THE BROMO-COMPOUNDS OF THE TRUE SPONGE VERONGIA AEROPHOBA1

G. Cimino^{*}, S. De Rosa^{*}, S. De Stefano^{*}, R. Self^{\diamond} and G. Sodano^{*}

^{*}Istituto per la Chimica di Molecole di Interesse Biologico del C.N.R. Via Toiano n. 6, 80072 Arco Felice, Napoli, Italy

 $^{\circ}$ Food Research Institute, Colney Lane, Norwich, NR4 7UA, U. K.

Abstract. - Three novel bromo-compounds have been isolated from the sponge <u>Verongia aerophoba</u>. The sponge previously identified as <u>V. aerophoba</u>, containing related but different bromocompounds, was indeed the related species <u>V. cavernicola</u>.

Two species of Verongia genus sponges are widespread along the Italian coasts²: Verongia aerophoba (= Aplysina aerophoba Schmidt, 1862) and Verongia cavernicola Vacelet, 1959. A number of interesting new metabolites were identified³ from a sponge collected in the bay of Naples area and reported as Verongia aerophoba in the chemical papers. However direct comparison⁴ of this sponge with the true Verongia aerophoba, eventually collected from the area near Gallipoli, has shown the previously examined sponge to be the related species Verongia cavernicola.

We report now that the true sponge Verongia aerophoba contains several new bromo-compounds, along with some others previously isolated from Verongia cavernicola.

V. aerophoba (800 g, dry weight after extraction) was extracted with acetone. The extract was evaporated under reduced pressure and the resulting aqueous suspension was extracted sequentially with diethyl ether and n-butyl alcohol.

The ethereal extract (22.5 g) was chromatographed on a silica gel column with CHCl₃ and increasing amounts of CH₃OH to afford *inter alia* the dienone 1^5 (0.27 g), aeroplysinin-1 (2)⁶ (0.38 g), both previously isolated from *V. cavernicola* and other *Verongia* species, and the new compound 3a (0.97 g) which was proved to be isomeric with fistularin-3⁷ and accordingly named isofistularin-3.

Isofistularin-3, $C_{31}H_{30}$ Br₆N₄O₁₁, $[\alpha]_D^+$ 108° (2.75, CH₃OH), shows IR and UV absorptions practically identical with those reported for fistularin-3⁷. Acetylation of 3*a* (Ac₂O, Py, reflux) gave a crystalline tetraacetate (3*b*), m.p. 194-97°C $[\alpha]_D^+$ 132° (1.6, CHCl₃), whose 270 MHz ¹H-NMR spectrum shows minor but significant differences when directly compared with the spectrum of fistularin-3 tetraacetate⁸:protons Ha and Ha' show two distinct signals in the spectrum of 3*b* at δ 5.82 and 5.86, while in the spectrum of fistularin-3 tetraacetate a unique broad singlet (2H) at δ 5.86 was observed; the signals due to the methylene protons of the two isoxazole rings were centred at δ 3.09 (2H), 3.41 and 3.46 in 3b, while the same protons resonate at 3.06, 3.08, 3.45 and 3.47 in the spectrum of fistularin-3 tetraacetate.

Treatment of 3b with methanolic KOH afforded a phenol (4a),FAB-mass spectrum⁹ 1109, 1111, 1113, 1115, 1117, 1119, 1121 (MH^+), which on reacetylation gave 4b whose 270 MHz ¹H-NMR spectrum is indistinguishable from the spectrum reported⁶ for the analogous compound obtained by similar treatment of fistularin--3 tetraacetate.

From these data it is argued that 3a is isomeric with fistularin-3, the difference between the two compounds lying in the stereochemistry of one (or more) of the chiral centres.

The butanolic extract (7.9 g) gave some polar compounds. From this mixture the purification of two new bromo-compounds, aerophobin-1 (5a) and -2 (8a), was achieved by a multiple-step procedure including chromatography on Sephadex LH-20 (CH₃OH) and silica gel (CHCl₃-CH₃OH).

The less polar compound, aerophobin-1 (δa ; 0.25 g), $C_{15}H_{16}N_4O_4Br_2$, FAB-mass spectrum⁹ 475, 477, 479 (MH⁺), $\left[\alpha\right]_D$ + 187° (2.0, CH₃OH), has a very simple ¹H-NMR spectrum^{1°} in which, besides the readily assigned signals^{7,11} due to the spirocyclohexadienylisoxazole moiety, there are two triplets at δ 3.67 and 2.99 and two singlets at δ 6.85 and 7.59 which suggest, in conjunction with the elemental composition of the molecule, the presence of a histamine residue.

Acetylation (Ac₂O, Py; r.t.) afforded a crystalline monoacetyl derivative 5b, m.p. 164-67°C, FAB-mass spectrum 517, 519, 521 (MH⁺), whose ¹³C-NMR spectrum (Table) when compared with the spectrum of the acetyl derivative (6) of the previously isolated aerothionin¹¹, which is the major bromo-compound of *V. cavernicola* and is absent in the extracts of *V. aerophoba*, confirmed the presence of the spiro-cyclohexadienylisoxazole moiety. Hydrolysis of 5a (6N HCl; reflux) afforded histamine (7) confirming the structure of aerophobin-1 as 5a.

The more polar compound, aerophobin-2 ($\beta\alpha$; 0.67 g), $C_{16}H_{19}N_5O_4Br_2$, FAB-mass spectrum 504, 506, 508 (MH⁺), $\left[\alpha\right]_D$ + 139⁰ (1.9, CH₃OH), displays a ¹H-NMR spectrum¹² similar to that of 5α , the major differences lying in the presence of an additional methylene signal at δ 2.01 and in the absence of the C-2' proton signal of the imidazole residue.

Acetylation of βa with acetic anhydride and pyridine affords a diacetate wich cannot be purified chromatographically from a persistent contaminant. However, acetylation of βa with acetic anhydride/pyridine in acetic acid (5-fold excess) at r.t., afforded the monoacetate βb in good yields, which was successfully purified by conventional silica gel chromatography. Also in this case the ¹³C-NMR spectrum (Table) confirmed the presence of the spirocyclohexadienylisoxazole moiety; hydrolysis of βa (6N HCl; reflux) yielded 2-amino-homoistamine (β), identified by



comparison with an authentic sample¹³. ϑ could be linked in aerophobin-2 through either the C-2' and the side chain amino groups; however, the chemical shift of C-10 in the ¹³C-NMR'spectrum of ϑb (Table), very close to the value of the corresponding carbon in the spectrum of aerothionin acetate (ϑ), strongly suggests the arrangement reported in $\vartheta a-b$.

Isofistularin-3 (3a) is cytotoxic in vitro (KB cells), the effective dose being 4 μ g/ml.

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- We thank Prof.F.J.Schmitz for the generous gift of a sample of fistularin-3 tetraacetate.
- 9. Details on the fragmentation pattern will be reported elsewhere.
- 10. δ (CDCl₃/CD₃OD): 4.26 (1H; s; Ha); 6.34 (1H; s; Hb); 3.77, 3.93 (2H; d, d, J 7.5 Hz; Hc; Hd); 3.67 (2H; t, J 7 Hz; He); 2.99 (2H; t, J 7Hz; Hf); 6.85 (1H; s; Hm); 7.59 (1H; s; Hn); 3.86 (3H; s; OCH₃).
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- 12. δ_(CD₃OD): 4.14 (1H; s; Ha); 6.35 (1H; s; Hb); 3.20, 3.85 (2H; d, d, J 17.5 Hz; Hc, Hd); 3.43 (2H; t, J 7 Hz; He); 2.01 (2H; t, J 7 Hz; Hf); 2.67 (2H; t, J 7 Hz; Hg); 6.47 (1H; s; Hm); 3.79 (3H; s; OCH₃).
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