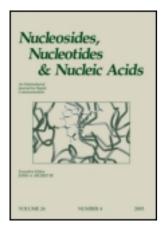
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PHOSPHORAMIDITES AND OLIGONUCLEOTIDES CONTAINING 7-DEAZAPURINES AND PYRIMIDINES CARRYING AMINOPROPARGYL SIDE CHAINS

F. Seela^a, N. Ramzaeva^a, P. Leonard^a, Y. Chen^a, H. Debelak^a, E. Feiling^a, R. Kröschel^a, M. Zulauf^a, T. Wenzel, T. Fröhlich & M. Kostrzewa
^a Institut für Chemie, Universität Osnabrück, Laboratorium für Organische und Bioorganische Chemie, Germany
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PHOSPHORAMIDITES AND OLIGONUCLEOTIDES CONTAINING 7-DEAZAPURINES AND PYRIMIDINES CARRYING AMINOPROPARGYL SIDE CHAINS

F. Seela,* N. Ramzaeva,¹ P. Leonard,¹ Y. Chen,¹
H. Debelak,¹ E. Feiling,¹ R. Kröschel,¹ M. Zulauf,¹
T. Wenzel,² T. Fröhlich,² and M. Kostrzewa²

¹Laboratorium für Organische und Bioorganische Chemie, Institut für Chemie, Universität Osnabrück, Germany
²Bruker Saxonia Analytik GmbH, Leipzig, Germany

ABSTRACT

The synthesis of phosphoramidites containing 7-deazaguanine, 7-deazaadenine, uracil and cytosine carrying aminopropargyl chains is described. The corresponding oligonucleotides are stabilized in duplexes thermally as well as against degradation by exonucleases.

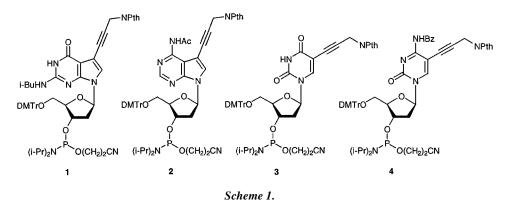
INTRODUCTION

In a series of manuscripts our laboratory has reported on oligonucleotides containing alkynyl- or aminoalkynyl side chains linked to the 7-position of 7-deazapurines or 8-aza-7-deazapurines (purine numbering is used throughout the manuscript) (1–3). A positively charged side chain transforms a negatively charged oligonucleotide in a zwitterion or even a positively charged species. As a result favorable properties are generated, such as duplex stabilization, resistance against enzymatic degradation, or an increased sensitivity of oligonucleotide detection by MALDI-TOF spectrometry.

^{*}Corresponding author.

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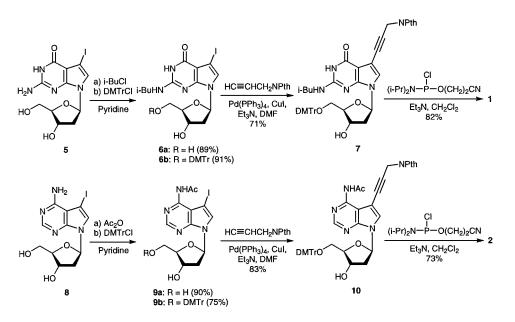


This manuscript reports on the synthesis of the phosphoramidites 1-4 (Scheme 1) carrying an aminopropargyl side chain protected by phthaloyl residues (4–6). This protecting group is removed with ammonia under standard conditions (25% ammonia, 60°C, 24 h) and is superior over the rather labile trifluoroacetyl group (7,8) which gives rise to side reaction (9).

RESULTS AND DISCUSSION

1. Synthesis of the 7-deazapurine Phosphoramidites 1 and 2

The synthesis of the phosphoramidites 1 and 2 was performed using the iodo compounds 5 (10) and 8 (11) as precursors (Scheme 2). While the 7-deazaguanine

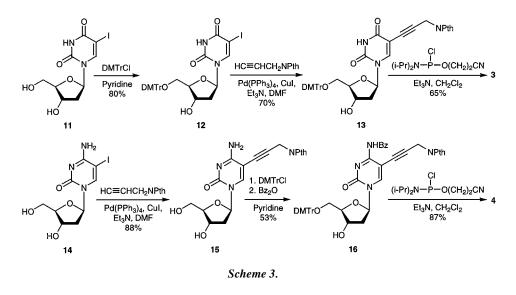




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PHOSPHORAMIDITES AND OLIGONUCLEOTIDES



base was protected with an isobutyryl residue, the 7-deazaadenine moiety was acetylated. Next, the DMTr residues were introduced. Compounds **6b** and **9b** were coupled with phthaloyl-propargylamine employing the Sonogashira reaction $(Pd(PPh_3)_4, CuI, Et_3N \text{ in DMF})$. The fully protected nucleosides **7** and **10** gave the phosphoramidites **1** and **2** applying standard methodology.

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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	piex	[C]	[KCal/III01]		[KCal/III01]
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(T-A-G-G-T-C-A-A-T-A-C-T) 17	7 50 ^a)	-90	-252	-11.8
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(A-T-C-C-A-G-T-T-A-T-G-A) 18	3 47 ^b)	-89	-253	-10.9
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(T-A-G-G-T-C-A-A-T-A-C-T) 17	7 54 ^a)	-80	-220	-12.0
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(A-T-C-C-A-G*-T-T-A-T-G*-A) 19) 52 ^b)	-95	-268	-12.0
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$d(T-A-G^*-G^*-T-C-A-A-T-A-C-T)$ 20) 54 ^a)	-71	-189	-11.9
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(A-T-C-C-A-G*-T-T-A-T-G*-A) 19) 54 ^b)	-98	-272	-13.0
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(T-A-G-G-T-C-A-A-T-A-C-T) 17	7 55 ^a)	-84	-232	-12.4
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	d(A-T-C-C-A*-G-T-T-A*-T-G-A) 21	1 53 ^b)	-85	-237	-11.8
$5'-d(T-A-G-G-U^*-C-A-U^*-A-C-T)$ 23 53^a) -86 -239 -12.0	$d(T-A^*-G-G-T-C-A^*-A^*-T-A^*-C-T)$ 22	2 59 ^a)	-82	-222	-12.9
	d(A-T-C-C-A*-G-T-T-A*-T-G-A) 21	1 56 ^b)	-84	-230	-12.7
	$d(T-A-G-G-U^*-C-A-A-U^*-A-C-T)$ 23	3 53 ^a)	-86	-239	-12.0
3'-d(A-T-C-C-A-G-T-T-A-T-G-A) 18 50°) -91 -254 -11.8	d(A-T-C-C-A-G-T-T-A-T-G-A) 18	3 50 ^b)	-91	-254	-11.8
5'-d(T-A-G-G-T-C*-A-A-T-A-C*-T) 24 57 ^a) -70 -187 -12.3	$d(T-A-G-G-T-C^*-A-A-T-A-C^*-T) \qquad 24$	1 57 ^a)	-70	-187	-12.3
3'-d(A-T-C*-C*-A-G-T-T-A-T-G-A) 25 56 ^b) -87 -239 -13.1	d(A-T-C*-C*-A-G-T-T-A-T-G-A) 25	5 56 ^b)	-87	-239	-13.1
5'-d(T-A-G-G-T-C*-A-A-T-A-C*-T) 24 54 ^a) -81 -222 -12.0	$d(T-A-G-G-T-C^*-A-A-T-A-C^*-T)$ 24	1 54 ^a)	-81	-222	-12.0
3'-d(A-T-C-C-A-G-T-T-A-T-G-A) 18 53 ^b) -84 -234 -11.8	d(A-T-C-C-A-G-T-T-A-T-G-A) 18	3 53 ^b)	-84	-234	-11.8

Table. Tm-Values and Thermodynamic Data of Base Modified Oligonucleotides

^a) 1 M NaCl, 100 mM MgCl₂, 60 mM Na-cacodylate (pH 7.0). ^b) 100 mM NaCl, 10 mM MgCl₂, and 10 mM Na-cacodylate (pH 7.0) with 5 μ M single strand concentration. The residues with a^{*} represent the modified constituents. Marcel Dekker, Inc.

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2. Synthesis of the Pyrimidine Phosphoramidites 3 and 4

The reaction sequence performed on the 7-deazapurine nucleosides was also used for the preparation of the phosphoramidite **3** (Scheme 3). The phosphoramidite **4** was prepared in a slightly different way. The 5-iodo-2'-deoxycytidine (14) was directly used for the cross coupling reaction yielding the derivative **15**. The crude reaction product was tritylated and benzoylated in a one-pot reaction to give the intermediate **16**. Afterwards, the phosphoramidite **4** was prepared as described before.

3. Synthesis and Properties of Oligonucleotides

The oligonucleotides shown in the Table were prepared by solid-phase synthesis in a 1- μ mole scale using the phosphoramidites **1**–**4**. The coupling yields were always higher than 96%. The oligonucleotides were deprotected and purified as DMTr derivatives. They were detrivated with 3% aq. trifluoroacetic acid on OPC cartridges. In the case of the dU*-derivatives a side product was formed during the solid-phase synthesis when multiple residues were incorporated. The T_m-values of the Table show that the positively charged aminopropargyl chain stabilizes the duplexes. Also degradation with exonucleases is retarded. The favorable properties of such oligonucleotides were used in MALDI-TOF spectrometry to increase the oligonucleotide detection sensitivity (12).

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