Design and Synthesis of Imidazo[4,5-*c*]pyridine Derivatives as Promising Aurora Kinase A (AURKA) Inhibitors

D. A. Lomov,* S. N. Lyashchuk, and M. G. Abramyants

Litvinenko Institute of Physical Organic Chemistry, ul. Rozy Lyuksemburg 70, Donetsk, 83000 Ukraine *e-mail: lomov_dmitrii@mail.ru

Received June 28, 2016

Abstract—Computer simulation at the PM7 level of theory of the structures of imidazo[4,5-c]pyridine derivatives (deaza analogs of purines) and their complexes with Aurora kinase A (AURKA) indicated prospects for their use as potential AURKA inhibitors in the treatment of oncological diseases. A number of new compounds of the selected imidazo[4,5-c]pyridine series, for which the highest inhibitory activity against AURKA was predicted, were synthesized in high yields for further biological testing.

DOI: 10.1134/S1070428016120198

About 6 million new cancer occurrences is registered worldwide annually, and this situation remains unchanged over the past two decades. Although cancer and other malignant oncological diseases constitute 5– 10% of all cases of somatic diseases, they follow next to cardiovascular disorders with respect to mortality.

In 1994–1997, three new kinases overexpressed in cancer have been identified in dividing cells of warmblooded by PCR (polymerase chain reaction) [1]. Aurora kinases belong to a small family of serine/ threonine protein kinases, which includes three enzymes: Aurora kinase A (AURKA), Aurora kinase B (AURKB), and Aurora kinase C (AURKC) [2-4]. These enzymes are involved in the most important molecular events in all stages of cell mitosis [5]. Considerable increase of the concentration of AURKA and AURKB induces nondisjunction, so that some cells degrade and die, while the others degenerate to tumor cells with a high probability [6]. It was shown experimentally that increased concentrations of Aurora kinases largely determine the development of many types of stomach, liver, and rectal cancers, some breast and pancreatic cancers, as well as of leukemia [2]. Thus, detection of increased AURKA and AURKB contents is an important diagnostic test for carcinogenic process or precancer state [7].

The new drug hesperadin (1) has recently been introduced into clinical practice; it is capable of suppressing excess activity of AURKA and AURKB in tumor tissues [8]. Study of the mechanism of hesperadin-induced inhibition of the catalytic activity of these enzymes has shown [9] that the drug blocks active site of the enzyme. At present, some leading pharmaceutical companies perform extensive search for new Aurora kinase inhibitors with the goal of creating new-generation anticancer drugs [10]. Currently known Aurora kinase inhibitors are represented by various heterocyclic structures; some compounds, e.g., **2–6**, are now under clinical trials [4, 11].

In recent years, while searching for new kinase inhibitors, researchers' attention has been focused on imidazo[4,5-*b*]pyridine derivatives [12, 13]. Compounds **5** and **6** showed a pronounced inhibitory effect against AURKA, and the magnitude of this effect may change by 1–2 orders upon insignificant structural modification of the inhibitor molecule [14, 15]. These data indicate high selectivity of binding to the enzyme active site. Analogous studies of imidazo[4,5-*c*]pyridine derivatives as Aurora kinase inhibitors have not been reported in the literature. Taking into account that imidazo[4,5-*b*]pyridine and imidazo[4,5-*c*]pyridine are the closest structural analogs, the latter are expected to exhibit similar biological properties, in particular with respect to various kinases.

The inhibitory activity of a series of imidazo-[4,5-c]pyridine derivatives against AURKA was quantitatively assessed by computational methods (*in silico*), followed by directed synthesis of compounds with the highest predicted activity.



In the first step, as subjects for the study we selected a sample consisting of 24 imidazo[4,5-c]pyridines 7–30 as potential ligands for AURKA inhibition. The choice was based on the drug likeness approach implying that structurally related compounds may possess similar biological properties. In our case, the closest analogs of compounds 7–30 are imidazo-[4,5-b]pyridine derivatives [12–15].

An empirical approach based on Lipinski's rules [16] has found wide application in the design of new drugs since it allows considerable contraction of the series of compounds to be studied. According to Lipinski's rules, a compound could be a drug if its molecule contains no more than 5 donor centers capable of forming hydrogen bonds (OH and NH groups), no more than 10 hydrogen bond acceptor centers (N, O), and no more than 15 nonterminal freely rotating bonds; furthermore, its molecular weight should be lower than 500, and the octanol–water partition coefficient log*P* should be lower than 5. All compounds 7–30 conform to Lipinski's rules, and there are some "reserves" regarding the molecular composition and properties, which provide the possibility of



obtaining more complex analogs through variation and modification of substituents therein.

An alternative approach to prediction of potential effect of compounds on cellular processes is provided by Molinspiration on-line service (version 2004.1) [17]. This software is based on a huge, continuously updated database, and it makes it possible not only to determine whether a compound fits Lipinski's rules but also to predict *in silico* with a high probability some



7, 10, 13, 16, 19, 22, 25, 28, X = NH; 8, 11, 14, 17, 20, 23, 26, 29, X = S; 9, 12, 15, 18, 21, 24, 27, 30, X = O; 7–9, 13–15, 19–21, 25–27, R^1 = H; 10–12, 16–18, 22–24, 28–30, R^1 = Me; 7–12, R^2 = NMe₂; 13–18, R^2 = MeO; 19–24, R^2 = Cl; 25–30, R^2 = Br.

RUSSIAN JOURNAL OF ORGANIC CHEMISTRY Vol. 52 No. 12 2016

biological properties of imidazo[4,5-c]pyridine derivatives. The results of the Molinspiration predictions for compounds 7–30 are given in Table 1.

All compounds 7–30 could both activate and inhibit proteases (IP -0.15 to 0.20), modulate and demodulate ion channels (MIC -0.21 to 0.20), and inhibit cellular enzymes in total (IE 0.02–0.40); however, the probability of these properties is not high ($\leq 40\%$). They are also capable of acting as medium-strength nuclear receptor activators (NRL -0.17 to -0.51) and inhibit transmembrane receptors (GPCR varies over a wide range from 0.06 to 0.58). The most probable property is the ability to inhibit kinases (IK 0.23-0.56).

By semiempirical quantum chemical calculations at the PM7 level of theory [18] using MOPAC2012 version 15.281W [19] we obtained more detailed information on the structure and electronic properties of molecules 7-30 and their complexes with AURKA. The calculated parameters given in Table 2 may be used as descriptors in the assessment of the inhibitory activity of the ligands against enzymes.

The dipole moments of 7-30 vary over a wide range (4.66–12 D) and strongly depend on the substituents in the imidazopyridine fragment. The negative charge is localized on the nitrogen atom of that fragment, and variation of the substituents exerts an appreciable effect only on the N¹ atom: $q(N^1) - 0.432$ to -0.295, $q(N^2) -0.437$ to -0.366, $q(N^3) -0.550$ to -0.509 a.u. The positive charge is localized on the carbon atoms. The absolute electronegativities EN 4.05-4.81 and the absolute hardnesses n 7.26-7.81 calculated from the HOMO and LUMO energies [20] are typical of nitrogen-containing heterocyclic systems capable of being involved in donor-acceptor interac-

G 1	117		CDCD	NDI	ID	Ш
Compound no.	IK	MIC	GPCK	NKL	IP	IE
7	0.53	0.06	0.42	-0.25	0.04	0.22
8	0.44	-0.07	0.17	-0.39	-0.06	0.12
9	0.56	0.16	0.54	-0.17	0.14	0.39
10	0.39	-0.04	0.39	-0.26	-0.03	0.17
11	0.27	-0.17	0.13	-0.39	-0.13	0.06
12	0.39	0.05	0.49	-0.18	0.07	0.32
13	0.49	0.02	0.41	-0.28	0.02	0.20
14	0.40	-0.10	0.15	-0.42	-0.09	0.10
15	0.52	0.13	0.53	-0.19	0.12	0.38
16	0.35	-0.07	0.37	-0.29	-0.05	0.15
17	0.23	-0.21	0.10	-0.42	-0.15	0.04
18	0.35	0.02	0.47	-0.20	0.05	0.31
19	0.52	0.08	0.46	-0.29	0.04	0.22
20	0.43	-0.05	0.19	-0.44	-0.08	0.12
21	0.56	0.20	0.58	-0.20	0.14	0.40
22	0.37	-0.02	0.41	-0.30	-0.04	0.16
23	0.25	-0.16	0.14	-0.44	-0.15	0.05
24	0.37	0.08	0.52	-0.21	0.07	0.33
25	0.50	0.03	0.38	-0.37	-0.02	0.19
26	0.41	-0.10	0.11	-0.51	-0.13	0.09
27	0.54	0.14	0.51	-0.28	0.09	0.37
28	0.35	-0.08	0.34	-0.37	-0.10	0.13
29	0.23	-0.21	0.06	-0.51	0.20	0.02
30	0.35	0.02	0.45	-0.28	0.01	0.30

Table 1. Some biological activities^a of compounds 7–30 predicted by Molinspiration (version 2004.1)

IK stands for kinase inhibition; MIC stands for ion channel modulation; GPCR (G protein-coupled receptors) stands for ligand binding to seven-transmembrane receptors; NRL stands for ligand binding to nuclear receptors; IP stands for protease inhibition; IE stands for enzyme inhibition. Negative sign of the biological activity indices indicates activation of the respective bioprocess.

Comp.	$\Delta H^{\rm f}$,	μ, D	Effective charges on atoms, a.u.						Energy, eV	
no.	kJ/mol		N^1	C^2	N ³	C^4	N^5	C ⁶	НОМО	LUMO
7	339.77	6.71	-0.422	0.299	-0.406	0.408	-0.517	0.139	-7.91	-0.36
8	353.27	7.24	-0.423	0.313	-0.437	0.409	-0.518	0.143	-8.12	-0.37
9	262.32	10.82	-0.432	0.292	-0.401	0.402	-0.527	0.147	-8.05	-0.50
10	357.65	9.91	-0.316	0.267	-0.384	0.419	-0.546	0.149	-7.90	-0.21
11	348.10	8.08	-0.306	0.298	-0.439	0.410	-0.518	0.138	-8.05	-0.25
12	259.52	12.00	-0.316	0.271	-0.390	0.406	-0.528	0.143	-8.06	-0.25
13	165.86	6.97	-0.427	0.267	-0.376	0.413	-0.531	0.147	-8.22	-0.67
14	223.52	9.22	-0.429	0.272	-0.394	0.403	-0.519	0.151	-8.34	-0.83
15	76.35	9.15	-0.428	0.267	-0.385	0.408	-0.529	0.154	-8.31	-0.70
16	145.39	7.32	-0.310	0.253	-0.383	0.410	-0.524	0.141	-8.24	-0.57
17	219.72	9.84	-0.316	0.264	-0.400	0.398	-0.509	0.141	-8.29	-0.63
18	72.78	9.60	-0.312	0.252	-0.385	0.408	-0.529	0.148	-8.22	-0.52
19	302.08	4.58	-0.419	0.244	-0.369	0.413	-0.524	0.153	-8.44	-1.08
20	375.39	7.42	-0.420	0.246	-0.379	0.406	-0.522	0.157	-8.46	-1.16
21	228.38	7.45	-0.420	0.242	-0.372	0.412	-0.530	0.160	-8.42	-1.05
22	321.79	6.22	-0.305	0.221	-0.366	0.428	-0.550	0.161	-8.14	-0.88
23	310.34	5.89	-0.295	0.255	-0.422	0.414	-0.518	0.145	-8.29	-0.72
24	224.04	8.50	-0.304	0.229	-0.376	0.411	-0.530	0.153	-8.32	-0.92
25	356.91	4.66	-0.418	0.242	-0.369	0.413	-0.521	0.153	-8.43	-1.09
26	430.14	7.42	-0.419	0.244	-0.378	0.407	-0.523	0.158	-8.46	-1.17
27	283.22	7.50	-0.420	0.241	-0.373	0.411	-0.529	0.159	-8.42	-1.06
28	352.25	5.54	-0.303	0.228	-0.372	0.413	-0.524	0.146	-8.33	-0.93
29	367.43	5.70	-0.296	0.256	-0.422	0.414	-0.519	0.145	-8.29	-0.75
30	278.86	8.55	-0.304	0.227	-0.376	0.411	-0.530	0.152	-8.32	-0.92

Table 2. Heats of formation, dipole moments, effective charges on some atoms, and frontier orbital energies of compounds 7–30, calculated by the PM7 method (MOPAC2012)

tions. According to the X-ray diffraction data for AURKA complexes with different inhibitors, Aurora kinase is an enzyme containing 402 amino acid residues and N- and C-terminal domains.

The AURKA complexes with potential inhibitors 7–30 were calculated in several steps. In the first step, the ligand was removed from the active site of the known AURKA complex with $4-\{[2-(4-\{[(4-fluoro$ $phenyl)carbonyl]amino\}-1H-pyrazol-3-yl)-1H-benz$ $imidazol-6-yl]methyl}morpholin-4-ium cation [21],$ whose structure was determined by X-ray analysis(RCSB Protein Data Bank ID: 2W1C), using AccelrysDiscovery Studio 3.5 program. The structure of theprotein molecule thus obtained was taken as the initialapproximation and was optimized by PM7 semiempirical quantum chemical calculations (MOPAC2012;MOZYME module for the calculation of protein molecules and complex macromolecular assemblies was applied). The corresponding inhibitor with an IK index of larger than 0.35 (which rendered it sufficiently active) was then placed into the active site, and the structure of the complex was optimized. All enthalpies of formation (Table 2) were calculated for the optimized structures.

As an example, Fig. 1 shows the calculated structure of the AURKA complex with compound **11**. The inhibitor molecule binds to the enzyme active site and is held thereon, thus leading to inactivation of the kinase in the cell division process. The stability of the protein–inhibitor complex is determined by the energy of its formation (exothermic process). Furthermore, ligand docking induces change of the secondary protein structure in the regions adjacent to the active site, which should also favor enzyme deactivation.





N-Terminal domain

C-Terminal domain

Fig. 1. Optimized structure of the Aurora kinase A complex with inhibitor **11** bound to the active site.

 $E_{\rm doc}$, kcal/mol



Fig. 2. Correlation of the docking energy (E_{doc}) with the kinase inhibitory activity (IK) of compounds 7–10, 12–15, 19–22, and 24–27, calculated by Molinspiration program (version 2004.1).

The docking energy E_{doc} for individual compounds was calculated as the difference in the enthalpy of formation of the complex $\Delta H_{AURKA-L}^{f}$ and the sum of the enthalpies of formation of unbound AURKA (ΔH_{AURKA}^{f}) and the corresponding ligand ΔH_{L}^{f} (1):

$$E_{\rm doc} = \Delta H_{\rm AURKA-L}^{\rm f} - (\Delta H_{\rm AURKA}^{\rm f} + \Delta H_{\rm L}^{\rm f}).$$
(1)

The calculated protein–ligand docking energy is linearly related to the kinase inhibition index IK determined by Molinspiration 2004.1 (Fig. 2) through Eq. (2):

$$E_{\rm doc} = (1.31 \pm 1.36) - (29.57 \pm 2.92) \,\text{IK};$$

$$r^2 = 0.871, s_0 = 8.32. \tag{2}$$

Our results indicated that all compounds 7-30 are capable of binding to the AURKA active site to greater or lesser extent. This especially applies to those possessing no methyl group on the nitrogen atom of the imidazopyridine fragment. Elongation of the alkyl substituent strongly reduces E_{doc} in absolute value due to considerable steric hindrances to approach of the inhibitor molecule to the active site and holding it therein.

For further biological testing we synthesized six imidazo[4,5-*c*]pyridine derivatives 7, 9, 15, 19, 21, and 27 for which the highest inhibitory activity against AURKA was predicted. The key intermediate products for the synthesis of these compounds were 2-chloropyridine-3,4-diamine (31) [22], *N*-(1*H*-imidazol-2-yl)-2-(piperazin-1-yl)acetamide (32), and *N*-(1,3-oxazol-2yl)-2-(piperazin-1-yl)acetamide (33). Acetamides 32 and 33 were synthesized in 72–83% yield by heating *N*-imidazolyl- and *N*-oxazolylchloroacetamides 34 and 35 with piperazine hydrochloride (prepared *in situ*) in ethanol (Scheme 1). In the ¹H NMR spectra of 32 and 33, signals of the piperazine fragment appeared as two singlets as δ 2.70–2.83 and 3.03–3.12 ppm.

2-Chloropyridine-3,4-diamine (**31**) reacted with *para*-substituted benzoic acids **36–39** in phosphoryl chloride at 130–140°C to give 4-chloro-2-phenyl-3*H*-imidazo[4,5-*c*]pyridines **40–43** in 50–64% yield (Scheme 2). Compounds **40–43** displayed in the ¹H NMR spectra signals from protons of the pyridine ring as two doublets at δ 7.57–7.62 and 7.93–8.25 ppm, and protons of the *para*-substituted benzene ring resonated at δ 6.83–7.81 and 8.04–8.24 ppm.

By reacting 4-chloro-2-phenylimidazo[4,5-*c*]pyridine derivatives 40–43 with acetamides 32 and 33 on heating in ethanol in the presence of triethylamine we obtained compounds 7, 9, 15, 19, 21, and 27 in 52–78% yield (Scheme 3). Analogous compounds were synthesized in [23] under similar conditions. In the ¹H NMR spectra of 7, 9, 15, 19, 21, and 27 we observed doublets at δ 7.58–8.28 and 7.59–7.94 ppm due to protons in the pyridine ring, doublets at δ 6.81–



7, 19, 32, X = NH; 9, 15, 21, 27, 33, X = O; 7, 9, 40, $R = Me_2N$; 15, 41, R = MeO; 19, 21, 42, R = CI; 27, 43, R = Br.

7.81 and 8.06–8.26 ppm due to protons of the benzene ring, and signals of protons in the piperidine and azole rings.

Thus, computer simulation of deaza analogs of purine bases, imidazo[4,5-c]pyridine derivatives, and their complexes with AURKA indicates prospects of their use as potential inhibitors of this key enzyme for oncological practice.

EXPERIMENTAL

The ¹H NMR spectra were recorded on a Bruker Avance II 400 spectrometer at 400 MHz using DMSO- d_6 as solvent and tetramethylsilane as internal standard. The purity of the isolated compounds was checked by TLC on Silufol UV-254 plates using methanol-chloroform (1:10) as eluent; spots were visualized by treatment with iodine vapor or under UV light.

Compounds 32 and 33 (general procedure). Piperazine hexahydrate, 10 mmol, was dissolved in 10 mL of propan-1-ol, 10 mmol of piperazine dihydrochloride monohydrate was added, and the mixture was heated with stirring until it became homogeneous. A solution of chloroacetamide **34** or **35** in 10 mL of propan-1-ol was added to the resulting solution of piperazine hydrochloride, and the mixture was heated for 2 h at 80–90°C and cooled. The precipitate of piperazine dihydrochloride was filtered off and washed on a filter with 5 mL of propan-1-ol. The filtrate was evaporated to 1/3 of the initial volume and filtered while hot. Compounds **32** and **33** crystallized from the filtrate on cooling.

N-(1*H*-Imidazol-2-yl)-2-(piperazin-1-yl)acetamide (32). Yield 83%, mp 107–109°C. ¹H NMR spectrum, δ , ppm: 2.83 s (4H, CH₂NHCH₂), 3.12 s (4H, CH₂NCH₂), 3.44 (2H, CH₂), 7.12 s (2H, 4-H, 5-H), 11.92 br.s (1H, NH). Found, %: C 51.61; H 7.25; N 33.41. C₉H₁₅N₅O. Calculated, %: C 51.66; H 7.23; N 33.47.

N-(1,3-Oxazol-2-yl)-2-(piperazin-1-yl)acetamide (33). Yield 72%, mp 79–81°C. ¹H NMR spectrum, δ , ppm: 2.70 s (4H, CH₂NHCH₂), 3.03 s (4H, CH₂NCH₂), 3.32 (2H, CH₂), 7.38 d (1H, 4-H, *J* = 4.0 Hz), 7.73 d (1H, 5-H, *J* = 4.0 Hz), 11.92 br.s (1H, NH). Found, %: C 51.37; H 6.72; N 26.47. C₉H₁₄N₄O₂. Calculated, %: C 51.42; H 6.71; N 26.65.

Compounds 40–43 (general procedure). A mixture of 3 mmol of 2-chloropyridine-3,4-diamine **31** and 3 mmol of *para*-substituted benzoic acid **36–39** in 4 mL of phosphoryl chloride was heated for 3–3.5 h at 130–135°C. Vigorous evolution of hydrogen chloride was observed. When the reaction was complete, excess POCl₃ was distilled off, the residue was treated with cold water, and the precipitate was filtered off, dried, and recrystallized from appropriate solvent.

4-(4-Chloro-3*H***-imidazo[4,5-***c***]pyridin-2-yl)-***N***,***N***dimethylaniline (40). Yield 64%, mp 245–247°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 3.01 s [6H, N(CH₃)₂], 6.83 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 7.62 d (1H, 7-H,** *J* **= 8.0 Hz), 8.04 d (2H, 2'-H, 6'-H,** *J* **= 8.0 Hz), 8.25 d (1H, 6-H,** *J* **= 8.0 Hz). Found, %: C 61.60; H 4.82; N 20.51. C₁₄H₁₃ClN₄. Calculated %: C 61.65; H 4.80; N 20.54.**

4-Chloro-2-(4-methoxyphenyl)-3*H*-imidazo-[**4,5-***c*]**pyridine (41).** Yield 54%, mp 163–165°C (from H₂O). ¹H NMR spectrum, δ , ppm: 3.85 s (3H, OCH₃), 7.15 d (2H, 3'-H, 5'-H, *J* = 8.0 Hz), 7.57 d (1H, 7-H, *J* = 8.0 Hz), 8.09 d (1H, 6-H, *J* = 8.0 Hz), 8.18 d (2H, 2'-H, 6'-H, *J* = 8.0 Hz). Found, %: C 60.10; H 3.92; N 16.14. C₁₃H₁₈ClN₃O. Calculated, %: C 60.12; H 3.88; N 16.18.

4-Chloro-2-(4-chlorophenyl)-3*H***-imidazo[4,5-***c***]pyridine (42). Yield 53%, mp 243–245°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 7.57 d (1H, 7-H, J = 8.0 Hz), 7.68 d (2H, 3'-H, 5'-H, J = 8.0 Hz), 7.93 d (1H, 6-H, J = 8.0 Hz), 8.24 d (2H, 2'-H, 6'-H, J = 8.0 Hz). Found, %: C 54.53; H 2.69; N 15.87. C₁₂H₇Cl₂N₃. Calculated, %: C 54.57; H 2.67; N 15.91.**

2-(4-Bromophenyl)-4-chloro-3*H***-imidazo[4,5-***c***]pyridine (43). Yield 50%, mp 268–270°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 7.60 d (1H, 7-H, J = 8.0 Hz), 7.81 d (2H, 3'-H, 5'-H, J = 8.0 Hz), 8.13 d (1H, 6-H, J = 8.0 Hz), 8.15 d (2H, 2'-H, 6'-H, J = 8.0 Hz). Found, %: C 46.68; H 2.31; N 13.58. C₁₂H₇BrClN₃. Calculated, %: C 46.71; H 2.29; N 13.62.**

Condensation of compounds 40–43 with acetamides 32 and 33 (*general procedure***).** A mixture of 0.4 mmol of compound **40–43**, 0.4 mmol of acetamide **32 or 33**, 0.5 mL of propan-1-ol, and 0.3 mL of triethylamine was heated for 4–5 h at 110–120°C. The mixture was evaporated to dryness, the residue was treated with water, and the precipitate was filtered off, dried, and recrystallized from appropriate solvent. **2-(4-{2-[(4-Dimethylamino)phenyl]-1***H*-imidazo-[4,5-*c*]pyridin-4-yl}piperazin-1-yl)-*N*-(1*H*-imidazol-**2-yl)acetamide (7)**. Yield 62%, mp 183–185°C (from EtOH). ¹H NMR spectrum, δ , ppm: 2.85 s (4H, CH₂NCH₂), 3.03 s [6H, N(CH₃)₂], 3.10 s (4H, CH₂NCH₂), 3.41 (2H, CH₂), 6.81 d (2H, 3'-H, 5'-H, *J* = 8.0 Hz), 7.10 s (2H, 4"-H, 5"-H), 7.60 d (1H, 7-H, *J* = 8.0 Hz), 8.07 d (2H, 2'-H, 6'-H, *J* = 8.0 Hz), 8.23 d (1H, 6-H, *J* = 8.0 Hz). Found, %: C 61.98; H 6.16; N 28.27. C₂₃H₂₇N₉O. Calculated, %: C 62.01; H 6.11; N 28.30.

2-(4-{2-[(4-Dimethylamino)phenyl]-1*H***-imidazo-[4,5-***c***]pyridin-4-yl}piperazin-1-yl)-***N***-(1,3-oxazol-2yl)acetamide (9). Yield 63%, mp 152–154°C (from EtOH). ¹H NMR spectrum, \delta, ppm: 2.75 s (4H, CH₂NCH₂), 2.99 s [6H, N(CH₃)₂], 3.05 s (4H, CH₂NCH₂), 3.37 (2H, CH₂), 6.87 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 7.37 d (1H, 4"-H,** *J* **= 4.0 Hz), 7.63 d (1H, 7-H,** *J* **= 8.0 Hz), 7.75 d (1H, 5"-H,** *J* **= 4.0 Hz), 8.06 d (2H, 2'-H, 6'-H,** *J* **= 8.0 Hz), 8.28 d (1H, 6-H,** *J* **= 8.0 Hz). Found, %: C 61.49; H 5.91; N 25.01. C₂₃H₂₆N₈O₂. Calculated, %: C 61.87; H 5.87; N 25.10.**

2-{4-[2-(4-Methoxyphenyl)-1*H***-imidazo[4,5-***c***]pyridin-4-yl]piperazin-1-yl}-***N***-(1,3-oxazol-2-yl)acetamide (15). Yield 78%, mp 193–195°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 2.72 s (4H, CH₂NCH₂), 3.03 s (4H, CH₂NCH₂), 3.37 (2H, CH₂), 3.85 s (3H, OCH₃), 7.14 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 7.37 d (1H, 4"-H,** *J* **= 4.0 Hz), 7.59 d (1H, 7-H,** *J* **= 8.0 Hz), 7.75 d (1H, 5"-H,** *J* **= 4.0 Hz), 8.04 d (1H, 6-H,** *J* **= 8.0 Hz), 8.14 d (2H, 2'-H, 6'-H,** *J* **= 8.0 Hz). Found, %: C 60.90; H 5.39; N 21.44. C₂₂H₂₃N₇O₃. Calculated, %: C 60.96; H 5.35; N 22.62.**

2-{4-[2-(4-Chlorophenyl)-1*H***-imidazo[4,5-***c***]pyridin-4-yl]piperazin-1-yl}-***N***-(1***H***-imidazol-2-yl)acetamide (19). Yield 52%, mp 207–209°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 2.83 s (4H, CH₂NCH₂), 3.12 s (4H, CH₂NCH₂), 3.47 (2H, CH₂), 7.16 s (2H, 4"-H, 5"-H), 7.72 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 7.94 d (1H, 6-H,** *J* **= 8.0 Hz), 8.26 d (2H, 2'-H, 6'-H,** *J* **= 8.0 Hz), 7.58 d (1H, 7-H,** *J* **= 8.0 Hz). Found, %: C 57.65; H 4.87; N 25.58. C₂₁H₂₁ClN₈O. Calculated, %: C 57.73; H 4.84; N 25.65.**

2-{4-[2-(4-Chlorophenyl)-1*H***-imidazo[4,5-***c***]pyridin-4-yl]piperazin-1-yl}-***N***-(1,3-oxazol-2-yl)acetamide (21). Yield 52%, mp 173–175°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 2.69 s (4H, CH₂NCH₂), 3.03 s (4H, CH₂NCH₂), 3.35 (2H, CH₂), 7.37 d (1H, 4"-H,** *J* **= 4.0 Hz), 7.59 d (1H, 7-H,** *J* **= 8.0 Hz), 7.68 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 7.71 d** (1H, 5"-H, J = 4.0 Hz), 7.95 d (1H, 6-H, J = 8.0 Hz), 8.23 d (2H, 2'-H, 6'-H, J = 8.0 Hz). Found, %: C 57.56; H 4.62; N 22.37. C₂₁H₂₀ClN₇O₂. Calculated, %: C 57.60; H 4.60; N 22.39.

2-{4-[2-(4-Bromophenyl)-1*H***-imidazo[4,5-***c***]pyridin-4-yl]piperazin-1-yl}-***N***-(1,3-oxazol-2-yl)acetamide (27). Yield 78%, mp 212–214°C (from PrOH). ¹H NMR spectrum, \delta, ppm: 2.72 s (4H, CH₂NCH₂), 3.05 s (4H, CH₂NCH₂), 3.34 (2H, CH₂), 7.37 d (1H, 4"-H,** *J* **= 4.0 Hz), 7.62 d (1H, 7-H,** *J* **= 8.0 Hz), 7.73 d (1H, 5"-H,** *J* **= 4.0 Hz), 7.81 d (2H, 3'-H, 5'-H,** *J* **= 8.0 Hz), 8.11 d (2H, 2'-H, 6'-H,** *J* **= 8.0 Hz), 8.15 d (1H, 6-H,** *J* **= 8.0 Hz). Found, %: C 52.21; H 4.23; N 20.29. C₂₁H₂₀BrN₇O₂. Calculated, %: C 52.29; H 4.18; N 20.33.**

REFERENCES

- Sen, S., Zhou, H., and White, R.A., *Oncogene*, 1997, vol. 14, p. 2195.
- 2. Libertini, S., Abagnale, A., Passaro, C., Botta, G., and Portella, G., *Recent Pat. Anti-Cancer Drug Discovery*, 2010, vol. 5, p. 219.
- Zhang, J., Yang, P.L., and Gray, N.S., *Nat. Rev. Cancer*, 2001, vol. 9, p. 28.
- Dar, A.A., Goff, L.W., Majid, Sh., Berlin, J., and El-Rifai, W., *Mol. Cancer Ther.*, 2010, vol. 9, p. 268.
- Murata-Hori, M., Tatsuka, M., and Wang, Y.L., *Mol. Biol. Cell*, 2002, vol. 13, p. 1099.
- Gürtler, U., Tontsch-Grunt, U., Jarvis, M., Zahn, S.K., Boehmelt, G., Quant, J., Adolf, G.R., and Solca, F., *J. Clin. Oncol.*, 2010, vol. 28, p. 1363.
- Sorrentino, R., Libertini, S., Pallante, P.L., Troncone, G., Palombini, L., Bavetsias, V., Spalletti-Cernia, D., Laccetti, P., Linardopoulos, S., Chieffi, P., Fusco, A., and Portella, G., *J. Clin. Endocrinol. Metab.*, 2005, vol. 90, p. 928.
- Ladygina, N.G., Latsis, R.V., and En, T., *Biomed. Khim.*, 2005, vol. 51, p. 170.
- Sessa, F., Mapelli, M., Ciferri, C., Tarricone, C., Areces, L.B., Schneider, Th.R., Stukenberg, P.T., and Musacchio, A., *Mol. Cell*, 2005, vol. 18, p. 379.
- 10. Koroleva, E.V., Ignatovich, Zh.I., Sinyutich, Yu.V., and Gusak, K.N., *Russ. J. Org. Chem.*, 2016, vol. 52, p. 139.

- 11. Yan, A., Wang, L., Xu, Sh., and Xu, J., *Drug Discovery Today*, 2011, vol. 16, p. 261.
- Bavetsias, V., Large, J.M., Sun, Ch., Bouloc, N., Kosmopoulou, M., Matteucci, M., Wilsher, N.E., Martins, V., Reynisson, J., Atrash, B., Faisal, A., Urban, F., Valenti, M., de Haven Brandon, A., Box, G., Raynaud, F.I., Workman, P., Eccles, S.A., Bayliss, R., Blagg, J., Linardopoulos, S., and McDonald, E., *J. Med. Chem.*, 2010, vol. 53, p. 5213.
- 13. Lan, P., Chen Wan-Na, and Chen Wei-Min, *Eur. J. Med. Chem.*, 2011, vol. 46, p. 77.
- Bavetsias, V., Crumpler, S., Sun, Ch., Avery, S., Atrash, B., Faisal, A., Moore, A.S., Kosmopoulou, M., Brown, N., Sheldrake, P.W., Bush, K., Henley, A., Box, G., Valenti, M., de Haven Brandon, A., Raynaud, F.I., Workman, P., Eccles, S.A., Bayliss, R., Linardopoulos, S., and Blagg, J., *J. Med. Chem.*, 2012, vol. 55, p. 8721.
- Bavetsias, V., Faisal, A., Crumpler, S., Brown, N., Kosmopoulou, M., Joshi, A., Atrash, B., Pérez-Fuertes, Y., Schmitt, J.A., Boxall, K.J., Burke, R., Sun, C., Avery, S., Bush, K., Henley, A., Raynaud, F.I., Workman, P., Bayliss, R., Linardopoulos, S., and Blagg, J., J. Med. Chem., 2013, vol. 56, p. 9122.
- 16. Lipinski, C.A., Lombardo, F., Dominy, B.W., and Feeney, P.J., *Adv. Drug Delivery Rev.*, 1997, vol. 23, p. 3.
- 17. *Molinspiration ver.2004.1: program (on-line service)*: Molinspiration Cheminformatics, Slovak Rep., 2015. http://www.molinspiration.com
- 18. Stewart, J.J.P., J. Mol. Model., 2013, vol. 19, p. 1.
- 19. Stewart, J.J.P., *MOPAC2012*, Colorado Springs: Stewart Computational Chemistry, 2013. http://openmopac.net
- Pearson, R.G., *Chemical Hardness*, Weinheim: Wiley-VCH, 1997.
- Howard, S., Berdini, V., Boulstridge, J.A., Carr, M.G., Cross, D.M., Curry, J., Devine, L.A., Early, T.R., Fazal, L., Gill, A.L., Heathcote, M., Maman, S., Matthews, J.E., Mcmenamin, R.L., Navarro, E.F., O'Brien, M.A., O'Reilly, M., Rees, D.C., Reule, M., Tisi, D., Williams, G., Vinkovic, M., and Wyatt, P.G., *J. Med. Chem.*, 2009, vol. 52, p. 379.
- 22. Bremer, O.J., Justus Liebigs Ann. Chem., 1935, vol. 518, p. 274.
- 23. Svertilova, I.A., Smolyar, N.N., and Yutilov, Yu.M., *Ukr. Khim. Zh.*, 1996, vol. 62, p. 64.