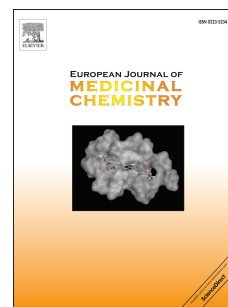


Accepted Manuscript

Synthesis, antiarrhythmic activity, and toxicological evaluation of mexiletine analogues

Mariagrazia Roselli, Alessia Carocci, Roberta Budriesi, Matteo Micucci, Maddalena Toma, Lorenzo Di Cesare Mannelli, Angelo Lovece, Alessia Catalano, Maria Maddalena Cavalluzzi, Claudio Bruno, Annalisa De Palma, Marialessandra Contino, Maria Grazia Perrone, Nicola Antonio Colabufo, Alberto Chiarini, Carlo Franchini, Carla Ghelardini, Solomon Habtemariam, Giovanni Lentini



PII: S0223-5234(16)30444-5

DOI: [10.1016/j.ejmech.2016.05.046](https://doi.org/10.1016/j.ejmech.2016.05.046)

Reference: EJMECH 8638

To appear in: *European Journal of Medicinal Chemistry*

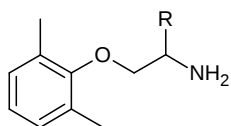
Received Date: 14 March 2016

Revised Date: 20 May 2016

Accepted Date: 21 May 2016

Please cite this article as: M. Roselli, A. Carocci, R. Budriesi, M. Micucci, M. Toma, L. Di Cesare Mannelli, A. Lovece, A. Catalano, M.M. Cavalluzzi, C. Bruno, A. De Palma, M. Contino, M.G. Perrone, N.A. Colabufo, A. Chiarini, C. Franchini, C. Ghelardini, S. Habtemariam, G. Lentini, Synthesis, antiarrhythmic activity, and toxicological evaluation of mexiletine analogues, *European Journal of Medicinal Chemistry* (2016), doi: 10.1016/j.ejmech.2016.05.046.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Table of Contents graphic**Synthesis, Antiarrhythmic Activity, and Toxicological****Evaluation of a Series of Mexiletine Analogues.**

mexiletine: R = Me, EC₅₀ = 11.6 μM

1b: R = *t*Bu, EC₅₀ = 2.1 μM

Synthesis, Antiarrhythmic Activity, and Toxicological Evaluation of Mexiletine Analogues.

Mariagrazia Roselli,[†] Alessia Carocci,^{*,†} Roberta Budriesi,[¥] Matteo Micucci,[¥] Maddalena Toma,[†] Lorenzo Di Cesare Mannelli,[^] Angelo Lovece,[†] Alessia Catalano,[†] Maria Maddalena Cavalluzzi,[†] Claudio Bruno,[†] Annalisa De Palma,[§] Marialessandra Contino,[†] Maria Grazia Perrone,[†] Nicola Antonio Colabufo,[†] Alberto Chiarini,[¥] Carlo Franchini,[†] Carla Ghelardini,[^] Solomon Habtemariam,[¶] Giovanni Lentini.[†]

[†]*Dipartimento di Farmacia-Scienze del Farmaco, Università degli Studi di Bari Aldo Moro, via E. Orabona n. 4, 70126 Bari, Italy.* [¥]*Dipartimento di Farmacia e Biotecnologie, Università di Bologna, Via Belmeloro 6, 40126 Bologna, Italy.* [^]*Dipartimento di Farmacologia Preclinica e Clinica, Università di Firenze, Viale Pieraccini 6, 50139 Firenze, Italy.* [§]*Dipartimento di Bioscienze, Biotecnologie e Biofarmaceutica, Università degli Studi di Bari Aldo Moro, Via Orabona 4, 70126 Bari, Italy.* [¶]*Pharmacognosy Research Laboratories & Herbal Analysis services, Chatham-Maritime, Kent ME4 4TB, UK.*

*Corresponding author: Dott.ssa Alessia Carocci,

Dipartimento di Farmacia-Scienze del Farmaco, Università degli Studi di Bari A. Moro, via Orabona 4, 70126 Bari, Italy, phone number: (+39) 080-5442745; fax number: (+39) 080-5442724, E-mail address: alessia.carocci@uniba.it

Abstract

Four mexiletine analogues have been tested for their antiarrhythmic, inotropic, and chronotropic effects on isolated guinea pig heart tissues and to assess calcium antagonist activity, in comparison with the parent compound mexiletine.¹ All analogues showed from moderate to high antiarrhythmic activity. In particular, three of them (**1b,c,e**) were more active and potent than the reference drug, while exhibiting only modest or no negative inotropic and chronotropic effects and vasorelaxant activity, thus showing high selectivity of action. All compounds showed no cytotoxicity and **1b,c,d** did not impair motor coordination. All in, these new analogues exhibit an interesting cardiovascular profile and deserve further investigation.

Keywords: Mexiletine; Antiarrhythmics; P-glycoprotein; Inotropism; MW-assisted synthesis

¹ Abbreviations used: CNS, central nervous system; DMEM, Dulbecco's modified eagle's medium; GPILSM, guinea pig ileum longitudinal smooth muscle; HepG2, human hepatocellular liver carcinoma cells; MDCK, Madin-Darby canine kidney; MDCK-MDR1, MDCK overexpressing P-gp; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; MW, microwave; P-gp, P-glycoprotein ; PSS, physiological salt solution.

1. Introduction

One of the major causes of mortality is represented by cardiovascular diseases and about 25% of cardiovascular-related adverse events are caused by arrhythmias. The term arrhythmia defines an abnormality in the sequence and/or shape of electrical impulses during the cardiac cycle. A critical role in the regulation of electrical activity of cardiac cells is played by ion channels such as sodium, calcium and potassium channels. Abnormalities in the expression and/or function of ion channels are responsible of many arrhythmias [1,2], thus, they represent one of the targets for antiarrhythmic therapy. Indeed, most of the existing antiarrhythmic agents, such as lidocaine, tocainide, and mexiletine [3,4] are ion channel blockers. Mexiletine (**1a**, Fig. 1) is an antiarrhythmic drug belonging to class IB. It is a voltage-gated sodium channel blocker whose mechanism of action is related to its preferential binding to the open and/or inactivated states of the channels. The selective action of mexiletine on pathologically hyperactive tissues seems to be related to the use-dependent blockade of sodium channels [5]. Mexiletine undergoes extensive metabolism; eleven phase I metabolites of mexiletine have been identified [6,7]. Mexiletine is predominantly used in the treatment of ventricular arrhythmias, however, recent studies have demonstrated its therapeutic value for treating many other disorders associated with voltage-gated sodium channel dysfunction including chronic painful diabetic neuropathy [8], neuropathic pain [9], myotonic syndromes [10], and Timothy syndrome [11]. Recently, mexiletine has been successfully used in the treatment of sporadic amyotrophic lateral sclerosis [12–14]. Despite all these reported therapeutic values, the use of mexiletine is characterized by several drawbacks including epigastric discomfort, atrioventricular heart block, and CNS disturbances [15]. Hence, it has now been discontinued in many countries including the US and the UK [16]. Consequently, numerous attempts have been made in recent years to develop an alternative to mexiletine, including the design of new analogues that offer the same pharmacological effect but without the unwanted side effects. In the past decade, our own laboratories have been focusing on the development of such analogues through which the structural determinants for blocking voltage-gated sodium channels of skeletal muscles have been established

[17–20]. As these identified compounds were more potent channel blockers than the parent compound, they have been proposed as mexiletine alternatives for the treatment of myotonias. We also established that the activity of mexiletine-like sodium channel blockers is strongly linked to the part of the molecule nearby the asymmetric carbon atom. In order to evaluate the potential of these compounds for treating arrhythmias, we selected the most interesting compounds from the previously reported series (**1c–e**, Fig. 1) that bear a bulky group at the stereogenic center. These compounds, along with a new additional compound, **1b**, bearing a *tert*-butyl group as substituent were prepared by alternative and efficient synthetic routes. The biological activity of all compounds was assessed with due comparison with mexiletine.

2. Results and discussion

2.1 Chemistry

Mexiletine (**1a**) and its analogues (**1b,c**) were prepared in two steps according to the synthetic route depicted in Scheme 1, carried out in both traditional conditions and microwave-assisted synthesis. In the first step, 2,6-dimethylphenol underwent Williamson reaction [21] with the suitable halo ketone (**2a–c**) to obtain aryloxyalkyl ketones (**3a–c**) which were converted into the corresponding amines **1a–c** by reductive amination under both traditional and microwave conditions [22]. The corresponding hydrochloride salts (**1a–c·HCl**) were obtained by treating **1a–c** with gaseous HCl. Mexiletine analogues **1d,e** were prepared as depicted in Scheme 2. Compounds **5d,e** were obtained by protecting the corresponding commercial valinol and phenylalaninol, respectively, with di-*tert*-butyl dicarbonate [23,24]. Compounds **5d,e** were subjected to Mitsunobu reaction with 2,6-dimethylphenol as previously described [25]. The so obtained aryl alkyl ethers **6d,e** were treated with gaseous HCl to give the corresponding amines as hydrochloride salts (**1d,e·HCl**).

2.2 Biological results

Mexiletine and its analogues (**1a–e**) were tested *in vitro* for their antiarrhythmic activity on guinea pig isolated left atria driven at 1 Hz. Compound **1b,c,e** increased the threshold of ac-arrhythmia more than mexiletine did, while **1d** showed an anti-arrhythmic activity comparable to that of the parent compound, mexiletine (Table 1). In particular, compound **1c** was found to be 27-fold more active than mexiletine. To better define the cardiac profile of mexiletine analogues, their influence on additional cardiac parameters were compared with that elicited by the reference compound, mexiletine (Table 2). All the tested compounds strongly decreased the developed tension on driven left atria with EC₅₀ values within the range of 0.012 and 0.042 mM. The most potent analogue was **1e** showing a negative inotropism 4-fold higher than that of mexiletine (Tab. 2), while its antiarrhythmic potency was 2.7-fold higher than that of mexiletine (Table 1). The effect of **1b** was comparable to that of mexiletine. It is noteworthy that the introduction of an aromatic moiety, in lieu of the methyl one, on the stereogenic center as in **1c** and **1e**, strongly increased antiarrhythmic and negative inotropic potency. On the other hand, the introduction of an isopropyl group, as in **1d**, while slightly affecting the antiarrhythmic activity, enhanced two times the negative inotropic effect. With regard to the negative chronotropic activity on spontaneously beating right atrium, only compound **1d** showed a weak activity, about 900-fold lower than that of mexiletine. Furthermore, **1b,c,e** showed negative inotropism on spontaneously beating isolated right atrium, too. It is noteworthy that this effect was not recorded for mexiletine and **1d** due to their insignificant inotropism resulting from chronotropic effect. Moreover, all the new mexiletine analogues were tested on K⁺-depolarized (80 mM KCl) guinea pig aortic strips to assess their vasorelaxant activity. Data are shown in Table 3 with mexiletine as the reference drug. For all compounds, the intrinsic vasorelaxant activity percentage on aorta was lower than 40% and thus unremarkable. In order to rule out a possible inhibitory effect on nonvascular smooth muscle, such as that displayed by Ca²⁺ channel antagonists, further investigation on relaxant activity using K⁺-depolarized (80 mM KCl) guinea pig ileum longitudinal smooth muscle (GPILSM) was pursued (Table 3). The observed profile of intrinsic activity and potency for **1b,c,d** was, however, substantially similar to that

displayed by mexiletine. Only compound **1e** showed a relaxant potency slightly higher than that of the parent compound, but in the micromolar range. Over all, all the compounds appears to display a higher intrinsic activity in nonvascular than vascular smooth muscle cells. Since mexiletine clinical use is often associated to CNS toxicity, a preliminary toxicological evaluation of mexiletine analogues was performed by assessing the motor coordination of treated animals using the rota-rod test (Table 4). All compounds at the dose of 25 mg/kg ip did not provoke any change in normal behavior of mice as demonstrated in rota-rod experiments. At the dose of 50 mg/kg ip, only compound **1e** showed a significant impairment of mouse motor coordination, with a slight increase in the number of falls being displayed. Since mexiletine is known to undergo extensive first-pass oxidative metabolism [16,26,27] that is linked to the generation of reactive species metabolites and/or cellular oxidative stress, the cytotoxicity of the four analogues was assessed using the MTT cell viability assay [28]. When the human hepatocellular liver carcinoma cells (HepG2) were challenged with the four mexiletine analogues at two concentrations (the first close to their corresponding EC₅₀ values, and the second 10-fold higher than the former), no cytotoxicity was observed (data not shown). This finding was in agreement with the previous report for mexiletine where cell viability was not affected at similar concentration range [16]. In addition, we tried to verify if these analogues may display differential ability to interact with P-glycoprotein (P-gp), an ATPase protein highly expressed in biological membranes, that generally affects pharmacokinetics, efficacy, safety, and tissue levels of drugs. For this, drug-protein interaction was assessed for each compound by using the Madin-Darby canine kidney (MDCK) cells overexpressing the ATPase protein P-gp (MDCK-MDR1). As shown in Table 5, only compounds that bear a phenyl ring near the basic center (**1c,e**) was able to bind P-gp. This data was in good agreement with a recent study on a series of structurally related rigid analogues [29].

3. Conclusions

Four mexiletine analogues differing for the substituent on the stereogenic center have been tested for their antiarrhythmic, inotropic, and chronotropic effects on isolated guinea pig heart tissues and for their calcium antagonist activity on GPILSM, in comparison with mexiletine. Even if with different degrees of potency, antiarrhythmic activity has been observed in all the studied compounds, conceivably suggesting the hypothesis that the substitution of the methyl group of mexiletine with a bulky one, strengthens the interaction with the biological structures involved in the regulation of heart activity. In particular, compounds **1b,c,e** showed high antiarrhythmic activity, exhibiting EC_{50} values in the range between 0.43 and 4.36 μ M, thus comparing favorably with the reference drug mexiletine ($EC_{50} = 11.6 \mu$ M). These compounds exhibit an interesting cardiovascular profile, being endowed with high antiarrhythmic activity and potency with only modest or no negative inotropic and chronotropic effects and vasorelaxant activity. When administered intraperitoneally (25 mg kg^{-1}), all compounds did not impair motor coordination as demonstrated in rota-rod experiments and showed no *in vitro* cytotoxicity at concentration 10 times higher than the corresponding antiarrhythmic EC_{50} values. On the basis that the observed favorable profile should not be affected by interaction with P-gp, compounds **1b** and **1d** could be considered as the preferred choices. Taken all the data together, this work afforded compounds endowed with better antiarrhythmic activity along with similar or less cardiovascular effects than mexiletine, thus presenting a higher selectivity of action and reduced side effects. In this respect, the *t*-butyl substituted mexiletine analogue **1b** seems to be the most interesting compound. Further investigation would allow more complete knowledge about its pharmacological potential and mechanism of action, which in turn might allow the design and development of even more active and potent antiarrhythmic agents.

4. Experimental Protocols

4.1 Chemistry.

Chemicals were purchased from Sigma-Aldrich or Lancaster. Yields refer to purified products and were not optimized. The structures of the compounds were confirmed by routine spectrometric and spectroscopic analyses. Only spectra for compounds not previously described are given. The reaction under microwaves irradiation were carried out at constant temperature in a CEM Discover BenchMate microwave reactor, with continuous stirring. The temperature was measured and controlled by a built-in infrared detector. Melting points were determined on a Gallenkamp apparatus in open glass capillary tubes and are uncorrected. Infrared spectra were recorded on a Perkin-Elmer (Norwalk, CT) Spectrum One FT spectrophotometer and band positions are given in reciprocal centimeters (cm^{-1}). ^1H NMR and ^{13}C NMR spectra were recorded on either a Varian VX Mercury spectrometer operating at 300 and 75 MHz for ^1H and ^{13}C , respectively, or an AGILENT 500 MHz operating at 500 and 125 MHz for ^1H and ^{13}C , respectively, using CDCl_3 and CD_3OD as solvents. Chemical shifts (δ) are reported in ppm relative to the residual non-deuterated solvent resonance: CDCl_3 , $\delta = 7.26$ (^1H NMR) and $\delta = 77.3$ (^{13}C NMR); CD_3OD , $\delta = 3.30$ (^1H NMR) and $\delta = 47.8$ (^{13}C NMR) as internal references. Coupling constants (J) are given in Hz. Gas chromatography (GC)/mass spectroscopy (MS) was performed on a Hewlett-Packard 6890–5973 MSD at low resolution. Liquid chromatography (LC)/mass spectroscopy (MS) was performed on a spectrometer Agilent 1100 series LC-MSD Trap System VL. Elemental analyses were performed on a Eurovector Euro EA 3000 analyzer and the data for C, H, N were within ± 0.4 of theoretical values. Chromatographic separations were performed on silica gel columns by column chromatography on silica gel (Kieselgel 60, 0.040–0.063 mm, Merck, Darmstadt, Germany) as described by Still et al [30]. TLC analyses were performed on precoated silica gel on aluminum sheets (Kieselgel 60 F_{254} , Merck).

4.1.1. 1-(2,6-Dimethylphenoxy)propan-2-amine (1a). The preparation of **1a** was performed under both traditional (method A) and microwaves conditions (method B) [31].

Method A

Ammonium acetate (23.8 g, 310 mmol) and sodium cyanoborohydride (1.4 g, 21.7 mmol) were added to a solution of 1-(2,6-dimethylphenoxy)propan-2-one (**3a**) (5.5 g, 31 mmol) in 40 mL of MeOH. The reaction mixture was stirred for four days at room temperature. 6 M HCl was added dropwise to the mixture reaction to destroy the excess hydride. The organic solvent was removed under vacuum and the residue aqueous phase was made alkaline with NaOH pellets. Then, the aqueous phase was extracted with AcOEt and the combined organic phases were dried (Na₂SO₄). The solvent was evaporated under reduced pressure to give the desired amine (**1a**) as a yellow oil (4.5 g, 81%). Spectroscopic data were in agreement with the literature [21]. *Method B.* In a microwave tube, ammonium acetate (7.35 g, 95.5 mmol) and sodium cyanoborohydride (0.42 g, 6.7 mmol) were added to a solution of 1-(2,6-dimethylphenoxy)propan-2-one (**3a**) (1.7 g, 9.55 mmol) in 15 mL of MeOH. The reaction mixture was stirred for 11 min at 110 °C in a microwave reactor at 95 watt as power max. The processing of reaction mixture was carried out as reported for method A, obtaining **1a** in 72% yield. Spectrometric and spectroscopic data were in agreement with the literature [21].

4.1.2. 1-(2,6-Dimethylphenoxy)propan-2-amine hydrochloride (1a·HCl). **1a** was treated with gaseous HCl to obtain the corresponding hydrochloride salt (**1a·HCl**) as a white solid which was recrystallized from EtOH/Et₂O to afford 2.9 g (40%) of white crystals: mp 215–216 °C; ¹H NMR (500 MHz, d₆-DMSO): δ 1.35 (d, *J* = 6.8 Hz, 2H, CH₃CH), 2.23 (s, 6H, CH₃Ar), 3.50–3.58 (m, 1H, CH), 3.82 (d, *J* = 5.4 Hz, 2H, CH₂), 6.91 (t, *J* = 6.8 Hz, 1H, Ar), 6.93 (t, *J* = 7.3 Hz, 1H, Ar), 7.00 (d, *J* = 7.8 Hz, 2H, Ar), 8.50 (br s, 3H, NH₃⁺); ¹³C NMR (125 MHz, d₆-DMSO): δ 15.5 (2C), 16.4 (1C), 47.3 (1C), 72.2 (1C), 124.7 (1C), 128.9 (2C), 129.3 (2C), 130.9 (2); 154.8 (1C); Anal. (C₁₁H₁₈NOCl) C, H, N.

4.1.3. 1-(2,6-Dimethylphenoxy)-3,3-dimethylbutan-2-amine (1b). Prepared as reported for **1a** (Method B) starting from **3b**. The synthetic pathway provided **1b** as a yellow oil in 34% yield. ¹H NMR (500 MHz, CDCl₃): δ 0.97 (s, 9H, (CH₃)₃C), 1.99 (br s, 2H, NH₂), 2.30 (s, 6H, CH₃Ar), 3.02 (dd, *J* = 9.3, 2.9 Hz, 1H, CHH), 3.63 (t, *J* = 9.3 Hz, 1H, CH), 3.84 (dd, *J* = 9.3, 2.9 Hz, 1H, CHH),

6.93 (t, $J = 7.3$ Hz, 1H, Ar) 7.02 (d, $J = 7.3$ Hz, 2H, Ar); ^{13}C NMR (125 MHz, CDCl_3): δ 15.3 (1C), 16.4 (1C), 26.5 (3C), 33.0 (1C), 60.5 (1C), 65.9 (1C), 123.9 (1C), 128.9 (2C), 130.9 (2C), 155.8 (1C); GC-MS (70 eV) m/z (%) 221 (M^+ , 1), 86 (100).

4.1.4. 1-(2,6-Dimethylphenoxy)-3,3-dimethylbutan-2-amine hydrochloride (**1b·HCl**). Obtained as reported above for **1a·HCl**: yield: 71%; white crystals; mp: 215–216 °C (abs EtOH/Et₂O). Anal. ($\text{C}_{14}\text{H}_{24}\text{NOCl}$) C, H, N.

4.1.5. 2-(2,6-Dimethylphenoxy)-1-phenylethanamine (**1c**). Prepared as reported for **1a** (Method B) starting from **3c**. The synthetic pathway provided **1c** as a yellow oil in 26% yield. ^1H NMR (500 MHz, CDCl_3): δ 2.17 (s, 6H, CH_3Ar), 3.88–4.08 (m, 2H, CH_2), 4.50–4.57 (m, 1H, CH), 6.88–7.03 (m, 3H, Ar), 7.13–7.22 (m, 3H, Ar), 7.24–7.38 (m, 2H, Ar), 7.50 (d, $J = 6.8$ Hz, 2H, CH_2); ^{13}C NMR (125 MHz, CDCl_3): δ 15.3 (1C), 16.3 (1C), 21.5 (1C), 65.9 (1C), 124.2 (1C), 125.3 (1C), 126.8 (1C), 127.4 (1C), 128.2 (1C), 128.6 (1C), 128.7 (1C), 128.9 (1C), 129.0 (2C), 130.8 (1C), 137.9 (1C); GC-MS (70 eV) m/z (%) 241 (M^+ , <1), 106 (100).

4.1.6 2-(2,6-dimethylphenoxy)-1-phenylethanamine hydrochloride (**1c·HCl**). Obtained as reported above for **1a·HCl**: yield: 80%; white crystals; mp: 259–261 °C (abs EtOH/Et₂O); Anal. ($\text{C}_{16}\text{H}_{20}\text{NOCl}$) C, H, N.

4.1.7. 1-(2,6-Dimethylphenoxy)-3-methylbutan-2-amine hydrochloride (**1d·HCl**). A solution of **5d** (2.5 g, 8.1 mmol) in anhydrous Et₂O (15 mL) was saturated with gaseous HCl and stirred at room temperature for 15 min. Removal of the solvent under reduced pressure gave a solid (1.5 g, 75%) which was recrystallized from EtOH/Et₂O: white crystals: mp 171–172 °C (abs EtOH/Et₂O); Anal. ($\text{C}_{13}\text{H}_{22}\text{NOCl} \cdot 0.17\text{H}_2\text{O}$) C, H, N.

4.1.8. 1-(2,6-Dimethylphenoxy)-3-methylbutan-2-amine (**1d**). Obtained as a yellow oil by extraction from the corresponding hydrochloride salt. ^1H NMR (500 MHz, CDCl_3): δ 0.98 (d, $J = 7.3$ Hz, 3H, CH_3CHCH_3), 0.99 (d, $J = 6.8$ Hz, 3H, CH_3CHCH_3), 1.73–1.82 (m, 1H, CHCH_3), 1.88 (br s, 2H, NH_2), 2.30 (s, 6H, CH_3Ar), 2.98–3.08 (m, 1H, CHCH_2), 3.64 (t, $J = 8.8$ Hz, 1H, CHHCH_2), 3.77

(dd, $J = 3.8, 8.8$ Hz, 1H, CHHCH₂), 6.92 (t, $J = 7.8$ Hz, 1H, Ar), 7.01 (d, $J = 7.8$ Hz, 2H, Ar); ¹³C (125 MHz, CDCl₃): δ 15.3 (1C), 16.4 (1C), 18.2 (1C), 19.4 (1C), 30.8 (1C), 57.1 (1C), 65.9 (1C), 123.8 (1C), 128.9 (2C), 129.0 (1C), 130.1 (1C), 155.7 (1C); MS (70 eV) m/z (%) 207 (M⁺, 3), 72 (100).

4.1.9. *1-(2,6-Dimethylphenoxy)-3-phenylpropan-2-amine hydrochloride (1e·HCl)*. Prepared as reported for **1d·HCl** starting from **5e**. The synthetic pathway provided **1e·HCl** as a white solid in 92% yield. The solid was recrystallized from abs EtOH/Et₂O to give white crystals (40%); mp: 218–219 °C. ¹H NMR (300 MHz, CD₃OD): δ 2.21 (s, 6H, CH₃Ar), 3.02–3.24 (m, 2H, CH₂Ar), 3.72–4.0 (m, 3H, CH₂CH), 6.95–7.05 (m, 3H, Ar), 7.25–7.45 (m, 5H, Ar); ¹³C NMR (100 MHz, CD₃OD): δ 15.3 (2C), 35.4 (1C), 53.5 (1C), 70.0 (1C), 124.6 (1C), 127.4 (1C), 129.0 (4C), 129.1 (2C), 130.5 (2C), 135.5 (1C), 154.6 (1C). Anal. (C₁₇H₂₂NOCl) C, H, N.

4.2. Pharmacology

4.2.1. Details for functional studies.

4.2.1.1. Guinea-Pig Atrial Preparations.

Female guinea pigs (300–400 g) were sacrificed by cervical dislocation. After thoracotomy, the heart was immediately removed and washed by perfusion through the aorta with oxygenated Tyrode solution of the following composition (mM): 136.9 NaCl, 5.4 KCl, 2.5 CaCl₂, 1.0 MgCl₂, 0.4 NaH₂PO₄·xH₂O, 11.9 NaHCO₃ and 5.5 glucose. The physiological salt solution (PSS) was buffered at pH 7.4 by saturation with 95% O₂-5% CO₂ gas, and the temperature was maintained at 35 °C. Isolated guinea pig heart preparations were used, spontaneously beating right atria and left atria driven at 1 Hz. For each preparation, the entire left and right atria were dissected from the ventricles, cleaned of excess tissue, and hung vertically in a 15 mL organ bath containing the PSS continuously bubbled with 95% O₂-5% CO₂ gas at 35 °C, pH 7.4. The contractile activity was recorded isometrically by means of a force transducer (FT 0.3, Grass Instruments Corporation,

Quincy, MA, USA) using Power Lab[®] software (AD-Instruments Pty Ltd, Castle Hill, Australia). The left atria were stimulated by rectangular pulses of 0.6–0.8 ms duration and about 50% threshold voltage through two platinum contact electrodes in the lower holding clamp (Grass S88 Stimulator). The right atrium was in spontaneous activity. After the tissue was beating for several minutes, a length-tension curve was determined, and the muscle length was maintained which elicited 90% of maximum contractile force observed at the optimal length. A stabilization period of 45–60 min was allowed before the atria were used to test compounds. During the equilibration period, the bathing solution was changed every 15 min and the threshold voltage was ascertained for the left atria. Atrial muscle preparations were used to examine the inotropic and chronotropic activity of the compounds (0.01, 0.05, 0.1, 0.5, 1, 5, 10, 50, and 100 μM) dissolved in PSS. During the construction of cumulative dose-response curves, the next higher concentration of the compounds was added only after the preparation reached a steady state.

4.2.1.2. Alternating Current Induced Arrhythmia in Guinea Pig Left Atrial Preparations [32,33].

Left atria from guinea pigs were fixed to platinum wire electrodes and placed between two parallel platinum field electrodes. The preparations were mounted vertically in a 15 mL organ bath containing a modified Krebs-Henselait solution of the following composition (mM): 118 NaCl, 4.7 KCl, 2.5 CaCl_2 , 1.2 MgSO_4 , 1.2 $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$, 24.9 NaHCO_3 , and 10.1 glucose. The solution was buffered at pH 7.4 by saturation with 95 % O_2 -5 % CO_2 gas, and the temperature was maintained at 35 °C. The preparations were connected to a force transducer. Isometric force contraction was recorded using Power Lab[®] software. Arrhythmias were induced by application of sinusoidal alternating current (50 Hz) of increasing strength to the isolated heart preparations, 2 and 4 $\text{mA}/(\text{cm}^2 \cdot \text{s})$ to induce arrhythmia corresponding to an increase in field intensity of 0.3 $\text{V}/(\text{cm} \cdot \text{s})$. The current intensity (mA/cm^2) at which extra beats occurred was called “threshold of ac-arrhythmia” [34]. The preparations were allowed to stabilize for 60 min. Then concentration-response curves for

threshold of ac-arrhythmia were recorded by cumulative application of compounds to the bathing solution every 30 min.

4.2.1.3. Guinea-Pig Aortic Strips. The thoracic aorta was removed and placed in Tyrode solution of the following composition (mM): 118 NaCl, 4.75 KCl, 2.54 CaCl₂, 1.20 MgSO₄, 1.19 KH₂PO₄, 25 NaHCO₃ and 11 glucose equilibrated with 95 % O₂-5 % CO₂ gas at pH 7.4. The vessel was cleaned of extraneous connective tissue. Two helicoidal strips (10 mm x 1 mm) were cut from each aorta beginning from the end most proximal to the heart. Vascular strips were then tied with surgical thread (6-0) and suspended in a jacketed tissue bath (15 mL) containing aerated PSS at 35 °C. Aortic strips were subjected to a resting force of 1 g. Strips were secured at one end to a force displacement transducer (FT 0.3, Grass Instruments Corporation) to monitor changes in isometric contraction and washed every 20 min with fresh PSS for 1 h after the equilibration period; guinea pig aortic strips were contracted by washing in PSS containing 80 mM KCl (equimolar substitution of K⁺ for Na⁺). After the contraction reached a plateau (about 45 min) the compounds (0.1, 0.5, 1, 5, 10, 50, and 100 µM) were added cumulatively to the bath allowing for any relaxation to obtain an equilibrated level of force. Addition of the drug vehicle had no appreciable effect on K⁺-induced contraction (PSS for all compounds).

4.2.1.4. Statistical analysis. Data were analyzed using Student's *t*-test and are presented as mean ± S.E.M. Since the drugs were added in cumulative manner, the difference between the control and the experimental values at each concentration were tested for a *P* value < 0.05. The potency of drugs defined as EC₅₀ and IC₅₀ was evaluated from log concentration-response curves (Probit analysis using Litchfield and Wilcoxon) [35] in the appropriate pharmacological preparations.

4.2.2. Details for motor coordination studies.

4.2.2.1 Rota-rod test. The apparatus consisted of a base platform and a rotating rod of 3 cm diameter with a non-slippery surface. The rod was placed at a height of 15 cm from the base. The rod, 30 cm in length, was divided into five equal sections by six disks. Thus, up to five mice were tested simultaneously on the apparatus, with a rod-rotating speed of 16 rpm. The integrity of motor coordination was assessed on the basis of the number of falls from the rod in 30 s according to Vaught et al. [36]

4.2.3. Details for cytotoxicity studies.

4.2.3.1. Cell cultures

HepG2 (Sigma-Aldrich, St. Louis, MO) cells were cultured in DMEM-Dulbecco's modified Eagle's medium (Sigma-Aldrich, St. Louis, MO) supplemented with 10% (v/v) inactivated fetal bovine serum (PAA Laboratories GmbH, Austria), 2 mM L-glutamine (Sigma-Aldrich, St. Louis, MO), 100 µg/mL penicillin and 100 µg/mL streptomycin (Sigma-Aldrich, St. Louis, MO) at 37°C in 5% CO₂. For experiments, the cells were grown to 70% confluence and then were treated with test compounds in serum free media for the indicated times.

4.2.3.2. Cell viability assay.

HepG2 cell viability was assessed using a conventional MTT reduction assay [37]. This method is based on the ability of viable cells to metabolize MTT, a water-soluble salt (yellow color), by cellular oxidoreductase into a water-insoluble blue formazan product. Therefore, the amount of formazan produced is proportional to the viable cells. Briefly, viable cells (10⁴/well) were plated into sterile 96-Well Cell Culture Cluster (Corning, NY 14831) and incubated with different concentrations (0.05–2 mM) of tested compounds for 2 h at 37 °C in 5% CO₂. At the end of incubation, the culture medium was replaced by a solution of MTT 0.5 mg/mL (Sigma-Aldrich, Milan, Italy) in PBS (Sigma-Aldrich, Milan, Italy). After 2 h incubation at 37 °C in 5% CO₂ this

solution was removed and 200 μ l of DMSO was added to each well to dissolve the formazan product. Absorbance values at 570 nm were measured using a plate reader Victor V³ (Perkin-Elmer) and DMSO medium was used as blank solution. Results are expressed as the percentage of MTT reduction respect to control cells. All experiments were carried out in sextuplicate and were repeated twice.

4.2.4. Details for P-gp interaction studies.

4.2.4.1. Cell cultures

MDCK-MDR1 cells are a gift of Prof. P. Borst, NKI-AVL Institute, Amsterdam, Nederland. MDCK cells were grown in DMEM high glucose supplemented with 10% fetal bovine serum, 2 mM glutamine, 100 U/mL penicillin, 100 μ g/mL streptomycin, in a humidified incubator at 37 °C with a 5 % CO₂ atmosphere.

4.2.4.2. Calcein-AM experiment

These experiments were carried out as described by Feng et al. with minor modifications [38]. Each cell line (50,000 cells per well) was seeded into black CulturePlate 96/wells plate with 100 μ l medium and allowed to become confluent overnight. 100 μ l of test compounds were solubilized in culture medium and added to monolayers. 96/Wells plate was incubated at 37 °C for 30 min. Calcein-AM was added in 100 μ l of Phosphate Buffered Saline (PBS) to yield a final concentration of 2.5 μ M and plate was incubated for 30 min. Each well was washed 3 times with ice cold PBS. Saline buffer was added to each well and the plate was read to Victor3 (PerkinElmer) at excitation and emission wavelengths of 485 nm and 535 nm, respectively. In these experimental conditions Calcein cell accumulation in the absence and in the presence of tested compounds was evaluated and fluorescence basal level was estimated by untreated cells. In treated wells the increase of

fluorescence with respect to basal level was measured. EC₅₀ values were determined by fitting the fluorescence increase percentage versus log [dose].

Acknowledgment. This work was accomplished thanks to the financial support of the Italian Ministero dell'Istruzione, dell'Università e della Ricerca.

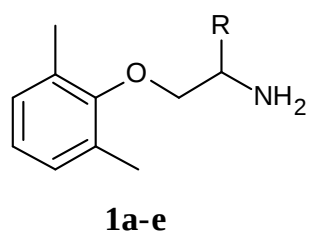
Supporting Information Available: Elemental analyses. Procedures for the synthesis of intermediates.

References

- [1] Nattel, S.; Duker, G.; Carlsson, L. Model systems for the discovery and development of antiarrhythmic drug. *Prog. Biophys. Mol. Biol.* **2008**, *98*, 328–339.
- [2] Lopshire, J. C.; Zipe, D. P. Sudden cardiac death: better understanding of risks, mechanisms, and treatment. *Circulation* **2006**, *114*, 1134–1136.
- [3] Muraglia, M.; De Bellis, M.; Catalano, A.; Carocci, A.; Franchini, C.; Carrieri, A.; Fortugno, C.; Bertucci, C.; Desaphy, J.F.; De Luca, A.; Conte Camerino, D.; Corbo F. *N*-aryl-2,6-dimethylbenzamides, a new generation of tocainide analogues as blockers of skeletal muscle voltage-gated sodium channels. *J. Med. Chem.* **2014**, *57*, 2589–2600.
- [4] Carocci, A.; Catalano, A.; Bruno, C.; Lentini, G.; Franchini, C.; De Bellis, M.; De Luca, A.; Conte Camerino, D. Synthesis and in vitro sodium channel blocking activity evaluation of novel homochiral mexiletine analogs. *Chirality* **2010**, *22*, 299–307.
- [5] Conte Camerino, D.; Tricarico, D.; Desaphy, J.-F. Ion channel pharmacology. *Neurotherapeutics* **2007**, *4*, 184–198.
- [6] Catalano, A.; Carocci, A.; Sinicropi, M. S. Mexiletine metabolites: a review. *Curr. Med. Chem.* **2015**, *22*, 1400–1413.
- [7] Catalano, A.; Carocci, A.; Fracchiolla, G.; Franchini, C.; Lentini, G.; Tortorella, V.; De Luca, A.; De Bellis, M.; Desaphy, J.-F.; Conte Camerino, D. Stereospecific synthesis of “*para*-hydroxymexiletine” and sodium channel blocking activity evaluation. *Chirality*, **2004**, *16*, 72–78.
- [8] Snedecor, S. J.; Sudharshan, L.; Cappelleri, J. C.; Sadosky, A.; Mehta, S.; Botteman, M. Systematic review and meta-analysis of pharmacological therapies for painful diabetic peripheral neuropathy. *Pain Practice* **2014**, *14*, 167–184.
- [9] Iose, S.; Misawa, S.; Sakurai, K.; Kanai, K.; Shibuya, K.; Sekiguchi, Y.; Nasu, S.; Noto, Y.; Fujimaki, Y.; Yokote, K.; Kuwabara, S. Mexiletine suppresses nodal persistent sodium currents in sensory axons of patients with neuropathic pain. *Clin. Neurophysiol.* **2010**, *121*, 719–724.
- [10] Desaphy, J.-F.; Carbonara, R.; Costanza, T.; Conte Camerino, D. Preclinical evaluation of marketed sodium channel blockers in a rat model of myotonia discloses promising antimyotonic drugs. *Exp. Neurol.* **2015**, *255*, 96–102.
- [11] Gao, Y.; Xue, X.; Hu, D.; Liu, W.; Yuan, Y.; Sun, H.; Li, L.; Timothy, K. W.; Zhang, L.; Li, C.; Yan, G.-X. Inhibition of late sodium current by mexiletine: a novel

- pharmotherapeutical approach in Timothy syndrome. *Circulation: Arrhythm. Electrophysiol.* **2013**, 6, 614–622.
- [12] Weiss, M.; Simmons, Z.; Atassi, N.; Graves, M.; Parziale, N.; Salameh J.; Quinn, C.; Brown, R.; Distad, B.; Trivedi, J.; Shefner, J.; Duleep, A.; Barohn, R.; Dimachkie, M.; McVey, A.; Pestronk, A.; Swenson, A.; Macklin, E.; Knox, A.; Gilardi, K.; Cudkowicz, M. A phase 2 study of mexiletine in sporadic amyotrophic lateral sclerosis (S50.004). *Neurology* **2005**, 84 (14 suppl S50.004).
- [13] Buccia, M.; Ramirez, A.; Parente, V.; Simone, C.; Nizzardo, M.; Magri, F.; Dametti, S.; Corti, S. Therapeutic development in amyotrophic lateral sclerosis. *Clin. Therapeut.* **2015**, 37, 668–680.
- [14] Shibuya, K.; Misawa, S.; Kimura, H.; Noto, Y. I. et al. A single blind randomized controlled clinical trial of mexiletine in amyotrophic lateral sclerosis: efficacy and safety of sodium channel blocker phase II trial. *Amyotroph. Lateral Scler. Frontotemporal. Degener.* **2015**, 16, 353–358.
- [15] Catalano, A.; Desaphy, J.-F.; Lentini, G.; Carocci, A.; Di Mola, A.; Bruno, C.; Carbonara, R.; De Palma, A.; Budriesi, R.; Ghelardini, C.; Perrone, M.G.; Colabufo, N.A.; Conte Camerino, D.; Franchini, C. Synthesis and toxicopharmacological evaluation of *m*-hydroxymexiletine, the first metabolite of mexiletine more potent than the parent compound on voltage-gated sodium channels. *J. Med. Chem.* **2012**, 55, 1418–1422.
- [16] Matthews, E.; Hanna, M.G. Repurposing of sodium channel antagonists as potential new anti-myotonic drugs. *Exp. Neurol.* **2014**, 261, 812–815.
- [17] Franchini, C.; Carocci, A.; Catalano, A.; Cavalluzzi, M.M.; Corbo, F.; Lentini, G.; Scilimati, A.; Tortorella, P.; Conte Camerino, D.; De Luca, A. Optically active mexiletine analogues as stereoselective blockers of voltage-gated Na⁺ channels. *J. Med. Chem.* **2003**, 46, 5238–5248.
- [18] Desaphy, J.-F.; Dipalma, A.; Costanza, T.; Carbonara, R.; Di Nardo, M. M.; Catalano, A.; Carocci, A.; Lentini, G.; Franchini, C.; Conte Camerino, D. Molecular insights into the local anesthetic receptor within voltage-gated sodium channels using hydroxylated analogs of mexiletine. *Front. Pharmacol.* **2012**, 3, 1–12.
- [19] De Luca, A.; Talon, S.; De Bellis, M.; Desaphy, J.-F.; Franchini, C.; Lentini, G.; Catalano, A.; Corbo, F.; Tortorella, V.; Conte Camerino, D. Inhibition of skeletal muscle sodium currents by mexiletine analogues: specific hydrophobic interactions rather than lipophilia per se account for drug therapeutic profile, *Naunyn Schmiedebergs Arch. Pharmacol.* **2003**, 367, 318–327.
- [20] De Bellis, M.; De Luca, A.; Desaphy, J.-F.; Carbonara, R.; Heiny, J.A.; Kennedy, A.; Carocci, A.; Cavalluzzi, M.M.; Lentini, G.; Franchini, C.; Conte Camerino, D. Combined Modifications of Mexiletine Pharmacophores for New Lead Blockers of Nav1.4 Channels. *Biophys. J.* **2013**, 104, 344–354.
- [21] Carocci, A.; Catalano, A.; Corbo, F.; Duranti, A.; Amoroso, R.; Franchini, C.; Lentini, G.; Tortorella, V. Stereospecific synthesis of mexiletine and related compounds: Mitsunobu versus Williamson reaction. *Tetrahedron: Asymmetry* **2000**, 11, 3619–3634.
- [22] Cavalluzzi, M. M.; Viale, M.; Bruno, C.; Carocci, A.; Catalano, A.; Carrieri, A.; Franchini, C.; Lentini, G. A convenient synthesis of lubeluzole and its enantiomer: Evaluation as chemosensitizing agents on human ovarian adenocarcinoma and lung carcinoma cells. *Bioorg. Med. Chem. Lett.* **2013**, 23, 4820–4823.
- [23] Carocci, A.; Lentini, G.; Catalano, A.; Cavalluzzi, M. M.; Bruno, C.; Muraglia, M.; Colabufo, N. A.; Galeotti, N.; Corbo, F.; Matucci, R.; Ghelardini, C.; Franchini, C. Chiral aryloxyalkylamines: selective 5-HT_{1B/D} activation and analgesic activity. *ChemMedChem* **2010**, 5, 696–704.
- [24] Catalano, A.; Carocci, A.; Cavalluzzi, M. M.; Di Mola, A.; Lentini, G.; Lovece, A.; Dipalma, A.; Costanza, T.; Desaphy, J.-F.; Conte Camerino, D.; Franchini, C. Hydroxylated

- analogs of mexiletine as tools for structural-requirements investigation of the sodium channel blocking activity. *Arch. Pharm. Chem. Life. Sci.* **2010**, *343*, 325–332.
- [25] Bruno, C.; Carocci, A.; Catalano, A.; Cavalluzzi, M. M.; Corbo, F.; Franchini, C.; Lentini, G.; Tortorella, V. Facile, alternative route to lubeluzole, its enantiomer, and the racemate. *Chirality*, **2006**, *18*, 227–231.
- [26] Catalano, A.; Budriesi, R.; Bruno, C.; Di Mola, A.; Defrenza, I.; Cavalluzzi, M. M.; Micucci, M.; Carocci, A.; Franchini, C.; Lentini, G. Searching for new antiarrhythmic agents: evaluation of *meta*-hydroxymexiletine enantiomers. *Eur. J. Med. Chem.* **2013**, *65*, 511–516.
- [27] De Bellis, M.; De Luca, A.; Rana, F.; Cavalluzzi, M.M.; Catalano, A.; Lentini, G.; Franchini, C.; Tortorella, V.; Conte Camerino, D. Evaluation of the pharmacological activity of the major mexiletine metabolites on skeletal muscle sodium currents. *Br. J. Pharmacol.* **2006**, *149*, 300–310.
- [28] Carocci, A.; Catalano, A.; Bruno, C.; Lovece, A.; Roselli, M.; Cavalluzzi, M. M.; De Santis, F.; De Palma, A.; Rusciano, M. R.; Illario, M.; Franchini, C.; Lentini, G. *N*-(Phenoxyalkyl)amides as MT₁ and MT₂ ligands: Antioxidant properties and inhibition of Ca²⁺/CaM-dependent kinase II. *Bioorg. Med. Chem.* **2013**, *21*, 847–851.
- [29] Carocci, A.; Catalano, A.; Turi, F.; Lovece, A.; Cavalluzzi, M. M.; Bruno, C.; Colabufo, N. A.; Contino, M.; Perrone, M. G.; Franchini, C.; Lentini, G. Stereoselective modulation of P-glycoprotein by chiral small molecules. *ChemMedChem* **2016**, *11*, 93–101.
- [30] Still, W. C.; Kahn, M.; Mitra, A. Rapid chromatographic technique for preparative separation with moderate resolution. *J. Org. Chem.* **1978**, *43*, 2923–2925.
- [31] Gualdani, R.; Tadini-Buoninsegna, F.; Roselli, M. G.; Defrenza, I.; Contino, M.; Colabufo, N. A.; Lentini, G. Inhibition of hERG potassium channel by the antiarrhythmic agent mexiletine and its metabolite *m*-hydroxymexiletine. *Pharmacol. Res. Perspect.* **2015**, *3*, e00160. Doi:10.1002/prp2.160
- [32] Borchard, U.; Böskén, R.; Greeff, K. Characterization of Antiarrhythmic Drugs by Alternating Current Induced Arrhythmias in Isolated Heart Tissues. *Arch. Int. Pharmacodyn.* **1982**, *256*, 253–268.
- [33] Borchard, U.; Clemm, S.; Hirth, C. Influence of Antiarrhythmogenic and Antiarrhythmic Agents on Alternating Current-Induced Arrhythmias in Guinea-pig Heart Preparations. *Arch. Int. Pharmacodyn.* **1984**, *269*, 83–93.
- [34] Borchard, U. Untersuchungen zum Wirkungsmechanismus und zur pharmakologischen Charakterisierung von Lokalanästhetika. Habilitationsschrift, Düsseldorf. 1978.
- [35] Tallarida, R. J.; Murray, R. B. *Manual of Pharmacologic Calculations with Computer Programs*, 2nd ed.; Springer-Verlag: New York, **1987**.
- [36] Vaught, J.; Pelley, K.; Costa, L. G.; Setter, P.; Enna, S. J. A comparison of the antinociceptive responses to GABA – receptor agonists THIP and baclofen. *Neuropharmacology* **1985**, *24*, 211–216.
- [37] Berridge, M. V.; Tan, A. S. Characterization of the cellular reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT): subcellular localization, substrate dependence, and involvement of mitochondrial electron transport in MTT reduction. *Arch. Biochem. Biophys.* **1993**, *303*, 474–482.
- [38] B. J. Feng, B. Mills, R. E. Davidson, R. J. Mireles, J. S. Janiszewski, M. D. Troutman, S. M. de Moraes, *Drug Metab. Dispos.* **2008**, *36*, 268–275.



a: R = Me

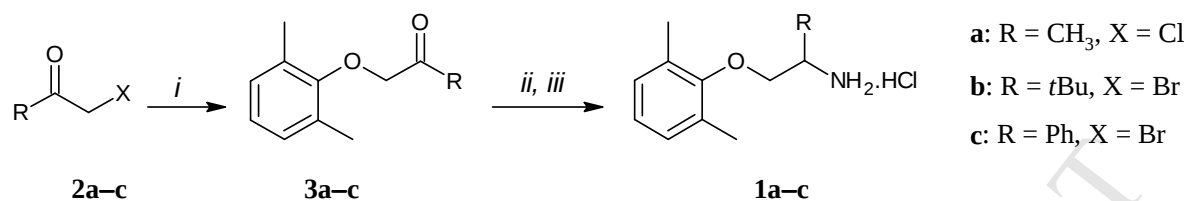
b: R = *t*Bu

c: R = Ph

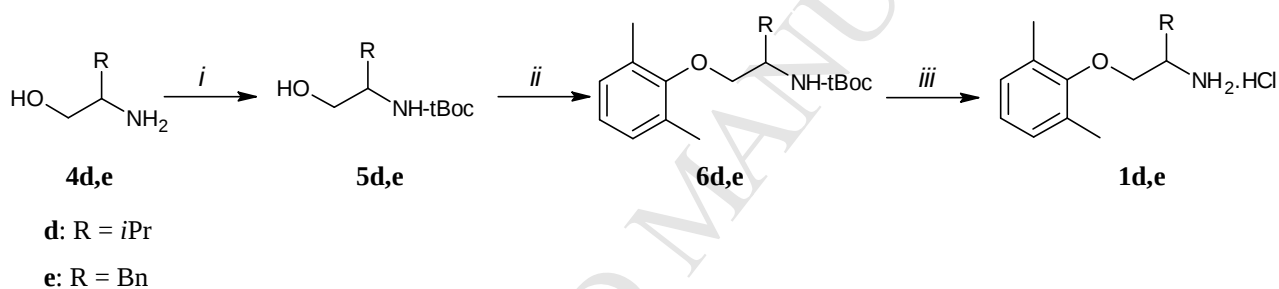
d: R = *i*Pr

e: R = Bn

Figure 1. Structures of mexiletine (**1a**) and its analogues (**1b–e**).



Scheme 1. Reagents and conditions: (i) 2,6-xyleneol, K₂CO₃, anhyd DMF, rt, 24h (for **2a**); 2,6-xyleneol, K₂CO₃, MeOH, MW, 100 °C, 20 min (for **2b**); 2,6-xyleneol, NaOH, DMSO, MW, 60 °C, 15 min (for **2c**); (ii) NaBH₃CN, AcONH₄, MeOH, rt (or MW, 110 °C, 11 min); (iii) anhyd Et₂O, gaseous HCl.



Scheme 2. Reagents and conditions: (i) Boc₂O, 1M NaOH, THF, rt; (ii) 2,6-xyleneol, DIAD, PPh₃, anhyd THF, rt; (iii) anhyd Et₂O, gaseous HCl.

Table 1. Antiarrhythmic Activity of Compounds **1a–e**.

Compd	Max % increase ^a (mean ± SEM)	EC ₅₀ ^b (μM)	95% confidence limit (x 10 ⁻⁶)
1a	64 ± 1.4	11.6	8.71–13.5
1b	71 ± 2.4 ^c	2.11	1.38–3.20
1c	135 ± 2.2 ^d	0.43	0.34–0.54
1d	145 ± 4	18.4	14.9–22.7
1e	169 ± 4 ^c	4.36	1.47–6.38

^aIncrease of threshold of ac-arrhythmia; increase in the intensity of 50 Hz alternating current required to produce arrhythmia in guinea pig left atria driven at 1 Hz in the presence of each tested compounds at 10⁻⁴ M. For all data $P < 0.05$. ^bCalculated from log concentration-response curves (Probit analysis according to Litchfield and Wilcoxon [35] with $n = 6–8$). When the maximum effect was $< 50\%$, the EC₅₀ values were not calculated. ^cAt 10⁻⁵ M. ^dAt 5x10⁻⁶ M.

Table 2. Influences of Tested Compounds on Cardiovascular Parameters.

Left atrium				Right atrium					
negative inotropy				negative inotropy			negative chronotropy		
Compd	Activity ^a (% ± SEM)	EC ₅₀ ^b (μM)	95% confidence lim (x10 ⁻⁶)	Activity ^c (% ± SEM)	EC ₅₀ ^b (μM)	95% confidence lim (x10 ⁻⁶)	Activity ^d (% ± SEM)	EC ₅₀ ^b (μM)	95% confidence lim (x10 ⁻⁶)
1a	90 ± 1 [*]	0.045	0.035–0.058				85 ± 3 ^g	0.014	0.009–0.023
1b	92 ± 1 [*]	0.042	0.027–0.060	77 ± 1	1.02	0.85–3.05	7.0 ± 0.3 ⁱ		
1c	94 ± 2 ^{*e}	0.029	0.017–0.052	54 ± 2 ^h	8.2	4.6–10.4	11 ± 1 ⁱ		
1d	84 ± 2 ^{*f}	0.021	0.012–0.037				55 ± 2 ^{*f}	13.1	10.1–15.0
1e	79 ± 2 ^{*g}	0.012	0.010–0.015	57 ± 3	0.066	0.048–0.091	24 ± 2 [*]		

^aDecrease in developed tension on isolated guinea-pig left atrium at 10⁻⁵ M, expressed as percentage change from the control ($n = 5–6$). The left atria were driven at 1 Hz. The 10⁻⁵ M concentration gave the maximum effect for most compounds. ^bCalculated from log concentration-response curves (Probit analysis by Litchfield and Wilcoxon [35] with $n = 6–7$). When the maximum effect was $< 50\%$, the EC₅₀ inotropic, EC₅₀ chronotropic and IC₅₀ vasorelaxant values were not calculated. ^cDecrease in developed tension on guinea-pig spontaneously beating isolated right atrium at 5x10⁻⁶ M, expressed as percent changes from the control ($n = 7–8$). The 5x10⁻⁶ M concentration gave the maximum effect for most compounds. ^dDecrease in atrial rate on guinea-pig spontaneously beating isolated right atria at 10⁻⁵ M, expressed as the percentage change from the control ($n = 7–8$). The 10⁻⁵ M concentration gave the maximum effect for most compounds. The pretreatment heart rate ranged from 170 to 195 beats/min. ^eAt 5x10⁻⁶ M. ^fAt 10⁻⁴ M. ^gAt 10⁻⁷ M. ^hAt 10⁻⁵ M. ⁱ $P > 0.05$. An asterisk indicates $P < 0.05$.

Table 3. Relaxant Activity of Compounds on K⁺-Depolarized Guinea Pig Vascular and non-vascular Smooth Muscle.

compd	Aorta		Ileum	
	Activity ^a	Activity ^a	IC ₅₀ ^b	95% confidence lim
	(% ± S.E.M.)	(% ± S.E.M.)	(μM)	(x10 ⁻⁶)
1a	5.0 ± 0.3	81 ± 2	8.5	6.5–11.1
1b	38 ± 2	73 ± 2 ^c	10.7	9.0–12.8
1c	31 ± 2	75 ± 3 ^d	13.8	10.8–17.5
1d	11 ± 1	88 ± 1	19.6	12.0–25.0
1e	33 ± 1	73 ± 2 ^d	3.6	2.8–4.6

^aPercent inhibition of calcium-induced contraction on K⁺-depolarized (80 mM) guinea pig aortic strips and longitudinal smooth muscle at 10⁻⁴ M. The 10⁻⁴ M concentration gave the maximum effect for most compounds. ^bCalculated from log concentration-response curves (Probit analysis by Litchfield and Wilcoxon [35] with *n* = 6–7). When the maximum effect was < 50%, the IC₅₀ values were not calculated. ^cAt 5x10⁻⁵ M. ^dAt 10⁻⁵ M.

Table 4. Effect of mexiletine and its analogues on motor coordination expressed as number of falls from Rota-rod.^a

Treatment	Dose, mg kg ⁻¹ ip	Falls from the rota-rod			
		Before treatment	After treatment		
			30 min	45 min	60 min
Saline		4.4 ± 1.1	2.5 ± 0.7	1.2 ± 0.5	0.9 ± 0.3
1a ^b	25	2.8 ± 0.5	1.3 ± 0.4	0.8 ± 0.3	-
1a ^b	50	2.4 ± 0.3	3.8 ± 0.3	3.5 ± 0.2	-
1b	25	4.6 ± 0.5	2.7 ± 0.4	1.7 ± 0.4	0.8 ± 0.2
1b	50	4.7 ± 0.4	3.1 ± 0.3	1.8 ± 0.5	1.2 ± 0.6
1c	25	4.3 ± 0.5	2.4 ± 0.3	1.8 ± 0.4	1.3 ± 0.3
1c	50	4.5 ± 0.6	3.2 ± 1.6	2.8 ± 0.7	1.0 ± 0.2
1d	25	4.8 ± 0.7	2.6 ± 0.8	2.2 ± 0.6	1.1 ± 0.3
1d	50	4.5 ± 0.6	3.0 ± 1.4	2.1 ± 1.3	1.0 ± 1.2
1e	25	4.5 ± 0.9	2.9 ± 0.8	1.9 ± 0.5	1.2 ± 0.5
1e	50	4.2 ± 0.8	4.4 ± 1.1 [*]	3.5 ± 0.9 [*]	1.3 ± 1.1

^aEach value represents the mean of 8 mice. ^bRef.[16] ^{*}P < 0.05 in comparison with saline-treated mice.

Table 5. P-gp interaction of mexiletine (**1a**) and its analogues (**1b–e**).

Compd	EC ₅₀ ± SEM, ^a μM
	P-gp
1a	NA ^{b,c}
1b	NA ^b
1c	27 ± 5
1d	NA ^b
1e	8.4 ± 1.5

^aThe value is the mean of three different experiments, samples in duplicate; ^bNA = No activity at 100μM; ^cRef. [31].

- Search for new antiarrhythmic agents.
- Antiarrhythmic effects evaluation of mexiletine analogues.
- Identification of potent and selective antiarrhythmic agents.