

Nucleophilic displacements on a cyclic sulfamidate derived from allosamine: application to the synthesis of thiooligosaccharides

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The cyclic sulfamidate of the allosamine derivative **11** is efficiently prepared by reaction with 1,1'-sulfuryl diimidazole; the regioselective opening of this compound by sulfur nucleophiles furnishes 3-thioglucoamine derivatives, useful intermediates in synthesis of thiooligosaccharides.

Sulfated tetrasaccharide **1** has been shown to be recognized by E-selectin,¹ a protein involved in the acute inflammatory process.² We have reported the synthesis of some oligosaccharides structurally related to E-selectin ligands that are inhibitors of neural cell division.^{3,4} Among the substrates tested, the sulfated trisaccharide **2**, analogue of **1**, showed the highest antimitotic activity on tumor C-6 glioma cells.⁴ A recent report showed that sulfated oligosaccharides of similar structure activate lymphocytes (natural killer cells) to destroy certain tumours and virally infected cells.⁵ Owing to the important biological interest of these compounds, we planned to prepare novel oligosaccharides stable at acidic pH and toward glycosidases. Since thioglycosides fulfill these requirements⁶ we designed the synthesis of sulfated thiotrisaccharide **3**, containing a sulfur atom linking the fucosyl and glucosaminyl moieties. In our synthetic scheme toward **3** the key step is the formation of the thioglycosidic bond. This reaction can be achieved by nucleophilic displacement at C-3 of an allosamine derivative with a fucose thiolate. In this work we have found that a cyclic sulfamidate derived from D-allosamine is regioselectively opened by nucleophiles in a reaction where conventional leaving groups, such as the triflate or tosylate, gave no reaction.

The allosamine **5**[†] (Scheme 1) was prepared in a simple and direct way from benzyl 2-acetamido-2-deoxy-β-D-glucopyranoside **4**.⁷ A selective inversion took place at C-3 of the glucose derivative. Debenzoylation of **5** followed by benzylation furnished **6**. Treatment of **6** with Tf₂O–pyridine did not afford the corresponding triflate at C-3, but instead a complex mixture of products was obtained. The reaction under the same conditions on the 2-phthalimide derivative **7** gave cleanly triflate **8**, however, when **8** was subjected to nucleophilic displacement with KSAc the elimination product **9** was

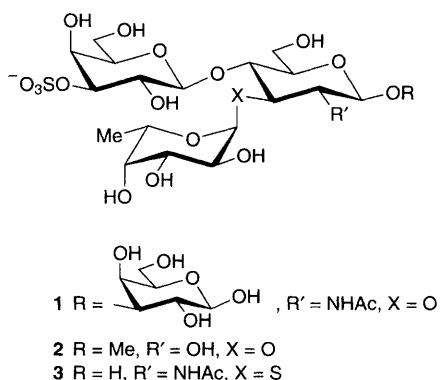
the main product. With the 3-tosylate **10** no reaction occurred with KSAc at 80 °C for 24 h.

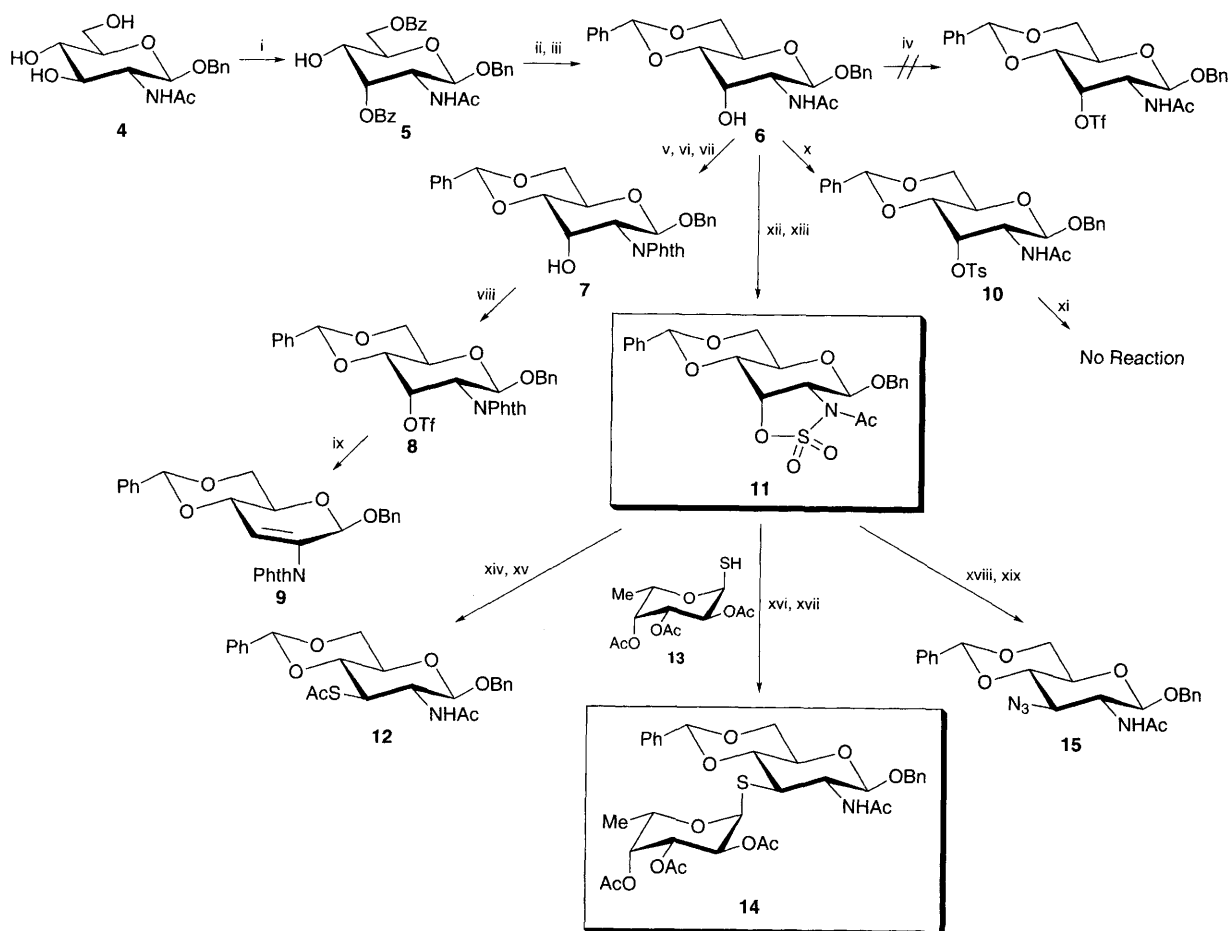
The recent application of cyclic sulfamidates derived from serine to activate the β-position to nucleophilic attack,^{8,9} suggested to us that such a methodology could be applicable to our problem. Direct formation of the cyclic sulfamidate **11** from **6** by reaction with sulfuryl chloride⁸ failed and the starting material remained unaltered. The two step procedure,⁹ *i.e.* reaction with thionyl chloride followed by oxidation, gave under the best conditions only a 45% yield of **11**. We found that **11** could be obtained in higher yield if **6** is treated with 1,1'-sulfuryl diimidazole followed by acetyl chloride to reinstall the acetyl group on the nitrogen atom that was cleaved under the basic conditions used. Without further optimization a 74% yield of **11** was obtained. The regioselective opening of **11** with KSAc afforded the 3-thio derivative **12** (82%) which is already an appropriate intermediate for the synthesis of thioglycosides. Nevertheless, by the reaction of the fucose 1-thiolate **13** with **11** the thiodisaccharide **14** is directly obtained in satisfactory yield (50%). To illustrate the potential of cyclic sulfamidates in substitution reactions, **11** was treated with a different nucleophile, sodium azide. The azide displacement took place at room temperature in only 2 h to give **15** in 92% yield. An azide displacement on a similar mesylated substrate has been reported¹⁰ to proceed in 85% yield after 4 d at 90 °C.

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Footnotes

[†] All new compounds gave satisfactory elemental analyses and the expected ¹³C NMR spectra. *Selected data (J/Hz) for 5*: mp 137 °C; [α]_D –52.0 (c 1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.9 (3 H, s, NHAc), 2.88 (1 H, d, J 3.4, OH), 4.1 (2 H, m, H-4, H-5), 4.4 (1 H, dt, J_{1,2} 7.9, J_{NH,2} 8.0, J_{2,3} 2.7, H-2), 4.87 (1 H, d, J_{1,2} 7.9, H-1), 5.85 (1 H, t, J 2.7, H-3). *For 6*: mp 265 °C, [α]_D –128.1 (c 0.1, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 1.99 (3 H, s, NHAc), 2.5 (1 H, d, J 1.5, OH), 3.7 (1 H, dd, J_{3,4} 2.3, J_{4,5} 9.2, H-4), 4.25 (2 H, m, H-2, H-3), 4.7 (1 H, d, J_{1,2} 7.96, H-1), 5.6 (1 H, s, CHPh), 5.87 (1 H, d, J 8.8, NH). *For 11*: mp 165 °C, [α]_D –55.2; ¹H NMR (300 MHz, CDCl₃) δ 2.38 (3 H, s, NAc), 3.83 (1 H, t, J 10.3, H-6_a), 4.0 (2 H, m, H-5, H-4), 4.49 (1 H, dd, J_{6a,6e} 10.3, J_{5,6e} 4.5, H-6_e), 4.64 (1 H, d, J 11.8, CH₂Ph), 4.71 (1 H, m, H-2), 4.93 (1 H, d, J 11.8, CH₂Ph), 4.97 (1 H, d, J_{1,2} 7.3, H-1), 5.22 (1 H, dd, J 2.5, J 4.3, H-3), 5.6 (1 H, s, CHPh). *For 12* ¹H NMR (500 MHz, CD₃COCD₃) δ 1.8 (3 H, s, NHAc), 2.24 (3 H, s, SAc), 3.68 (1 H, dd, J_{4,5} 8.97, J_{3,4} 10.8, H-4), 3.87 (1 H, t, J 10.8, H-3), 4.09 (1 H, m, H-2), 4.85 (1 H, d, J_{1,2} 8.2, H-1), 5.6 (1 H, d, J_{NH,2} 7.1, NH). *For 14*: mp 132 °C, [α]_D –122.3 (c 1, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.63 (3 H, d, J 6.6, CH₃, Fuc), 3.38 (2 H, m, H-3, H-4), 3.58 (2 H, m, H-2, H-5), 3.74 (1 H, t, J 10.4, H-6_a), 4.34 (1 H, dd, J_{5,6e} 4.7, J_{6a,6e} 10.4, H-6_e), 4.41 (1 H, q, H-5'), 4.56 (1 H, d, J 11.9, CH₂Ph), 4.83 (1 H, d, J_{1,2} 7.87, H-1), 4.88 (1 H, d, J 11.9, CH₂Ph), 5.03 (1 H, dd, J_{2',3'} 10.9, J_{1',2'} 5.7, H-2'), 5.08 (1 H, d, J 3.0, H-4'), 5.14 (1 H, dd, J_{2',3'} 10.9, J_{3',4'} 3.0, H-3'), 5.52 (2 H, d, J_{NH,2} 6.96, NH, CHPh). *For 15*: ¹H NMR (300 MHz, CDCl₃) δ 1.97 (3 H, s, NHAc), 3.20 (1 H, m, H-2), 3.81 (1 H, t, J 10.4, H-4), 4.44 (1 H, dd, J_{2,3} 10.8, J_{3,4} 9.18, H-4), 5.1 (1 H, d, J_{1,2} 8.3, H-1), 5.62 (1 H, d, J_{NH,2} 7.5, NH).





Scheme 1 Reagents and conditions: i, BzOH, Ph₃P, DEAD, THF, 65 °C, 1.5 h, 85%; ii, NaOMe (0.1 mol dm⁻³), MeOH, room temp., 1 h; iii, PhCH(OMe)₂, *p*-TsOH, MeCN, room temp., 2 h, 94% (2 steps); iv, Tf₂O, Py, CH₂Cl₂, -30 °C, 30 min; v, KOH, MeOH (32% *m/m*), 125 °, 20 h; vi, phthalic anhydride, CHCl₃, room temp., 1 h; vii, Ac₂O, Pyridine, reflux, 5 h, 70% (3 steps); viii, Tf₂O, Py, CH₂Cl₂, -30 °C, 30 min; ix, KSac, DMF, 0 °C, 1 h, 75%; x, TsCl, Py, CH₂Cl₂, room temp., 4 h, 65%; xi, KSac, DMF, 80 °C, 24 h; xii, 1,1'-sulfonyl-diimidazole, NaH, DMF, -40 °C → room temp.; xiii, AcCl, Py, CH₂Cl₂, room temp., 2 h, 74% (2 steps); xiv, KSac, DMF, room temp., 30 min; xv, H₂SO₄, H₂O, THF, room temp., 30 min, 82%; xvi, **13**, NaH, DMF, 0 °C, 15 min; xvii, H₂SO₄, H₂O, THF, room temp., 30 min, 50% (2 steps); xviii, NaN₃, DMF, room temp., 2 h; xix, H₂SO₄, H₂O, THF, room temp., 30 min, 92% (2 steps). Py = pyridine.

‡ Preparation of **11**: To a stirred solution of **6** (300 mg, 0.75 mmol) in dry DMF (2 cm³) was added sodium hydride (54 mg, 2.25 mmol) at 0 °C under argon. When the hydrogen bubbling ceased, the reaction mixture was cooled at -40 °C, and 1,1'-sulfonyl diimidazole (222 mg, 1.12 mmol) in dry DMF (1 cm³) was added dropwise. After being warmed to room temperature over a period of 1 h, the solvent was removed. The residue was dissolved in CH₂Cl₂ (9 cm³), then acetyl chloride (64 mm³ 0.9 mmol) and pyridine (146 mm³ 1.8 mmol) were added, and the solution stirred at room temperature for 2 h. The mixture was diluted with CH₂Cl₂, washed successively with saturated aq. NaHCO₃ and water, dried (Na₂SO₄) and concentrated. Flash chromatography of the residue (CH₂Cl₂-AcOEt 110:1) gave the sulfamidate **11** (256 mg, 74%).

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