

reasons for this speculation are the distinct tetrahedral distortions in the basal planes of the square pyramids, which are quite common in this type of copper surrounding and the fact that Cu(I) is known to interact with  $\pi$  systems but not Cu(II). It is hoped that additional experimental observations which can be expected from our structure determinations of Cu(II) (*l*-tyrosine)<sub>2</sub> and the peptide glycyl-*l*-leucyl-*l*-tyrosine will throw more light on these questions.

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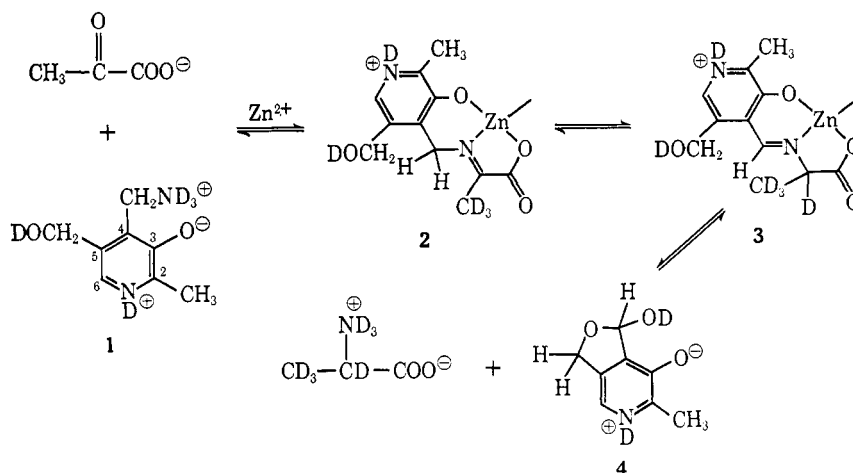
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### Detection and Identification of Intermediates and Products of a Nonenzymatic Transamination Reaction by Proton Resonance

Sir:

We are currently investigating by pmr the widely quoted mechanism<sup>1</sup> for transamination of  $\alpha$ -amino acids and  $\alpha$ -keto acids involving pyridoxal cofactors and metal ions<sup>2</sup> in aqueous solutions. Systems studied include 0.1 *M* pyridoxal-0.1 *M* alanine and 0.1 *M* pyridoxamine-0.1 *M* pyruvate in the presence and absence of 0.05 *M* Zn<sup>2+</sup> over the pD range 1-13.<sup>3</sup> The purpose here is to demonstrate the direct detection and identification of the species implicated in that part of the over-all transamination sequence represented by the reaction pyridoxamine + pyruvate  $\rightleftharpoons$  pyridoxal + alanine occurring at pD  $\sim$ 7. Previous pmr studies<sup>4</sup> serve to identify the signals and predominant forms of pyridoxamine (1) and pyridoxal (4) at this pD.



In the pyridoxamine-pyruvate-zinc system transamination in D<sub>2</sub>O solution occurs rapidly but can be conveniently followed by pmr. Spectra taken before and after reaction ( $\sim$ 20 min) are given in Figure 1; those recorded at intermediate times are superpositions of the two shown. Interpretation of Figure 1a requires

(1) D. E. Metzler, M. Ikawa, and E. E. Snell, *J. Am. Chem. Soc.*, **76**, 648 (1954).

(2) For extended discussions of the transamination reaction, cf. T. C. Bruice and S. J. Benkovic, "Bioorganic Mechanisms," Vol. II, W. A. Benjamin, Inc., New York, N. Y., 1966, Chapter 8; B. M. Guirard and E. E. Snell, "Comprehensive Biochemistry," Vol. 15, M. Florkin and E. H. Stoltz, Ed., Elsevier Publishing Co., New York, N. Y., 1964, Chapter V.

(3) pD = pH + 0.40: P. K. Glasoe and F. A. Long, *J. Phys. Chem.*, **64**, 188 (1960).

(4) O. A. Gansow and R. H. Holm, *Tetrahedron*, **24**, 4477 (1968).

brief discussion of the spectra of solutions containing pyridoxamine and pyruvate (both at 0.1 *M*) in the absence of Zn<sup>2+</sup>. The solutions do not transaminate rapidly at room temperature, but their spectra reveal formation of  $\alpha$ -pyridoximinopyruvate (ketimine) at pD  $\sim$ 7-10. Ketimine formation results in labilization of the pyruvate methyl protons and substantial H-D exchange in D<sub>2</sub>O solution. At pH 7.4 in H<sub>2</sub>O solution a new signal appears at 50 cps corresponding to the condensed pyruvate methyl group.<sup>5</sup> In D<sub>2</sub>O (pD 7.4) new signals are observed at 658, 276, and 116 cps, near to those of 6-H, 4-CH<sub>2</sub>, and 2-CH<sub>3</sub> of free pyridoxamine, respectively, and are assigned to the corresponding protons of the ketimine. Extent of ketimine formation and ketimine chemical shifts are functions of pD; it suffices here to note that the former reaches a maximum of  $\sim$ 30% (by signal integration) at pD 9.8 and decreases to zero in strongly basic solutions (pD  $>$ 11.5). Before transamination is observable, pmr spectra of D<sub>2</sub>O solutions containing Zn<sup>2+</sup> are rather similar to the metal-free solutions at a given pD except that the ketimine 2-CH<sub>3</sub> signal is broadened and shifted up-field (cf. Figure 2), indicating complexation of Zn<sup>2+</sup> and slow exchange between coordinated and free ketimine, and the per cent solute in the ketimine form is more than 20% greater than in the metal-free solutions. All features of the pmr spectra before transamination are consistent with formation of the labile 1:1 and 1:2 Zn(II):ketimine complexes 2 with the concentration of the latter increasing with increasing pD.

The pmr spectrum of the pyridoxamine:pyruvate: Zn<sup>2+</sup> solution after transamination is shown in Figure 1b. In order to identify reaction products a complete pmr study of D<sub>2</sub>O solutions of pyridoxal and alanine

(0.1 *M*), both metal-free and with Zn<sup>2+</sup> added, has been carried out. In the metal-free solutions formation of N-pyridoxylidenealanine (aldimine) was detectable at pD  $\geq$ 7 by the appearance of the characteristic low-field azomethine proton signal at  $\sim$ 760 cps. In addition, two 6-H and 2-CH<sub>3</sub> signals and two alanine methyl doublets and CH quartets were observed. Features due to free pyridoxal, which exists mainly in the dipolar hemiacetal form 4 at pD 4.4-9,<sup>4</sup> and free alanine were readily identified; those remaining must

(5) All chemical shifts refer to 100 Mc and a *t*-butyl alcohol internal standard; the chemical shift of this signal is close to those observed for  $\alpha$ -oximinopropionic acid and acetaldoxime.

(6) M. J. O'Connor, R. E. Ernst, J. E. Schoenborn, and R. H. Holm, *J. Am. Chem. Soc.*, **90**, 1744 (1968).

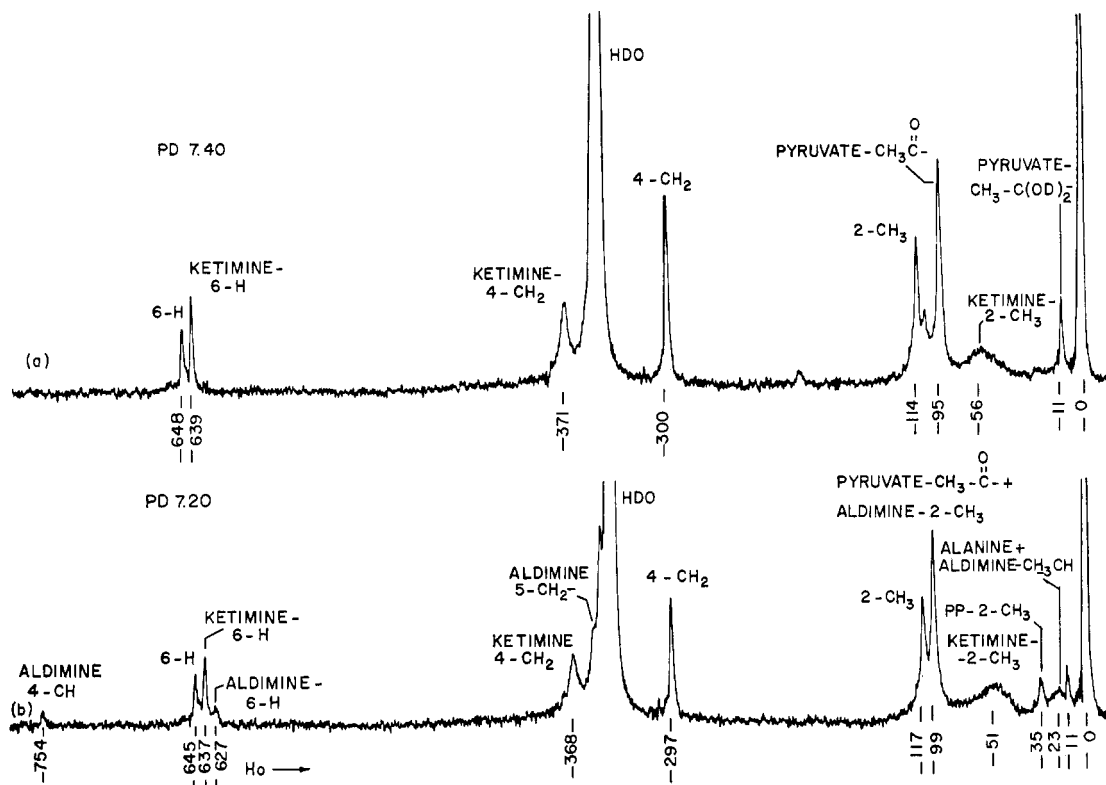


Figure 1. 100-Mcps pmr spectra of a solution initially containing 0.1 *M* pyridoxamine, 0.1 *M* sodium pyruvate, and 0.05 *M* Zn<sup>2+</sup> at 33°: (a) before transamination, illustrating formation of ketimine species; (b) after transamination, revealing the presence of ketimine and aldimine species. Signals labeled 6-H, 4-CH<sub>2</sub>, and 2-CH<sub>3</sub> refer to free pyridoxamine; PP = N-pyridoxylidenepyridoxamine. Chemical shifts are relative to a *t*-butyl alcohol internal standard.

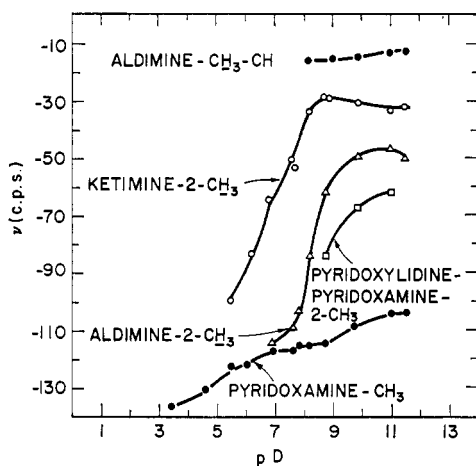


Figure 2. 100-Mcps chemical shifts as a function of pD in the methyl region of transaminated solutions at 33° initially containing 0.1 *M* pyridoxamine, 0.1 *M* sodium pyruvate, and 0.05 *M* Zn<sup>2+</sup>. Chemical shifts are relative to a *t*-butyl alcohol internal standard.

arise from the aldimine, whose chemical shifts and extent of formation, like those of the ketimine, are pD dependent. Spectra of 0.05 *M* Zn<sup>2+</sup> solutions were similar to the metal-free solutions except that aldimine formation was first detectable at pD 4.2 and at pD 9.8 was complete, but only 25% complete in the metal-free case. Further, the observed broadening and upfield shifting of the aldimine 2-CH<sub>3</sub> signal with increasing pD (cf. Figure 2) indicates formation of labile 1:1 and 1:2 Zn(II):aldimine complexes **3** with the latter predominating above pD ~8.5. Similar studies were performed

on solutions containing pyridoxal, pyridoxamine, and Zn<sup>2+</sup>.

Plots of chemical shift vs. pD for each signal in all of the solutions described above has permitted certain identification of all signals in the spectra of transaminated solutions having pD 5–10. As an example, a plot of chemical shift in the methyl region is given in Figure 2. Signal assignments in the transaminated solution having pD 7.2 are indicated in Figure 1b. Several salient factors pertinent to the mechanism of nonenzymatic transamination at pD 4–9.5 emerge from this work: (i) ketimine and aldimine complexes are definitely produced in a sequential fashion in the reaction; (ii) pyruvate methyl protons of **2** and the free ketimine exchange rapidly with solvent at a rate comparable to or exceeding that of transamination; (iii) alanine methyl and CH protons of **3** and free aldimine do not exchange with solvent; (iv) ketimine  $\rightleftharpoons$  aldimine conversion (**2**  $\rightleftharpoons$  **3**) does not occur as a consequence of rapid, complete exchange of both 4-CH<sub>2</sub> protons of **2** with solvent accompanied by protonation of an intermediate<sup>1,2</sup> to yield **3**, since the latter is not deuterated at the azomethine carbon. Observation i is completely consistent with the proposed mechanism,<sup>1</sup> whereas factors ii and iv are not anticipated by it. The origin of (iv) presumably rests in the chelate ring conformations of **2** which confer a selective lability on one of the two methylene protons. The significance of (iii) in connection with the racemization of amino acids effected by pyridoxal and metal ions is being investigated and the results of this work together with the full details of the pmr study of transamination will be presented shortly.

