

Synthesis of N^4 - β -D-glycoside cytosines and sugar N^4 -acetylcytosin-1-ylmethylhydrazones as antiviral agents

Omar M. Ali, Hamada H. Amer and Adel A.-H. Abdel-Rahman*

Department of Chemistry, Faculty of Science, Menoufia University, Shebin El-Koam, Egypt

Reaction of monosaccharide aldoses with cytosine (**1**) gave stereoselectively β - N -glycosides **2a–d**, which were treated with acetic anhydride in pyridine to afford the corresponding acetylated derivatives **3a–d**. N^4 -Acetylcytosine (**4**) was synthesised and treated with ethyl chloroacetate to give 1-(ethoxycarbonylmethyl)- N^4 -acetylcytosine (**5**). Hydrolysis of the latter ester with hydrazine hydrate afforded the hydrazide derivative **6**. Condensation of the hydrazide with monosaccharide aldoses gave the corresponding sugar hydrazones **7a–f**. Acetylation of the hydrazones afforded the per- O -acetyl derivatives **8a–f**. The prepared compounds were tested for antiviral activity against hepatitis B virus (HBV) which showed moderate activities.

Keywords: cytosine, N^4 -acetylcytosin, N -glycosides, antiviral activity

Glycosyl amines^{1–6} are important because these occur as junctures in glycoproteins.^{7,8} The chemical and structural nature of the derivatives formed from the reaction of a monosaccharide and different nitrogen bases depends upon the reaction conditions and the base used.⁹ N -Glycosyl amines have better advantage in binding to metal ions over their saccharide counter parts, as these compounds provide additional binding centres. In view of such important aspects of N -glycosyl amines, herein we report the synthesis and characterisation of different N -glycosyl amines of simple saccharides using cytosine as an amine. Recently, the synthesis of acyclic nucleosides has attracted much attention.¹⁰ Consequently, this significance and possible enhancement of biological activity resulting from the attachment of carbohydrate moieties to heterocycles and our interest in the synthesis of heterocyclic derivatives of carbohydrates,¹¹ attracted our attention to synthesise C -nucleosides using monosaccharide (N^4 -acetylcytosine-1-yl)acetohydrazides.

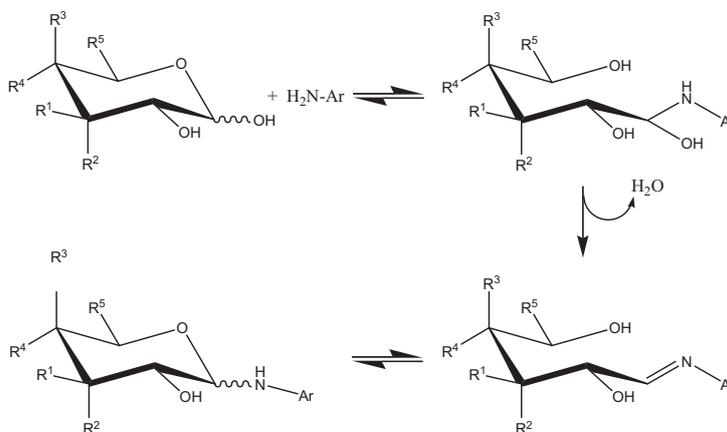
Results and discussion

Condensation of cytosine (**1**) with a number of monosaccharides (D-glucose, D-galactose, D-ribose, and D-xylose) in boiling ethanol and in the presence of a catalytic amount of glacial acetic acid gave stereoselectively 4-(β -D-glycopyranosylamino)pyrimidin-2(1*H*)-ones **2a–d** in 85–88% yields. In the ¹H NMR spectra of **2a–d**, the anomeric protons appear as doublet in the range δ 7.16–7.20 ppm, in which $J_{(1,2)} = 8.0$ Hz. This value is consistent with the β -configuration of ⁴C₁ (D) conformation.¹²

The reaction of **2a–d** with acetic anhydride in pyridine at room temperature led to the acetylation of sugar hydroxyl groups without affecting the NH group to give tri- or tetraacetoxy- β -D-glycosides **3a–d** in 95–98% yields. The IR spectra of **3a–d** are characterised by two absorption bands at 3380–3395 and 1745–1755 cm⁻¹ indicating the presence OAc and NH groups. In addition, the ¹H NMR spectra showed triplets at low field δ 5.15–5.20 ppm assigned as H-1' with $J_{(1,2)} = 9.0$ Hz, which confirm the expected β -configuration of ⁴C₁ (D) conformation.

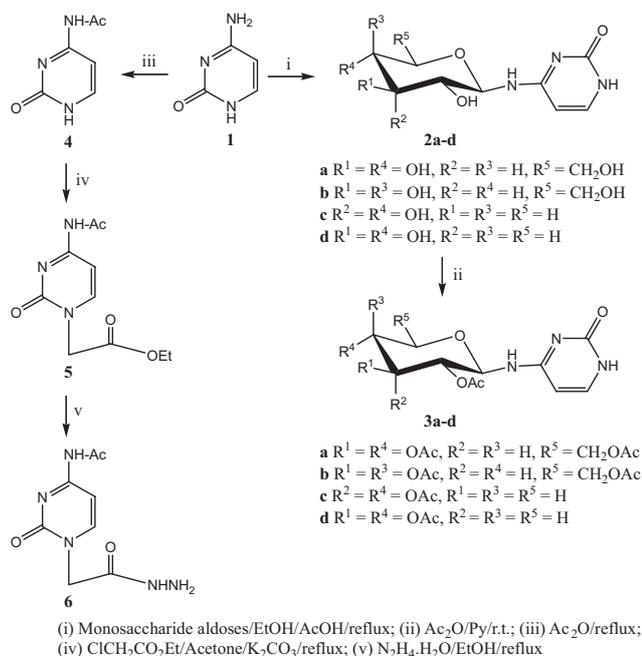
Refluxing of cytosine in acetic anhydride afforded the corresponding N^4 -acetylcytosine (**4**),¹³ which was treated with ethyl chloroacetate in acetone and anhydrous K₂CO₃ to afford 1-(ethoxycarbonylmethyl)- N^4 -acetylcytosine (**5**) in 75% yield. Hydrazinolysis of the ethyl ester **5** gave (N^4 -acetylcytosin-1-yl)acetohydrazide (**6**) in 89% yield.

Condensation of the hydrazides **6** with L-arabinose, D-ribose, D-xylose, D-glucose, D-galactose, and D-mannose gave the corresponding sugar hydrazones **7a–f** in 80–88% yield. The ¹H NMR spectra of the hydrazones **7a–f** confirmed the presence of sugar protons in the range δ 3.14–4.27 ppm and a doublet at δ 7.20–7.30 ppm corresponding to the proton at C-1 of the sugar. The assignments of NH and OH groups in these compounds were achieved by D₂O exchange. Acetylation of **7a–f** using acetic anhydride in pyridine at room temperature gave the corresponding acetylated derivatives **8a–f** 80–85% yields. The ¹H NMR spectra of the acetyl derivatives **8a–f** showed the O -acetyl-methyl groups at δ 1.95–2.10 ppm and the methine proton as doublet at δ 7.25–7.30 ppm.



Scheme 1

* Correspondent. E-mail: adelnassar63@hotmail.com


Scheme 2

Preliminary viral screening against hepatitis B virus (HBV) (Hep G2 2.2.15 cell method)¹⁴ indicated that compounds **2a-c**, **3a**, **3b**, **7a-d**, **8a** and **8b** were found to be active against HBV replication with IC_{50} 80–90 μ M and CC_{50} 85–95 μ M. Compounds **2d**, **3c**, **7e**, and **8d** showed moderate viral replication inhibition and moderate cytotoxicity, while compounds **3d**, **7f**, **8c**, **8e**, and **8f** showed very low inhibition and high cytotoxicity.

Experimental

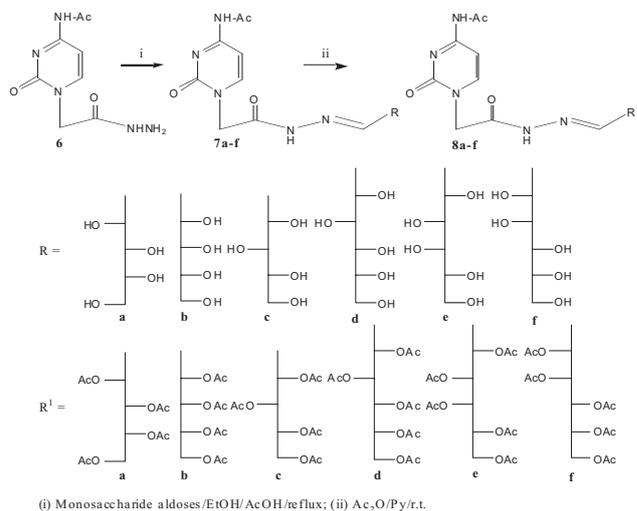
Melting points were determined using a Kofler block instrument. TLC was performed on plastic plates Silica Gel 60 F₂₅₄ (E. Merck, layer thickness 0.2 mm). The detection was achieved by treatment with a solution of 15% H₂SO₄ in methanol, and heating at 150°C. IR spectra were recorded with a Perkin-Elmer model 1720 FTIR (KBr). ¹H NMR spectra were recorded on a Bruker AC 250 FT NMR spectrometer, 250 MHz with TMS as an internal standard. MALDI-MS were measured with a KRATOS Analytical Compact, using 2,5-dihydroxybenzoic acid (DHB) as matrix. The (M + Na)⁺ ion was peak matched using ions derived from the 2,5-dihydroxybenzoic acid matrix. All commercially available reagents were used without further purification. Pyridine was distilled from CaH₂ and stored over molecular sieves. The microanalyses were performed at the microanalytical unit, Universität Konstanz, Germany. Viral screening against HBV was conducted at the National Liver Institute, Menoufia University, Shebin El Koam, Egypt.

4-(β-D-Glycopyranosylamino)pyrimidin-2(1H)-ones **2a-d**

A solution of the respective sugar (5 mmol) in ethanol (5 ml) was treated with **1** (0.55 g, 5 mmol) in ethanol (40 ml), and glacial acetic acid (0.1 ml). The mixture was heated under reflux for 2 h (TLC). The solid, which separated out on cooling, was filtered, washed with ethanol, and crystallised from ethanol to afford **2a-d** in 85–88% yields.

4-(β-D-Glucopyranosylamino)pyrimidin-2(1H)-one (2a): Yield 87%. M.p. 202–204°C. ¹H NMR (DMSO-*d*₆) δ: 3.25–3.60 (m, 2H, H-6'), 4.30–4.90 (m, 8H, H-2', H-3', H-4', H-5', 4xOH), 7.20 (d, 1H, *J* = 8.0 Hz, H-1'), 7.25 (d, 1H, *J* = 5.5 Hz, H-5), 7.72 (d, 1H, *J* = 5.5 Hz, H-6), 9.52 (brs, 1H, NH). MS: *m/z* (%) = 296 [(M⁺ + Na), 39]. Anal. Calcd. for C₁₀H₁₅N₃O₆: C, 43.96; H, 5.53; N, 15.38. Found: C, 43.88; H, 5.23; N, 15.19%.

4-(β-D-Galactopyranosylamino)pyrimidin-2(1H)-one (2b): Yield 85%. M.p. 181–183°C. ¹H NMR (DMSO-*d*₆) δ: 3.25–3.60 (m, 2H, H-6'), 4.30–4.80 (m, 8H, H-2', H-3', H-4', H-5', 4xOH), 7.19 (d, 1H, *J* = 8.0 Hz, H-1'), 7.26 (d, 1H, *J* = 5.5 Hz, H-5), 7.88 (d, 1H, *J* = 5.5 Hz, H-6), 9.59 (brs, 1H, NH). MS: *m/z* (%) = 296 [(M⁺ + Na), 44]. Anal. Calcd. for C₁₀H₁₅N₃O₆: C, 43.96; H, 5.53; N, 15.38. Found: C, 43.75; H, 5.29; N, 15.21%.


Scheme 3

4-(β-D-Ribopyranosylamino)pyrimidin-2(1H)-one (2c): Yield 88%. M.p. 167–169°C. ¹H NMR (DMSO-*d*₆) δ: 3.20–3.50 (m, 2H, H-5'), 4.35–4.95 (m, 7H, H-2', H-3', H-4', 4xOH), 7.18 (d, 1H, *J* = 8.0 Hz, H-1'), 7.24 (d, 1H, *J* = 5.5 Hz, H-5), 7.89 (d, 1H, *J* = 5.5 Hz, H-6), 9.55 (brs, 1H, NH). MS: *m/z* (%) = 266 [(M⁺ + Na), 30]. Anal. Calcd. for C₉H₁₃N₃O₅: C, 44.44; H, 5.39; N, 17.28. Found: C, 44.17; H, 5.22; N, 17.14%.

4-(β-D-Xylopyranosylamino)pyrimidin-2(1H)-one (2d): Yield 86%. M.p. 145–147°C. ¹H NMR (DMSO-*d*₆) δ: 3.10–3.40 (m, 2H, H-5'), 4.45–4.90 (m, 7H, H-2', H-3', H-4', 4xOH), 7.16 (d, 1H, *J* = 8.0 Hz, H-1'), 7.23 (d, 1H, *J* = 5.5 Hz, H-5), 7.81 (d, 1H, *J* = 5.5 Hz, H-6), 9.52 (brs, 1H, NH). MS: *m/z* (%) = 266 [(M⁺ + Na), 27]. Anal. Calcd. for C₉H₁₃N₃O₅: C, 44.44; H, 5.39; N, 17.28. Found: C, 44.15; H, 5.24; N, 17.18%.

4-(2,3,4-Tri-O-acetyl)- and 4-(2,3,4,6-Tetra-O-acetyl)-β-D-glycopyranosylamino)pyrimidin-2(1H)-ones **3a-d**

A mixture of **2a-d** (0.2 g), dry pyridine (3 ml) and acetic anhydride (3 ml) was stirred for 15 min at 0°C, then kept overnight at room temperature with stirring. The mixture was poured onto crushed-ice (30 g) and the precipitate was collected by filtration, washed repeatedly with water, dried, and recrystallised from ethanol–water (2: 8, v/v) to afford **3a-d** in 95–98% yields.

4-(2,3,4,6-Tetra-O-acetyl-β-D-glucopyranosylamino)pyrimidin-2(1H)-one (3a): Yield 98%. M.p. 140–142°C. IR (cm⁻¹): 3390 (NH), 1750 (C=O). ¹H NMR (DMSO-*d*₆) δ: 1.95, 1.97, 1.99, 2.09 (4 s, 12H, 4xCH₃CO), 3.75–3.90 (m, 2H, H-6'), 4.60–5.00 (m, 4H, H-2', H-3', H-4', H-5'), 5.15 (d, 1H, *J* = 9.0 Hz, H-1'), 7.20 (d, 1H, *J* = 5.5 Hz, H-5), 7.70 (d, 1H, *J* = 5.5 Hz, H-6), 9.58 (brs, 1H, NH). MS: *m/z* (%) = 464 [(M⁺ + Na), 25]. Anal. Calcd. for C₁₈H₂₃N₃O₁₀: C, 48.98; H, 5.25; N, 9.52. Found: C, 48.77; H, 5.17; N, 9.46%.

4-(2,3,4,6-Tetra-O-acetyl-β-D-galactopyranosylamino)pyrimidin-2(1H)-one (3b): Yield 95%. M.p. 129–131°C. IR (cm⁻¹): 3370 (NH), 1745 (C=O). ¹H NMR (DMSO-*d*₆) δ: 1.94, 1.96, 1.99, 2.08 (4 s, 12H, 4xCH₃CO), 3.95–4.15 (m, 2H, H-6'), 4.70–5.00 (m, 4H, H-2', H-3', H-4', H-5'), 5.20 (d, 1H, *J* = 9.0 Hz, H-1'), 7.21 (d, 1H, *J* = 5.5 Hz, H-5), 7.73 (d, 1H, *J* = 5.5 Hz, H-6), 9.58 (brs, 1H, NH). MS: *m/z* (%) = 464 [(M⁺ + Na), 22]. Anal. Calcd. for C₁₈H₂₃N₃O₁₀: C, 48.98; H, 5.25; N, 9.52. Found: C, 48.73; H, 5.11; N, 9.39%.

4-(2,3,4-Tri-O-acetyl-β-D-ribofuranosylamino)pyrimidin-2(1H)-one (3c): Yield 97%. M.p. 135–137°C. IR (cm⁻¹): 3396 (NH), 1755 (C=O). ¹H NMR (DMSO-*d*₆) δ: 1.97, 1.99, 2.05 (3 s, 9H, 3xCH₃CO), 3.80–4.00 (m, 2H, H-5'), 4.80–5.08 (m, 3H, H-2', H-3', H-4'), 5.17 (d, 1H, *J* = 9.0 Hz, H-1'), 7.25 (d, 1H, *J* = 5.5 Hz, H-5), 7.70 (d, 1H, *J* = 5.5 Hz, H-6), 9.60 (brs, 1H, NH). MS: *m/z* (%) = 392 [(M⁺ + Na), 56]. Anal. Calcd. for C₁₅H₁₉N₃O₈: C, 48.78; H, 5.19; N, 11.38. Found: C, 48.66; H, 5.05; N, 11.22%.

4-(2,3,4-Tri-O-acetyl-β-D-xylofuranosylamino)pyrimidin-2(1H)-one (3d): Yield 96%. M.p. 120–122°C. IR (cm⁻¹): 3395 (NH), 1755 (C=O). ¹H NMR (DMSO-*d*₆) δ: 1.96, 1.98, 2.11 (3 s, 9H, 3xCH₃CO), 3.78–4.05 (m, 2H, H-5'), 4.75–4.95 (ms, 3H, H-2', H-3', H-4'), 5.18 (d, 1H, *J* = 9.0 Hz, H-1'), 7.20 (d, 1H, *J* = 5.5 Hz, H-5), 7.72 (d, 1H, *J* = 5.5 Hz, H-6), 9.61 (brs, 1H, NH). MS: *m/z* (%) = 392 [(M⁺ + Na), 49]. Anal. Calcd. for C₁₅H₁₉N₃O₈: C, 48.78; H, 5.19; N, 11.38. Found: C, 48.70; H, 5.03; N, 11.20%.

1-(Ethoxycarbonylmethyl)-N⁴-acetylcytosine (5)

A solution of **4**¹³ (1.53 g, 10 mmol), dry acetone (100 ml), K₂CO₃ (1.38 g, 10 mmol) and ethyl chloroacetate (1.23 g, 10 mmol) was refluxed for 5 h (TLC) and then cooled to room temperature. The resulting precipitate was filtered and the filtrate was evaporated under reduced pressure. The residue was recrystallised from ethanol to give 1.79 g (75%) as yellow crystals. M.p. 180–181°C. ¹H NMR (DMSO-*d*₆) δ: 1.20 (t, 3H, *J* = 8.1 Hz, CH₃CH₂), 2.11 (s, 3H, CH₃CO), 4.18 (q, 2H, *J* = 8.1 Hz, CH₂CH₂), 4.61 (s, 2H, CH₂), 7.20 (d, 1H, *J* = 5.5 Hz, H-5), 8.05 (d, 1H, *J* = 5.5 Hz, H-6), 10.84 (brs, 1H, NH). Anal. Calcd. for C₁₀H₁₃N₃O₄: C, 50.21; H, 5.48; N, 17.56. Found: C, 50.04; H, 5.18; N, 17.33%.

(N⁴-Acetylcytosin-1-yl)acetohydrazide (6)

A solution of **5** (2.39 g, 10 mmol) in ethanol (30 ml) was refluxed with hydrazine hydrate (1.25 g, 25 mmol) for 4 h. After cooling it to room temperature, a white solid appeared. This was recrystallised from ethanol to afford **6** (2.0 g, 89%). M.p. 258–260°C. ¹H NMR (DMSO-*d*₆) δ: 2.15 (s, 3H, CH₃CO), 4.20 (brs, 2H, NH₂), 4.34 (s, 2H, CH₂), 7.20 (d, 1H, *J* = 5.5 Hz, H-5), 7.62 (d, 1H, *J* = 5.5 Hz, H-6), 9.50 (brs, 1H, NH). Anal. Calcd. for C₈H₁₁N₃O₃: C, 42.67; H, 4.92; N, 31.10. Found: C, 42.50; H, 4.81; N, 30.94%.

Sugar 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazones 7a-f

A solution of the respective sugar (10 mmol) in water (3 ml) was treated with a solution of **6** (2.25 g, 10 mmol) in ethanol (100 ml) and few drops of glacial acetic acid. The mixture was boiled under reflux for 4–5 h (TLC). The excess of ethanol was removed under reduced pressure and the residue was triturated with diethyl ether (15 ml). The product was filtered, washed with ether, and recrystallised from ethanol to give **7a-f** in 80–88% yield.

L-(+)-Arabinose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7a): Yield 88%. M.p. 150–151°C. ¹H NMR (DMSO-*d*₆) δ: 2.15 (s, 3H, CH₃CO), 3.14–3.40 (m, 3H, H-4', H-5'), 3.49 (s, 1H, H-3'), 3.60–3.65 (m, 1H, H-2'), 4.39–4.51 (m, 6H, CH₂, 4xOH), 7.22 (d, 1H, *J* = 5.5 Hz, H-5), 7.30 (d, 1H, *J* = 2.8 Hz, H-1'), 8.00 (d, 1H, *J* = 5.5 Hz, H-6), 9.88 (brs, 1H, NH). MS: *m/z* (%) = 380 [(M⁺ + Na), 12].

D-(–)-Ribose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7b): Yield 80%. M.p. 140–142°C. ¹H NMR (DMSO-*d*₆) δ: 2.18 (s, 3H, CH₃CO), 3.14–3.52 (m, 5H, H-2', H-3', H-4', H-5'), 3.99–4.21 (brs, 4H, 4xOH), 4.37 (s, 2H, CH₂), 7.15 (d, 1H, *J* = 5.5 Hz, H-5), 7.25 (d, 1H, *J* = 2.8 Hz, H-1'), 8.02 (d, 1H, *J* = 5.5 Hz, H-6), 9.85 (brs, 1H, NH). MS: *m/z* (%) = 380 [(M⁺ + Na), 16].

D-(+)-Xylose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7c): Yield 87%. M.p. 169–171°C. ¹H NMR (DMSO-*d*₆) δ: 2.14 (s, 3H, CH₃CO), 3.41–3.69 (brm, 3H, H-4', H-5'), 3.59–3.61 (m, 2H, H-2', H-3'), 4.35–4.88 (m, 6H, CH₂, 4xOH), 7.20–7.25 (m, 2H, H-5, H-1'), 8.01 (d, 1H, *J* = 5.5 Hz, H-6), 9.90 (brs, 1H, NH). MS: *m/z* (%) = 380 [(M⁺ + Na), 34].

D-(+)-Glucose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7d): Yield 85%. M.p. 122–123°C. ¹H NMR (DMSO-*d*₆) δ: 2.18 (s, 3H, CH₃CO), 3.44–3.89 (m, 5H, H-3', H-4', H-5', H-6'), 4.27–4.88 (m, 8H, CH₂, H-2', 5xOH), 7.18 (d, 1H, *J* = 5.5 Hz, H-5), 7.27 (d, 1H, *J* = 2.8 Hz, H-1'), 8.07 (d, 1H, *J* = 5.5 Hz, H-6), 9.89 (brs, 1H, NH). MS: *m/z* (%) = 410 [(M⁺ + Na), 11].

D-(+)-Galactose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7e): Yield 85%. M.p. 144–146°C. ¹H NMR (DMSO-*d*₆) δ: 2.13 (s, 3H, CH₃CO), 3.34–3.93 (m, 3H, H-5', H-6'), 4.12–4.23 (m, 3H, H-2', H-3', H-4'), 4.33–4.80 (m, 7H, CH₂, 5xOH), 7.15 (d, 1H, *J* = 5.5 Hz, H-5), 7.23 (d, 1H, *J* = 2.8 Hz, H-1'), 8.00 (d, 1H, *J* = 5.5 Hz, H-6), 9.88 (brs, 1H, NH). MS: *m/z* (%) = 410 [(M⁺ + Na), 16].

D-(+)-Mannose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (7f): Yield 82%. M.p. 166–167°C. ¹H NMR (DMSO-*d*₆) δ: 2.16 (s, 3H, CH₃CO), 3.44–3.78 (m, 6H, H-2', H-3', H-4', H-5', H-6'), 4.36–4.50 (m, 7H, CH₂, 5xOH), 7.17 (d, 1H, *J* = 5.5 Hz, H-5), 7.25 (d, 1H, *J* = 2.8 Hz, H-1'), 8.01 (d, 1H, *J* = 5.5 Hz, H-6), 9.86 (brs, 1H, NH). MS: *m/z* (%) = 410 [(M⁺ + Na), 19].

Sugar tetra-O-acetyl- and Sugar penta-O-acetyl-2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazones 8a-f

A cold solution of sugar hydrazones **7a-f** (1 mmol) in dry pyridine (3 ml) was treated with acetic anhydride (3 ml). The reaction mixture was left overnight with stirring, then poured onto crushed ice and the separated product was filtered off, washed with water, dried, and crystallised from ethanol-water (2: 8, v/v) to afford **8a-f** in 80–85% yields.

2,3,4,5-Tetra-O-acetyl-L-(+)-arabinose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8a): Yield 88%. M.p. 150–151°C. ¹H NMR (DMSO-*d*₆) δ: 1.95, 2.04, 2.10, 2.13, 2.15 (5 s, 15H,

5xCH₃CO), 4.17 (m, 2H, H-5'), 4.25 (m, 1H, H-4'), 4.50 (s, 2H, CH₂), 5.60 (m, 1H, H-3'), 5.75 (m, 1H, H-2'), 7.15 (d, 1H, *J* = 5.5 Hz, H-5), 7.25 (d, 1H, *J* = 2.5 Hz, H-1'), 8.05 (d, 1H, *J* = 5.5 Hz, H-6), 10.10 (brs, 1H, NH). MS: *m/z* (%) = 548 [(M⁺ + Na), 22]. Anal. Calcd. for C₂₁H₂₇N₃O₁₁: C, 48.00; H, 5.18; N, 13.33. Found: C, 47.80; H, 5.00; N, 13.21%.

2,3,4,5-Tetra-O-acetyl-D-(–)-ribose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8b): Yield 80%. M.p. 140–142°C. ¹H NMR (DMSO-*d*₆) δ: 1.95, 2.03, 2.11, 2.13, 2.15 (5 s, 15H, 5xCH₃CO), 4.19 (m, 2H, H-5'), 4.33 (m, 1H, H-4), 4.53 (s, 2H, CH₂), 5.69 (m, 1H, H-3), 5.77 (m, 1H, H-2), 7.13 (d, 1H, *J* = 5.5 Hz, H-5), 7.27 (d, 1H, *J* = 2.5 Hz, H-1'), 8.00 (d, 1H, *J* = 5.5 Hz, H-6), 10.07 (brs, 1H, NH). MS: *m/z* (%) = 548 [(M⁺ + Na), 31]. Anal. Calcd. for C₂₁H₂₇N₃O₁₁: C, 48.00; H, 5.18; N, 13.33. Found: C, 47.88; H, 5.07; N, 13.23%.

2,3,4,5-Tetra-O-acetyl-D-(+)-xylose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8c): Yield 87%. M.p. 169–171°C. ¹H NMR (DMSO-*d*₆) δ: 1.95, 2.03, 2.11, 2.12, 2.15 (5 s, 15H, 5xCH₃CO), 4.29 (m, 2H, H-5), 4.39 (m, 1H, H-4), 4.58 (s, 2H, CH₂), 5.74 (m, 1H, H-3), 5.88 (m, 1H, H-2), 7.12 (d, 1H, *J* = 5.5 Hz, H-5), 7.30 (d, 1H, *J* = 2.5 Hz, H-1'), 8.03 (d, 1H, *J* = 5.5 Hz, H-6), 10.00 (brs, 1H, NH). MS: *m/z* (%) = 548 [(M⁺ + Na), 19]. Anal. Calcd. for C₂₁H₂₇N₃O₁₁: C, 48.00; H, 5.18; N, 13.33. Found: C, 47.77; H, 5.04; N, 13.19%.

2,3,4,5,6-Penta-O-acetyl-D-(+)-glucose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8d): Yield 85%. M.p. 122–123°C. ¹H NMR (DMSO-*d*₆) δ: 1.95, 2.00, 2.04, 2.10, 2.13, 2.15 (6 s, 18H, 6xCH₃CO), 4.00–4.11 (m, 2H, H-6), 4.50 (m, 1H, H-5'), 4.60 (s, 2H, CH₂), 4.70 (m, 1H, H-4), 5.15 (m, 1H, H-3), 5.45 (m, 1H, H-2), 7.12 (d, 1H, *J* = 5.5 Hz, H-5), 7.27 (d, 1H, *J* = 2.5 Hz, H-1'), 8.03 (d, 1H, *J* = 5.5 Hz, H-6), 10.00 (brs, 1H, NH). MS: *m/z* (%) = 620 [(M⁺ + Na), 17]. Anal. Calcd. for C₂₄H₃₁N₃O₁₃: C, 48.24; H, 5.23; N, 11.72. Found: C, 48.09; H, 5.11; N, 11.53%.

2,3,4,5,6-Penta-O-acetyl-D-(+)-galactose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8e): Yield 85%. M.p. 144–146°C. ¹H NMR (DMSO-*d*₆) δ: 1.95, 2.00, 2.04, 2.10, 2.12, 2.15 (6 s, 18H, 6xCH₃CO), 4.08–4.22 (m, 2H, H-6), 4.59 (m, 1H, H-5'), 4.69 (s, 2H, CH₂), 4.76 (m, 1H, H-4), 5.18 (m, 1H, H-3), 5.40 (m, 1H, H-2), 7.13 (d, 1H, *J* = 5.5 Hz, H-5), 7.30 (d, 1H, *J* = 2.5 Hz, H-1), 8.03 (d, 1H, *J* = 5.5 Hz, H-6), 10.03 (brs, 1H, NH). MS: *m/z* (%) = 620 [(M⁺ + Na), 19]. Anal. Calcd. for C₂₄H₃₁N₃O₁₃: C, 48.24; H, 5.23; N, 11.72. Found: C, 48.13; H, 5.10; N, 11.59%.

2,3,4,5,6-Penta-O-acetyl-D-(+)-mannose 2-[4-acetamido-2-oxopyrimidin-1(2H)-yl]acetylhydrazide (8f): Yield 82%. M.p. 166–167°C. ¹H NMR (DMSO-*d*₆) δ: 1.98, 2.02, 2.04, 2.11, 2.13, 2.15 (6 s, 18H, 6xCH₃CO), 4.13–4.33 (m, 2H, H-6), 4.57 (m, 1H, H-5'), 4.70 (s, 2H, CH₂), 4.79 (m, 1H, H-4), 5.20 (m, 1H, H-3), 5.47 (m, 1H, H-2), 7.14 (d, 1H, *J* = 5.5 Hz, H-5), 7.30 (d, 1H, *J* = 2.5 Hz, H-1), 8.06 (d, 1H, *J* = 5.5 Hz, H-6), 10.05 (brs, 1H, NH). MS: *m/z* (%) = 620 [(M⁺ + Na), 24]. Anal. Calcd. for C₂₄H₃₁N₃O₁₃: C, 48.24; H, 5.23; N, 11.72. Found: C, 48.08; H, 5.14; N, 11.57%.

Received 27 March 2007; accepted 17 May 2007

Paper 07/4564 doi: 10.3184/030823407X215889

References

- G.P. Ellis and J. Honeyman, *Adv. Carbohydr. Chem.*, 1955, **10**, 95.
- R. Kuhn, *Angew. Chem.*, 1957, **69**, 23.
- P.L. Durette, R.L. Bugianesi, M.M. Nonpipom, T.Y. Shen, M.A. Cascieri, M.S. Glitzer and H.M. Katzen, *J. Med. Chem.*, 1978, **21**, 854.
- H.M. Katzen, *J. Biol. Chem.*, 1979, **254**, 2983.
- M.A. Cascieri, R.A. Mumford and H.M. Katzen, *Arch. Biochem. Biophys.*, 1979, **195**, 30.
- S.J. Danishefsky and J.R. Allen, *Angew. Chem. Int. Ed.*, 2000, **39**, 836.
- P.W. Kent, *Essays Biochem.*, 1967, **3**, 105.
- R.C. Hughes, *Progr. Biophys. Mol. Biol.*, 1973, **26**, 189.
- W.H. Ojala, J.M. Ostman and C.R. Ojala, *Carbohydr. Res.*, 2000, **326**, 104.
- E.S.H. El Ashry and Y.E. Kilany, in *Advances in Heterocyclic Chemistry* (Ed.: A.R. Katritzky), Academic press, New York, Acyclonucleosides: Part 1. 1996, **67**, p. 391; Part 2. 1997, **68**, p. 1.; Part 3. 1998, **69**, p. 129.
- F.A. El-Essawy, A.F. Khattab and A.A.-H. Abdel-Rahman, *Monatsh. Chem. In Press*.
- P.A. Gorin, M. Mazurek, *Can. J. Chem.*, 1975, **53**, 1212; M.F. Abdel-Megeed, M.A. Saleh, M.A. Abdo and G.A. El-Hiti, *Collect. Czech. Chem. Commun.*, 1995, **60**, 1016.
- F. Sun, T. Darbre and R. Keese, *Tetrahedron*, 1999, **55**, 9777.
- M.A. Sells, A.Z. Zelent, M. Shvartsman and G. Acs, *J. Virol.*, 1988, **62**, 2836; M.A. Sells, M.L. Chen and G. Acs, *Proc. Nat. Acad. Sci. USA*, 1987, **84**, 1005; B.E. Korba and J.L. Gerin, *Antiviral Res.*, 1992, **19**, 55; T. Fouad, C. Nielsen, L. Brunn and E.B. Pederson, *J. Az. Med. Fac. (GIRLS)*, 1998, **19**, 1173.