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Voltammetric determination of ascorbic acid in the presence of acetaminophen and tryptophan using an improved carbon nanotube paste electrode

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1. Introduction

Electrochemical methods have traditionally been applied in sample analysis and organic and inorganic synthesis because of their low cost, ease of use, and reliability [1,2]. Most recent developments in electroanalytical chemistry include advances in sensor design, chemical modification, and functionalization of electrodes for enhanced sensitivity and selectivity [3]. Electrochemical methods using chemically modified electrodes (CMEs) have been widely used for sensitive and selective detection of trace amounts of important biological compounds, particularly for vitamins such as ascorbic acid (vitamin C, AA) [4–7]. CMEs can catalyze the electrode process at significantly lower overpotential through the selective interaction of an electron mediator with a target analyte [8,9].

Since the discovery of carbon nanotubes (CNTs) in 1991

ABSTRACT

A carbon paste electrode (CPE) modified with carbon nanotubes and 5-amino-3',4'-dimethyl- biphenyl-2-ol (5ADB) is prepared. Under the optimum pH of 7.0, the oxidation of ascorbic acid (AA) on the modified CPE occurs at a potential about 280 mV less positive than that on the unmodified CPE. Some kinetic and thermodynamic parameters for electrocatalytic oxidation of AA, including electron transfer coefficient (α = 0.58) and diffusion coefficient (D = 2.2 × 10⁻⁶ cm²/s), are also determined. AA, acetaminophen (AC), and tryptophan (TRP) were detected simultaneously using the modified CPE. The peak potentials recorded using the modified CPE in phosphate-buffered solution at pH 7.0 were 265, 465, and 780 mV for AA, AC, and TRP, respectively. The modified CPE was successfully used to determine the concentrations of AA, AC, and TRP in real samples.

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[10], their properties and applications have been investigated extensively. CNTs are important nanomaterials because of their high chemical stability and surface area, excellent mechanical properties, unique electrical conductivity, and metallic structural characteristics. Their special tube structure endows CNTs with these unique properties. In addition, the subtle electronic behavior of CNTs means that they have the ability to promote electron transfer reactions, and have a strong electrocatalytic effect when used as electrode materials [11–17].

AA is a six-carbon lactone that is synthesized from glucose in the liver of most mammalian species except humans [18]. AA is an electron donor, and this property accounts for all of its known functions, including acting as a potent water-soluble antioxidant in humans [19]. AA is present in many biological systems and multivitamin preparations. It is commonly used to prevent and treat the common cold, mental illness, infertility,

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cancer, photoaging, skin disorders, and AIDS [20,21]. Therefore, detecting and determining the concentration of AA in samples are of great importance for pharmaceutical, clinical, and food industries. To date, many methods have been used to detect AA including spectroscopy [22], enzymatic analysis [23], and electrochemistry [24]. Importantly, AA is electroactive at carbon and platinum electrodes. However, its oxidation requires undesirably high overvoltages. To improve the sensitivity and selectivity of AA detection, the operating potentials should be lowered and the oxidation currents should be increased. This can be achieved by modifying the surface of electrodes with suitable electrocatalysts [25–27].

Acetaminophen (N-acetyl-P-aminophenol or paracetamol, AC) is a common antipyretic and analgesic drug typically used to reduce mild to moderate pain or fever [28]. It is frequently recommended when aspirin may present problems to a patient, particularly in pediatrics or after surgery [28]. Ingestion of excess AC causes toxic metabolites to accumulate, which may cause severe and sometimes fatal hepatotoxicity and nephrotoxicity, and in some cases, renal failure [29]. Until 2010, AC was believed safe in pregnancy. However, a recent study linked it to infertility in adults exposed to AC as fetuses [30]. Many methods have been used to determine the concentration of AC, such as titrimetry, spectrophotometry, flow injection, and chromatography. Electrochemical methods have also been used extensively because of their elegance and sensitivity [31–36]. Tryptophan (TRP) is one of the 20 standard amino acids, and is essential in many organisms, including humans. TRP cannot be synthesized by organisms, so it must be part of their diet. TRP is a building block in protein biosynthesis, and also functions as a biochemical precursor for biologically important molecules, such as the neurotransmitter serotonin and the neurohormone melatonin [37]. It has been reported that inadequate serotonin and melatonin levels in human brains are partially responsible for depression [38], and Alzheimer's and Parkinson's diseases [39], respectively. TRP is currently prescribed for a wide range of disorders including anxiety, insomnia, addiction, and obesity. As a result, analysis of TRP is of great importance in the fields of biochemistry, pharmaceuticals, and dietetics. Methods to determine the concentration of TRP are mainly based on HLPC and spectrophotometry, although electroanalytical methods have received interest in recent years because of their sensitivity, accuracy, and simplicity [40-43].

The adjunct presence of AA with AC increases the positive effects of AC and decreases its toxicity [19,44]. High doses of AA may lower the amount of AC passed in urine, which could cause the level of this drug in blood to rise [45]. Lactose and AA are not contaminated by TRP when used as anti-oxidizing reagents. Both improve TRP recovery, but AA is more effective [46]. Therefore, determination of these compounds in the presence of each other is important. However, most previous electrochemical studies have only considered individual determination of AA, AC, and TRP, or with other substrates. A single study has reported the simultaneous determination of AA, AC, and TRP using a modified carbon nanotube paste electrode (CNPE) [24]. In the present work, we prepare a novel electrode com-

posed of a CNPE modified with 5-amino-3',4'-dimethyl- biphenyl-2-ol (5ADB-CNPE), and investigate its performance for the electrocatalytic determination of AA in aqueous solution. We also evaluate the analytical performance of the modified electrode for quantification of AA in the presence of AC and TRP.

2. Experimental

2.1. Apparatus and chemicals

Electrochemical measurements were performed with an Autolab potentiostat/galvanostat (PGSTAT 12, Eco Chemie, The Netherlands). Experimental conditions were controlled with General Purpose Electrochemical System (GPES) software. A conventional three electrode cell was used at (25±1) °C. An Ag/AgCl/KCl (3.0 mol/L) electrode, platinum wire, and 5ADB-CNPE were used as the reference, auxiliary, and working electrodes, respectively. pH was monitored with a pH/ion meter (Metrohm 827).

All solutions were freshly prepared with double distilled water. AA, AC, TRP, and all other reagents were of analytical grade and were purchased from Merck (Darmstadt, Germany). Graphite powder and paraffin oil (DC 350, density = 0.88 g/cm³) as binding agents (both from Merck) were used to prepare pastes. Multi-walled CNTs (purity > 95%) with outer and inner diameters of 10–20 and 5–10 nm, respectively, and lengths of 0.5–200 μ m were purchased from Nanostructured & Amorphous Materials, Inc. Buffer solutions were prepared from orthophosphoric acid and its salts in the pH range of 2.0–9.0.

2.2. Synthesis of 5ADB

To prepare 5ADB, *N*-(2-hydroxy-phenyl)-acetamide (3.02 g, 20 mmol), 4-bromo-*o*-xylene (3.70 g, 20 mmol), and PdCl₂(PPh₃)₄ (2.81 g) were added to a mixture of dimethylacetamide (20 ml) and *t*-BuOK (6.72 g). The mixture was stirred and heated at about 90 °C for 12 h. The progress of the reaction was monitored by TLC. After completion of reaction, the resulting mixture was allowed to cool for a few minutes, and then Pd(PPh₃)₄ was collected by vacuum filtration using a Hirsch funnel. Chloroform was added to the mixture, which was then filtered to recover the catalyst. The resulting acetamide was readily cleaved by barium hydroxide with a yield of > 90%. The crude product was recrystallized from isopropanol and chloroform (20:80) to afford 5ADB in 83% yield.

2.3. Preparation of 5ADB-CNPE

5ADB-CNPE was prepared by mixing 5ADB (0.01 g) with graphite powder (0.89 g) and CNTs (0.1 g) using a mortar and pestle. Paraffin oil (\sim 0.7 ml) was added and the mixture was ground for 20 min until a uniform paste was obtained. The paste was packed into the end of a glass tube with an internal diameter of 3.4 mm and length of 15 cm. A copper wire was inserted into the carbon paste as an electrical contact. When necessary, a new surface was obtained by pushing some of the

paste out of the tube and polishing the end with weighing paper.

For comparison, a 5ADB-modified CPE lacking CNTs (5ADB-CPE), a CNPE lacking 5ADB, and an unmodified CPE lacking both 5ADB and CNTs were also prepared in the same manner.

3. Results and discussion

3.1. Electrochemical properties of 5ADB-CNPE

The electrochemical properties and, in particular, the electrocatalytic activity of 5ADB in aqueous media have not been reported. Therefore, we studied the electrochemical properties of 5ADB-CNPE in phosphate-buffered solution (PBS) at pH 7.0 using cyclic voltammetry (CV), as shown in Fig. 1. One of the advantages of 5ADB as an electrode modifier is its insolubility in aqueous media. 5ADB-CNPE exhibited reproducible, well-defined anodic, and cathodic peaks with E_{pa} , E_{pc} , and $E^{o'}$ of 0.28, 0.18, and 0.23 V vs. Ag/AgCl/KCl (3.0 mol/L), respectively. The observed peak separation potential ΔE_p (= E_{pa} - E_{pc}) of 100 mV, was greater than the value of 59/n mV expected for a reversible system [47], suggesting that the redox couple of 5ADB in 5ADB-CNPE shows quasi-reversible behavior in an aqueous medium. The effect of the potential scan rate (ν) on the electrochemical properties of 5ADB-CNPE was also studied by CV. Plots of both the anodic and cathodic peak currents (I_p) depended linearly on ν in the range of 10 to 800 mV/s (Fig.

1(b)), indicating that the redox process of 5ADB at the modified electrode is diffusionless.

The apparent charge transfer rate constant, k_s , and charge transfer coefficient, α , of a surface-confined redox couple can be determined from CV experiments using the variation of anodic and cathodic peak potentials with logarithm of scan rate according to the procedure of Laviron [48]. Figure 1(c) shows such plots, indicating that E_p is proportional to the logarithm of scan rate for ν higher than 3 V/s. The slopes of the plots in Fig. 1(c) can be used to extract the kinetic parameters α_c and α_a (cathodic and anodic transfer coefficients, respectively). The slopes of the linear segments are equal to $-2.303RT/\alpha nF$ and $2.303RT/(1-\alpha)nF$ for the cathodic and anodic peaks, respectively, so $\alpha = 0.5$.

Equation (1) can be used to determine the electron transfer rate constant between the modifier (5ADB) and CNPE:

 $\log k_{\rm s} = \alpha \log(1-\alpha) + (1-\alpha)\log\alpha - \log(RT/nFv)$

$$\alpha(1-\alpha)nF\Delta E_{\rm p}/2.3RT$$
(1)

(4)

where $(1-\alpha)n_{\alpha} = 0.5$, ν is the sweep rate, and all other symbols have their conventional meanings. k_s was calculated to be 23.9 s⁻¹ using Eq. (1).

3.2. Influence of pH

The electrochemistry of 5ADB is generally pH dependent, so the electrochemical behavior of 5ADB-CNPE was studied at different pH using CV (Fig. 2(a)). The anodic and cathodic peak potentials of 5ADB-CNPE shifted to less positive values with



Fig. 1. (a) CVs of 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) at scan rates of 10 (1), 20 (2), 50 (3), 100 (4), 200 (5), 300 (6), 400 (7), 500 (8), 600 (9), 700 (10), and 800 mV/s (11); (b) *I*_p vs. scan rate; (c) Variation of *E*_p versus the logarithm of high scan rates.



Fig. 2. (a) CVs of 5ADB-CNPE at a scan rate of 20 mV/s and pH of 2.0 (1), 3.0 (2), 4.0 (3), 5.0 (4), 6.0 (5), 7.0 (6), 8.0 (7), and 9.0 (8); (b) Plots of *E*_{pa}, *E*_{pc}, and *E*_{1/2} vs. pH.



Fig. 3. CVs of unmodified CPE in 0.1 mol/L PBS (pH 7.0) (1) and 0.4 mmol/L AA (2), 5ADB-CNPE in 0.1 mol/L PBS (3), CNPE in 0.4 mmol/L AA (4), 5ADB-CPE in 0.4 mmol/L AA (5), and 5ADB-CNPE in 0.4 mmol/L AA (6). In all cases the scan rate was 10 mV/s.

increasing pH. Figure 2(b) shows potential-pH diagrams constructed by plotting anodic, cathodic, and half-wave potentials as functions of pH. The slopes of these plots are 47.84, 49.26, and 48.29 mV/pH for E_{pa} , E_{pc} , and $E_{1/2}$, respectively, indicating that the system obeys the Nernst equation with equal electron and proton transfer reaction [47].

3.3. Electrocatalytic oxidation of AA at a 5ADB-CNPE

Figure 3 depicts the CV responses for the electrochemical oxidation of 0.4 mmol/L AA at the unmodified CPE, CNPE, 5ADB-CPE, and 5ADB-CNPE. While the anodic peak potentials for AA oxidation at the CNPE and unmodified CPE are 500 and 560 mV, respectively, the corresponding potentials at both 5ADB-CNPE and 5ADB-CPE are ~280 mV. These results indicate that the peak potential for AA oxidation at 5ADB-CNPE and 5ADB-CPE shift to a more negative potential by ~220 and 280 mV compared with those of CNPE and unmodified CPE, respectively. However, 5ADB-CNPE shows much higher anodic peak current for the oxidation of AA than 5ADB-CPE, indicating that the combination of CNTs and 5ADB significantly improves the performance of the electrode toward AA oxidation. In fact, 5ADB-CNPE in the absence of AA exhibited a stable redox reaction (Fig. 3(3)) in 0.1 mol/L PBS at pH 7.0. However, a large increase in the anodic peak current was observed in the presence of 0.4 mmol/L AA (Fig. 3(6)), which is attributed to the

strong electrocatalytic effect of 5ADB-CNPE towards AA [47].

The effect of scan rate on the electrocatalytic oxidation of AA at 5ADB-CNPE was investigated by CV (Fig. 4(a)). The oxidation peak potential shifted to a more positive value with increasing scan rate, confirming the kinetic limitation of the electrochemical reaction. Also, a plot of peak height (I_p) vs. the square root of scan rate ($\nu^{1/2}$) was linear in the range of 2–20 mV/s, suggesting that at sufficient overpotential, the process is diffusion rather than surface controlled (Fig. 4(b)).

The Tafel slope (*b*) can be obtained from the slope of a plot of E_p vs. logv using Eq. (2) [47]:

$$E_{\rm p} = b/2 \log v + {\rm constant}$$
 (2)

The Tafel slope was found to be 0.14 V (Fig. 4(c)), which indicates that a one-electron transfer process is the rate-limiting step assuming a transfer coefficient (α) of about 0.58.

3.4. Chronoamperometric measurements

Chronoamperometric measurements of AA at 5ADB-CNPE were carried out by setting the working electrode potential at 0.35 V at the first potential step and 0.05 V at second potential step vs. Ag/AgCl/KCl (3.0 mol/L) for various concentrations of AA in PBS (pH 7.0), as presented in Fig. 5. For an electroactive material (AA in this case) with a diffusion coefficient *D*, the current observed for the electrochemical reaction under mass transport-limited conditions can be described by the Cottrell equation [47]. Experimental plots of *I* vs. $t^{-1/2}$ were drawn, and the best fits for different concentrations of AA were determined (Fig. 5(b)). The slopes of the resulting straight lines were then plotted against AA concentration (Fig. 5(c)). From the resulting slope and Cottrell equation, the mean value of *D* was found to be 2.2 × 10⁻⁶ cm²/s.

Chronoamperometry can also be used to evaluate the catalytic rate constant, *k*, for the reaction between AA and 5ADB-CNPE according to the method of Galus [49]:

 $I_C/I_L = \gamma^{1/2} [\pi^{1/2} \operatorname{erf} (\gamma^{1/2}) + \exp(-\gamma)/\gamma^{1/2}]$ (3) where I_C is the catalytic current of AA at 5ADB-CNPE, I_L is the limited current in the absence of AA, and $\gamma = kC_bt$ is the argument of the error function (C_b is the bulk concentration of AA). When γ exceeds 2, the error function is almost equal to 1, so Eq. (3) can be reduced to:

$$I_{\rm C}/I_{\rm L} = \pi^{1/2} \gamma^{1/2} = \pi^{1/2} (kC_{\rm b}t)^{1/2}$$
(4)

where t is the time elapsed. Equation (4) can be used to calcu-



Fig. 4. (a) CVs of 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) containing 10.0 μmol/L AA at scan rates of 2 (1), 4 (2), 6 (3), 8 (4), 10 (5), 15 (6), and 20 mV/s (7); (b) Anodic peak current vs. ν^{1/2}; (c) Anodic peak potential vs. logv.



Fig. 5. (a) Chronoamperograms obtained at 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) for concentrations of AA of 0.0 (1), 0.1 (2), 0.2 (3), 0.4 (4), 0.6 (5), 0.8 (6), 1.0 (7), 1.2 (8), 1.6 (9), and 1.8 mmol/L (10); (b) Plots of *I* vs. $t^{-1/2}$ obtained from chronoamperograms (2)–(10) in (a); (c) Plot of the slope of the straight lines against AA concentration; (d) Dependence of I_{cat}/I_1 on $t^{1/2}$ derived from the data in chronoamperogram (2)–(10) in (a).

late *k* of the catalytic process from the slope of I_C/I_L vs. $t^{1/2}$ at a given AA concentration. The average value of *k* was calculated to be 2.4 × 10⁴ L/(mol·s).

3.5. Electrocatalytic determination of AA

The electrocatalytic peak current of AA oxidation at the surface of the modified electrode can be used to determine the concentration of AA in solution. Therefore, square wave voltammetry (SWV) experiments were performed using the modified electrode in PBS containing different concentrations of AA (Fig. 6). The mediated oxidation peak currents of AA at the surface of the modified electrode were proportional to the concentration of AA within the range 6.0×10^{-7} – 1.0×10^{-3} mol/L in the SWV measurements. The obtained detection limit (3σ) was 3.0×10^{-7} mol/L.

3.6. Simultaneous determination of AA, AC, and TRP

The simultaneous determination of AA, AC, and TRP using a 5ADB-CNPE has not been reported, so the main objective of this study was to detect AA, AC, and TRP simultaneously using 5ADB-CNPE. This was achieved by simultaneously changing the concentrations of AA, AC, and TRP, and recording SWVs, as shown in Fig. 7. SWVs showed well-defined anodic peaks at potentials of 265, 465, and 780 mV, corresponding to the oxi-

dation of AA, AC, and TRP, respectively. These peaks indicate that simultaneous determination of these compounds at 5ADB-CNPE is feasible.

The sensitivity of the modified electrode for the oxidation of AA was 0.019 (μ A·L)/ μ mol, which is equal to that obtained in the absence of AC and TRP (see section 3.5). This indicates that the oxidation processes of these compounds at 5ADB-CNPE are independent and, therefore, simultaneous determination of their concentrations in mixtures without significant interference is possible.

3.7. Sample analysis using 5ADB-CNPE

The catalytic oxidation of AA by 5ADB-CNPE was also investigated using real samples. The concentration of AA in pharmaceutical samples, such as vitamin C ampoules and effervescent tablets, was determined by recording SWVs of 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) containing real samples.

3.7.1. Determination of the concentration of AA in an ampoule

AA from an ampoule (1 ml) was diluted to 10 ml with PBS (0.1 mol/l, pH 7.0) and then different amounts of the diluted solution were transferred into a series of volumetric flasks and diluted to 10 ml with PBS. Each sample solution was transferred into an electrochemical cell and SWVs were recorded between 0.0 and 0.5 V at a scan rate of 10 mV/s. I_{Pa} was meas-



Fig. 6. (a) SWVs of 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) containing different concentrations of AA. AA concentrations (µmol/L): (1) 0.6; (2) 2.5; (3) 10.0; (4) 20.0; (5) 35.0; (6) 40.0; (7) 60.0; (8) 80.0; (9) 100.0; (10) 150.0; (11) 200.0; (12) 300.0; (13) 500.0; (14) 800.0; (15) 1000.0. (b) A plot of electrocatalytic peak current as a function of AA concentration in the range of 0.6–1000.0 µmol/L.



Fig. 7. (a) SWVs of 5ADB-CNPE in 0.1 mol/L PBS (pH 7.0) containing different concentrations of AA, AC, and TRP. Concentrations of AA+AC+TRP (μ mol/L): (1) 0.6 + 35.0 + 50.0; (2) 15.0 + 75.0 + 100.0; (3) 175.0 + 125.0 + 175.0; (4) 225.0 + 175.0 + 200.0; (5) 300.0 + 250.0 + 250.0; (6) 375.0 + 300.0 + 300.0; (7) 450.0 + 350.0 + 350.0; (8) 550.0 + 425.0 + 400.0; (9) 650.0 + 475.0 + 475.0. (b)–(d) are plots of I_p vs. concentration of AA, AC, and TRP, respectively.

ured at the oxidation potential of AA and its concentration was obtained from the calibration plot. This procedure was repeated three times for each sample. The average amount of AA in the ampoule was determined to be 500.5 mg AA per 5 ml, consistent with the value on the label (500 mg AA per 5 ml). Also, different amounts of the diluted AA solutions together with standard AC and TRP solutions were added to a series of volumetric flasks and diluted to 10 ml with PBS. SWVs were recorded and the anodic peak currents for each of AA, AC, and TRP were measured at their own oxidation potentials. According to the results listed in Table 1, the concentrations of AA, AC, and TRP were accurately determined with high reproducibility, which indicates that the sensor can be used to analyze these compounds with no significant interference.

3.7.2. Determination of the amount of AA in effervescent tablets

To evaluate the analytical applicability of 5ADB-CNPE, it was also used to determine the amount of AA in effervescent

tablets. Based on the repeated SWV responses (n = 3) of the diluted analytes and samples that were spiked with specified concentrations of AA, measurements were made to determine the amount of AA in effervescent tablets. The results presented in Table 2 show that the RSD and the recovery rates of the spiked samples were acceptable. The average amount of AA in each effervescent tablet was found to be 553.7 mg, which agrees with that reported on the label (550 mg). Thus, the modified electrode can be used to efficiently determine the amounts of AA, AC, and TRP in effervescent tablets without significant interference from the other compounds.

4. Conclusions

A novel modified CNPE was constructed to detect AA. The results show that the oxidation of AA was catalyzed at pH 7.0, and the peak potential of AA was shifted to less positive potential at 5ADB-CNPE than at an electrode lacking 5ADB.

Table 1

Determination of AA, AC, and TRP in an ampoule containing AA.

			-	•							
AA	AC	TRP	AA			AC			TRP		
ampoule	added	added	Found	Recovery	RSD	Found	Recovery	RSD	Found	Recovery	RSD
(µmol/L)	(µmol/L)	(µmol/L)	(µmol/L)	(%)	(%)	(µmol/L)	(%)	(%)	(µmol/L)	(%)	(%)
100.0	0.0	0.0	103.0	103.0	1.5	_		_	_		_
100.0	30.0	30.0	99.7	99.7	1.6	29.2	97.3	1.8	30.1	100.3	3.5
150.0	0.0	0.0	149.6	99.7	2.8	_	_	_	_	_	_
150.0	50.0	50.0	150.2	100.1	1.3	50.7	101.4	3.2	51.0	102.0	1.3
230.0	0.0	0.0	228.4	99.3	1.5	_	_	_	_	_	_
230.0	100.0	200.0	227.7	99.0	1.8	100.9	100.9	1.7	196.3	98.1	1.1
D a D											

RSD: relative standard deviations.

Table 2

Determination of the amounts of AA, AC, and TRP in AA tablets.

AA	AC	TRP		AA			AC			TRP		
tablet	added	added	Found	Recovery	RSD	Found	Recovery	RSD	Found	Recovery	RSD	
(µmol/L)	(µmol/L)	(µmol/L)	(µmol/L)	(%)	(%)	(µmol/L)	(%)	(%)	(µmol/L)	(%)	(%)	
50.0	0.0	0.0	49.5	99.0	1.89	—	—	_	_	_	_	
50.0	70.0	50.0	50.1	100.2	2.4	67.3	96.1	2.3	50.8	101.6	2.8	
100.0	0.0	0.0	102.1	102.1	1.1	_	—	_	_	—	_	
100.0	100.0	80.0	99.4	99.4	3.2	100.1	100.1	1.6	80.2	100.2	2.1	
200.0	0.0	0.0	198.5	99.2	1.8	_		_	_	—	_	
200.0	130.0	150.0	204.3	102.1	2.4	128.9	99.1	1.5	147.1	98.1	1.6	

Graphical Abstract

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Voltammetric determination of ascorbic acid in the presence of acetaminophen and tryptophan using an improved carbon nanotube paste electrode

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A new electrochemical sensor based on a modified carbon nanotube paste electrode is constructed for simultaneous determination of ascorbic acid (AA), acetaminophen (AC), and tryptophan (TRP).

5ADB-CNPE exhibited a linear response over a wide concentration range of 6.0 × 10⁻⁷–1.0 × 10⁻³ mol/L with a detection limit of 3.0 × 10⁻⁷ mol/L. This modified electrode separated the anodic oxidation peak potentials of AA, AC, and TRP in a well defined manner, allowing the concentrations of AA, AC, and TRP to be determined without apparent intermolecular effects. The values of *k*, *D*, and *α* for 5ADB-CNPE were found to be 2.4 × 10⁴ L/(mol·s), 2.2 × 10⁻⁶ cm²/s, and 0.58, respectively. 5ADB-CNPE was also successfully used for determination of AA, AC, and TRP in actual samples.

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