CYCLOFUNCTIONALISATION REACTIONS OF EPOXYALCOHOL DERIVATIVES. 3. CYCLISATION-ACYL MIGRATION OF N-BENZOYLCARBAMATES TO STEREODEFINED OXAZOLIDINONES. A NEW, DIASTEREOSPECIFIC ROUTE TO THIAMPHENICOL.

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<u>Abstract</u>: N-Benzoylcarbamates formed in situ from 2,3-epoxyalcohols and PhCONCO undergo clean N to C-2 cyclisation followed by N to O acyl migration on treatment with catalytic sodium imidazolide or other bases. Subsequent benzoate cleavage (NaOMe) is accompanied by equilibration of the N-unsubstituted oxazolidinones; cleavage without significant isomerisation is achieved with MeLi or Zn(BH4)2. This methodology is applied in a diastereospecific, 6-step conversion of methyl 4-bromophenyl sulfone to racemic Thiamphenicol.

In recent papers , we have described methods for intramolecular delivery of carbon¹ and oxygen² to C-2 of epoxy-

m alcohol derivatives under basic conditions where strict $m S_N2$ behaviour was obtained . To deliver a nitrogen atom , two

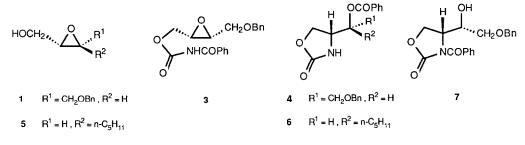
groups³ have reported the cyclisation of anions derived from N-benzylcarbamates . This route has been used to

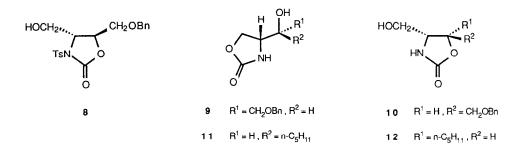
prepare aminopolyols⁴, the sequence being completed by oxazolidinone hydrolysis followed by reductive cleavage

(H₂/Pd or Na/NH₃) of the benzyl group ; attempts to cyclise unsubstituted or silylated carbamates failed .

In connection with studies on beta-lactam and chloramphenicol antibiotics, we required a method which would effect nitrogen delivery under less basic conditions, and also avoid problems associated with reductive debenzylation. In this paper, we report the base catalysed cyclisation/isomerisation of 2,3-epoxyalcohol N-benzoylcarbamates, and a new route to Thiamphenicol based on this chemistry.

Treatment of the <u>cis</u> epoxyalcohol (1) with PhCONCO⁵ (2, 1.05 eq, THF, 0°) rapidly gave a solution of (3) to which was added 10% w/v DMSO and 0.1 to 0.2 eq of NaH and imidazole. After 24h at RT, workup and chromatography provided benzoate (4)⁶ in 86% yield. A similar sequence on <u>trans</u> epoxyalcohol (5) gave (6)⁶ (79%, mp 91-93°). The cyclisation/ rearrangement of (3) to (4) also proceeded efficiently in MeCN with potassium carbonate and 5 mol % of methyl trioctylammonium chloride.





Presumed intermediate (7) was not detected during these reactions, suggesting rapid N to O migration of the activated benzoyl group. When an N-sulfonyl group was present, oxazolidinone equilibration occurred : reaction of (1) with 4-TsNCO followed by Na imidazolide gave 83% of (8), arising from epoxide opening followed by C=O migration to afford the more stable trans disubstituted product.

Standard ester cleavage (K_2CO_3 -MeOH, 25°, 4h) of (4) provided quantitatively a readily separable mixture of regioisomers (9) and (10) in 1:6 ratio . Oxazolidinone equilibration proceeded at a rate comparable to ester cleavage , although a higher proportion of (9) was found (TLC) at partial conversion of (4) .Similar treatment of (6) produced a 3:2 mixture of $(11)^6$, mp 75-77° and $(12)^6$, mp 95-96° . Cleavage of (6) to (11) exclusively occured with CH₃Li(4 eq, -70°, 1 min) or Zn(BH₄)₂ (THF , 25°) .The structures of the isomeric oxazolidinones were evident from the ¹H NMR spectra⁶ of the alcohols and their derived adducts with CCl₃CONCO . As expected , the primary alcohols were more polar ; benzoylation of the less polar isomers regenerated samples of (4) and (6) .

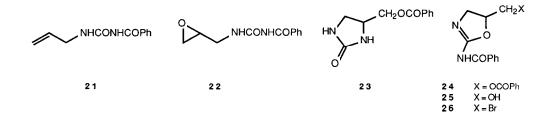
The <u>Scheme</u> shows the use of this methodology in a new , diastereospecific synthesis of the antibiotic Thiamphenicol (19) in racemic form . Syntheses of Chloramphenicol-type antibiotics frequently rely on either a diastereoselective condensation of ArCHO with a glycine anion equivalent⁷(followed by ester reduction) , or a diastereoselective Al(Oi-Pr)₃ reduction of the appropriate 2-acetamido-3-hydroxypropiophenone⁸. For this conceptually different route , methyl-4-bromophenyl sulfone (13)⁹ was converted in good yield to the arylpropynol (14) by a known Cu(I) / Pd(0) - catalysed process¹⁰ using CuI and Cl₂Pd(PPh₃)₂ in Et₂NH . Reduction with Zn-HOAc in MeOH¹¹ followed by mCPBA oxidation then gave the racemic¹² cis epoxyalcohol (16)⁶ , mp 72-74°. Reaction with PhCONCO followed by NaH-imidazole (THF-DMSO , 48h at 25°) gave (17)¹³, mp 181-183°, which was stirred with NaOMe (0.05 eq) in MeOH , resulting in the precipitation of pure (18), mp 165-167°. (18) is a convenient substrate for -CH₂OH modification¹⁴, and was converted to a sample of racemic Thiamphenicol by acidic hydrolysis followed by reaction with methyl dichloroacetate .

Br CCCH₂OH CH₂OH CH2OH b а с (73%) (86% from 14) SO₂Me SO₂Me SO₂Me SO₂Me (87%) d 13 14 15 16 ОН OCOPh HOCH2 ... н e HN n CI₂CHCONH NH (90%) SO₂Me 0 X=OH 19 18 17 X=F 20

Scheme

REAGENTS a : HCCCH₂OH , HNEt₂ , cat.Cul + PdCl₂(PPh₃)₂ , 55° , 5h. b : Zn - HOAc , MeOH , 65° , 4h. c : mCPBA , CH₂Cl₂ , 25° , 8h. d : PhCONCO - THF , 0° , 0.25h ; 0.1eq NaH / imidazole , THF - DMSO , 25° , 48h. e : 0.05 eq NaOMe , MeOH , 25° , 6h.

This sequence illustrates the application of the methodology to the preparation of a relatively base-sensitive, benzylic aminodiol, where the benzylcarbamate method could not be used. Preliminary studies indicate that imidazolidinones (and derived 1,2-diamines) may be obtained in an analogous manner: reaction of allylamine with (2) gave (21), which was epoxidised (mCPBA, CH_2Cl_2) to (22). Treatment of this material with NaH-imidazole afforded (23)⁶ as the major product, with a lesser quantity of (24)⁶, from O-attack and intermolecular acyl transfer. Exclusive attack by the urea oxygen took place under neutral conditions: protic cyclisation of (21) [0.2 eq MsOH, CH_2Cl_2 , 0.5h] gave only (25)⁶, and bromomethyl compound (26)⁶ was the sole product from (21) and N-bromosuccinimide.



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- 5. Prepared from PhCONH₂ and (COCl)₂ according to A. J. Speziale and L. R. Smith , <u>J. Org. Chem.</u>, 1962, <u>27</u>, 3742. Stored at 5° after vacuum distillation. All new compounds gave correct microanalyses. Selected PMR values (ppm, in CDCl₃ unless otherwise indicated) : (4) : $3.18(d_2,J=4.5)$, 3.5-3.9(m,3), 3.96(s,2), 4.58(m,1) and 5.43,s,1, exch. D_2O). (6) : 0.85(t,3), 1.2-1.4(m,6), 1.5-1.8(m,2), 4.08(m,1), 4.3-4.6(m,2), 5.22(m,1) and $6.24(s,1,exch. <math>D_2O)$ (8) : 2.36(s,3), $2.8(br.s,1,exch. <math>D_2O)$, $3.52(d_2,J=3.0)$, $3.7-4.2(br.m,becomes ABX,J=3.3,4.5 and 12 after <math>D_2O$ exch.), 4.35(s,2), 4.3(m,1) and 4.61(br.q,J=3). (11) : 0.88(t,3), 1.2-1.6(m,8), $3.55(br.s,1,exch. <math>D_2O)$, $3.69(m,1, sharpened by <math>D_2O$ exch.), 3.84(m,1), 4.3-4.5(m,2) and $6.65(s,1,exch. <math>D_2O)$. (12) : 0.88(t,3), 1.2-1.4(m,6), 1.5-1.9(m,2), $3.0(br.s,1,exch. <math>D_2O)$, 3.7(m,2), 3.80(m,1), 4.64(m,1) and $6.43(s,1,exch. <math>D_2O)$. (16) : $1.85(br.s,1,exch. D_2O)$, 3.07(s,3), 3.4-3.6(m,3) and 4.25(d,1,J=4.0). (18) [in DMSO-d_6] : 3.21(s,3), $3.35(s,1,exch. <math>D_2O)$, 3.45-3.6(m,3), $5.2(br.s,1,exch. <math>D_2O)$ and 5.44(d,1,J=4.5). (23) : 3.36(d,1,J=5.5 and 11), 3.71(t,1,J=11), 4.1-4.5(m,3), $5.14(br.s,1,exch. <math>D_2O)$, and $5.37(br.s,1,exch. <math>D_2O)$. (24) : 3.7-4.2(m,2), 4.54(br.d,2), 5.05(m,1), 7.2-7.6(m,6), 8.00 (m,2), 8.23(m,2) and $9.6(br.m,1,exch. <math>D_2O)$. (25) [in DMSO-d_6] : 3.4-3.9(m,4), 4.75(m,1), $5.19(t,1,J=6,exch. D_2O)$, 7.3-7.55(m,3), 8.05(m,2) and $9.5(br.s,1,exch. <math>D_2O)$. (26) : 3.5-3.75(ABX,2,J=4.5,8.0 and 11), 3.83(dd,1,J=6.8 and 9.2), 4.06(t,1,J=9.2) and 5.01(m,1).
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- 12. Hindered, <u>cis</u> alcohols including Z-3-Aryl-2-propenols such as (15) represent one of the few classes of allylic alcohols which give epoxides with relatively low (60-70%) e.e. when subjected to the Sharpless asymmetric epoxidation.
- 13. On occasion, small amounts of the isomeric ester (18-O-benzoate) were produced, presumably via intramolecular acyl transfer reactions. Separation was unnecessary for the subsequent hydrolysis / equilibration to (18).
- 14. The 3-fluoro-3-deoxy analog (20) is a potent, broad spectrum compound : T. L. Nagabhushan, D. Kandasamy, H. Tsai, W. N. Turner and G. H. Miller, <u>Current Chemotherapy and Infectious Disease Proceedings of the 11th ICC</u> and 19th ICAAC, American Society of Microbiology, <u>1980</u>, p442. See also T. W. Schafer, E. L. Moss, Jr., T. L. Nagabhushan and G. H. Miller, <u>Ibid.</u>, <u>1980</u>, p444.

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