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STUDIES RELATED TO THE ABSOLUTE CONFIGURATION OF CYCLOCINAMIDE A: TOTAL SYNTHESIS OF 4(R),11(R)-CYCLOCINAMIDE A

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corresponding methyl ester 3 (R = Me)³ via a two-step sequence. Hydrolysis [1.0 N NaOH, MeOH] of the methyl ester followed by esterification⁴ with 9-fluorenylmethanol [DCC (N,N'-dicyclohexylcarbodiimide), CH₂Cl₂] provided fluorenylmethyl ester 3 (R = Fm) in 98% overall yield. The synthesis of N-BOC-O-benzoyl-(R)-isoserine 4 via a six-step sequence originated with (R)-malic acid employing the protocol of Milewska and Polonski⁵ developed for the conversion of (S)-malic acid into (S)-isoserine. (R)-Malic acid was converted [(CH₂O)_X, CHCl₃, TsOH, reflux] in quantitative yield into dioxolone 5 which was subsequently transformed via a Curtius reaction [(a) SOCl₂, reflux; (b) NaN₃, acetone, H₂O, -20 °C; (c) t-BuOH, reflux] into the N-BOC-dioxolone 6 in 41% overall yield. Cleavage [1.0 N NaOH, MeOH] of the dioxolone followed by benzoylation [BzCl, pyr, 0 °C] provided (71%) the (R)-isoserine fragment 4. The fully protected, activated (S)-asparagine residue 7 is commercially available.⁶ The (R)-2,3-diaminopropanoic acid fragment 8 was prepared from N_{\alpha}-BOC-(R)-asparagine via a Hoffman rearrangement.⁷ Thus, treatment of N_{\alpha}-BOC-(R)-asparagine with iodosobenzene diacetate in *n*-propanol:methyl acetate:water (8:5:1) [0 °C (1h) \rightarrow 50 °C (2h)] and subsequent exposure of the resulting amine to N-(9-fluorenylmethoxycarbonyloxy)succinimide⁸ in aqueous acetone afforded the fully protected (R)-2,3-diaminopropanoic acid moiety 8 in 75% overall yield.



With the requisite amino acid fragments in hand, assembly of the linear tetrapeptide 11 was undertaken. It was anticipated that tetrapeptide 11, needed for construction of the tetracyclic peptidic core 12 of 4(R), 11(R)-cyclocinamide A (2), could be assembled by conventional BOP [(1*H*-benzotriazol-1-yloxy)tris(dimethylamino)-phosphonium hexafluorophosphate] promoted coupling protocols.⁹ In an effort to minimize protecting group manipulations, the *O*-benzoyl-(*R*)-isoserine, derived from fragment 4, was coupled with the commercially available (*S*)-asparagine moiety 7.⁶ Thus, the *N*-BOC protected (*R*)-isoserine 4 was deprotected [30% TFA in CH₂Cl₂] and the resulting crude trifluoroacetate salt was treated [HOBt (1-hydroxybenzotriazole), DIEA (*N*,*N*-diisopropylethylamine), DMF] with the activated 4-nitrophenyl ester 7 of N_α-BOC-(*S*)-asparagine giving rise to dipeptide 9 as a single diastereomer in 65% overall yield. Removal [TMSI, CHCl₃, reflux] of the methyl carbamate protecting group in 3 provided the corresponding fluorenylmethyl ester of (*S*)-5-bromotryptophan which, upon coupling [BOP, HOBt, DIEA, DMF] with dipeptide 9, afforded tripeptide 10 in 57% overall yield. Cleavage [30% TFA in CH₂Cl₂] of the *N*-BOC group in tripeptide 10 gave rise to the corresponding trifluoroacetate salt which was dissolved in DMF and coupled [BOP, HOBT, DIEA] with the differentially protected (*R*)-2,3-diaminopropanoic acid 8 giving rise to tetrapeptide 11 in 76% yield.



With the linear tetrapeptide 11 in hand, the ring closure reaction to form the core of 4(R), 11(R)-cyclocinamide A (2) was examined. Simultaneous deprotection [20% TEA in CH₂Cl₂, DMF, 2h] of the termini of tetrapeptide 11 provided, after evaporation of the volatile components *in vacuo*, a solid residue which, upon dissolution in DMF and treatment with N,N-diisopropylethylamine and pentafluorophenyldiphenylphosphinate, ¹⁰ afforded in 62% overall yield the cyclized tetrapeptide core 12 of 2.



Prior to completing the total synthesis of 2, a synthesis of the glycine derived fragment 14 needed to be developed. This was accomplished via a two-step protocol. Condensation of methyl glycinate with 1-methyl-2-trichloroacetyl-4-chloropyrrole $(13)^{11}$ and subsequent hydrolysis of the methyl ester provided dipeptide fragment 14 in 90% overall yield.



Completion of the total synthesis of 4(R),11(R)-cyclocinamide A (2) necessitated cleavage of the BOC group on the nitrogen at C(11) in 12 followed by coupling with the dipeptide side chain 14. Thus, cleavage (30% TFA in CH₂Cl₂) of the BOC group on the cyclic tetrapeptide core 12 provided the crude trifluoroacetate salt which was dissolved in *N*,*N*-dimethylformamide and coupled [BOP, HOBT, DIEA] with dipeptide 14. Deprotection of the C(4) hydroxyl group in the coupled material employing 1% sodium hydroxide in methanol gave rise to 4(R),11(R)-cyclocinamide A (2) in 41% overall yield. A comparison of the ¹H NMR spectrum (500 MHz, MeOH-

d4) of 2 with that of natural cyclocinamide A provided by Professor Crews revealed that the spectra were similar, but not identical. Biogenetic considerations would suggest that the absolute configuration at C(4) and C(11) are both S. Efforts are underway to prepare the remaining diastereomers [4(S),7(S),11(S),14(S);4(S),7(S),11(R),14(S) and 4(R),7(S),11(S),14(S)] of cyclocinamide A.

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