

# First synthesis of L-ascorbic acid (vitamin C) from a non-carbohydrate source

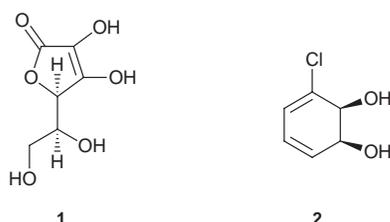
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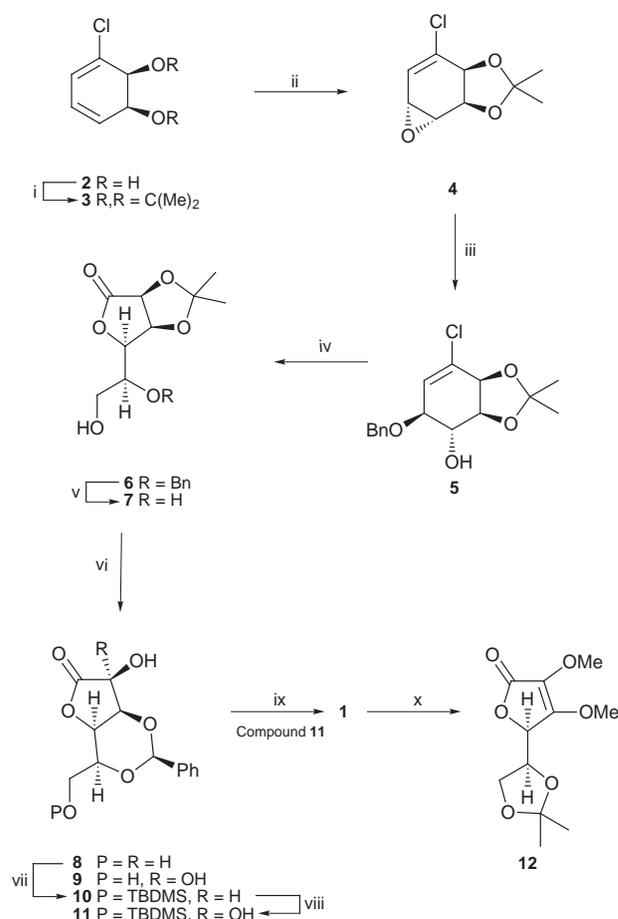
The enantiopure *cis*-1,2-dihydrocatechol **2**, which is obtained by microbial oxidation of chlorobenzene, has been converted, *via* 3,5-*O*-benzylidene-L-gulonolactone (**8**), into L-ascorbic acid (**1**).

L-Ascorbic acid (vitamin C, **1**), a biologically significant



reducing agent, is implicated in a variety of key physiological processes<sup>1</sup> including, for example, the production of collagen.<sup>2</sup> It is also considered to be important in the prevention of various chronic diseases such as cancer, cerebral apoplexy, diabetes, atopic dermatitis, myocardial infarction and AIDS.<sup>3</sup> The compound has been the subject of many synthetic studies<sup>4,5</sup> and is produced commercially from D-glucitol *via* a six step reaction sequence including an initial microbiological oxidation process.<sup>6</sup> In connection with other work, we required access to certain <sup>17</sup>O-, <sup>13</sup>C- and/or <sup>2</sup>H-labelled samples of L-ascorbic acid and none of the existing syntheses, all of which involve the use of carbohydrate-based starting materials, seemed suitable for our purposes. Consequently, we now report the first total synthesis of the title compound from non-carbohydrate sources.<sup>7,8</sup> The reaction sequence described herein should be especially amenable to the preparation of labelled ascorbates which could be used for probing their *in vivo* functions.

The present synthesis (Scheme 1) employs enantiopure (>99% ee) *cis*-1,2-dihydrocatechol **2**† (obtained by microbial oxidation of chlorobenzene) as starting material and was inspired by the seminal contributions of Hudlicky and co-workers who have pioneered the use of such microbial oxidation products in the chemoenzymatic synthesis of, *inter alia*, various monosaccharides.<sup>7,9</sup> Thus, the known<sup>10</sup> acetonide derivative, **3**, of diol **2** was converted into the previously reported<sup>10,11</sup> mono-epoxide **4** by standard methods. Reaction of this last compound with benzyl alcohol and a catalytic amount of trifluoromethanesulfonic acid (TfOH) afforded a stereochemically pure nucleophilic ring-cleavage product which, by analogy with related reactions of this epoxide,<sup>11</sup> is assigned structure **5**‡ {50%, mp 93–94 °C, [ $\alpha$ ]<sub>D</sub> +23.2 (*c* 0.2, CHCl<sub>3</sub>)}.<sup>12</sup> Confirmation of this assignment followed from the conversion of compound **5** into the known<sup>13</sup> 2,3-*O*-isopropylidene-L-gulonono-1,4-lactone (**7**). Thus, subjecting of chloroalkene **5** to reaction with ozone in methanol and subsequent *in situ* reduction of the intermediate hydroperoxide with sodium cyanoborohydride at pH 3 gave the L-gulonolactone derivative **6** {74%, mp 120–122 °C, [ $\alpha$ ]<sub>D</sub> +53.4 (*c* 0.8, CHCl<sub>3</sub>)}. Compound **6** could not be de-benzylated under standard conditions but by using transfer hydrogenolysis techniques<sup>14</sup> (with sonication) conversion into compound **7** {53%; mp 143–146 °C; lit.,<sup>13</sup> mp



**Scheme 1** Reagents and conditions: (i) (CH<sub>3</sub>)<sub>2</sub>C(CH<sub>3</sub>)<sub>2</sub> (1.5 mol equiv.), CH<sub>2</sub>Cl<sub>2</sub>, *p*-TsOH (1.5 mol%), 20 °C, 0.33 h; (ii) MCPBA, NaHCO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 18 °C, 5 h; (iii) C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>OH (1.05 mole equiv.), TfOH (5 mol%), CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 0.25 h; (iv) O<sub>3</sub> (excess), CH<sub>3</sub>OH, –78 °C → 20 °C then HCl (2 M solution in CH<sub>3</sub>OH), NaBH<sub>3</sub>CN (8 mole equiv.), 20 °C, 3 h; (v) cyclohexa-1,4-diene (10 mole equiv.), 10% Pd on C, CH<sub>3</sub>CH<sub>2</sub>OH, sonication, 18 °C, 8 h; (vi) C<sub>6</sub>H<sub>5</sub>CHO (4 mole equiv.), TfOH (5 mol%), 18 °C, 4 h; (vii) TBDMSCl (1.1 mole equiv.), imidazole (3.0 mole equiv.), DMAP (3 mol%), THF, 0 °C → 18 °C, 19 h; (viii) Dess–Martin periodinane (1.1 mole equiv.), CH<sub>2</sub>Cl<sub>2</sub>, 18 °C, 1.5 h; (ix) CH<sub>3</sub>CO<sub>2</sub>H–H<sub>2</sub>O (7:3), 70–75 °C, 4 h; (x) CH<sub>3</sub>COCl (5 mol%), (CH<sub>3</sub>)<sub>2</sub>CO, 18 °C, 2 h then K<sub>2</sub>CO<sub>3</sub> (5 mole equiv.), (CH<sub>3</sub>O)<sub>2</sub>SO<sub>2</sub> (5 mole equiv.), 56 °C, 3 h.

143–146 °C; [ $\alpha$ ]<sub>D</sub> +74 (*c* 0.5, EtOH); lit.,<sup>13</sup> [ $\alpha$ ]<sub>D</sub> +30 (*c* 2, EtOH)}§ was achieved. Reaction of acetonide **7** with neat benzaldehyde and TfOH as catalyst resulted in a *trans*-acetalisation reaction and formation of the benzylidene acetal **8** {47%; mp 188–190 °C; lit.,<sup>5b</sup> mp 188–189 °C; [ $\alpha$ ]<sub>D</sub> +57 (*c* 1.0, DMF); lit.,<sup>5b</sup> [ $\alpha$ ]<sub>D</sub> +61.1 (DMF)} which was identical, in all respects, with an authentic sample<sup>5,15</sup> prepared (in <10% yield) from commercially available L-gulonono- $\gamma$ -lactone. Theoretically, the acquisition of acetal **8** constitutes a formal total synthesis of

L-ascorbic acid since Crawford and Breitenbach<sup>5</sup> have converted the former compound into the latter *via* sequential oxidation and acetal hydrolysis steps. However, despite numerous attempts we were not able to effect the previously reported<sup>5</sup> manganese dioxide<sup>16</sup>-promoted oxidation of compound **8** to compound **9**. In order to examine alternate oxidants, the primary hydroxy group of diol **8** was protected as the corresponding TBDMS-ether so as to give compound **10** {74%; mp 176 °C;  $[\alpha]_{\text{D}} +55.5$  (*c* 0.4, CHCl<sub>3</sub>)}. After extensive experimentation with a variety of potential oxidants it was established that the Dess–Martin periodinane<sup>17</sup> effects smooth conversion of compound **10** into a variable mixture of ketone hydrate **11** and a regioisomer which is presumed to derive from silyl-group migration. Acetic acid promoted hydrolysis of this unstable mixture then provided L-ascorbic acid (**1**) in 66% yield (from compound **10**). For the purposes of comprehensive spectroscopic characterisation, compound (**1**) was converted, *via* a simple one-pot procedure,<sup>18</sup> into the stable derivative (**12**) {80%, mp 99–100 °C; lit.,<sup>18</sup> mp 100–101 °C;  $[\alpha]_{\text{D}} +9.1$  (*c* 0.6, CHCl<sub>3</sub>);  $[\alpha]_{\text{D}}$  (authentic sample) +9.3 (*c* 0.6, CHCl<sub>3</sub>)}. This latter material was identical, in all respects, with an authentic sample prepared from commercial L-ascorbic acid.

## Experimental

### Compound 6

A solution of chloroalkene **5** (100 mg, 0.32 mmol) in methanol (15 cm<sup>3</sup>) was cooled to –78 °C and a stream of ozone was bubbled through the solution until a blue colouration persisted. The solution was then purged with dioxygen for 0.25 h before being allowed to warm to room temperature. The resulting solution was concentrated to about one-third of the original volume then treated with one drop of methyl orange indicator followed by sufficient HCl (2 M solution in methanol) to establish pH 3. Sodium cyanoborohydride (160 mg, 2.5 mmol) was added in four roughly equal portions over a period of 3 h while ensuring pH 3 was maintained by appropriate additions of HCl. The reaction was quenched with acetone (10 cm<sup>3</sup>) then filtered through Celite<sup>TM</sup> and the filtrate concentrated under reduced pressure to give a light yellow oil. Subjection of this material to flash chromatography<sup>19</sup> (1:1 hexane–ethyl acetate elution) and concentration of the appropriate fractions (*R<sub>f</sub>* 0.2 in 3:2 hexane–ethyl acetate) afforded a white solid. Recrystallisation (CH<sub>2</sub>Cl<sub>2</sub>–hexane) of this material then gave the *title compound* **6** (73 mg, 74%) as white needles, mp 118–120 °C (Found: M<sup>+</sup> 308.1262; C, 62.0; H, 6.6. C<sub>16</sub>H<sub>20</sub>O<sub>6</sub> requires M<sup>+</sup> 308.1260; C, 62.3; H, 6.5%);  $\nu_{\text{max}}$  (KBr)/cm<sup>-1</sup> 3473 and 1788;  $\delta_{\text{H}}$  (300 MHz, CDCl<sub>3</sub>) 7.42–7.28 (5H, complex m), 4.85 (1H, d, *J* 11.4 Hz), 4.80 (2H, broadened s), 4.68 (1H, m), 4.64 (1H, d, *J* 11.4 Hz), 3.91 (1H, dd, *J* 12.1 and 2.7 Hz), 3.84 (1H, dt, *J* 8.2 and 3.1 Hz), 3.77 (1H, dd, *J* 12.1 and 3.1 Hz), 2.04 (1H, br s), 1.45 (3H, s, CH<sub>3</sub>), 1.37 (3H, s, CH<sub>3</sub>);  $\delta_{\text{C}}$  (75 MHz, CDCl<sub>3</sub>) 173.8 (C), 137.6 (C), 128.5 (CH), 128.0 (CH), 127.9(7) (CH), 114.1 (C), 80.3 (CH), 78.7 (CH), 75.9 (CH), 75.8 (CH), 73.4 (CH<sub>2</sub>), 61.3 (CH<sub>2</sub>), 26.7 (CH<sub>3</sub>), 25.8 (CH<sub>3</sub>); *m/z* (EI, 70 eV) 308 (8%), 250 [5, (M – CH<sub>3</sub>COCH<sub>3</sub>)<sup>+</sup>] and 91 (100, C<sub>7</sub>H<sub>7</sub><sup>+</sup>).

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### Notes and references

† Available commercially from Genencor International Inc., 925 Page Mill Road, Palo Alto, CA 94304-1013, USA.

‡ All new compounds had spectroscopic data (IR, <sup>1</sup>H and <sup>13</sup>C NMR, *m/z*) consistent with the assigned structure. Satisfactory combustion analysis data were obtained for new compounds.

§ The specific rotation recorded for an authentic sample of compound (**7**) { $[\alpha]_{\text{D}} +74$  (*c* 0.5, EtOH)} matched that obtained on our material thus suggesting that the value recorded in the literature<sup>13</sup> may be in error.

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