Hydroxy-Directed Regio- and Diastereoselective Ene Reaction of Singlet Oxygen with Chiral Allylic Alcohols

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Abstract: The photooxygenation of chiral allylic alcohols 1a, (Z)-1f-k, ethers 1b-d, and acetate 1e gave the corresponding hydroperoxy homoallylic alcohols and derivatives 2 through the ene reaction with singlet oxygen. While the reaction of the acetate 1e proceeded *erythro*-diastereoselectively as a result of the classical *cis* effect, for the allylic alcohols 1a and (Z)-1f-k, in which an alkyl group is located *cis* to the hydroxy-bearing substituent, high *threo* selectivity was observed. This finding is explained in terms of coordination of the nucleophilic hydroxy functionality of the stereogenic center with the incoming electrophilic singlet oxygen enophile. The stereodifferentiation is a consequence of the preferred conformation of the allylic alcohol for oxygen transfer, which is mainly determined by 1,3-allylic strain, while the influence of 1,2-allylic strain is small. A similar sensitivity toward both types of allylic strain is observed in epoxidations with *m*-CPBA, for which stereocontrol by cooperation of hydroxy-coordination and allylic strain is established. These similarities were convincingly demonstrated for the chiral allylic alcohol (Z)-1g as a novel stereochemical probe. Moreover, from these results it can be concluded that the optimal C=C-C-O dihedral angle of the allylic alcohol in the transition state for the singlet oxygen ene reaction lies between 90° and 130°. In addition to the *threo* selectivity with which the hydroperoxy moiety is introduced, the newly formed allylic double bond in the hydroperoxide is exclusively formed in the *E* configuration, as exemplified for the chiral allylic alcohol (Z)-1k; again, allylic strain in the ${}^{1}O_{2}$ ene reaction is responsible.

The prototropic ene reaction of singlet oxygen $({}^{1}O_{2})$ with alkenes represents a convenient route to allylic hydroperoxides, a class of compounds of versatile synthetic utility.¹ Much effort has been invested to achieve regio- and diasterocontrol in this prototropic ene reaction, and the by now classical *cis* effect² constitutes one of the prominent factors of attaining this goal with some success. More recently, the *gem*-directing effect³ and the "nonbonding large group" interactions⁴ have been recognized. Additionally, excellent regiocontrol is exercised by silyl^{4b,5} substituents on the double bond, which direct oxyfunctionalization to the α -allylic site. Furthermore, although diastereofacial differentiation in cyclic and bicyclic substrates has been abundantly documented in ${}^{1}O_{2}$ ene reactions,⁶ for acyclic olefins such control was only recently accomplished.^{2c,d} Again, it is the *cis* effect² which provides the opportunity for diastereomeric selection.

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Previously we reported⁷ that the photooxygenation of chiral allylic alcohols proceeds with a high degree of regio- and diastereoselectivity which was attributed to coordination of the hydroxy functionality with the incoming ${}^{1}O_{2}$. Such directing effects, which are well-recognized in peracid⁸ and transition metal-catalyzed^{8c,d,9} epoxidations as well as in nitrile oxide cycloadditions,¹⁰ were hitherto unknown for ${}^{1}O_{2}$ reactions. Herewith we report the full details on the synthetic and mechanistic features of this useful regio- and diastereoselectively controlled ${}^{1}O_{2}$ ene reaction.

Results

Photooxygenation of the allylic alcohols 1a, (Z)-1f-k, and (E)-1f,g, ethers 1b-d, and the acetate 1e afforded the corresponding β -hydroperoxy homoallylic alcohols and derivatives 2 as a result of the ene reaction with singlet oxygen (Scheme I). The observed product ratios, which were determined by ¹H NMR spectroscopy directly on the crude reaction mixture after evaporation of the solvent, are given in Tables I and II.

The conversion of the allylic alcohols 1a and (Z)-1f-k, which bear an alkyl substituent *cis* to the hydroxy group, proceeded wth high regio- and diastereoselectivity to afford predominantly the *threo*-configurated hydroperoxides $(S^*, S^*)-2a, f-k$ (entries 1 and 8-13, Table I). The degree of *threo* selectivity for these allylic alcohols is, within error limits, independent of the alkyl substituents

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Scheme I



Table I: Regio- and Diastereoselectivity^a in the Photooxygenation of Chiral Allylic Alcohols 1a, (Z)-1f-k, Ethers 1b-d, and Acetate 1e

		\mathbb{R}^1			R4	x	solvent	diastereoselectivity		regioselectivity	
			R ²	R ³				(S*,S*)-2	(S*,R*)-2	2	3
1	1a	Me	Н	Me	Н	Н	CCl ₄	93	7	96	46
2	1a	Me	н	Me	н	Н	MeOH	73	27	96	4 ^b
3	1a	Me	Н	Me	Н	Н	MeCN ^c	76	24	96	4 ^b
4	1b	Me	Н	Me	н	SiMe ₃	CCl4	80	20	74	26 ^d
5	1c	Me	Н	Me	н	SiMe ₂ -t-Bu	CCl ₄	83	17	67	334
6	1d	Me	н	Me	н	Me	CCl4	72	28	80	20ª
7	1e	Me	н	Me	н	Ac	CCl ₄	39	61	82	18e
8	(Z)-1f	Me	н	Н	Н	Н	CCl ₄	93	7	>97	<¥
9	(Z)-1g	Me	Н	Н	Me	н	CCl ₄	93	7	>97	<358
10	(Z)-1h	Et	Н	Н	Н	Н	CCl4	94	6	>97	<¥
11	(Z)-1i	i-Pr	н	Н	Н	н	CCl ₄	94	6	>97	<¥
12	(Z)-1j	t-Bu	н	н	н	н	CCl ₄ ^h	95	5	>97	<3∕
13	(Z)-1k	Me	n-Pr	Н	н	Н	CCl4	94	6'	>97	<3⁄

^a Error was $\pm 5\%$ of stated value. ^b The regioisomer 3a was observed in the form of its ring tautomer 4a. ^c Rose bengal was used as sensitizer. ^d The intermediary enol ethers 3b-d were unstable under the reaction conditions. ^e (*E*)-3e:(*Z*)-3e = 38:62. ^f No products 3f-k could be detected. ^g 8% of [(S^{*}, S^{*}), (S^{*}, R^{*})]-5 was formed as side products in a 23:77 ratio. ^h The photooxygenation was conducted in the presence of 10 mol % 2,6-di-*tert*-butyl-4-methylphenol as radical scavenger. ⁱ Only *E*-configurated hydroperoxides 2k were detected.

 Table II:
 Regio- and Diastereoselectivity^a in the Photooxygenation of Chiral Allylic Alcohols (E)-1f,g

reaction proceeded erythro-selectively (eq 1). In the photo-

		R4		diastereo	regioselectivity		
			solvent	(S*,S*)-2	(S*,R*)-2	2	3
1	(E)-1f	н	CCl₄	54	46	96	46
2	(E)-1g	Me	CCl₄	66	34	>97	<3 ^{b,c}

^a Error was $\pm 5\%$ of stated value. ^b The regioisomer **3f** was observed in the form of its ring tautomer **4f**. ^c 45% of [(S^{*},S^{*}),(S^{*},R^{*})]-5 was formed as side products in a 66:34 ratio.

located in the *cis* position [Me as in 1a, (Z)-1f-j and *n*-Pr as in (Z)-1k] and at the stereogenic unit [R¹ = Me as in 1a, (Z)-1f,g; Et as in (Z)-1h; *i*-Pr as in (Z)-1i; and *t*-Bu as in (Z)-1j]. In the case of (Z)-1k, additional to the large *threo* selectivity, the new double bond is selectively introduced to give exclusively *E*-configurated hydroperoxide 2k (entry 13, Table I). On the contrary, the extent of stereoselection was rather low for the allylic alcohols (*E*)-1f,g which lack such a *cis* substituent (Table II). A methyl group located in the *gem* position to the hydroxy-bearing substituent has only a small influence on the stereochemical course of the reaction (entries 8 and 9, Table I; entry 2, Table II). While the conversion of the allylic alcohol (*E*)-1g proceeded in low selectivity (entry 2, Table II), the degree of diastereoselection for (Z)-1f and (Z)-1g, the latter additionally possessing a *gem* methyl group, is high and identical (entries 8 and 9, Table I).

Furthermore, (Z)-1g was allowed to react with *m*-CPBA and $VO(acac)_2/t$ -BuOOH.¹¹ In the peracid epoxidation high *threo* diastereoselectivity was observed, whereas the vanadium-catalyzed



oxygenations of 1a and (E)-1f the dioxolanes 4a,f were detected in minor amounts, which arise from ketonization of the initial enols 3a,f followed by cyclization.¹² Moreover, in the photooxygenation of allylic alcohols (E)- and (Z)-1g the diastereomeric hydroperoxides (S^*, S^*) - and (S^*, R^*) -5g were formed as regioisomeric side products.

The degree of stereoselection depends on the solvent in which the photooxygenation is performed. As demonstrated for allylic alcohol **1a**, for example, in methanol and acetonitrile significantly lower stereoselectivities were obtained than in CCl₄ (entries 1–3, Table I). Compared to the allylic alcohol **1a**, the corresponding allylic ethers **1b-d** gave smaller selectivities (cf. entries 1 *versus* **4–6**, Table I). Furthermore, in the latter photooxygenations

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several unidentified products were formed, presumably by subsequent reactions of the intermediary enol ethers 3b-d with singlet oxygen;^{3d,6d} the latter result from regioisomeric photooxygenation.

An opposite sense of stereocontrol was observed in the ene reaction of the allylic acetate 1e, for which the erythroconfigurated hydroperoxide (S^*, R^*) -2e dominates (entry 7, Table I). In this case the regioisomeric products (Z,E)-3e also were detected in a ratio which is within error limits identical to that of (S^*, R^*) - and (S^*, S^*) -2e.

Control experiments, i.e. irradiation of allylic alcohols 1a-k in the absence of sensitizer and prolonged photooxygenation of the corresponding hyroperoxides, established that, except for derivative 2j, all starting materials and products were stable under the reaction conditions. In the case of 2j, rearrangement¹³ to the terminal hydroperoxide 6j was observed (Scheme I), which could be suppressed by addition of 2,6-di-tert-butyl-4-methylphenol as radical scavenger.

Diastereomeric Assignments

The relative configurations of 2a, f-h,k and 5g were determined by chemical correlation through catalytic reduction to the corresponding literature-known¹⁴ saturated diols 7a,f-h,k and 8g (Scheme I). For hydroperoxide 2k the large coupling constant¹⁵ of the olefinic protons (J = 15.5 Hz) establishes the E configuration of the double bond. Desilylation of the silyl ether 2b by silica gel in methanol at room temperature (9 h) gave allylic hydroperoxide 2a, which allowed assessment of the relative configuration of 2b.

The stereochemistry of the acetate 2e was established by conversion to the diol 9a by LiAlH₄ reduction (eq 2). The latter



was alternatively prepared by Ph₃P treatment of the allylic hydroperoxide 2a (eq 2). The configurational assignment of the diastereomeric enol acetates 3e is based on the fact¹⁶ that the olefinic proton of (E)-3e ($\delta = 5.20$) is shifted downfield relative to the Z-isomer (Z)-3e ($\delta = 5.11$). More convincingly, irradiation of the allylic methyl protons of (Z)-3e at $\delta = 1.90$ gave a significant NOE enhancement (1.2%) of the olefinic proton at $\delta = 5.11$.

All secondary allylic hydroperoxides 2 displayed similar spectral characteristics: (a) the resonances of both OCH protons of the S^*, S^* hydroperoxides are shifted upfield relative to the S^*, R^* isomers, (b) the vicinal coupling constants of these protons are higher for (S^*, S^*) -2 versus (S^*, R^*) -2, (c) the signals of the hydroxy-bearing carbon atoms are shifted downfield for (S^*, S^*) -2 relative to their S^*, R^* -configurated counterparts, and (d) with the exception of 2j, the latter trend is also observed for the resonances of the carbon atoms which carry the hydroperoxy functionality. On the basis of these spectral criteria and the Scheme II



chemical correlation of 2a,b,e-h,k, the stereochemistry of 2c,d,i,j was assessed.

The stereochemistry of the epoxy alcohols 10 was assigned by comparison of the ¹H NMR spectra with known examples.¹⁷ Thus, for the diastereomeric epoxy alcohols the carbinol proton of the erythro isomer ($\delta = 3.77$) is generally observed at lower field than the corresponding proton of the *threo* isomer ($\delta = 3.69$).

Discussion

The above results provide compelling evidence that the ene reaction of singlet oxygen with allylic alcohols is governed by an interaction between the enophile ${}^{1}O_{2}$ and the allylic hydroxy functionality of the chiral alcohol. The stereodifferentiation is then a consequence of the preferred conformation of the allylic alcohol in the transition state (the C=C-C-O dihedral angle should be of the order of 90-130°), which is determined predominantly by 1,3-allylic strain¹⁸ (Scheme II). Side products may arise from competing ene reactions without involvement of the allylic HO group. Experimental features which are in favor of this mechanism include the following:

(a) The classical cis effect² is not operating in the ene reactions of the chiral allylic alcohols 1a and (Z)-1f-k. Had a cis effect² operated, the enophilic attack of ${}^{1}O_{2}$ would be for optimal intramolecular assistance directed to that side of the substrate which aligns two allylic hydrogens perpendicular to the olefin plane to flank the terminal perepoxide oxygen. Of the two possible geometries for the transition states which fulfill this condition, namely threo- versus erythro-A*, the threo-A* arrangement should be destabilized due to severe 1,3-allylic strain due to methyl-methyl interactions (Scheme III). As a consequence, the photooxygenation would be expected to proceed mainly through the erythro-A* transition state and, thus, the erythroconfigurated hydroperoxides (S^*, R^*) -2 should dominate. Instead, the preferred formation of the threo hydroperoxides suggests that the stereochemical course of this ene reaction is efficiently controlled by a mechanism not subject to the classical cis effect.²

(b) The presence of a free HO group is essential to achieve high degrees of selectivity. That the allylic hydroxy functionality is directly involved is inferred from the solvent dependence on the diastereoselectivity, as exemplified for allylic alcohol 1a (entries 1-3, Table I). In methanol and acetonitrile, solvents which can

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 Chapter 4. Additionally, for derivative [S*, S*-(E)]-2k this assignment was confirmed by an NOE exteriment

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Scheme III



interact with the HO group by hydrogen bonding, a significant drop in the diastereoselectivity is observed. Moreover, functionalization of the HO group also leads to decreased selectivity. While the allylic ethers **1b-d** still react with appreciable *threo* selectivity (entries 4-6, Table I), for the acetate **1e** an inverted sense of diastereoselection is observed, i.e. the *erythro*-configurated hydroperoxide (S^*, R^*) -**2e** dominates (entry 7, Table I); thus, for the acetate the classical *cis* effect² operates. This happenstance is further substantiated by the fact that the regioisomeric ene products, i.e. the enol acetates (Z, E)-**3e**, are formed with the same diasteromeric ratio (dr) value as the *gem*-selective $[(S^*, R^*),$ $(S^*, S^*)]$ -**2e** regioisomers. Therefore, both sets of regioisomers must each derive from a common precursor, namely *erythro*-A* for the major diastereomers (S^*, R^*) -**2e**, (Z)-**3e** and *threo*-A* for the minor (S^*, S^*) -**2e**, (E)-**3e** (X = OAc, Scheme III).

From these substituent effects the nature of the directing effect of the HO group can be inferred. In view of the electrophilic properties¹⁹ of ${}^{1}O_{2}$, it is reasonable to assume that for the alcohols **1a** and (Z)-**1f-k**, as well as the ethers **1b-d**, the nucleophilic allylic oxygen functionalities of these substrates initially interact with the electrophilic ${}^{1}O_{2}$ thereby directing the attack of this enophile mainly to give the *threo*-configurated products. In the case of the corresponding acetate **1e**, the less nucleophilic oxygen functionality at the chiral allylic position does not interact as effectively with ${}^{1}O_{2}$; hence, the reaction is governed by the classical *cis* effect² and *erythro* selectivity is preferred.

The controlling action of the ether functionalities is smaller than that of the HO group, as indicated by the reduced degree of *threo* selectivity in the conversions of derivatives **1b-d** (entries 4-6, Table I) compared to that of the free alcohol **1a** (entry 1, Table I). This may derive from steric hindrance by the silyl and methyl substituents which hinders the approach of ${}^{1}O_{2}$ to the allylic oxygen so that the classical *cis* effect² may compete.

Our results imply that during the early stage of the photooxygenation, i.e. during exciplex formation, but well before the perepoxide transition state is reached, the HO group interacts through its nucleophilic oxygen atom with the electrophilic $^{1}O_{2}$. Once this diastereofacial control has occurred, the HO group can then through hydrogen bonding interact with the negatively charged terminal oxygen atom of the subsequently formed perepoxide transition state and further stabilize the aggregate; however, the stereoselective control appears to derive from the initial interaction between the electrophilic $^{1}O_{2}$ and the nucleophilic oxygen atom of the HO group in the exciplex.

(c) 1,3-Allylic strain¹⁸ is responsible for the three diastereoselectivity. If one accepts the premise that the enophilic attack of ${}^{1}O_{2}$ is directed by the allylic hydroxy functionality, the stereochemical differentiation is then a consequence of the preferred conformation of the hydroxy group of the chiral allylic alcohol in the transition state of the ene reaction. Thus, the diastereoselectivity presumably originates from allylic strain¹⁸ by substituents at the double bond. The importance of such 1,3allylic strain¹⁸ caused by a substituent *cis* to the hydroxy-bearing substituent is recognized by comparison of the photooxygenations of the chiral allylic alcohols **1a**, (Z)-**1f**, and (E)-**1f**,g (compare entries 1 and 8, Table I *versus* entries 1 and 2, Table II). The absence of a methyl group *cis* to the hydroxy-bearing substituent in derivative (E)-**1f** would suggest little 1,3-allylic strain, and consequently almost complete loss of diastereoselectivity is observed (entry 1, Table II). On the other hand, the presence of an additional methyl group in the *gem* position, which introduces 1,2-allylic strain, causes only a minor effect as best revealed by the very similar stereoselectivities observed in the photooxygenation of derivatives (Z)-**1f**,g (entries 8 and 9, Table I) and (E)-**1f**,g (entries 1 and 2, Table II).

(d) The C=C-C-O dihedral angle of the allylic alcohol in the transition state of the ${}^{1}O_{2}$ ene reaction is of the order of ca. $90-130^{\circ}$. Control of stereoselectivity by cooperation of hydroxycoordination and allylic strain is well-established for epoxidations of allylic alcohols with *m*-CPBA and VO(acac)₂/t-BuOOH (Table III). Nevertheless, both reactions exhibit a different sensitivity toward 1,2- and 1,3-allylic strain,¹⁸ which is explained in terms of the proposed^{8d-f,9} transition-state geometries *threo*-1^{*} and *erythro*-1^{*}. While for *m*-CPBA epoxidations a C=C-C-O



threo-1[≢]

preferred transition state for *m*-CPBA epoxidations



erythro-1'

preferred transition state for vanadium-catalyzed epoxidations

dihedral angle of ca. 120° appears to be optimal,^{8d} for the vanadium-catalyzed reaction an angle of ca. 50° applies.^{8d} Dominating control through 1,3-allylic strain¹⁸ accounts for the results in the former (compare entries 1 and 2 with 4 for *m*-CPBA, Table III) and 1,2-allylic strain¹⁸ in the latter (compare entries 1 and 2 with 3 for VO(acac)₂/*t*-BuOOH, Table III). These differences are most convincingly demonstrated in the epoxidations of (Z)-1g (eq 1; entry 6, Table III), a substrate which serves as a versatile stereochemical probe¹¹ since it has the advantage of possessing both a *cis* methyl group (Z = Me) for 1,3-allylic strain and a *gem* methyl group ($R^4 = Me$) for 1,2-allylic strain and thus allows assessment of the relative importance of such steric interactions.

As expected on the basis of coordination by the hydroxy group and allylic strain, the *m*-CPBA epoxidation of (Z)-1g exhibits a high degree of *threo* diastereoselectivity due to 1,3-allylic interactions, while the vanadium-catalyzed reaction proceeds

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Table III: Stereoselectivities in the Epoxidations and Singlet Oxygen Ene Reactions of Chiral Allylic Alcohols 1 (X = H)

		Zª		R4	strain ^b	threo/erythro selectivity			
			R ³			m-CPBA	VO(acac) ₂ /tBuOOH	ene reaction ${}^{1}O_{2}$	
1		Н	Н	Н		60:40¢	20:80°		
2	(<i>E</i>)-1f	н	Me	н		64:36 ^c	29 :71°	54:46ª	
3		н	н	Me	1,2	45:55°	5:95°		
4	(Z)-1f	Me	н	н	1,3	95:5°	71:29°	93:7e	
5	Ìa	Me	Me	н	1.3	95:5°	86:14 ^c	93:7*	
6	(Z)-1g	Me	H	Me	1,2 + 1,3	90:10	33:67/	93:7*	

 $^{a}Z = CH_{2}R^{2}$ in structure 1 of Scheme I. b Type of allylic strain involved in the diasteroselectivity. c Values from ref 8d. d Cf. Table II. e Cf. Table II. f Cf. eq 1.

Scheme IV



erythro-selectively due to control by 1,2-allylic strain. Nonetheless, that 1,3-allylic interactions can be important for VO- $(acac)_2/t$ -BuOOH epoxidations is reflected in the small degree of erythro diastereoselection in the epoxidation of (Z)-1g. This is also evident for 1a and (Z)-1f as substrates, in which 1,2-allylic strain is small while 1,3-allylic strain becomes more significant (entries 4 and 5 for VO(acac)_2/t-BuOOH, Table III). Similarly, the slight reduction in the threo selectivity for the m-CPBA epoxidations of 3-methyl-3-buten-2-ol and (Z)-1g (entries 3 and 6 for m-CPBA, Table III) derives presumably from increased 1,2-allylic strain.

At this point a comparison of the stereochemical course of the singlet oxygen ene reaction with the *m*-CPBA and vanadiumcatalyzed epoxidations is instructive. As can be seen in Table III, not only the sense but also the extent of diastereoselection of the singlet oxygen ene reaction and the *m*-CPBA epoxidation are very similar, i.e. *threo* control, whereas large differences are evident for the vanadium-catalyzed epoxidation. The oxidations of our stereochemical probe (Z)-1g (entry 6, Table III), for which the stereochemical course is affected by 1,2- and 1,3-allylic strain,¹⁸ especially underline these differences. These results imply that the singlet oxygen ene reaction is subject to 1,2- and 1,3allylic strain in the same manner as the *m*-CPBA epoxidation; hence, the preferred transition-state geometries of the allylic alcohols must be very similar. Since the optimal C=C-C-O dihedral angle (α) in the transition state for the *m*-CPBA epoxidation^{8d} is ca. 120°, it follows that for the singlet oxygen ene reaction α should lie between 90° and 130° (structure A, Scheme II).

(e) Ene reactions without participation of the HO group may compete. To a minor extent singlet oxygen may react with allylic alcohols by alternative pathways, similar to reactions of unfunctionalized olefins, i.e. without involvement of the allylic HO group. The formation of the hydroperoxides $\mathbf{5g}$ in the photooxygenation of (Z)-1g demonstrates this possibility hydrogen abstraction from the gem methyl group cannot be assisted by coordination with the HO group because of lack of proximity (structures F and H, Scheme IV). In this context, the high regioselectivity which results from threo attack to afford the allylic hydroperoxides (S^*, S^*) -2g and (S^*, S^*) -5g in a ratio of 98:2 is remarkable and accentuates the importance of the directing effect of the hydroxy functionality. On the other hand, erythro attack involves rather unselective hydrogen abstraction $[(S^*, R^*)-2g:(S^*, R^*)-5g = 47:53]$, which confirms that for both erythro diastereomers $(S^*, R^*)-2g$ and

Scheme V



 (S^*, R^*) -5g control by the hydroxy group is not operating since otherwise (S^*, R^*) -2g should have been preferentially formed relative to (S^*, R^*) -5g.

The finding that the erythro-configurated hydroperoxides are formed mainly without participation of the HO group seems to be general. Were transition states A (threo product) and B (erythro product) the only precursors for (S^*, S^*) -2 and (S^*, R^*) -2 (Scheme II), the 1,3-allylic strain¹⁸ should grow with increasing size of the alkyl group R^1 at the chiral hydroxy-bearing carbon. Hence, structure A should become more decisive relative to B and thus an increase in the *threo* diastereoselectivity would have been expected. The experimental results are not in accord with this expectation since the degree of stereodifferentiation for the alcohols 1f,h-j possessing R¹ groups of different size (Scheme I, $R^1 = Me, Et, i-Pr, t-Bu$) is identical within experimental error (entries 8 and 10-12, Table I). Also these findings speak against significant involvement of the hydroxy functionality in the formation of the erythro hydroperoxides. More likely, these diastereomers arise from the classical singlet oxygen ene reaction (structures C and D, Scheme I) as competing pathway.

(f) The double bond of the allylic hydroperoxides is formed stereoselectively. This is demonstrated in the photooxygenation of 1k (entry 13, Table I). As expected, the hydroperoxy functionality is introduced highly threo-selectively. The advantageous feature of this case is that the *E*-configurated hydroperoxides are exclusively formed; none of the corresponding *Z*-isomers were detected.

Again, the concept of allylic strain¹⁸ offers an explanation for this double stereoselective reaction (Scheme V). The conformer $1k(Z^*)$, from which the Z-configurated hydroperoxides $[S^*, S^*-(Z)]$ - and $[S^*, R^-(Z)]$ -2k would arise, is destabilized by severe 1,3-allylic strain¹⁸ due to interactions between the *n*-Pr group and the chiral substituent. On the other hand, such allylic interactions are minimized in the conformer $1k(E^*)$, which leads to the *E*-configurated hydroperoxides $[S^*, S^*-(E)]$ - and $[S^*, R^*-(E)]$ -2k. Of the latter, the *E*-threo-configurated hydroperoxide $[S^*, S^*-(E)]$ -2k is preferentially formed by the favored HOassisted pathway (entry 13, Table I).

Conclusions

The ene reaction of ${}^{1}O_{2}$ with chiral allylic alcohols 1 proceeds in high regio- and diastereoselectivity through control by 1,3allylic strain, provided efficient coordination between ${}^{1}O_{2}$ and the hydroxy group of the substrate operates. Similar directive effects are expected to occur for other nucleophilic functional groups which are capable of interacting with the electrophilic singlet oxygen as enophile. Such control of stereoselectivity should not be limited to the ene reaction, but should also be applicable to [4 + 2] cycloadditions of ${}^{1}O_{2}$. Moreover, by employing enantiomerically pure chiral substrates, a variety of optically active oxyfunctionalized products would become conveniently available, which should constitute valuable building blocks for asymmetric synthesis.

Experimental Section

General Aspects. IR spectra were recorded with a Perkin-Elmer infrared recording spectrometer 1420 and ¹H and ¹³C NMR spectra with a Bruker AC 200, a Bruker AC 250, or a Bruker WM 400 spectrometer. Elemental analyses were performed in the Analytical Laboratory of the Institute of Inorganic Chemistry, University of Würzburg. For TLC runs Machery und Nagel Polygram SIL G/UV₂₅₄ plates were used; spots were detected with phosphomolybdic acid test spray; for hydroperoxides the KI test was additionally employed. Column-chromatographical purifications were performed on silica gel (63–200 μ m) from Woelm, Erlangen. Commercial reagents and solvents were purified according to literature procedures to match reported physical and spectral data. Allylic alcohols and derivatives 1 were prepared according to literature procedures; cf. references given for each particular compound.

CAUTION! Hydroperoxides are potentially dangerous and should be handled with care!

General Procedure for the Photooxygenation of Allylic Alcohols and Derivatives. A solution of the particular substrate (2.0-5.2 mmol) and ca. 2 mg of tetraphenylporphine in 80 mL of CCl₄ was irradiated with two external Philips G/98/2 SON 150-W sodium lamps at 0 °C while a stream of dry oxygen gas was passed continuously through the reaction mixture. The course of the reaction was monitored by means of TLC; after all starting material had been consumed, the solvent was evaporated (20 °C/18 Torr) and the composition of the crude reaction mixture was determined by ¹H and ¹³C NMR (for product ratios cf. Tables I and II, error ±5%). Further purification by column chromatography on silica gel eluting with mixtures of petroleum ether (50–60 °C) (PE) and ether afforded analytically pure samples of the hydroperoxides 2 as colorless oils.

Photooxygenation of 4-Methyl-3-penten-2- ol^{20} (1a). From 401 mg (4.00 mmol) of 1a was obtained after 4 h and chromatography (PE:ether, 60:40) 471 mg (89%) of 2a.

3-Hydroperoxy-4-methyl-4-penten-2-ol [(S^*, S^*)-**2a**]: ¹H NMR (CDCl₃, 250 MHz) δ 1.09 (d, 3 H, J = 6.4 Hz), 1.70 (dd, 3 H, J = 1.3 Hz, J = 1.0 Hz), 3.63 (br s, 1 H), 3.84 (dq, 1 H, J = 8.6 Hz, J = 6.4 Hz), 4.12 (d, 1 H, J = 8.6 Hz), 5.08–5.10 (m, 2 H), 9.44 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 18.4 (q), 19.1 (q), 67.7 (d), 95.3 (d), 117.2 (t), 141.6 (s). (S^*, R^*)-**2a**: ¹H NMR (CDCl₃, 250 MHz) δ 1.18 (d, 3 H, J = 6.4 Hz), 1.78 (dd, 3 H, J = 1.2 Hz, J = 1.0 Hz), 2.54 (br s, 1 H), 3.95 (dq, 1 H, J = 6.4 Hz, J = 4.9 Hz), 4.28 (d, 1 H, J = 4.9 Hz), 5.04–5.06 (m, 2 H), 9.17 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 18.6 (q), 19.6 (q), 67.4 (d), 92.8 (d), 115.9 (t), 141.6 (s); IR (cm⁻¹) 3700–3040, 1660.

The formation of 2a in the photooxygenation of 1a with methanol as solvent has already been described;²¹ unfortunately no dr values were reported.

3-Hydroxy-3,5,5-trimethyl-1,2-dioxolane (4a): ¹H NMR (CDCl₃, 250 MHz) δ 1.35 (s, 3 H), 1.38 (s, 3 H), 1.53 (s, 3 H), 2.39 (d, 1 H, J = 12.8 Hz), 2.49 (d, 1 H, J = 12.8 Hz), 3.00 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 23.2 (q), 24.8 (q), 27.3 (q), 58.6 (t), 84.0 (s), 106.1 (s); IR (cm⁻¹) 3640–3280, 1660.

Dioxolane 4a could not be isolated due to the too low concentration; it was identified in the crude reaction mixture by comparison of the NMR spectral data with those of an authentic sample.¹²

Photooxygenation of 2-Methyl-4-[(trimethylsilyl)oxy]-2-pentene²² (1b). From 900 mg (5.22 mmol) of 1b was obtained after 8 h and chromatography (PE:ether, 80:20) 672 mg (63%) of 2b.

2-Methyl-4-[(trimethylsilyl)oxy]-1-penten-3-yl hydroperoxide [(S^{\bullet}, S^{\bullet})-2b]: ¹H NMR (CDCl₃, 250 MHz) δ 0.15 (s, 9 H), 1.10 (d, 3 H, J = 6.3 Hz), 1.74 (dd, 3 H, J = 1.3 Hz, J = 1.0 Hz), 3.91 (dq, 1 H, J = 7.7 Hz, J = 6.3 Hz), 4.19 (d, 1 H, J = 7.7 Hz), 4.98–5.05 (m, 2 H), 9.13 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz), δ 0.6 (q), 19.2 (q), 20.8 (q), 69.7 (d), 94.1 (d), 116.1 (t), 141.9 (s). (S^{*}, R^{*})-2b: ¹H NMR (CDCl₃, 250 MHz) δ 0.19 (s, 9 H), 1.16 (d, 3 H, J = 6.3 Hz), 1.77 (dd, 3 H, J =1.1 Hz, 1.0 Hz), 3.95 (dq, 1 H, J = 6.3 Hz, J = 4.6 Hz), 4.27 (d, 1 H, J = 4.6 Hz), 4.98–5.05 (m, 2 H), 8.88 (br s, 1 H); ¹³C NMR (CDCl₃,

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63 MHz) δ 0.4 (q), 19.6 (q), 20.2 (q), 69.4 (d), 92.7 (d), 116.1 (t), 141.9 (s); IR (cm^{-1}) 3650–3250, 1660, 1270. Anal. Calcd for C_9H_{20}O_3Si: C, 52.90; H, 9.87. Found: C, 53.21; H, 10.03.

Photooxygenation of 4-[[(Dimethyl(1,1-dimethylethyl)sily]oxy]-2methyl-2-pentene²³ (1c). From 994 mg (4.65 mmol) of 1c was obtained after 8 h and chromatography (PE:ether, 80:20) 668 mg (58%) of 2c.

4-[[(Dimethyl(1,1-dimethylethyl)sily]oxy]-2-methyl-1-penten-3-yl hydroperoxide [(S^{\bullet}, S^{\bullet})-2c]: ¹H NMR (CDCl₃, 250 MHz) δ 0.11 (s, 3 H), 0.14 (s, 3 H), 0.91 (s, 9 H), 1.10 (d, 3 H, J = 6.3 Hz), 1.75 (dd, 3 H, J = 1.3 Hz, J = 1.0 Hz), 3.95 (dq, 1 H, J = 7.6 Hz, J = 6.3 Hz), 4.21 (d, 1 H, J = 7.6 Hz), 4.98–5.06 (m, 2 H), 8.85 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ -4.3 (q), -4.2 (q), 18.5 (s), 19.4 (q), 20.7 (q), 26.2 (q), 69.8 (d), 94.2 (d), 115.9 (t), 141.9 (s). (S^{*}, R^{*})-2c: ¹H NMR (CDCl₃, 250 MHz) δ 0.08 (s, 3 H), 0.12 (s, 3 H), 0.88 (s, 9 H), 1.18 (d, 3 H, J = 6.3 Hz), 1.78 (dd, 3 H, J = 1.1 Hz, J = 1.0 Hz), 4.00 (dq, 1 H, J = 6.3 Hz, J = 4.6 Hz), 4.30 (d, 1 H, J = 4.6 Hz), 4.98–5.06 (m, 2 H), 8.78 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ -4.6 (q), -4.2 (q), 18.5 (s), 19.7 (q), 20.1 (q), 26.1 (q), 69.7 (d), 92.7 (d), 115.0 (t), 114.9 (s); IR (cm⁻¹) 3660–3320, 1680, 1270. Anal. Calcd for C₁₂H₂₆O₃Si: C, 58.49; H, 10.63. Found: C, 58.76; H, 10.64.

Photooxygenation of 4-Methoxy-2-methyl-2-pentene²⁴ (1d). From 579 mg (5.07 mmol) of 1d was obtained after 8 h and chromatography (PE: ether, 70:30) 423 mg (57%) of 2d.

4-Methoxy-2-methyl-1-penten-3-yl hydroperoxide $[(S^*, S^*)-2d]$: ¹H NMR (CDCl₃, 250 MHz) δ 1.08 (d, 3 H, J = 6.3 Hz), 1.74 (dd, 3 H, J = 1.2 Hz, J = 1.0 Hz), 3.41 (s, 3 H), 3.48 (dq, 1 H, J = 8.3 Hz, J = 6.3 Hz), 4.28 (d, 1 H, J = 8.3 Hz), 5.02–5.06 (m, 2 H), 9.64 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 15.5 (q), 17.9 (q), 59.6 (q), 77.3 (d), 92.9 (d), 116.0 (t), 141.4 (s); IR (cm⁻¹) 3600–3160, 1650. Anal. Calcd for C₇H₁₄O₃: C, 57.51; H, 9.65. Found: C, 57.15; H, 9.64. (S^{*}, R^{*})-2d: ¹H NMR (CDCl₃, 250 MHz) δ 1.16 (d, 3 H, J = 6.4 Hz), 1.74–1.77 (m, 3 H), 3.37 (s, 3 H), 3.49 (dq, 1 H, J = 6.4 Hz, J = 4.5 Hz), 4.39 (d, 1 H, J = 4.5 Hz), 5.02–5.08 (m, 2 H), 9.04 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 14.6 (q), 18.6 (q), 55.7 (q), 76.7 (d), 90.6 (d), 114.5 (t), 141.3 (s); IR (cm⁻¹) 3580–3180, 1645. Anal. Calcd for C₇H₁₄O₃: C, 57.51; H, 9.65. Found: C, 56.82; H, 10.18.

Photooxygenation of 4-Methyl-3-penten-2-ol Acetate²⁵ (1e). From 498 mg (3.50 mmol) of 1e was obtained after 8 h a mixture of 2e and 3e. Chromatography (PE:ether, 80:20) gave 668 mg (58%) of 2e, of which 104 mg of (S^*,S^*) -2e and 176 mg of (S^*,R^*) -2e were isolated in diastereomerically pure form.

4-Acetoxy-2-methyl-1-penten-3-yl hydroperoxide $[(S^*, S^*)-2e]$: ¹H NMR (CDCl₃, 400 MHz) δ 1.21 (d, 3 H, J = 6.5 Hz), 1.82 (dd, 3 H, J = 1.5 Hz, J = 0.9 Hz), 2.10 (s, 3 H), 4.20 (d, 1 H, J = 8.1 Hz), 5.07-5.14 (m, 2 H), 5.15 (dq, 1 H, J = 8.1 Hz, J = 6.5 Hz), 9.44 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 16.5 (q), 18.1 (q), 20.7 (q), 68.8 (d), 90.7 (d), 116.3 (t), 140.8 (s), 171.0 (s); IR (cm⁻¹) 3620-3260, 1740, 1650. Anal. Calcd for C₈H₁₄O₄: C, 55.16; H, 8.10. Found: C, 55.54; H, 8.23. (S^{*}, R^{*})-2e: ¹H NMR (CDCl₃, 400 MHz) δ 1.23 (d, 3 H, J =3.7 Hz, J = 0.5 Hz), 5.00-5.02 (m, 1 H), 5.10-5.12 (m, 1 H), 5.38 (dq, 1 H, J = 6.6 Hz, J = 3.7 Hz), 9.44 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 15.5 (q), 18.8 (q), 20.9 (q), 68.5 (d), 89.9 (d), 116.1 (t), 140.0 (s), 171.5 (s); IR (cm⁻¹) 3600-3400, 1720, 1640. Anal. Calcd for C₈H₁₄O₄: C, 55.16; H, 8.10. Found: C, 55.47; H, 8.30.

(*E*)-4-Hydroperoxy-4-methyl-2-penten-2-olacetate[(*E*)-3e]: ¹H NMR (CDCl₃, 400 MHz) δ 1.42 (s, 6 H), 2.04 (d, 3 H, *J* = 1.1 Hz), 2.13 (s, 3 H), 5.20 (d, 1 H, *J* = 1.1 Hz), 9.44 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 20.2 (q), 22.2 (q), 25.7 (q), 80.2 (s), 121.9 (d), 148.2 (s), 170.2 (s).

(Z)-4-Hydroperoxy-4-methyl-2-penten-2-ol acetate [(Z)-3e)]: ¹H NMR (CDCl₃, 400 MHz) δ 1.36 (s, 6 H), 1.90 (d, 3 H, J = 1.1 Hz), 2.31 (s, 3 H), 5.11 (d, 1 H, J = 1.1 Hz), 9.44 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 20.0 (q), 20.7 (q), 25.1 (q), 80.7 (s), 120.3 (d), 146.8 (s), 171.2 (s).

The labile enol acetates 3e could not be isolated and purified by chromatographic methods, so that the NMR data were extracted from the crude reaction mixture by comparison with those of the isolated and fully characterized hydroperoxides 2e. **Photooxygenation of (Z)-3-Penten-2-ol**²⁶ (1f). From 440 mg (5.11 mmol) of 1f was obtained after 6 h and chromatography (PE:ether, 80: 20) 443 mg (73%) of (Z)-2f.

3-Hydroperoxy-4-penten-2-ol $[(S^*, S^*)-1f]$: ¹H NMR (CDCl₃, 250 MHz) δ 1.16 (d, 3 H, J = 6.4 Hz), 2.92 (br s, 1 H), 3.86 (dq, 1 H, J = 7.9 Hz, J = 6.4 Hz), 4.19 (dd, 1 H, J = 7.9 Hz, J = 7.8 Hz), 5.38 (dd, 1 H, J = 10.3 Hz, J = 1.6 Hz), 5.42 (dd, 1 H, J = 17.4 Hz, J = 1.6 Hz), 5.78 (ddd, 1 H, J = 17.4 Hz, J = 10.3 Hz, J = 1.6 Hz), 5.42 (dd, 1 H, J = 7.8 Hz), 9.00 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 18.7 (q), 68.0 (d), 91.6 (d), 120.9 (t), 133.5 (d). (S*, R*)-1f: ¹H NMR (CDCl₃, 250 MHz) δ 4.20 (dd, 1 H, J = 6.6 Hz, J = 2.9 Hz), 4.30 (dd, 1 H, J = 7.7 Hz, J = 2.9 Hz), the signals are partially overlapped by those of the S*, S* diastereomer, and therefore, only the separated resonances are given; ¹³C NMR (CDCl₃, 63 MHz) δ 20.7 (q), 67.1 (d), 90.0 (d), 121.6 (t), 131.5 (d); IR (cm⁻¹) 3600-3040, 1635. Anal. Calcd for C₅H₁₀O₃: C, 50.84; H, 8.53. Found: C, 50.43; H, 8.80.

Photooxygenation of (E)-3-Penten-2-ol²⁷[(E)-1f)]. Photooxygenation (20 h) of a solution of 105 mg (1.22 mmol) of (E)-1f in 20 mL of CCl₄ gave in addition to 2f also 4% of 4f (1:1 mixture of both diastereomers).

3-Hydroxy-3,5-dimethyl-1,2-dioxolane (4f): ¹H NMR (CDCl₃, 250 MHz) δ 2.15 (dd, 1 H, J = 12.6 Hz, J = 7.5 Hz), 2.25 (dd, 1 H, J = 12.7 Hz, J = 8.3 Hz), 2.71 (dd, 1 H, J = 12.7 Hz, J = 7.5 Hz), 2.84 (dd, 1 H, J = 12.6 Hz, J = 7.0 Hz), the signals are partially overlapped by those of other compounds, and therefore, only the separated resonances ae given; ¹³C NMR (CDCl₃, 63 MHz) δ 20.4 (q), 20.9 (q), 22.9 (q), 23.4 (q), 53.7 (t), 53.9 (t), 77.2 (d), 78.5 (d), 105.1 (s), 106.3 (s).

The dioxolanes 4f could not be isolated due to too low concentration. The NMR data were extracted with the crude reaction mixture by comparison with those of the isolated and fully characterized hydroperoxides 2f. Moreover, the spectral characteristics were similar to those observed for dioxolane 4a.

Photooxygenation of 3-Methyl-3-penten-2-ol^{15d} (1g). The conversion (5 h) of 501 mg (5.00 mmol) of 1g (Z:E = 85:15) afforded a mixture of 87% of 2g [$(S^*, S^*):(S^*, R^*) = 88:12$] and 13% of 5g [$(S^*, S^*):(S^*, R^*) = 31:69$]. By chromatographic purification (PE:ether, 80:20) 380 mg (58%) of 2g and 60 mg (9%) of 5g were isolated. The photooxygenation (5 h) of 200 mg (2.00 mmol) of (Z)-1g in 40 mL of CCl₄ gave the crude product, which consisted of 92% of 2g [$(S^*, S^*):(S^*, R^*) = 93:7$] and 8% of 5g [$(S^*, S^*):(S^*, R^*) = 23:77$]. When the reaction was repeated with 202 mg (2.02 mmol) of 1g (Z:E = 36:64), a mixture of 68% of 2g [$(S^*, S^*):(S^*, R^*) = 76:24$] and 32% of 5g [$(S^*, S^*):(S^*, R^*) = 49:51$] was obtained. From these product ratios the product distribution for the conversion of (E)-1g was extrapolated.

3-Hydroperoxy-3-methyl-4-penten-2-ol [(S^*, S^*)-2g]: ¹H NMR (CDCl₃, 40 MHz) δ 1.14 (d, 3 H, J = 6.5 Hz), 1.28 (s, 3 H), 2.55 (br s, 1 H), 4.02 (q, 1 H, J = 6.5 Hz), 5.35 (dd, 1 H, J = 11.0 Hz, J = 1.1 Hz), 5.38 (dd, 1 H, J = 17.7 Hz, J = 1.1 Hz), 5.92 (dd, 1 H, J = 17.7 Hz, J =11.0 Hz), 8.15 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 15.0 (q), 17.3 (q), 70.1 (d), 87.4 (s), 117.6 (t), 138.2 (d). (S^*, R^*)-2g: ¹H NMR (CDCl₃, 400 MHz) δ 1.14 (d, 3 H, J = 6.6 Hz), 1.34 (s, 3 H), 2.55 (br s, 1 H), 4.01 (q, 1 H, J = 6.6 Hz), 5.36 (dd, 1 H, J = 17.7 Hz, J = 1.1 Hz), 5.39 (dd, 1 H, J = 11.2 Hz, J = 1.1 Hz), 5.99 (dd, 1 H, J = 17.7 Hz, J =11.2 Hz), 8.15 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 17.4 (q), 18.5 (q), 71.1 (d), 87.4 (s), 118.7 (t), 135.9 (d); IR (cm⁻¹) 3700–3050, 1650. Anal. Calcd for C₆H₁₂O₃: C, 54.53; H, 9.15. Found: C, 54.58; H, 9.24.

4-Hydroperoxy-3-methylenepentan-2-ol [(S^*, S^*)-**5g**]: ¹H NMR (CDCl₃, 400 MHz) δ 1.39 (d, 3 H, J = 6.6 Hz), 1.40 (d, 3 H, J = 6.5 Hz), 2.83 (br s, 1 H), 4.44 (dq, 1 H, J = 6.5 Hz, J = 0.9 Hz), 4.64 (dq, 1 H, J = 6.6 Hz, J = 0.8 Hz), 5.22 (m, 1 H), 5.32 (m, 1 H), 9.18 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 19.1 (q), 22.4 (q), 69.3 (d), 81.8 (d), 112.8 (t), 152.1 (s). (S^*, R^*)-**5g**: ¹H NMR (CDCl₃, 400 MHz) δ 1.30 (d, 3 H, J = 6.6 Hz), 1.40 (d, 3 H, J = 6.5 Hz), 2.83 (br s, 1 H), 4.50 (dq, 1 H, J = 6.5 Hz, J = 0.7 Hz), 4.72 (q, 1 H, J = 6.6 Hz), 5.22 (m, 1 H), 5.32 (m, 1 H), 9.18 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 18.4 (q), 22.1 (q), 67.1 (d), 82.6 (d), 114.5 (t), 151.4 (s); IR (cm⁻¹) 3700–3080, 1630. Anal. Calcd for C₆H₁₂O₃: C, 54.53; H, 9.15. Found: C, 53.95; H, 9.43.

Photooxygenation of (Z)**-4-Hexen-3-ol**²⁸ [(Z)**-1h**]. From 346 mg (3.45 mmol) of Z-1h was obtained after 5.5 h and chromatography (PE: ether, 50:50) 392 mg (85%) of 2h.

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4-Hydroperoxy-5-hexen-3-ol [(S^*, S^*)-**2h**]: ¹H NMR (CDCl₃, 250 MHz) δ 0.96 (t, 3 H, J = 7.4 Hz), 1.28–1.64 (m, 2 H), 2.80 (br s, 1 H), 3.61 (dt, 1 H, J = 7.9 Hz, J = 3.5 Hz), 4.21 (dd, 1 H, J = 7.9 Hz, J = 7.8 Hz), 5.38 (dd, 1 H, J = 10.3 Hz, J = 1.6 Hz), 5.42 (dd, 1 H, J = 17.4 Hz, J = 1.6 Hz), 5.81 (ddd, 1 H, J = 17.4 Hz, J = 10.3 Hz, J = 7.8 Hz), 8.97 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 9.5 (q), 25.5 (t), 73.1 (d), 90.2 (d), 120.8 (t), 133.5 (d). (S^*, R^*)-**2h**: ¹H NMR (CDCl₃, 14, J = 7.8 Hz), δ 3.93 (dt, 1 H, J = 6.8 Hz, J = 2.8 Hz), 4.36 (dd, 1 H, J = 7.8 Hz), 4.36 (dd, 1 H, J = 6.8 Hz, J = 2.8 Hz), 4.36 (dd, 1 H, J = 6.8 Hz, J = 2.8 Hz), 4.36 (dd, 1 H, J = 7.8 Hz, J = 2.8 Hz), the signals are partially overlapped by those of the S^*, S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 63 MHz) δ 10.2 (q), 25.1 (t), 72.5 (d), 89.2 (d), 121.6 (t), 131.4 (d); IR (cm⁻¹) 3600–3040, 1635. Anal. Calcd for C₆H₁₂O₃: C, 54.53; H, 9.15. Found: C, 54.42; H, 9.51.

Photooxygenation of (Z)-2-Methyl-4-hexen-3-ol²⁹[(Z)-1i)]. From 245 mg (1.74 mmol) of (Z)-1i (40 mL of CCl₄) was obtained after 5.5 h and chromatography (PE:ether, 50:50) 202 mg (79%) of 2i.

4-Hydroperoxy-2-methyl-5-hexen-3-ol[(S^*, S^*) -2i]: ¹H NMR (CDCl₃, 250 MHz) δ 0.89 (d, 3 H, J = 6.8 Hz), 1.00 (d, 3 H, J = 6.9 Hz), 1.70–1.86 (m, 1 H), 2.44 (br s, 1 H), 3.48 (dd, 1 H, J = 7.8 Hz, J = 3.5 Hz), 4.32 (dd, 1 H, J = 7.9 Hz, J = 7.8 Hz), 5.39 (d, 1 H, J = 10.2 Hz), 5.47 (d, 1 H, J = 17.3 Hz), 5.82 (ddd, 1 H, J = 17.3 Hz, J = 10.2 Hz, J = 7.9 Hz), 8.76 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 15.0 (q), 19.9 (q), 29.2 (d), 76.0 (d), 88.5 (d), 120.5 (t), 133.6 (d). (S^*, R^*)-2i: ¹H NMR (CDCl₃, 250 MHz) δ 3.65 (dd, 1 H, J = 8.3 Hz, J = 3.1 Hz), 4.66 (dd, 1 H, J = 7.9 Hz, J = 3.1 Hz), the signals are partially overlapped by those of the S^*, S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 63 MHz) δ 14.8 (q), 18.6 (q), 30.0 (d), 75.6 (d), 87.6 (d), 121.7 (t), 131.1 (d); IR (cm⁻¹) 3660–3060, 1640. Anal. Caled for C₇H₁₄O₃: C, 57.51; H, 9.65. Found: C, 57.86; H, 9.87.

Photooxygenation of (Z)-2,2-Dimethyl-4-hexen-3-ol³⁰[(Z)-1j)]. After photooxygenation of a solution of 200 mg (1.56 mmol) of 1j and 35 mg (0.16 mmol) of 2,6-di-*tert*-butyl-4-methylphenol in 40 mL of CCl₄ for 7 h, 200 mg (80%) of 2j was isolated on chromatography (PE:ether, 80:20). In the absence of 2,6-di-*tert*-butyl-4-methylphenol, 2j is unstable under photooxygenation conditions; in this case 2j is converted to 6j.

2,2-Dimethyl-4-hydroperoxy-5-hexen-3-ol $[(S^*,S^*)-2j]$: ¹H NMR (CDCl₃, 250 MHz) δ 1.00 (s, 9 H), 2.46 (br s, 1 H), 3.37 (d, 1 H, J = 5.2 Hz), 4.43 (dd, 1 H, J = 7.8 Hz, J = 5.2 Hz), 5.40 (dd, 1 H, J = 10.3, J = 0.9 Hz), 5.43 (dd, 1 H, J = 17.4 Hz, J = 0.9 Hz), 6.03 (ddd, 1 H, J = 17.4 Hz, J = 10.3 Hz, J = 7.8 Hz), 9.14 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 26.6 (q), 34.7 (s), 80.2 (d), 86.8 (d), 119.9 (t), 135.6 (d). (S^*,R^*) -2j: ¹H NMR (CDCl₃, 250 MHz) δ 0.97 (s, 9 H), 3.55 (d, 1 H, J = 2.5 Hz), 4.48 (dd, 1 H, J = 8.2 Hz, J = 2.5 Hz), the signals are partially overlapped by those of the S^*,S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 63 MHz) δ 26.4 (q), 34.1 (s), 77.2 (d), 87.8 (d), 122.2 (t), 132.4 (d); IR (cm⁻¹) 3680–3160, 1640. Anal. Calcd for C₈H₁₆O₃: C, 59.98; H, 10.07. Found: C, 59.91; H, 10.08.

(*E*)-2,2-Dimethyl-6-hydroperoxy-4-hexen-3-ol (6j): ¹H NMR (CDCl₃, 250 MHz) δ 0.90 (s, 9 H), 2.46 (br s, 1 H), 3.82 (d, 1 H, J = 5.5 Hz), 4.51 (d, 2 H, J = 5.3 Hz), 5.77–5.96 (m, 2 H), 9.26 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 25.5 (q), 34.7 (s), 76.9 (t), 80.1 (d), 126.6 (d), 135.9 (d); IR (cm⁻¹) 3680–3120, 1640. Anal. Calcd for C₈H₁₆O₃: C, 59.98; H, 10.07. Found: C, 60.34; H, 9.73.

Photooxygenation of (Z)-3-Octen-2-ol³¹[(Z)-1k)]. From 256 mg (2.00 mmol) of (Z)-1k was obtained after 4 h and chromatography (PE:ether, 50:50) 224 mg (70%) of 2k.

3-Hydroperoxy-4-octen-2-ol[[**5***,**S***-(**E**)]-**2k**]: ¹H NMR (CDCl₃, 250 MHz) δ 0.91 (t, 3 H, J = 7.3 Hz), 1.16 (d, 1 H, J = 6.4 Hz), 1.44 (tq, 2 H, J = 7.5 Hz, J = 7.3 Hz), 2.02–2.15 (m, 2 H), 2.90 (br s, 1 H), 3.87 (dq, 1 H, J = 8.2 Hz, J = 6.4 Hz), 4.12 (dd, 1 H, J = 8.4 Hz, J = 8.2 Hz), 5.39 (ddt, 1 H, J = 15.5 Hz, J = 6.4 Hz), 4.12 (dd, 1 H, J = 8.4 Hz, J = 8.2 Hz), 5.39 (ddt, 1 H, J = 15.5 Hz, J = 8.4 Hz, J = 1.5 Hz), 5.84 (dt, 1 H, J = 15.5 Hz, J = 6.7 Hz), 8.83 (br s, 1 H); ¹³C NMR (CDCl₃, 63 MHz) δ 13.8 (q), 19.1 (q), 25.2 (t), 34.7 (t), 68.7 (d), 91.7 (d), 125.3 (d), 138.5 (d). [**S***,**R***-(**E**)]-**2k**: ¹H NMR (CDCl₃, 250 MHz) δ 4.18 (dq, 1 H, J = 6.6 Hz, J = 3.0 Hz), 4.27 (dd, 1 H, J = 8.3 Hz, J = 3.0 Hz), 5.49 (ddt, 1 H, J = 15.5 Hz, J = 8.3 Hz, J = 1.5 Hz), the signals are partially overlapped by those of the **S***,**S*** diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 63 MHz) δ 13.8 (q), 17.9 (q), 22.2 (t), 34.9 (t), 67.3 (d), 90.1 (d), 122.7 (t), 139.6 (d); IR

 (cm^{-1}) 3660–3080, 1665. Anal. Calcd for C₈H₁₆O₃: C, 59.98; H, 10.07. Found: C, 60.23; H, 10.40.

General Procedure for the Catalytic Hydrogenation of Allylic Hydroperoxides. A solution of the allylic hydroperoxide (0.45-1.6 mmol) in methanol (5-15 mL) was hydrogenated at room temperature and normal pressure of hydrogen gas with ca. 5 mg of PtO₂. The catalyst was removed by filtration and the solvent rotoevaporated (20 °C/18 Torr). In each case the dr values of the saturated diol and the corresponding allylic hydroperoxide were identical.

Hydrogenation of 3-Hydroperoxy-4-methyl-4-penten-2-ol (2a). From 215 mg (1.63 mmol) of 2a $[(S^*, S^*): (S^*, R^*) = 93:7]$ in 15 mL of methanol was isolated after 2 h 179 mg (93%) of 7a as colorless prisms, mp 49-52 °C (methanol).

4-Methylpentane-2,3-diol[(S^*,S^*) -7a]: ¹H NMR (CDCl₃, 250 MHz) δ 0.82 (d, 3 H, J = 6.8 Hz), 0.91 (d, 3 H, J = 6.8 Hz), 1.12 (d, 3 H, J= 6.4 Hz), 1.70 (dsept, 1 H, J = 6.8 Hz, J = 4.7 Hz), 3.02 (dd, 1 H, J = 6.2 Hz, J = 4.7 Hz), 3.41 (br s, 2 H), 3.69 (dq, 1 H, J = 6.4 Hz, J = 6.2 Hz); ¹³C NMR (CDCl₃, 63 MHz) δ 16.5 (q), 20.0 (q), 20.4 (q), 30.2 (d), 68.9 (d), 80.9 (d). (S^*,R^*)-7a: ¹H NMR (CDCl₃, 250 MHz) δ 0.79 (d, 3 H, J = 6.8 Hz), 0.94 (d, 3 H, J = 6.7 Hz), 1.07 (d, 3 H, J= 6.4 Hz), 1.53 (dqq, 1 H, J = 8.4 Hz, J = 6.8 Hz, J = 6.7 Hz), 3.20 (dd, 1 H, J = 8.4 Hz), ¹³C NMR (CDCl₃, 63 MHz) δ 16.1 (q), 19.1 (q), 19.7 (q), 30.7 (d), 68.7 (d), 80.4 (d); IR (cm⁻¹) 3660–3140.

The spectral data matched those reported in refs 14b,c,e,g.

Hydrogenation of 3-Hydroperoxy-4-penten-2-ol (2f). Conversion of 105 mg(0.889 mmol) of $2f[(S^*,S^*):(S^*,R^*) = 93:7]$ in 15 mL of methanol gave after 2 h 88 mg (95%) of 7f as a colorless oil.

Pentane-2,3-diol [(S^*, S^*)-7f]: ¹H NMR (CDCl₃, 250 MHz) δ 0.99 (t, 3 H, J = 7.4 Hz), 1.18 (d, 3 H, J = 6.3 Hz), 1.31–1.65 (m, 2 H), 2.70 (br s, 2 H), 3.26 (ddd, 1 H, J = 8.4 Hz, J = 6.4 Hz, J = 3.8 Hz), 3.60 (dq, 1 H, J = 6.4 Hz, J = 6.3 Hz); ¹³C NMR (CDCl₃, 50 MHz) δ 9.9 (q), 19.4 (q), 26.0 (t), 70.5 (d), 77.5 (d). (S^*, R^*)-7f: ¹H NMR (CDCl₃, 250 MHz) δ 1.14 (d, 3 H, J = 6.5 Hz), 3.80 (dq, 1 H, J = 6.5 Hz, J = 3.2 Hz), the signals are partially overlapped by those of the S^*, S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 50 MHz) δ 10.4 (q), 18.3 (q), 24.7 (t), 70.1 (d), 76.4 (d); IR (cm⁻¹) 3650–3040.

The spectral data matched those reported in ref 14j.

Hydrogenation of 3-Hydroperoxy-3-methyl-4-penten-2-ol (2g). From 167 mg (1.26 mmol) of 2g $[(S^*,S^*):(S^*,R^*) = 76:24]$ in 15 mL of methanol was obtained after 70 min 140 mg (94%) of 7g as a colorless oil.

3-Methylpentane-2,3-diol[(S^*, R^*)-7g]: ¹H NMR (CDCl₃, 250 MHz) δ 0.89 (t, 3 H, J = 7.5 Hz), 1.03 (s, 3 H), 1.09 (d, 3 H, J = 6.5 Hz), 1.28-1.64 (m, 2 H), 2.74 (br s, 2 H), 3.61 (q, 1 H, J = 6.5 Hz); ¹³C NMR (CDCl₃, 50 MHz) δ 7.8 (q), 17.6 (q), 20.1 (q), 31.7 (t), 72.8 (d), 75.4 (s). (S^*, R^*)-7f: ¹H NMR (CDCl₃, 250 MHz) δ 1.11 (d, 3 H, J = 6.4 Hz), 3.59 (q, 1 H, J = 6.4 Hz), the signals are partially overlapped by those of the S^*, S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 50 MHz) δ 7.8 (q), 17.5 (q), 22.9 (q), 28.4 (t), 74.1 (d), 75.1 (s); IR (cm⁻¹) 3640–3080.

The spectral data matched those reported in ref 14f.

Hydrogenation of 4-Hydroperoxy-5-hexen-3-ol (2h). From 112 mg (0.874 mmol) of 2h $[(S^*,S^*):(S^*,R^*) = 94:6]$ in 10 mL of methanol was obtained after 2 h 94 mg (94%) of 7h as a colorless oil.

Hexane-3,4-diol [(S^*,S^*) -7h]: ¹H NMR (CDCl₃, 250 MHz) $\delta 0.95$ (t, 6 H, J = 7.4 Hz), 1.33–1.64 (m, 4 H), 2.28 (br s, 2 H), 3.29–3.36 (m, 2 H); ¹³C NMR (CDCl₃, 50 MHz) $\delta 10.0$ (q), 26.3 (t), 75.4 (d). (S^*,R^*)-7h: ¹H NMR (CDCl₃, 250 MHz) $\delta 0.96$ (t, 6 H, J = 7.4 Hz), 3.50–3.56 (m, 2 H), the signals are partially overlapped by those of the S^*,S^* diastereomer, and only the separated resonances are given; ¹³C NMR (CDCl₃, 50 MHz) $\delta 10.4$ (q), 24.0 (t), 76.0 (d); IR (cm⁻¹) 3700–3060.

The spectral data matched those reported in ref 14g.

Hydrogenation of 3-Hydroperoxy-4-octen-2-ol (2k). The conversion of 72 mg (0.449 mmol) of 2k $[[S^*,S^{*-}(E)]:[S^*,R^{*-}(E)] = 94:6]$ in 5 mL of methanol gave after 1 h 63 mg (96%) of 7k as a colorless oil.

Octane-2,3-diol [(S^*, S^*) -7k]: ¹H NMR (CDCl₃, 250 MHz) $\delta 0.88$ (t, 3 H, J = 6.5 Hz), 1.17 (d, 3 H, J = 6.3 Hz), 1.20–1.52 (m, 8 H), 2.93 (br s, 2 H), 3.31 (dt, 1 H, J = 6.5 Hz, J = 6.3 Hz), 3.57 (quin, 1 H, J = 6.3 Hz); ¹³C NMR (CDCl₃, 50 MHz) δ 14.0 (q), 19.4 (q), 22.5 (t), 25.2 (t), 31.8 (t), 33.3 (t), 70.9 (d), 76.2 (d). (S*, R*)-7f: ¹H NMR (CDCl₃, 250 MHz) δ 1.13 (d, 3 H, J = 6.4 Hz), 3.77 (dq, 1 H, J = 6.4 Hz, J = 3.2 Hz), the signals are partially overlapped by those of the S*, S* diastereomer, and only the separated resonances are given; ¹³C

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NMR (CDCl₃, 50 MHz) δ 13.8 (q), 19.8 (q), 23.2 (t), 25.4 (t), 31.5 (t), 32.7 (t), 70.4 (d), 74.9 (d); IR (cm⁻¹) 3680–3220.

The spectral data matched those reported in refs 14b,h,i.

Hydrogenation of 4-Hydroperoxy-3-methylenepentan-2-ol (5g). From 60 mg (0.454 mmol) of 5g [(S^*, S^*) : $(S^*, R^*) = 68:32$] in 5 mL of methanol was obtained after 1 h 49 mg (91%) of 8g [(S^*, R^*, R^*) : (S^*, S^*, R^*) : $(S^*, S^*) = 49:20:31$] as a colorless oil.

3-Methylpentane-2,4-diol [(S^*, R^*, R^*)-8g]: ¹H NMR (CDCl₃, 250 MHz) δ 0.91 (d, 3 H, J = 6.1 Hz), 4.08 (dq, 2 H, J = 6.4 Hz, J = 2.2 Hz). (S^*, S^*, R^*)-8g: ¹H NMR (CDCl₃, 250 MHz) δ 0.76 (d, 3 H, J = 6.9 Hz), 3.73 (dq, 2 H, J = 8.5 Hz, J = 6.2 Hz). (S^*, S^*)-8g: ¹H NMR (CDCl₃, 250 MHz) δ 0.85 (d, 3 H, J = 6.1 Hz), 3.85 (dq, 1 H, J = 6.8 Hz, J = 6.3 Hz), 4.12 (dq, 1 H, J = 6.4 Hz, J = 3.1 Hz). The other observed resonances listed below could not unequivocally be assigned to one of the diastereomers: ¹H NMR (CDCl₃, 250 MHz) δ 1.14 (d, 3 H, J = 6.3 Hz), 1.17 (d, 3 H, J = 6.4 Hz), 1.19 (d, 3 H, J = 6.2 Hz), 1.21 (d, 3 H, J = 6.3 Hz), 1.28–1.48 (m, 3 H), 3.12 (br s, 6 H); ¹³C NMR (CDCl₃, 63 MHz) δ 4.0 (q), 12.5 (q), 13.4 (q), 19.1 (q), 21.6 (q), 22.2 (q), 22.3 (q), 43.6 (d), 44.6 (d), 47.3 (d), 70.0 (d), 71.1 (d), 72.9 (d), 73.5 (d); IR (cm⁻¹) 3640–3080.

The spectral data matched those reported in refs 14a,d.

Desilylation of 2b. To a solution of 261 mg (1.28 mmol) of **2b** [(S^*, S^*) : $(S^*, R^*) = 80:20$] in 5 mL of methanol was added 250 mg of silica gel, and the mixture was stirred at room temperature. After 9 h the silica gel was removed by filtration and the solvent removed (20 °C/18 Torr) to afford 160 mg (95%) of **2a** [$(S^*, S^*):(S^*, R^*) = 80:20$] as a colorless oil. This material was identical with that obtained by photooxygenation of **1a**.

Ph₃P Reduction of 2a. To a solution of 237 mg (1.79 mmol) of **2a** $[(S^*,S^*):(S^*,R^*) = 93:7]$ in 5 mL of CH₂Cl₂ was added while stirring at 0 °C a solution of 510 mg (1.94 mmol) of Ph₃P in 5 mL of CH₂Cl₂. The reaction mixture was warmed up to room temperature, stirring was continued for 30 min, and the solvent was rotoevaporated (20 °C/18 Torr). Purification of the residue by Kugelrohr distillation at 140 °C (18 Torr) yielded 168 mg (81%) of **9a** $[(S^*,S^*):(S^*,R^*) = 93:7]$ as a colorless oil.

4-Methyl-4-penten-2,3-ol[(S^*, S^*) -9a]: ¹H NMR (CDCl₃, 250 MHz) δ 1.08 (d, 3 H, J = 6.5 Hz), 1.68 (d, 3 H, J = 0.6 Hz), 3.13 (br s, 2 H), 3.67–3.78 (m, 2 H), 4.86–4.88 (m, 1 H), 4.95 (d, 1 H, J = 0.7 Hz); ¹³C NMR (CDCl₃, 63 MHz) δ 17.7 (q), 18.9 (q), 68.6 (d), 80.6 (d), 113.7 (t), 144.5 (s). (S^*, R^*)-9a: ¹H NMR (CDCl₃, 250 MHz) δ 1.07 (d, 3 H, J = 6.3 Hz), 1.71 (s, 3 H), 3.13 (br s, 2 H), 3.85 (dq, 1 H, J = 6.3 Hz, J = 4.5 Hz), 4.01 (d, 1 H, J = 4.5 Hz), 4.90 (s, 1 H), 4.99 (s, 1 H); ^{13}C NMR (CDCl₃, 63 MHz) δ 18.6 (q), 19.6 (q), 67.4 (d), 92.8 (d), 115.9 (t), 141.6 (s); IR (cm⁻¹) 3640–3000, 1650.

The spectral data matched those reported in ref 32.

LiAlH₄ Reduction of (S^*, R^*) -3e. To a suspension of 191 mg (5.02 mmol) of LiAlH₄ in 5 mL of dry ether was added a solution of 350 mg (2.01 mmol) of (S^*, R^*) -3e in 5 mL of ether. After 30 min were added in succession 0.2 mL of water, 0.2 mL of 15% aqueous NaOH, and 0.6 mL of water by means of a syringe (CAUTION! Vigorous evolution of hydrogen gas!). The precipitate was removed by filtration, the solvent rotoevaporated (20 °C/18 Torr), and the residue recrystallized from ether to afford 164 mg (70%) of (S^*, R^*) -9a as colorless needles, mp 47 °C (ref 33, 48-49 °C). The spectral data were identical with those above.

m-CPBA Epoxidation of (Z)-1g.¹¹ To a solution of 100 mg (1.00 mmol) of (Z)-1g in 20 mL of CH₂Cl₂ was added at 0 °C portionwise 207 mg (1.20 mmol) of *m*-CPBA. After 3 h, the solid matter was removed by filtration and the solvent removed (20 °C/18 Torr). The resulting colorless oil contained *threo*- and *erythro*-10 in a 90:10 ratio.

3,4-Epoxy-3-methylpentan-2-ol[(S^*, R^*, S^*) -10 (*threo*-10)]: ¹H NMR (CDCl₃, 250 MHz) δ 1.20 (d, 3 H, J = 6.7 Hz), 1.29 (s, 3 H), 1.30 (d, 3 H, J = 5.7 Hz), 2.02 (br s, 1 H), 2.96 (q, 1 H, J = 5.7 Hz), 3.69 (q, 1 H, J = 6.7 Hz); ¹³C NMR (CDCl₃, 63 MHz) δ 13.7 (q), 15.8 (q), 19.0 (q), 60.8 (d), 64.4 (s), 69.0 (d). [(S^*, S^*, R^*) -10 (*erythro*-10)]: ¹H NMR (CDCl₃, 250 MHz) δ 1.27 (s, 3 H), 1.27 (d, 3 H, J = 6.3 Hz), 1.42 (d, 3 H, J = 5.8 Hz), 1.83 (br s, 1 H), 2.95 (q, 1 H, J = 5.8 Hz), 3.77 (q, 1 H, J = 6.3 Hz); ¹³C NMR (CDCl₃, 63 MHz) δ 13.5 (q), 17.7 (q), 18.6 (q), 61.3 (d), 62.8 (s), 67.8 (d); IR (cm⁻¹) 3640–3240.

VO(acac)₂/t-BuOOH Epoxidation of (Z)-1g¹¹: A mixture of 0.48 mL of 3.1 M t-BuOOH (1.50 mmol) in CH₂Cl₂, 100 mg (1.00 mmol) of (Z)-1g, and 2.7 mg (1 mol %) of VO(acac)₂ in 20 mL of CH₂Cl₂ was stirred at 0 °C for 12 h. The solution was washed with 1 mL of saturated, aqueous Na₂SO₃ solution and water (2 × 2 mL) and dried over MgSO₄ and the solvent rotoevaporated (20 °C/18 Torr). The residual oil consisted of *threo*- and *erythro*-10 in a 33:67 ratio. The spectral data were identical with those above.

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