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Synthesis of the key intermediate of ramelteon

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Abstract

Asymmetric conjugated addition of allylcopper reagents derived from an allyl Grignard reagent and CuBr·Me₂S to chiral α , β unsaturated *N*-acyl oxazolidinones has been achieved. The synthetic procedure was applied to the preparation of the key intermediate of the novel nonbenzodiazepine hypnotic drug, ramelteon.

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Keywords: Ramelteon; Asymmetric Michael addition; Synthesis

Ramelteon, marketed as Rozerem by Takeda Pharmaceuticals North America, is the first in a new class of sleep agents that selectively binds to the MT1 and MT2 receptors in the suprachiasmatic nucleus (SCN) instead of binding to GABA_A receptors, such as occurs with zolpidem, eszopiclone, and zaleplon [1]. Ramelteon has been approved by the US Food and Drug Administration (FDA) for long-term use for the treatment of insomnia characterized by the difficulty of sleep onset. It is also the first and only prescription sleep medication that is not designated as a controlled substance by the US Drug Enforcement Administration (DEA).

Ramelteon has a unique chemical structure, comprising a three-ring system with an asymmetric center at the benzylic position. Due to the challenging structure, ramelteon has been a popular compound for synthetic chemists. (S)-N-(2-(6-Methoxy-2,3-dihydro-1H-inden-1-yl)ethyl)propionamide **1** (Fig. 1) is the key intermediate for the synthesis of ramelteon. So far, the reported synthesis of compound **1** has mainly been based on catalytic asymmetric hydrogenation [2,3] or resolution [4] of the corresponding racemic mixtures involving difficult procedures. However, known synthetic methods about compound **1** have many defects, involving the use of expensive metal catalysts and phosphorus ligands, or low yield, or low *ee* value. As a part of our research program, we have directed our efforts towards a more practical and efficient strategy to synthesize compound **1** using simple starting materials and reactions.

After having reviewed lots of literature, we found that conjugated addition of various nucleophiles to α , β unsaturated acid derivatives attached to a chiral auxiliary is an important methodology in asymmetric synthesis. For example, oxazolidinones, which are easily obtained, have been extensively used in related asymmetric fields [5]. Thus, we expected to introduce a stereogenic center *via* an asymmetric Michael addition using (*S*)-4-benzyloxazolidin-2-one **2** as a chiral auxiliary. Herein, we report our synthetic approach to the enantiomerially pure intermediate **1** based on a copper-assisted conjugate addition of a Grignard reagent to chiral α , β -unsaturated *N*-acyl oxazolidinones.

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Since we were not able to find any chiral 1-substituted indans with a known absolute configuration prepared from 3substituted indanones in the literature, we focused our attention on developing a novel synthetic strategy that would allow for the synthesis of the enantiopure intermediate **1** with a predetermined absolute configuration. The synthetic procedure we designed consisted of the decarbonylation of chiral 3-substituted indanones (Fig. 2). Optically active 3-



Scheme 1. (a) Malonic acid, piperidine, pyridine. (b) $(COCl)_2$, DMF, CH_2Cl_2 ; (*S*)-4-benzyloxazolidin-2-one **2**, LiCl, Et₃N, CH_2Cl_2 . (c) Allylmagnesium chloride, CuBr. Me₂S, THF. (d) LiAlH₄, THF. (e) AcCl, Et₃N, CH_2Cl_2 . (f) NaIO₄, KMnO₄, Me₂CO, H₂O. (g) $(COCl)_2$, DMF, CH_2Cl_2 ; SnCl₄, CH₂Cl₂. (h) Pd/C, H₂, H₂SO₄, EtOH. (i) MsCl, Et₃N, CH₂Cl₂; phthalimide potassium salt, DMF. (j) NH₂NH₂, EtOH. (k) $(CH_3CH_2CO)_2O$, NaOH, THF, H₂O.

substituted indanones with a known stereochemistry have been reported in the literature, and have been prepared using a classical cyclohydration from the corresponding chiral 3-substituted cinnamic acid, which can be obtained using asymmetric synthesis procedures with a high enantiomeric purity [6–8]. The synthesis of (S)-N-(2-(6-methoxy-2,3-dihydro-1H-inden-1-yl)ethyl)propionamide using this methodology is outlined in Scheme 1.

As shown in Scheme 1, the cinnamic acid 4 was easily obtained from 3-methoxybenzaldehyde 3 in a 95% yield, and was condensed with (*S*)-4-benzyloxazolidin-2-one 2 through cinnamoyl chloride to form the chiral unit 5. Then, the asymmetric conjugate addition of an allylcopper reagent derived from commercially available allylmagnesium chloride and CuBr·Me₂S (allylMgCl/Cu 1:1) to the chiral cinnamic unit 5 furnished the intermediate 6 with a high enantiomeric purity. High-pressure liquid chromatography (HPLC) analysis of the reaction mixture suggested a high diastereomeric excess (>99%). This was confirmed in the final step. Removal of the chiral auxiliary (LiAlH₄ in THF) yielded the alcohol 7, which was protected by the acetyl group to result in compound 8 in a 90% yield. Subsequent oxidation of the allyl compound with NaIO₄/KMnO₄ (6:1) gave the acid 9 in high yields without further purification. Treating 9 with oxalyl chloride, followed by a Lewis acid-promoted Friedel–Crafts acylation afforded the indanone 10. Then, pure indanol 11 was obtained through deprotection and reduction in a one-pot procedure with Pd/C catalytic hydrogenation in concentrated sulfuric acid in ethanol. Subsequent amination of the indanol 11 *via* a Gabriel synthesis gave the indanamine 12, which, after salification (4 M HCl/EtOH), yielded the indanamine hydrochloride 13. Finally, the reaction of this compound with propionic anhydride under alkaline conditions afforded the key target 1. HPLC analysis [2,9] revealed an *ee* > 99%. At last, all the prepared compounds gave satisfactory analytical data [10].

In conclusion, an efficient enantioselective synthesis of (*S*)-*N*-(2-(6-methoxy-2,3-dihydro-1*H*-inden-1-yl)ethyl)propionamide with an absolute configuration has been achieved. Our methodology appears to be suitable for preparing various chiral 3-substituted indanones and 1-substituted indan derivatives.

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- [10] 5: $[\alpha]_D^{20}$ +39.1 (c, 1.0, CHCl₃); mp 69–71 °C; ¹H NMR (500 MHz, CDCl₃): δ 2.86 (q, 1H, J = 9.5 Hz), 3.37 (dd, 1H, J = 3.1 Hz, J = 13.4 Hz), 3.85 (s, 3H), 4.22 (m, 2H), 4.81 (m, 1H), 6.96 (m, 1H), 7.14 (s, 1H), 7.24–7.36 (m, 7H), 7.90 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 165.1, 159.8, 153.4, 146.2, 135.8, 135.3, 129.8, 129.4, 128.9, 127.2, 121.3, 117.2, 116.6, 113.3, 66.1, 55.3, 55.2, 37.8. EI-MS: 337 ([M]⁺); HR-MS 337.1314 ([M]⁺, C₂₀H₁₉NO₄; Calcd. 337.1315). **6**: $[\alpha]_D^{20}$ +61.6 (*c*, 1.0, CHCl₃); mp 66–67 °C; ¹H NMR (500 MHz, CDCl₃): δ 2.45 (m, 2H), 2.66 (m, IH), 3.29 (dd, 2H, J = 4.0 Hz, J = 15.0 Hz), 3.20–3.41 (m, 2H), 3.79 (s, 3H), 4.00 (m, 1H), 4.07 (m, 1H), 4.51 (m, 1H), 5.02 (m, 2H), 5.70 (m, 1H), 6.74 (d, 1H, J = 8.0 Hz), 6.79 (s, 1H), 6.84 (d, 1H, J = 8.0 Hz), 7.16–7.33 (m, 6H). ¹³C NMR (100 MHz, CDCl₃): δ 172.8, 159.6, 153.4, 145.3, 136.0, 135.3, 129.3, 128.9, 127.3, 120.0, 116.9, 113.4, 113.3, 111.8, 60.1, 55.2, 55.1, 41.4, 40.9, 40.8, 37.8. EI-MS: 379 ([M]⁺); HR-MS 379.1783 ([M]⁺, C₂₃H₂₅NO₄; Calcd. 379.1784). 7: [α]_D²⁰ +0.3 (*c*, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 1.79 (m, 1H), 1.97 (m, 1H), 2.37 (t, 2H, J = 7.0 Hz), 3.47 (m, 1H), 3.54 (m, 1H), 3.80 (s, 3H), 4.97 (m, 2H), 5.66 (m, 1H), 6.72–6.78 (m, 3H), 7.21 (t, 1H, J = 8.0 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 159.6, 146.2, 136.6, 129.3, 120.0, 116.1, 113.6, 111.1, 60.8, 55.0, 42.2, 42.8, 38.5. HR-MS 245.1519 ([M+Na]⁺, C₁₄H₂₂O₂Na; Calcd. 245.1517). 8: [α]_D²⁰ +20.0 (c, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 1.86 (m, 1H), 1.99 (s, 3H), 2.35 (m, 1H), 1.99 (s, 2H), 2.35 (m, 1H), 2.38 (m, 2H), 2.72 (m, 1H), 3.80 (s, 3H), 3.87 (m, 1H), 3.96 (m, 1H), 4.97 (m, 2H), 5.65 (m, 1H), 6.70 (d, 1H, J = 1.4 Hz), 6.74 (m, 2H), 7.21 (t, 1H, J = 7.9 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 170.9, 159.7, 145.6, 136.4, 129.4, 119.9, 116.3, 113.6, 111.3, 62.8, 55.1, 42.5, 41.1, 34.3, 20.8. EI-MS: 248 ([M]⁺); HR-MS 248.1411 ([M]⁺, C₁₅H₂₀O₃; Calcd. 248.1412). 9: [α]_D²⁰ +11.0 (*c*, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): § 1.87 (m, 1H), 1.96 (s, 3H), 2.01 (m, 1H), 2.65 (d, 2H, J = 7.4 Hz), 3.20 (m, 1H), 3.79 (s, 3H), 3.85 (m, 1H), 3.99 (m, 1H), 6.72 (s, 1H), 6.75 (m, 2H), 7.21 (t, 1H, J = 7.9 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 177.1, 171.1, 159.7, 144.0, 129.7, 119.5, 113.3, 111.8, 62.2, 55.0, 41.0, 38.5, 34.6, 20.7. HR-MS 289.1051 ([M+Na]⁺, $C_{14}H_{18}NaO_5$; Calcd. 289.1052). **10**: $[\alpha]_D^{20} - 26.0$ (*c*, 1.0, CHCl₃); mp 67–68 °C; ¹H NMR (500 MHz, CDCl₃): δ 1.79 (m, 1H), 2.05 (s, 3H), 2.24 (m, 1H), 2.41 (dd, 1H, *J* = 3.4 Hz, *J* = 19.0 Hz), 2.86 (dd, 1H, *J* = 7.6 Hz, *J* = 19.0 Hz),

3.39 (m, 1H), 3.88 (s, 3H), 4.18 (m, 2H), 6.91 (m, 2H), 7.67 (t, 1H, J = 4.6 Hz). ¹³C NMR (100 MHz, CDCl₃): δ 203.7, 170.9, 165.4, 160.5, 130.0, 125.4, 115.5, 108.9, 62.6, 55.7, 43.1, 35.3, 34.7, 20.9. EI-MS: 248 ([M]⁺); HR-MS 248.1047 ([M]⁺, C₁₄H₂₆O₄; Calcd. 248.1049). 11: [α]_D²⁰ –15.0 (c, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 1.70 (m, 2H), 2.05 (s, 1H), 2.13 (m, 1H), 2.32 (m, 1H), 2.76 (m, 1H), 2.85 1H), 3.21 (m, 1H), 3.78 (s, 3H), 3.80 (m, 2H), 6.71 (d, 1H, J = 8.2 Hz), 6.77 (s, 1H), 7.26 (d, 1H, J = 8.2 Hz), ¹³C NMR (100 MHz, CDCl₃); δ 158.7, 148.6, 135.8, 124.9, 112.1, 109.3, 61.5, 55.4, 41.7, 37.8, 32.7, 30.6. EI-MS: 192 ([M]⁺); HR-MS 215.1040 ([M+Na]⁺, C₁₂H₁₆NaO₂; Calcd. 215.1043). 12: ¹H NMR (500 MHz, CDCl₃): δ 1.83 (m, 2H), 2.24 (s, 1H), 2.41 (m, 1H), 2.78 (m, 1H), 2.86 (m, 1H), 3.11 (m, 1H), 3.77 (s, 3H), 3.81 (m, 2H), 6.68 (dd, 1H, J = 1.8 Hz, 8.2 Hz), 6.79 (s, 1H), 7.08 (d, 1H, J = 8.2 Hz), 7.71 (m, 2H), 7.84 (m, 2H). ¹³C NMR (100 MHz, $CDCl_3$: δ 168.4, 158.7, 147.8, 135.8, 133.9, 132.2, 124.9, 123.2, 112.6, 108.9, 55.4, 42.6, 36.5, 33.4, 32.4, 30.6. **13**: $[\alpha]_D^{20} - 27.0$ (c, 0.20, 1.20, H₂O); mp 174–175 °C; ¹H NMR (500 MHz, DMSO-d₆): δ 1.66 (m, 2H), 2.09 (m, 1H), 2.24 (m, 1H), 2.70 (m, 1H), 2.80 (m, 3H), 3.12 (m, 1H), 3.71 (s, 3H), 6.71 (d, 1H, J = 8.2 Hz), 6.76 (s, 1H), 7.11 (d, 1H, J = 8.2 Hz), 8.18 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆): δ 158.3, 147.4, 135.0, 124.9, 112.2, 109.1, 55.1, 41.6, 37.1, 31.9, 31.7, 29.9. 1: [α]_D²⁰ -10.0 (*c*, 0.20, EtOH); mp 76-77 °C; ¹H NMR (500 MHz, CDCl₃): δ 1.15 (t, 3H, J = 7.5 Hz), 1.60 (m, 1H), 1.70 (m, 1H), 2.02 (m, 1H), 2.19 (q, 2H, J = 7.5 Hz), 2.32 (m, 1H), 2.76 (m, 1H), 2.85 (m, 1H), 3.11 (m, 1H), 2.16 (m, 1H), 2.17 (m, 1H), 2.19 (m, 1H), 2.19 (m, 2H), 2.19 (m, 2H), 2.10 (1H), 3.41 (m, 2H), 3.79 (s, 3H), 5.48 (s, 1H), 6.71 (dd, 1H, J = 2.0 Hz, 8.5 Hz), 6.75 (s, 1H), 7.11 (d, 1H, J = 8.0 Hz). ¹³C NMR (100 MHz, 100 MHz), 100 MHz, DMSO-d₆): δ 173.7, 158.7, 148.1, 135.8, 124.9, 112.3, 109.2, 55.5, 42.7, 37.9, 34.8, 32.5, 30.6, 29.8, 9.9. EI-MS: 247 ([M]⁺); HR-MS 247.1572 ([M]⁺, C₁₅H₂₁NO₂; Calcd. 247.1571). The enantiomeric excess of (S)-1 was determined by HPLC as >99.9% [column, CHIRALPAK AS-H $(4.6 \text{ mm} \times 250 \text{ mm})$, room temperature; eluent, hexane-2-propanol-trifluoroacetic acid (90:10:0.1); flow rate, 1.0 mL/min; detect, 290 nm; $t_{\rm R}$ of (S)-1, 30.7 min; t_{R} of (R)-1 (enantiomer of (S)-1), 37.1 min].