

# Total synthesis of viridifungin A†

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Viridifungin A, a member of amino alkyl citrate antibiotics from *Trichoderma viride*, was enantioselectively synthesized in naturally occurring form for the first time, employing regio- and stereoselective opening of the chiral glycidate with vinylmagnesium bromide and alkene cross metathesis of the citric acid core and hexadec-15-en-8-one as key steps.

Viridifungin A (**1**) was isolated from a strain of *Trichoderma viride* Pers., together with viridifungin B and C, by Harris *et al.*<sup>1</sup> The viridifungins have potent, broad spectrum antifungal activity which arises from the nanomolar level of inhibition of serine palmitoyltransferase.<sup>2</sup> These compounds were also found to inhibit Ras farnesyl transferase<sup>3</sup> as well as squalene synthase<sup>4</sup> *in vitro* at the micromolar level. The viridifungins have interesting structures consisting of a common citric acid moiety having a C-16 long chain and an aromatic amino acid residue such as tyrosine, phenylalanine, and tryptophan.<sup>1</sup> The absolute structure of viridifungin A (**1**) was determined by our enantiocontrolled synthesis of viridifungin A trimethyl ester (**2**).<sup>5</sup> However, we could not achieve the synthesis of viridifungin A (**1**) from trimethyl ester **2** because saponification of **2** caused decomposition, possibly *via* a retro-aldol process. Recently, Hiersemann *et al.*<sup>6</sup> reported an efficient synthesis of viridifungin A triester (**3**) based on ester dienolate [2,3]-Wittig rearrangement; however, they have not synthesized viridifungin A (**1**). We describe herein the first total synthesis of viridifungin A (**1**).

Taking into account the labile nature of **1** under basic conditions, we selected tri-*tert*-butyl ester **4** as a precursor with anticipation of its successful deprotection under mild acidic conditions in the final step of the synthesis. To access **4**, we envisaged the strategy which relies on alkene cross metathesis<sup>7</sup> between the citric acid core **5** and hexadec-15-en-8-one (**6**) as indicated in Fig. 1.

Our synthesis began with the stereoselective six-step preparation of *Z*-allylic alcohol **7** from 3-butyne-1-ol according to our previously established procedure (Scheme 1).<sup>5</sup> Katsuki–Sharpless asymmetric epoxidation<sup>8</sup> of **7** afforded epoxide **8** in 87% ee.<sup>9</sup> Successive Parikh–Doering oxidation,<sup>10</sup> NaClO<sub>2</sub> oxidation,<sup>11</sup> esterification using *N,N'*-diisopropyl-*O*-2-*tert*-butylisourea,<sup>12</sup> and removal of the THP protecting group converted **8** into glycidate **9** in good yield. Upon exposure of **9** to vinylmagnesium bromide in the presence of CuI, highly regio- and stereoselective nucleophilic opening of the epoxide occurred to give diol **10** exclusively. After protection of **10** as its cyclic carbonate,<sup>13</sup> removal of the *p*-methoxybenzyl protecting group afforded **11** which was then

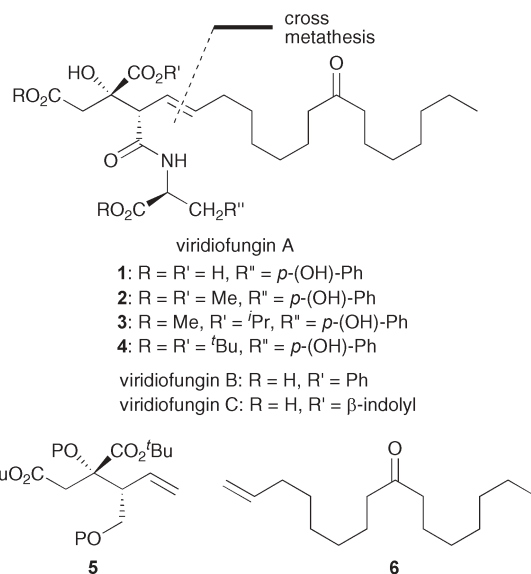
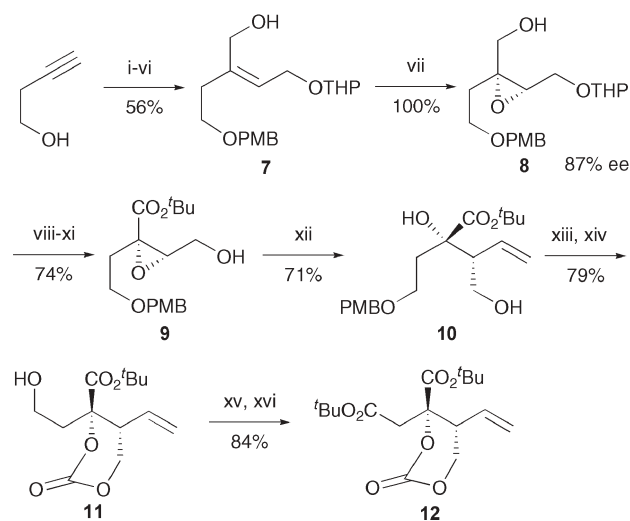


Fig. 1



**Scheme 1** Reagents and conditions: (i) *p*-(MeO)C<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>Cl, NaH, <sup>t</sup>Bu<sub>4</sub>NI, THF; (ii) <sup>t</sup>BuLi, (CH<sub>2</sub>O)<sub>n</sub>, THF; (iii) Red-Al, Et<sub>2</sub>O, 0 to 25 °C, then I<sub>2</sub>, -50 to 25 °C; (iv) PPTS, DHP, CH<sub>2</sub>Cl<sub>2</sub>; (v) <sup>t</sup>BuLi, CO<sub>2</sub>, Et<sub>2</sub>O, -78 °C, then MeI, DMF; (vi) DIBAL, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C; (vii) diethyl D-tartrate (0.3 eq.), Ti(O<sup>i</sup>Pr)<sub>4</sub> (0.25 eq.), <sup>t</sup>BuOOH (2 eq.), molecular sieves, CH<sub>2</sub>Cl<sub>2</sub>, -20 °C; (viii) SO<sub>3</sub>·pyridine, Et<sub>3</sub>N, DMSO, CH<sub>2</sub>Cl<sub>2</sub>; (ix) NaClO<sub>2</sub>, NaH<sub>2</sub>PO<sub>4</sub>, 2-methyl-2-butene, <sup>t</sup>BuOH–H<sub>2</sub>O (4:1); (x) *N,N'*-diisopropyl-*O*-2-*tert*-butylisourea, CH<sub>2</sub>Cl<sub>2</sub>; (xi) PPTS, MeOH; (xii) CH<sub>2</sub>=CHMgBr (10 eq.), CuI (1 eq.), THF, -26 °C; (xiii) triphosgene, pyridine, THF; (xiv) DDQ, CH<sub>2</sub>Cl<sub>2</sub>–H<sub>2</sub>O (20:1); (xv) H<sub>2</sub>CrO<sub>4</sub>, aq. acetone, -10 °C; (xvi) *N,N'*-diisopropyl-*O*-2-*tert*-butylisourea, CH<sub>2</sub>Cl<sub>2</sub>.

† Electronic supplementary information (ESI) available: experimental details. See <http://www.rsc.org/suppdata/cc/b5/b500660k/>

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**Table 1** Alkene cross metathesis of **12** with hexadec-15-en-8-one (**6**)<sup>a</sup>

Entry	Catalyst	Solvent	Temperature (°C)	Time (h)	Yield	
					<b>15</b> + Z-isomer (%) <sup>b</sup> [E:Z] <sup>c</sup>	<b>12</b> (%)
1	<b>13</b>	CH <sub>2</sub> Cl <sub>2</sub>	40	84	65 (86) [88:12]	24
2	<b>13</b>	Toluene	100	72	33 (65) [70:30]	49
3	<b>14</b>	CH <sub>2</sub> Cl <sub>2</sub>	40	16	48 (100) [85:15]	52
4	<b>14</b>	Toluene	100	72	57 (66) [89:11]	14

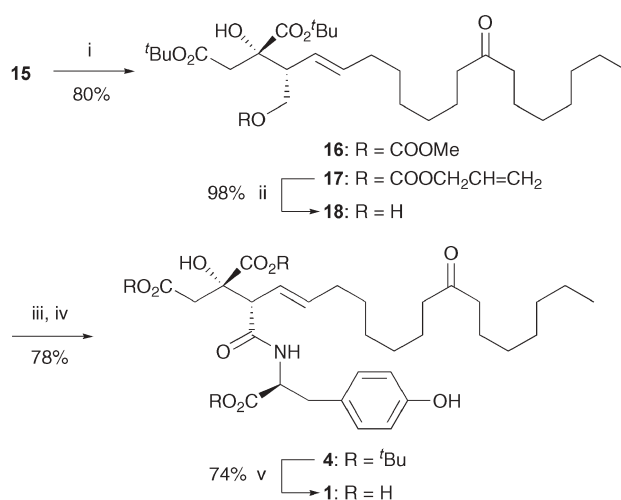
<sup>a</sup> All reactions were carried out using **12** (1 eq.), **13** or **14** (0.2 eq.) and **6** (2 eq.). <sup>b</sup> The yields in the parentheses were calculated based on recovered **12**. <sup>c</sup> Determined by <sup>1</sup>H NMR analysis.

subjected to Jones oxidation followed by *tert*-butyl esterification to give diester **12**.

The crucial alkene cross metathesis of **12** with hexadec-15-en-8-one (**6**)<sup>14</sup> was then examined under various conditions using ruthenium carbene complexes.<sup>15</sup> Although Grubbs' first generation catalyst<sup>16</sup> gave unsuccessful results, it was gratifyingly found that Grubbs' second generation catalyst **13**<sup>17</sup> and Hoveyda's catalyst **14**<sup>18</sup> effectively promoted the desired cross metathesis reaction (Table 1). Thus, upon reaction of **12** with 2 equivalents of **6** in the presence of 20 mol% of **13** in boiling CH<sub>2</sub>Cl<sub>2</sub> for 3.5 days, **15** and its Z-isomer were obtained in a ratio of 88:12 in 65% yield, together with unreacted **12** (24%) (entry 1). When this reaction was conducted using **14** with a shorter reaction time, both the E/Z-ratio and the total yield became lower [E:Z = 85:15 (48%)] although unreacted **12** was recovered without loss (entry 3). It was observed that catalyst **14** is more effective than catalyst **13** at higher temperature although both reactions were accompanied by appreciable decomposition (entries 2 and 4).

Having constructed the required carbon skeleton as **15**, we then investigated its conversion into viridifungin A (**1**). Methanolysis of **15** in the presence of K<sub>2</sub>CO<sub>3</sub> at -20 °C initially gave carbonate **16** but the prolonged reaction time caused decomposition of **16** rather than producing the desired diol **18**. However, this result allowed us to come up with the following transformations for the preparation of **18** (Scheme 2). Thus, **15** was first subjected to alcoholysis with K<sub>2</sub>CO<sub>3</sub> in allyl alcohol to give allyl carbonate **17** which, upon palladium-catalyzed reductive deallylation,<sup>19</sup> afforded **18** in excellent yield. Jones oxidation of **18** gave the corresponding carboxylic acid which was then directly reacted with L-tyrosine *tert*-butyl ester<sup>20</sup> using EDCI as a dehydrating agent in the presence of *N*-methylmorpholine and 1-hydroxybenzotriazole<sup>21</sup> to give viridifungin A tri-*tert*-butyl ester (**4**). Finally, cleavage of all *tert*-butyl ester groups in formic acid<sup>22</sup> followed by reverse phase column chromatography furnished (-)-viridifungin A (**1**) in good yield. The spectral data (<sup>1</sup>H and <sup>13</sup>C NMR, IR, MS) and specific rotation<sup>‡</sup> were identical with those reported for natural viridifungin A.<sup>1</sup>

In conclusion, we have accomplished the first total synthesis of (-)-viridifungin A from 3-buten-1-ol in 22 steps in 5% overall



**Scheme 2** Reagents and conditions: (i) K<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>=CHCH<sub>2</sub>OH, -20 °C; (ii) HCO<sub>2</sub>NH<sub>4</sub> (3 eq.), Ph<sub>3</sub>P (0.3 eq.), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.1 eq.), THF; (iii) H<sub>2</sub>CrO<sub>4</sub>, aq. acetone, -10 °C; (iv) L-tyrosine *tert*-butyl ester, Me<sub>2</sub>N(CH<sub>2</sub>)<sub>3</sub>N=C=NEt·HCl (EDCI), *N*-methylmorpholine, 1-hydroxybenzotriazole, DMF; (v) HCO<sub>2</sub>H.

yield. This synthesis also provides a flexible route to various viridifungin analogues required for biological testing.

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## Notes and references

<sup>‡</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> -15.2° (c 0.93, MeOH) {lit.<sup>1</sup> [ $\alpha$ ]<sub>D</sub><sup>25</sup> -18.2° (c 2.37, MeOH)}; <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD)  $\delta$  0.88 (t, *J* = 7.0 Hz, 3H), 1.28 (br s, 14H), 1.53 (m, 4H), 1.97 (m, 2H), 2.42 (t, *J* = 7.3 Hz, 3H), 2.43 (t, *J* = 7.3 Hz, 3H), 2.62 (d, *J* = 16.3 Hz, 1H), 2.86–2.92 (m, 3H), 3.10 (dd, *J* = 4.6, 14.1 Hz,

1H), 3.21 (d,  $J = 8.0$  Hz, 1H), 4.60 (dd,  $J = 4.8, 8.5$  Hz, 1H), 5.53 (m, 2H), 6.67 (d,  $J = 8.5$  Hz, 2H), 7.02 (d,  $J = 8.5$  Hz, 2H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  14.4, 23.7, 24.8, 24.9, 29.8, 29.9, 30.0, 30.2, 30.3, 32.9, 33.5, 37.5, 43.1, 43.5 (2), 55.1, 57.7, 80.0, 116.2 (2), 124.5, 128.7, 131.4 (2), 137.6, 157.4, 173.6, 173.9, 174.5, 175.8, 214.6; FTIR (neat) 3748, 3656, 3363, 1712, 1523, 1454, 1232  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  164 (100), 465, 511  $[(M - 2\text{H}_2\text{O} - \text{CO}_2)^+]$ ; HRMS (EI) calcd for  $\text{C}_{30}\text{H}_{41}\text{NO}_6$   $[(M - 2\text{H}_2\text{O} - \text{CO}_2)^+]$  511.2934, found 511.2922; HRMS (FAB, NBA) calcd for  $\text{C}_{31}\text{H}_{46}\text{NO}_{10}$   $[(M + \text{H})^+]$  592.3121, found 592.3110.

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