (23) providing potent and selective FFA4 agonists. In select cases incorporation of *ortho-meta*-disubstitution (2,5-orientation) and *meta-meta*-disubstitution (3,5-orientation) afforded equipotent agonists with the mono-substituted comparators (24 vs. 23; 27 vs. 18 & 23) while for the majority of exemplars improved potency was realized (25 vs. 19; 26 vs. 23; 28 vs. 23; 29 vs. 21) at FFA4. Benzyl disubstitution provided weak activity at FFA1. Unmasking polar functionality (30 vs. 29) provided a dramatic decrease in activity at FFA4, consistent with the lipophilic nature of the binding pocket and structures of the natural ligands.

hFFA1 EC50 (% Max Resp.)

³⁸ for assay details.

1423 (77)

13,142 (109)

1544 (108)

21.777 (115)

8514 (81)

>30,000

>30,000

>10,000

>30.000

>30,000

>30.000

>30,000

N.T.

Please cite this article in press as: Sparks S.M., et al. Bioorg. Med. Chem. Lett. (2017), http://dx.doi.org/10.1016/j.bmcl.2017.01.034

Values are means of three experiments, see the supporting information for standard deviations (N.T. = not tested). EC₅₀ reported in nM. See Ref.

Х

0

 CH_2

CHOH

CHCH₃

 CH_2

CH₂

 CH_2

CH₂

 CH_2

 CH_2

 CH_2

 CH_2

CH₂

ARTICLE IN PRESS

S.M. Sparks et al./Bioorganic & Medicinal Chemistry Letters xxx (2017) xxx-xxx

4

Table 2

In vitro FFA4 agonism: benzyl substitution.



Cmpd	R	hFFA4 EC ₅₀ ª (% Max Resp)	hFFA1 EC ₅₀ ª (% Max Resp)
15	Н	750 (83)	>30,000
16	2-Cl	972 (130)	11,588 (80)
17	2-CF ₃	531 (103)	8600 (95)
18	2-OiPr	271 (118)	>30,000
19	2-Br	236 (116)	>30,000
20	3-Cl	362 (104)	10,268 (60)
21	3-CF ₃	898 (117)	>30,000
22	3-OPh	556 (95)	3837 (90)
23	3-Me	275 (100)	>30,000
24	2-Cl-5-Me	188 (115)	18,051 (243)
25	2-Br-5-Me	53 (118)	>30,000
26	3,5-diMe	60 (66)	3782 (97)
27	2-OiPr-5-Me	172 (106)	19,588 (154)
28	3-OiPr-5-Me	60 (112)	>30,000
29	2-MeO-5-	299 (124)	11,803 (105)
	CF ₃		
30	2-0H-5-CF ₃	10,000 (78)	N.T.
31	3-Me-5-OPh	40 (97)	1631 (85)

^a Values are means of three experiments, see the supporting information for standard deviations (N.T. = not tested). EC_{50} reported in nM. See Ref. ³⁸ for assay details.

Interestingly, combination of the biphenyl ether moiety of **1** with the *meta*-methyl (**22**) provided the most potent FFA4 agonist of the series (**31**).

With a series of FFA4 agonists with varying degrees of selectivity over FFA1 in hand, examination of the pharmacokinetic (PK) properties of select compounds in C57Bl6 mice was initiated (Table 3). In general, the compounds were characterized with low to moderate IV clearance, low volumes of distribution (V_{ss} < 2 L/kg), and rapid oral absorption (T_{max} values of 0.25–0.5 h). Compound **29** showed good oral exposure and high oral bioavailability with moderate half-life (t½ = 1.7 h). Examination of the protein binding of the compounds revealed all to be >99% bound in human plasma with 99.9% protein binding measured for compound **29**. Despite high protein binding, compound **29** was chosen as a tool molecule for in vivo studies based upon its superior oral exposure (C_{max} = 6779 ng/mL), potency at FFA4 (EC₅₀ = 299 nM), and selectivity over human FFA1 (EC₅₀ = 11.8 µM).

Prior to in vivo studies, the in vitro selectivity of compound **29** was evaluated in a panel of targets in both full curve binding and functional assays including the rodent ortholog assays for FFA1 and FFA4 and additional members of the free fatty acid receptor family (Fig. 2). Increased selectivity (>85-fold) for both rat and mouse FFA4 over FFA1 was realized relative to the human

Table 3					
DMPK profiles of selected	FFA4	agonists	in	C57BL/6 I	mic



FFA4 (GPR120): Human EC₅₀ = 299 nM, % Max Resp = 124 ± 24 % Mouse EC₅₀ = 70 nM, Max Resp = 108 ± 32 % Rat EC₅₀ = 230 nM, Max Resp = 110 ± 41 %

 $FFA1 \text{ (GPR40):} \\ Human EC_{50} = 11,803 \text{ nM}, \text{ Max Resp} = 105 \pm 36 \% \\ Mouse EC_{50} = 6,213 \text{ nM}, \text{ Max Resp} = 83 \pm 2 \% \\ Rat EC_{50} > 30,000 \text{ nM} \\ \end{cases}$

FFA2 (GPR43) human $EC_{50} = 25,312$ nM, Max Resp = 59 ± 37 % FFA3 (GPR41) human $EC_{50} > 50,000$ nM

Fig. 2. In vitro profile of FFA1 agonist 29.



Fig. 3. Effect of FFA4 agonist 29 on plasma glucose in 8-week old male ZDF rats (n = 8) treated orally twice a day at the indicated doses for 14 days. \dot{p} < 0.05 vs vehicle.

receptors (40-fold). With respect to activity on the remaining members of the free fatty acid family, compound **29** showed weak partial agonism of the FFA2 receptor and profiled as inactive against FFA3 in FLIPR based assays. The remainder of the panel included a mixture of GPCR, ion channel, and enzyme assays with compound **29** showing robust selectivity against the panel.

Investigation of the anti-diabetic properties of compound **29** was undertaken in Zucker Diabetic Fatty (ZDF) rats, an established preclinical model of type 2 diabetes. Compound **29** was administered to 8-week old male ZDF rats at doses of 10 and 100 mg/kg twice daily for 14 days with plasma glucose concentrations determined at baseline, 7 and 14 days.

Cmpd	IV Cl (mL/min/kg)	IV AUC (hr*ng/mL)	PO Cmax (ng/mL)	PO AUC (hr*ng/mL)	% F
23	12	4105	3083	5353	39
24	18	2859	3028	4403	46
26	35	1416	2627	2596	55
28	43	1171	687	767	20
29 *	20	3618	6779	9438	110

^a C57BL/6 J Mice (n = 2), Oral dose = 10 mg/kg, IV dose = 3 or 4.2* mg/kg. PO formulation: 0.5% HMPC: 0.1% Tween suspension; IV formulation: 2% DMSO, 5% solutol.

Please cite this article in press as: Sparks S.M., et al. Bioorg. Med. Chem. Lett. (2017), http://dx.doi.org/10.1016/j.bmcl.2017.01.034

S.M. Sparks et al./Bioorganic & Medicinal Chemistry Letters xxx (2017) xxx-xxx



Fig. 4. Effect of compound **29** on blood glucose in db/db mice (n = 10) dosed at 100 mg/kg BID as compared to vehicle. p < 0.05 vs vehicle.

All groups of ZDF rats had comparable levels of plasma glucose, insulin, and HbA_{1c} prior to treatment. As illustrated in Fig. 3, the vehicle-treated animals showed a progressive hyperglycemia with plasma glucose rising from baseline values of 188 to 245 mg/dL. In contrast, plasma glucose for compound **29** treated animals was reduced to 151 and 139 mg/dL at the 10 and 100 mg/kg doses, respectively, from a starting average of 192 mg/dL.

With positive effects on plasma glucose measured in the ZDF rat, compound **29** was progressed into a second rodent model of type 2 diabetes, the db/db mouse. In this model fed db/db mice were orally administered either vehicle or compound **29** (100 mg/kg) twice a day and blood glucose was measured before and after 1 and 2 weeks of treatment. In addition, HbA_{1c} levels were measured before and after 2 weeks of treatment. As shown in Fig. 4, compound **29** gave a robust response, normalizing blood glucose after 1 week of treatment. Glucose lowering was maintained through the end of the study with levels for the compound **29** treated animals finishing at 106 mg/dL versus 276 mg/dL for vehicle controls. Compound **29** also robustly lowered HbA_{1c} levels by 1.6% to a final value of 5.8% with the vehicle control group finishing the study at 7.8%.

In summary, the SAR of a series of phenylpropanoic acid-free fatty acid receptor agonists is described which resulted in the identification of potent FFA4 agonists. The superior pharmacokinetic properties of compound **29** led to its selection as an in vivo tool compound which afforded robust efficacy in two rodent models of type 2 diabetes. In vivo results with compound **29** detail the potential for FFA4 agonists as treatments for type 2 diabetes and additional factors associated with metabolic syndrome.

A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl.2017.01. 034.

References

- 1. Hue L, Taegtmeyer H. The Randle cycle revisited: a new head for an old hat. *Am J Endocrinol MeTable*. 2009;297:E578–E591.
- Layden BT, Angueira AR, Brodsky M, Durai V, Lowe Jr WL. Short chain fatty acids and their receptors: new metabolic targets. *Transl Res.* 2013;161:131–140.
- Stoddart LA, Smith NJ, Milligan G. International union of pharmacology. LXXI. Free fatty acid receptors FFA1, -2, and -3: pharmacology and pathophysiological functions. *Pharmacol Rev.* 2008;60:405–417.
- Vangaveti V, Shashidhar V, Jarrod G, Baune BT, Kennedy RL. Free fatty acid receptors: emerging targets for treatment of diabetes and its complications. *Ther Adv Endocrinol MeTable*. 2010;1:165–175.
- 5. Vinolo MAR, Hirabara SM, Curi R. G-protein-coupled receptors as fat sensors. *Curr Opin Clin Nutr Metab Care*. 2012;15:112–116.

- 6. Yonezawa T, Kurata R, Yoshida K, Murayama MA, Cui X, Hasegawa A. Free fatty acids-sensing G protein-coupled receptors in drug targeting and therapeutics. *Curr Med Chem.* 2013;20:3855–3871.
- Burns RN, Moniri NH. Agonism with the omega-3 fatty acids α-linolenic acid and docosahexaenoic acid mediates phosphorylation of both the short and long isoforms of the human GPR120 receptor. *Biochem Biophys Res Commun.* 2010;396:1030–1035.
- (a) Oh DY, Talukdar S, Bae EJ, et al. GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell.* 2010;142:687–698;
 (b) DY, Wiley E, Aligner TF, et al. A Control of the sensitivity of the senset of the sensitivity of the sensitity of the sensitity of th

(b) Oh DY, Walenta E, Akiyama TE, et al. A Gpr120-selective agonist improves insulin resistance and chronic inflammation in obese mice. *Nat Med.* 2014;20:942–947.

- 9. Garrido DM, Corbett DF, Dwornik KA, et al. Synthesis and activity of small molecule GPR40 agonists. *Bioorg Med Chem Lett.* 2006;16:1840–1845.
- McKeown SC, Corbett DF, Goetz AS, et al. Solid phase synthesis and SAR of small molecule agonists for the GPR40 receptor. *Bioorg Med Chem Lett.* 2007;17:1584–1589.
- Briscoe CP, Peat AJ, McKeown SC, et al. Pharmacological regulation of insulin secretion in MIN6 cells through the fatty acid receptor GPR40: identification of agonist and antagonist small molecules. *Br J Pharmacol.* 2006;148:619–628.
- 12. Briscoe CP, Tadayyon M, Andrews JL, et al. The orphan G protein-coupled receptor GPR40 is activated by medium and long chain fatty acids. *J Biol Chem.* 2003;278:11303–11311.
- Itoh Y, Kawamata Y, Harada M, et al. Free fatty acids regulate insulin secretion from pancreatic l² cells through GPR40. *Nature*. 2003;422:173–176.
- 14. Burant CF. Activation of GPR40 as a therapeutic target for the treatment of type 2 diabetes. *Diabetes Care*. 2013;36:S175–S179.
- Choi YJ, Shin D, Lee JY. G-protein coupled receptor 40 agonists as novel therapeutics for type 2 diabetes. Arch Pharmacal Res. 2013. Ahead.
- Feng XT, Leng J, Xie Z, Li SL, Zhao W, Tang QL. GPR40: a therapeutic target for mediating insulin secretion (review). Int J Mol Med. 2012;30:1261–1266.
- Kaku K. Fasiglifam as a new potential treatment option for patients with type 2 diabetes. Expert Opin Pharmacother. 2013;14:2591–2600.
- Poitout V, Lin DCH. Modulating GPR40: therapeutic promise and potential in diabetes. Drug Discovery Today. 2013;18:1301–1308.
- Ichimura A, Hirasawa A, Poulain-Godefroy O, et al. Dysfunction of lipid sensor GPR120 leads to obesity in both mouse and human. *Nature*. 2012;483: 350–354.
- Mo XL, Wei HK, Peng J, Tao YX. Free fatty acid receptor GPR120 and pathogenesis of obesity and type 2 diabetes mellitus. *Prog Mol Biol Transl Sci.* 2013;114:251–276.
- Talukdar S, Olefsky JM, Osborn O. Targeting GPR120 and other fatty acidsensing GPCRs ameliorates insulin resistance and inflammatory diseases. *Trends Pharmacol Sci.* 2011;32:543–550.
- 22. Hirasawa A, Tsumaya K, Awaji T, et al. Free fatty acids regulate gut incretin glucagon-like peptide-1 secretion through GPR120. *Nat Med*. 2005;11:90–94.
- 23. Gotoh C, Hong YH, Iga T, et al. The regulation of adipogenesis through GPR120. Biochem Biophys Res Commun. 2007;354:591–597.
- 24. Miyauchi S, Hirasawa A, Iga T, et al. Distribution and regulation of protein expression of the free fatty acid receptor GPR120. *Naunyn-Schmiedeberg's Arch Pharmacol*. 2009;379:427–434.
- Parker HE, Habib AM, Rogers GJ, Gribble FM, Reimann F. Nutrient-dependent secretion of glucose-dependent insulinotropic polypeptide from primary murine K cells. *Diabetologia*. 2009;52:289–298.
- Tanaka T, Katsuma S, Adachi T, Koshimizu TA, Hirasawa A, Tsujimoto G. Free fatty acids induce cholecystokinin secretion through GPR120. Naunyn-Schmiedeberg's Arch Pharmacol. 2008;377:523–527.
- Martin C, Passilly-Degrace P, Chevrot M, et al. Lipid-mediated release of GLP-1 by mouse taste buds from circumvallate papillae: putative involvement of GPR120 and impact on taste sensitivity. J Lipid Res. 2012;53:2256–2265.
- Ichimura A, Hirasawa A, Hara T, Tsujimoto G. Free fatty acid receptors act as nutrient sensors to regulate energy homeostasis. *Prostaglandins Other Lipid Mediators*. 2009;89:82–88.
- Nguyen CA, Akiba Y, Kaunitz JD. Recent advances in gut nutrient chemosensing. Curr Med Chem. 2012;19:28–34.
- Oh D, Lagakos WS. The role of G-protein-coupled receptors in mediating the effect of fatty acids on inflammation and insulin sensitivity. *Curr Opin Clin Nutr Metab Care*. 2011;14:322–327.
- Im DS. Omega-3 fatty acids in anti-inflammation (pro-resolution) and GPCRs. Prog Lipid Res. 2012;51:232–237.
- Morishita M, Tanaka T, Shida T, Takayama K. Usefulness of colon targeted DHA and EPA as novel diabetes medications that promote intrinsic GLP-1 secretion. J Controlled Release. 2008;132:99–104.
- Cornall LM, Mathai ML, Hryciw DH, McAinch AJ. GPR120 agonism as a countermeasure against metabolic diseases. *Drug Discovery Today*. 2014;19:670–679.
- **34.** Halder S, Kumar S, Sharma R. The therapeutic potential of GPR120: a patent review. *Expert Opin Ther Pat.* 2013;23:1581–1590.
- **35.** Hara T, Hirasawa A, Ichimura A, Kimura I, Tsujimoto G. Free fatty acid receptors FFAR1 and GPR120 as novel therapeutic targets for metabolic disorders. *J Pharm Sci.* 2011;100:3594–3601.
- **36.** Hudson BD, Shimpukade B, Mackenzie AE, et al. The pharmacology of TUG-891, a potent and selective agonist of the free fatty acid receptor 4 (FFA4/GPR120), demonstrates both potential opportunity and possible challenges to therapeutic agonism. *Mol Pharmacol.* 2013;84:710–725.

5

6

ARTICLE IN PRESS

S.M. Sparks et al./Bioorganic & Medicinal Chemistry Letters xxx (2017) xxx-xxx

- 37. Medina JC, Houze JB. GPR40 (FFAR1) modulators. Annu Rep Med Chem. 2008;43:75–85.
- **38.** Sparks SM, Chen G, Collins JL, et al. Identification of diarylsulfonamides as agonists of the free fatty acid receptor 4 (FFA4/GPR120). *Bioorg Med Chem Lett.* 2014;24:3100–3103.
- Azevedo CM, Watterson KR, Wargent ET, et al. Non-acidic free fatty acid receptor 4 agonists with antidiabetic activity. J Med Chem. 2016;59:8868–8878.
- **40.** Lombardo M, Bender K, London C, et al. Discovery of benzofuran propanoic acid GPR120 agonists: from uHTS hit to mechanism-based pharmacodynamic effects. *Bioorg Med Chem Lett.* 2016;26:5724–5728.
- **41.** Burgos CH, Barder TE, Huang X, Buchwald SL. Significantly improved method for the Pd-catalyzed coupling of phenols with aryl halides: understanding ligand effects. *Angew Chem Int Ed.* 2006;45:4321–4326.