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ABSTRACT

Gemcitabine, 2'-deoxy-2',2'-difluorocytidine, is currently prescribed against a number of cancers. Here we report a linear synthesis of gemcitabine with a high-yielding direct conversion of 3,5-di-O-benzoyl-2-deoxy-2,2-difluororibose into the corresponding glycosyl urea as the key step, followed by conventional conversion to the cytosine base via the uracil derivative. The process proceeded with modest anomeric selectivity.

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1. Introduction

Gemcitabine **1** (Scheme 1), marketed by its discoverer Lilly as the HCl salt under the trade name of Gemzar, is a widely prescribed anticancer drug against pancreatic, ovarian and breast cancer. Before its recent patent expiry, it was a \$1 bn dollar per annum drug, and while sales have now dropped below that figure, its market continues to grow. It is a fluorinated nucleoside^{1–6} prodrug, undergoing intracellular phosphorylation to its active diphosphate and triphosphate form, which inhibits DNA synthesis leading to apoptosis.^{7–11}

The synthesis of gemcitabine¹² has been subject to tremendous and—spurred by the Lilly patent expiry—still continuing efforts.¹³ In general, a convergent synthesis has been applied, in which a suitably protected and activated 2-deoxy-2,2-difluororibofuranose derivative **2** (Scheme 1) is combined with an activated cytosine base, e.g. **3**. Much optimisation has been devoted to an efficient 2deoxy-2,2-difluororibofuranose synthesis, a feat which has now largely been achieved through use of specific alcohol protecting groups allowing efficient and high-yielding diastereomer separation by crystallisation(s). The nucleobase introduction reaction has also been subject to intensive research. This process is made difficult by the difluorination at the sugar 2-position, and typically leads



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Scheme 1. Convergent nucleobase introduction in the synthesis of gemcitabine 1.

to an anomeric mixture. Again, selective crystallisation procedures have been developed to obtain anomerically pure gemcitabine.¹³

While alternative linear nucleoside syntheses are common in the carbanucleoside field, $^{14-16}$ this strategy has to our knowledge been applied only once for the gemcitabine synthesis (Scheme 2a).¹⁷

Here the sugar donor **5** was reacted with the N-2-cyanovinyl amide **4** to give the intermediate **6**, after which the cytosine synthesis was completed by base-induced cyclisation. An overall yield of 12% was reported. A linear synthesis is also possible—but not reported—from the corresponding amino-2-deoxy-2,2-difluoroglycoside **9**, which has been synthesised in two steps by a Lilly group¹⁸ (Scheme 2b). However, while **9** was obtained in high yield, no anomeric selectivity was obtained, and the anomeric ratio of **9** was independent of the anomeric ratio of the azide precursor **8**, suggesting an equilibration process.

Herein we describe our efforts leading to an alternative linear gemcitabine synthesis. In order to avoid the abovementioned

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Scheme 2. Linear gemcitabine synthesis.

anomerisation at the aminoglycoside stage, a nucleobase construction strategy starting from the corresponding glycosyl urea **10** was envisioned (Scheme 3). It was hoped that the possible crystallinity of this urea derivative would offer anomeric purification prospects.



Scheme 3. Retrosynthetic analysis.

In addition, it was planned to introduce the urea by a direct condensation reaction with the known 3,5-di-O-benzoyl-2-deoxy-2,2-difluororibose **11**, thus bypassing the need for anomeric activation. This protected difluororibose derivative was first described by a Lilly group,¹⁹ but obtained by us by reduction of the commercially available dibenzoylated difluororibonolactone.

Table 1

Optimisation of the urea condensation reaction

2. Results and discussion

2.1. Urea condensation

The work commenced by investigating the direct urea introduction. The acid-catalysed condensation of aldehydes with urea has been reported in high yields,^{20,21} and there were few reports available of this process with unprotected (and unfluorinated) carbohydrates.^{22,23} In recent years, the reaction of unprotected and non-activated carbohydrates with ureas has received renewed attention, and has typically been achieved with an acid-catalyst.²⁴ Hence, it was first investigated whether the acid-catalysed condensation of a difluorinated sugar such as furanose **11** was possible with ureas. Initially, the 5-O-TBDPS lactol **12** was used as substrate (Table 1), and to avoid complication by the urea reacting at both ends, *N*,*N*-dimethyl urea **12** was used.

Reaction of 12 with 13b using a variety of acid catalysts (TMSOTf, Ti(Oi-Pr)₄, 4 N HCl, AcOH, TFA, p-TsOH) in a range of solvents (THF, DCM, toluene, trifluoroethanol, hexafluoroisopropanol) all gave no or only trace amounts of product 14 (not shown), except the combination of *p*-TsOH and toluene at reflux (entry 1). While, when using 2 equiv of urea the presence/absence of molecular sieves had no significant influence (entries 1, 2), when 1 equiv of urea was used, the use of molecular sieves did lead to a higher yield (entries 3, 4). Due to the possible lability of the TBDPS ether in acidic medium, further optimisation experiments were carried out with the 3.5-di-O-benzovl-2-deoxy-2.2-difluororibose **11**. obtained bv reduction of the commercially available lactone. Indeed, reaction with **13b** now gave a much improved 65% yield (entry 5), although a further increase in amount of urea was used. At this stage, it was decided to investigate the non-methylated urea 13a, which turned out to be unproblematic with regard to a sequential condensation process. The best solvent proved to be 1,4-dioxane (entry 6), giving a yield of 40%, with 39% recovered starting material, when using

Entry	Lactol	13 (equiv)	Solvent	PTSA (equiv)	Drying agent	Time (h)	Work-up	Et ₃ N Column?	Yield ^a (%)
1	12	b (2)	Toluene	0.05	MS	18	b	N	17
2	12	b (2)	Toluene	0.05	_	5.5	b	Ν	22
3	12	b (1)	Toluene	0.05	_	5	b	Ν	29
4	12	b (1)	Toluene	0.05	MS	5.5	b	N	38
5	11	b (3.3)	Toluene	1.1	MS	20	с	Y	65
6	11	a (3)	Dioxane	1	MS	18	Aqueous	Y	40
7	11	a (5)	Dioxane	1	MS	44	Aqueous	Y	59
8	11	a (3)	Dioxane	1	Na ₂ SO ₄	18	Aqueous	Y	61
9	11	a (3)	Dioxane	1	Na ₂ SO ₄	36	Column	Y	62
10	11	a (5)	Dioxane	1	Na ₂ SO ₄	72	Aqueous	Y	64
11	11	a (3)	Dioxane	1	Na ₂ SO ₄	66	с	Y	65
12	11	a (3)	Dioxane	1	Na ₂ SO ₄	36	d	N	79
13	11	a (3)	Dioxane	1	_	36	d	N	76
14	11	a (5)	Dioxane	1	Na ₂ SO ₄	36	d	N	79
15 ^e	11	a (3)	Dioxane	1	Na ₂ SO ₄	36	d	Ν	88

^a Isolated yield.

^b Remove reaction solvent by evaporation in vacuo.

^c Filtration over silica.

^d Filtration over Celite.

e 2.3 mmol scale.

1 equiv of *p*-TsOH. An increase in the relative amount of urea increased the yield to 59% (entry 7). As good conversion was seen by NMR analysis of crude material, losses during the workup due to difficult filtration of the molecular sieves were thought to be responsible for the reduced yield. However, a change of drying agent to Na₂SO₄ (entries 8–11), whilst facilitating the work-up, did not have a significant effect upon the yield, regardless of urea stoichiometry. Changing the workup to filtration over Celite instead of silica gel did improve the yield (entry 12), though, surprisingly, the reaction without Na₂SO₄ worked equally well (entry 13). Increasing the number of urea equivalents to 5 had no further beneficial effect (entry 14). Finally, reaction on 2.3 mmol scale under the best conditions led to an excellent 88% yield (entry 15) of the glycosyl ureas in a 1:1.8 anomeric ratio. Unfortunately, assignment of the anomers could not be achieved.

2.2. Formation of the cytosine nucleobase

Next, acylation of the urea group with acyl chloride **16**, synthesised by the method of Tietze,²⁵ to give the nucleobase cyclisation precursor was investigated. Interestingly, this glycosyl urea acylation does not have a precedent for O-nucleosides, but has been demonstrated with carbanucleosides using pyridine with DMAP.^{26,27} However, reaction of **14a** with **16** under those conditions (Table 2, entry 1), only afforded 30% of the desired product **17**. Unexpectedly, significant anomerisation was observed, which led to a decrease in anomeric ratio (1:2.4 to 1:1). This anomerisation may be due to urea deprotonation by pyridine (Scheme 4), facilitated by the electron withdrawing fluorines at the 2-position, to give the conjugate base **18**. Anomerisation can then be envisaged by ring opening to **19** followed by non-selective ring closure.

Table 2

Optimisation of the urea acylation reaction



Entry	/ Equiv 16	Reagents	Solvent	Temp (°C)	Time Yield ^a	Anomeric ratio ^b
1	1.5	0.25 equiv DMAP	Pyridine	Reflux	40 h 30%	1:1
2	1	1 equiv DMAP	DCM	RT	3.5 d 0%	_
3	2	None	MeCN	Reflux	18 h 45%	1:1.5
4	4	None	MeCN	Reflux	18h 71%	1:1.5
5	4	None	Dioxane	Reflux	18h 61%	1:1.15
6	3	0.25 equiv ZnCl ₂	CHCl ₃	Reflux	24 h 45%	1:1.9

^a Isolated yield.

^b Determined by NMR analysis on the crude reaction mixture.

Thus, the basic pyridine solvent was replaced by DCM, but even with 1 equiv of DMAP, none of the desired product was observed after 3.5 d (entry 2). Pleasingly, refluxing in acetonitrile with 2 equiv of **16** (entry 3) did yield the product in 45% yield, with a further improvement to 71% seen when the amount of **16** was increased to 4 equiv (entry 4). These conditions also resulted in less anomerisation (1:2.4 to 1:1.5). The same conditions with 1,4-dioxane as solvent gave a slightly lower yield, but more anomerisation was observed (entry 5). The least anomerisation was seen when ZnCl₂ was employed, with a 1:1.9 anomeric ratio seen in the isolated product (entry 6), but a lower yield was obtained.

With the acyl carbamate **17** in hand, cyclisation to the protected difluorouridine **20** was studied (Scheme 5) using methodology originally developed by Shaw.²⁸ Among the various acidic protocols screened, HCl/AcOH emerged as the most efficient, leading to quantitative conversion to **20** in a 1:1.2 α/β mixture. Next, conversion of uridine to cytidine was effected using described methodology^{29,30} via the triazole intermediate **21** by treatment with 2-chlorophenyl phosphorodichloridate and 1,2,4-triazole in pyridine, followed by reaction with ammonia. The ammonia treatment also caused concomitant benzoate deprotection, which then furnished gemcitabine **1** as a 1:1.3 mixture of α : β anomers.



Scheme 5. Conversion to gemcitabine 1 via the corresponding uridine 20.

3. Conclusion

The linear synthesis of gemcitabine was achieved in 5 steps from a 2-deoxy-2,2-difluororibose substrate with moderate anomeric selectivity. The demonstration of a high-yielding direct glycosyl urea formation through reaction of a reducing 2-deoxy-2,2difluorinated sugar derivative with urea will be of interest for other applications.

4. Experimental section

4.1. Synthesis of the urea derivative 14a

Lactol **11** (870 mg, 2.30 mmol), urea (3 equiv, 6.90 mmol, 414 mg), PTSA (1 equiv, 2.30 mmol, 437 mg) and Na_2SO_4 (2 equiv, 4.60 mmol, 653 mg) were stirred in 1,4-dioxane (6.5 mL) at reflux for 36 h. The reaction mixture was cooled to RT, diluted with DCM and filtered through Celite. The solvents were reduced in vacuo to yield a crude residue, which was purified by column chromatography on silica gel



Scheme 4. Possible anomerisation mechanism.

 $(10:90 \rightarrow 40:60 \text{ acetone/petrol})$ to yield the desired glycosylurea **14a** as a white foam, as a 1:1.8 anomeric mixture (853 mg, 88%). IR (film) $3366 (br), 2925 (w), 1723 (m), 1678 (m), 1266 (s) cm^{-1}$. LRMS (ESI⁺) m/ z 443.2 (M+Na)⁺ (100). **HRMS** (ESI⁺) for $C_{20}H_{18}F_2N_2O_6$ (M+Na)⁺ calcd 443.1025, found 443.1018. ¹H NMR (400 MHz, CDCl₃) δ 8.04 (4H, m, $4 \times CH_{Ar}$), 7.61–7.33 (6H, m, $6 \times CH_{Ar}$), 6.69 (²/₃H, d, J=10.0 Hz, NH_{Major}), 6.54 (¹/₃H, d, J=8.8 Hz, NH_{Minor}), 5.94 (¹/₃H, q, J=8.2 Hz, CHNH_{Minor}), 5.78 (²/₃H, ddd, J=14.4, 10.1, 5.4 Hz, CHNH_{Maior}), 5.67 (¹/₃H, ddd, *J*=11.5, 8.1, 6.6 Hz, CHOBz_{Minor}), 5.46 (²/₃H, m, CHOBz_{Major}), $5.34(^{2}/_{3}H, \text{ br s, NH}_{2\text{Minor}}), 5.32(^{4}/_{3}H, \text{ br s, NH}_{2\text{Major}}), 4.65-4.51(^{7}/_{3}H,$ m, CHCH_{2Minor}, CH₂), 4.36 (²/₃H, q, J=4.3 Hz, CHCH_{2Major}) ppm. ¹³C NMR+DEPT (100 MHz, CDCl₃) δ 166.3 (C=O), 165.0 (C=O_{Major}), 164.9 (C=O_{Minor}), 157.8 (C=O_{Minor}), 157.7 (C=O_{Major}), 134.0 (CH_{Ar}), 133.9 (CH_{Ar}), 133.3 (CH_{Ar}), 130.0 (CH_{Ar}), 129.7 (CH_{Ar}), 129.2 (C_{Ar}), 128.6 (CH_{Ar}), 128.5 (CH_{Ar}), 128.43 (CH_{Ar}), 128.38 (CH_{Ar}), 128.1 (C_{Ar}), 121.4 (dd, J=262.4, 257.6 Hz, CF_{2Minor}), 120.4 (dd, J=266.3, 253.7 Hz, CF_{2Maior}), 81.9 (dd, J=37.9, 19.4 Hz, CHN_{Minor}), 81.2 (dd, J=34.0, 20.4 Hz, CHN_{Major}), 77.2 (d, J=5.8 Hz, CHCH_{2Minor}), 76.0 (CHCH_{2Major}), 72.3 (dd, J=31.1, 17.5 Hz, CHOBz_{Minor}), 72.0 (dd, J=36.0, 16.5 Hz, CHOBz_{Maior}), 63.4 (CH_{2Minor}), 63.3 (CH_{2Major}) ppm (some CH_{Ar} overlap).

Major Anomer: ¹⁹F NMR (282 MHz, CDCl₃) δ –117.9 (1F, dt, *J*=246.6, 14.7 Hz, CFF), –122.9 (1F, dd, *J*=247.4, 5.2 Hz, CFF) ppm. Minor Anomer: ¹⁹F NMR (282 MHz, CDCl₃) δ –113.2 (1F, d,

J=243.1 Hz, *CF*F), –125.1 (1F, dt, *J*=244.0, 7.8 Hz, *CFF*) ppm.

4.2. Synthesis of the N,N-dimethyl urea derivative 14b

To lactol **11** (35 mg, 0.09 mmol) in toluene (1 mL) was added 3 Å molecular sieves (53 mg), N.N-dimethylurea (3 equiv, 0.30 mmol, 27 mg) and PTSA (1 equiv, 0.09 mmol, 19 mg). The reaction mixture was stirred at reflux for 20 h then cooled to RT and diluted with DCM before filtration through a plug of silica (pre-treated with Et₃N), NaHCO₃ and NaSO₄. The solvents were reduced in vacuo to yield a crude residue, which was purified by column chromatography on silica gel (15:85 \rightarrow 25:75 acetone/hexane+0.5% Et₃N) to yield the desired glycosylurea 14b as an oil, as a 1:2.7 anomeric mixture (27 mg, 65%). IR (film) 3307 (br), 2923 (w), 2853 (w), 1724 (s), 1659 (m), 1522 (m), 1266 (s), 1095 (s) cm⁻¹. **LRMS** (ESI⁺) m/z 919.5 (2M+Na)⁺ (100), 471.1 $(M+Na)^+$ (71). **HRMS** (ESI⁺) for C₂₂H₂₂F₂N₂O₆ $(M+Na)^+$ calcd 471.1338, found 471.1342. ¹H NMR (400 MHz, CDCl₃) δ 8.09–8.03 (4H, m, 4×CH_{Ar}), 7.65–7.39 (6H, m, 6×CH_{Ar}), 6.12 (0.25H, q, J=8.2 Hz, CHN_{Minor}), 5.89 (0.75H, ddd, J=14.9, 9.9, 5.2 Hz, CHN_{Maior}), 5.72 (0.25H, ddd, J=10.1, 8.3, 5.8 Hz, CHOBz_{Minor}), 5.51 (0.75H, dd, J=15.6, 5.0 Hz, CHOBz_{Maior}), 5.41 (0.25H, d, J=9.5 Hz, NH_{Minor}), 5.28 (0.75H, d, J=9.8 Hz, NH_{Maior}), 4.67–4.56 (2.25H, m, CH₂OBz, CHCH_{2Minor}), 4.40 (0.75H, q, J=4.4 Hz, CHCH_{2Major}), 2.98 (6H, br s, N(CH₃)₂) ppm. ¹³C **NMR**+**DEPT** (100 MHz, CDCl₃) δ 166.0 (PhC=O), 164.9 (PhC=O), 156.0 (NHC=0), 134.02 (CH_{Ar}), 133.98 (CH_{Ar}), 133.3 (CH_{Ar}), 130.0 (CH_{Ar}), 129.9 (CH_{Ar}), 129.8 (CH_{Ar}), 129.4 (CH_{Ar}), 129.3 (CH_{Ar}), 128.6 (CH_{Ar}), 128.4 (CH_{Ar}), 128.3 (CH_{Ar}), 128.2 (2×C_{Ar}), 82.1 (dd, J=36.9, 18.5 Hz, CHN_{Minor}), 81.6 (dd, J=34.0, 19.4 Hz, CHN_{Major}), 77.1 (CHCH_{2Minor}), 76.2 (t, J=2.9 Hz, CHCH_{2Major}), 72.6 (dd, J=31.1, 17.5 Hz, CHOBz_{Minor}), 72.0 (dd, J=36.0, 17.5 Hz, CHOBz_{Major}), 63.8 (CH_{2Minor}), 63.3 (CH_{2Major}), 36.3 (N(CH₃)_{2Major}), 36.2 (N(CH₃)_{2Minor}) ppm (some CH_{Ar} overlap, CF₂ not visible).

Major anomer: ¹⁹**F NMR** (282 MHz, CDCl₃) δ –118.4 (1F, dt, *J*=247.1, 15.0 Hz, CFF), –123.3 (1F, dd, *J*=247.1, 5.4 Hz, CFF) ppm.

Minor anomer: ¹⁹**F** NMR (282 MHz, CDCl₃) δ –114.9 (1F, dt, *J*=243.9, 9.7 Hz, CFF), –126.4 (1F, dt, *J*=242.8, 8.6 Hz, CFF) ppm.

4.3. Synthesis of the N,N-dimethyl urea derivative 15b

To lactol **12** (102 mg, 0.20 mmol) in toluene (0.8 mL) was added N,N-dimethylurea (1 equiv, 0.16 mmol, 18 mg), a catalytic amount of PTSA and molecular sieves (90 mg). The reaction mixture was

stirred at reflux for 5.5 h, cooled to RT and the solvents removed in vacuo to yield a crude residue. This residue was first purified by column chromatography on silica gel (10:90 acetone/hexane) followed by HPLC (10:90 acetone/hexane) to yield **15b** as a colourless oil as a 1:2.9 anomeric mixture (44 mg, 38%). **IR** (film) 3070 (w), 2930 (m), 2857 (m), 1472 (m), 1456 (m) cm⁻¹.

Major anomer: ¹H NMR (400 MHz, acetone- d_6) δ 7.76–7.67 (4H, m, 4×ArH), 7.49–7.31 (11H, m, 11×ArH), 4.87 (1H, d, *J*=11.8 Hz, OCHHPh), 4.66 (1H, d, *J*=11.6 Hz, OCHHPh), 4.63 (1H, m, CHN), 4.23 (1H, dd, *J*=12.3, 11.2, 8.3 Hz, CHOBn), 3.97 (1H, m, CHCH₂OSi), 3.84 (1H, dd, *J*=11.6, 3.5 Hz, CHHOSi), 3.75 (1H, m, CHHOSi), 2.47 (6H, s, N(CH₃)₂), 1.01 (9H, s, C(CH₃)₃) ppm (NH not visible). ¹³C NMR+DEPT (100 MHz, acetone- d_6) δ 182.2 (C=O), 138.6 (C_{Ar}), 136.5 (2×CH_{Ar}), 134.1 (2×C_{Ar}), 130.84 (2×CH_{Ar}), 130.76 (2×CH_A), 129.3 (2×CH_{Ar}), 128.8 (5×CH_{Ar}), 128.7 (2×CH_{Ar}), 120.7 (2×CH_A), 129.3 (2×CH_A), 128.8 (5×CH_A), 128.7 (2×CH_A), 125.1 (dd, *J*=259.5, 255.1 Hz, CF₂), 96.6 (dd, *J*=31.8, 24.3 Hz, CHN), 81.7 (CHCH₂), 77.7 (m, CHOBn), 73.3 (CH₂), 63.3 (CH₂), 41.0 (N(CH₃)₂), 27.3 (C(CH₃)₃), 19.9 (C(CH₃)₃) ppm. ¹⁹F NMR (376 MHz, acetone- d_6) δ –112.3 (1F, ddd, *J*=240.3, 14.3, 11.0 Hz, CFF), –113.0 (1F, dt, *J*=240.3, 11.8 Hz, CFF) ppm.

Minor anomer: ¹H NMR (400 MHz, acetone-*d*₆) δ 7.76–7.67 (4H, m, 4×ArH), 7.49–7.31 (11H, m, 11×ArH), 4.86 (1H, d, *J*=11.9 Hz, OCHHPh), 4.74 (1H, t, *J*=11.1 Hz, CHN), 4.65 (1H, d, *J*=11.7 Hz, OCHHPh), 4.39 (1H, ddd, *J*=12.2, 9.2, 7.1 Hz, CHOBn), 4.09 (1H, dtd, *J*=6.9, 3.4, 1.3 Hz, CHCH₂OSi), 3.86 (1H, m, CHHOSi), 3.76 (1H, m, CHHOSi) 2.44 (6H, s, N(CH₃)₂), 1.02 (9H, s, C(CH₃)₃) ppm (NH not observed). ¹³C NMR+DEPT (100 MHz, acetone-*d*₆) δ 182.2 (C=O), 138.6 (C_{Ar}), 136.5 (2×CH_{Ar}), 134.0 (2×C_{Ar}), 130.9 (2×CH_{Ar}), 130.8 (2×CH_{Ar}), 129.4 (2×CH_{Ar}), 128.8 (5×CH_{Ar}), 128.7 (2×CH_{Ar}), 96.1 (dd, *J*=36.4, 18.0 Hz, CHN), 81.6 (CHCH₂), 77.9 (m, CHOBn), 73.4 (CH₂), 64.4 (CH₂), 41.4 (N(CH₃)₂), 27.3 (C(CH₃)₃), 19.9 (C(CH₃)₃) ppm (CF₂ not observed). ¹⁹F NMR (376 MHz, acetone-*d*₆) δ –105.4 (1F, dt, *J*=240.8, 11.8 Hz, CFF), -126.4 (1F, dt, *J*=240.3, 11.0 Hz, CFF) ppm.

4.4. Synthesis of 17

To glycosylurea 14a (93 mg, 0.22 mmol) stirring at reflux in MeCN (0.6 mL) was added dropwise a solution of acyl chloride 16 (4 equiv, 0.88 mmol, 119 mg) in MeCN (0.4 mL). The reaction mixture was stirred at reflux for 18 h, then quenched with water (0.1 mL). The solvents were reduced in vacuo to yield a crude residue, which was taken up in DCM (15 mL), washed with water (5 mL), then brine and dried over anhydrous MgSO₄. The solvents were reduced in vacuo to yield a crude residue, which was purified by flash chromatography on silica gel $(10:90 \rightarrow 40:60 \text{ acetone})$ petrol) to yield the desired product 17 as a colourless oil as a 1:1.5 mixture of anomers (82 mg, 71%). IR (film) 3246 (br), 2981 (w), 1717 (s), 1680 (s), 1603 (m), 1537 (s), 1492 (w), 1452 (m), 1378 (w), 1316 (m), 1247 (s), 1176 (m), 1093 (s) cm⁻¹. **LRMS** (ESI⁺) m/z 1059.8 $(2M+Na)^+$ (50), 541.3 $(M+Na)^+$ (100). **HRMS** (ESI⁺) for C₂₅H₂₄F₂N₂O₈ (M+Na)⁺ Calcd: 541.1393; Found: 541.1411. ¹H NMR (400 MHz, CDCl₃) δ 10.01 (0.5H, d, *J*=9.2 Hz, 0.5×CHNH), 9.79 (0.5H, d, J=9.5 Hz, 0.5×CHNH), 9.65 (0.5H, s, 0.5×O=CNHC=0), 9.28 (0.5H, s, 0.5×0=CNHC=0), 8.22-8.20 (1H, m, CH_{Ar}), 8.08-8.02 (3H, m, 3×CH_{Ar}), 7.69 (1H, d, *J*=11.9 Hz, HC=CHOEt), 7.65–7.40 (6H, m, $6 \times CH_{Ar}$), 6.08 (0.5H, t, J=9.3 Hz, 0.5×CHN), 5.90 (0.5H, ddd, J=12.9, 9.5, 6.0 Hz, 0.5×CHN), 5.70–5.67 (0.5H, m, 0.5×CHOBn), 5.57 (0.5H, ddd, J=14.7, 5.1, 1.0 Hz, 0.5×CHOBn), 5.37 (0.5H, d, J=12.1 Hz, 0.5×CHOEt), 5.34 (0.5H, d, J=12.0 Hz, 0.5×CHOEt), 4.69-4.58 (2.5H, m, CHCH₂, 0.5×CHCH₂), 4.43 (0.5H, q, J=4.5 Hz, 0.5×CHCH₂), 3.98 (2H, q, *J*=7.1 Hz, CH₂CH₃), 1.34 (1.5H, t, *J*=7.0 Hz, 0.5×CH₃), 1.33 (1.5H, t, *J*=7.1 Hz, 0.5×CH₃) ppm. ¹³C NMR+DEPT $(100 \text{ MHz}, \text{CDCl}_3) \delta$ 168.3 $(0.5 \times C = \text{OCH} = \text{CH})$, 168.1 $(0.5 \times C = \text{OCH} =$ CH), 166.07 (0.5×PhCOCH₂), 166.05 (0.5×PhCOCH₂), 164.9 (0.5×PhCOCH), 164.7 (0.5×PhCOCH), 164.0 (0.5×CH=CHOEt), 163.6 (0.5×CH=CHOEt), 155.1 (0.5×NC=ON), 154.9 (0.5×NC=ON), 134.0 (CH_Ar), 133.3 (CH_Ar), 133.2 (CH_Ar), 130.4 (CH_Ar), 130.0 (CH_Ar), 129.8 (CH_Ar), 128.6 (CH_Ar), 128.5 (CH_Ar), 128.4 (CH_Ar), 129.33 (0.5×C_Ar), 129.30 (0.5×C_Ar), 128.2 (0.5×C_Ar), 128.0 (0.5×C_Ar), 97.5 (HC=CHOEt), 81.2 (dd, *J*=39.5, 22.0 Hz, 0.5×CHN), 80.5 (dd, *J*=36.6, 22.0 Hz, 0.5×CHN), 79.3 (0.5×CHCH₂), 76.7 (0.5×CHCH₂), 72.2 (dd, *J*=33.7, 17.6 Hz, 0.5×CHOBz), 72.0 (dd, *J*=35.1, 17.6 Hz, 0.5×CHOBz), 68.0 (0.5×CH₂CH₃), 67.7 (0.5×CH₂CH₃), 63.3 (0.5×CHCH₂), 63.2 (0.5×CHCH₂), 14.43 (0.5×CH₃), 14.36 (0.5×CH₃) ppm (some CH_Ar overlap, CF₂ not visible). ¹⁹F NMR (282 MHz, CDCl₃) δ Anomer 1: -109.5 (1F, ddd, *J*=251.4, 16.1, 6.4 Hz, CFF), -125.4 (1F, d, *J*=252.5 Hz, CFF) ppm; Anomer 2: -123.6 (1F, dd, *J*=241.8, 9.7 Hz, CFF), -125.5 (1F, ddd, *J*=241.8, 10.7, 6.4 Hz, CFF) ppm. (Data assigned from an earlier experiment, which gave a 1:1 mixture of anomers).

4.5. Synthesis of the uridine 20

Acyl carbamate **17** (136 mg, 0.26 mmol) was stirred in 1:10 HCl/ AcOH (2.75 mL) at RT in a stoppered flask for 24 h. The solvents were reduced in vacuo to yield the desired protected difluorouracil **20** as a yellow oil as a 1:1.2 α/β mixture of anomers (125 mg, quantitative yield).

¹H NMR (400 MHz, CDCl₃) δ 8.91 (1H, br s, NH), 8.11–8.02 (4H, m, CH_{Ar}), 7.69–7.39 (7H, m, $6 \times CH_{Ar}$, NCH=CH), 6.55 (0.45H, t, *J*=7.5 Hz, CHN_α), 6.40 (0.55H, dd, *J*=11.8, 6.4 Hz, CHN_β), 5.82 (0.45H, d, *J*=8.1 Hz, NCH=CH_α), 5.79 (0.45H, m, CHOBz_α), 5.68 (0.55H, d, *J*=8.3 Hz, NCH=CH_β), 5.65 (0.55H, m, CHOBz_β), 4.89–4.81 (1H, m, CHCHH), 4.73–4.59 (2H, m, CHO, CHCHH) ppm. ¹⁹F NMR (282 MHz, CDCl₃) δ –109.2 (0.45F, d, *J*=249.3 Hz, CFF), –115.5 (0.55F, dt, *J*=247.4, 12.9 Hz, CFF), –120.3 (0.5F, d, *J*=246.6 Hz, CFF), –122.0 (0.5F, d, *J*=250.9 Hz, CFF) ppm.

4.6. Synthesis of gemcitabine 1

To crude protected difluorouridine 20 (378 mg, 0.75 mmol) in pyridine (9 mL) was added 1,2,4-triazole (3 equiv, 2.26 mmol, 156 mg) and 2-chlorophenyl phosphorodichloridate (6 equiv, 4.51 mmol, 0.74 mL). The reaction mixture was stirred at RT for 5 d. The solvents were reduced in vacuo to yield a crude residue, which was taken up in DCM and washed with satd NaHCO₃ (aq), then brine and dried over anhydrous Na₂SO₄. The solvents were reduced in vacuo to yield a crude oil, which was passed through a short silica column (10:90 acetone/petrol) to yield the impure triazole 21 (94 mg, ~24%). This triazole (76.5 mg, 0.17 mol) was stirred in 7 N NH₃ in MeOH (5 mL) at RT for 36 h. The solvents were reduced in vacuo and the resultant residue evaporated onto silica gel for purification by column chromatography (10:90-20:80 MeOH/DCM) to yield the desired difluorocytidine **1** as a 1:1.3 α/β mixture of anomers (18.7 mg, 49%). ¹**H NMR** (400 MHz, DMSO-*d*₆) δ 7.69 (0.6H, d, I=7.6 Hz, NCH=CH_{β}), 7.52 (0.4H, dd, I=7.5, 1.6 Hz, NCH=CH_{α}), 7.37-7.30 (2H, m, NH₂), 6.30-6.26 (0.8H, m, CHN_α, CHOH_α), 6.22 (0.6H, d, *J*=6.6 Hz, CHOH_β), 6.13 (0.6H, t, *J*=8.3 Hz, CHN_β), 5.79 (0.4H, d, *J*=7.5 Hz, NCH=CH_α), 5.78 (0.6H, d, *J*=7.6 Hz, NCH=CH_β), 5.19 $(0.6H, t, J=5.4 \text{ Hz}, CH_2OH_\beta)$, 5.07 $(0.4H, t, J=5.7 \text{ Hz}, CH_2OH_\alpha)$, 4.34 (0.4H, m, CHOH_α), 4.18–4.07 (1.2H, m, CHOH_β, CHCH_{2(β)}), 3.81–3.74 (1H, m, CHCH_{2α}, CHCHH_β), 3.64–3.59 (1H, m, CHCHH), 3.53 (0.4H, m, CHCHH_α) ppm. ¹³**C** NMR+DEPT (100 MHz, DMSO-*d*₆) δ 165.7 (0.5×NCON), 165.6 (0.5×NCON), 154.8 (0.5×N=C), 154.7 (0.5×N=C), 141.3 (0.5×N=C), 140.8 (0.5×N=C), 123.1 (t, *J*=252.6 Hz, CF₂), 94.6 (0.5×N=C), 94.4 (0.5×N=C), 83.9–83.2 (m, CHN), 80.4 (CHCH₂), 69.6 (dd, *J*=26.3, 17.6 Hz, 0.5×CHOH), 68.7 (t, *J*=22.0 Hz, CHOH), 60.0 (0.5×CH₂), 59.0 (0.5×CH₂) ppm. ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ –114.3 (0.4F, d, *J*=232.8 Hz, CFF_α), –115.5 to –117.3 (1.2F, m, CF_{2(β)}), –124.3 (0.4F, d, *J*=234.5 Hz, CFF_α) ppm.

Data consistent with the literature.¹²

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Supplementary data

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