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SAR Studies of 3-Cyclopropanecarbonyloxy-2-cyclohexen-1-one as Inhibitors of 4-Hydroxyphenylpyruvate Dioxygenase

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Abstract—Various 3-cyclopropanecarbonyloxy-2-cyclohexen-1-one 1 derivatives have been synthesized and tested as inhibitors of 4-hydroxyphenylpyruvate dioxygenase (4-HPPD) from pig liver. The inhibition results indicated that well-positioned dicarbonyl groups as well as the cyclopropyl group of 1 were essential for potent inhibition. Substitution at the 2-position of the ring system has a significant effect on inhibitor potency, while the 5-position can undergo substantial variations and retain inhibitor potency. In the compounds examined, 2-chloro substituted 12 is the best inhibitor of all with IC₅₀ of 15 nM, the rest of the synthesized analogues were less potent inhibitors than the parent compound. © 2002 Elsevier Science Ltd. All rights reserved.

Introduction

4-Hydroxyphenylpyruvate dioxygenase (4-HPPD)¹ is an important enzyme involved in the catabolism of tyrosine and phenylalanine in most organisms which catalyzes the conversion of 4-hydroxyphenylpyruvate to homogentisate. The biotransformations catalyzed by 4-HPPD require an oxidative decarboxylation of the 2-oxoacid side chain of the substrate with an accompanying hydroxylation of the aromatic ring and a 1,2-migration of the carboxymethyl group.² In the past 10 years, 4-HPPD has been the target site to develop effective drug therapy for the fatal hereditary disease tyrosinemia type I³ which is characterized by a deficiency of fumarylacetoacetase activity that leads to the accumulation of hepatotoxic and nephrotoxic compounds like succinylacetone.⁴ Inhibition of liver 4-HPPD activity will prevent the formation of homogentisate, thereby blocking catabolism. Currently, 2-(2-(nitro-4-trityrosine fluoromethylbenzoyl)-cyclohexane-1,3-dione⁵ (NTBC), a potent 4-HPPD inhibitor with a triketone type structure, has been on the clinical trial as the first effective drug therapy to treat tyrosinemia type I patients.



NTBC

Recently, we reported⁶ the discovery of a new potent, low molecular weight, non-triketone type 4-HPPD inhibitor 3-cyclopropanecarbonyloxy-2-cyclohexen-1-one **1**. Inhibition studies indicated that compound **1** was a potent 4-HPPD inhibitor with $IC_{50} = 30 \text{ nM}$, which is comparable to that of NTBC ($IC_{50} = 40 \text{ nM}$). Here we report the structure–activity relationships of the parent compound **1** as inhibitors of 4-HPPD in an effort to provide insights into its mode of action at the molecular level.



Results and Discussion

The molecules designed and synthesized for SAR are compounds 2--16, shown in Figure 1, Scheme 1 and 3. Vinyl ether 2 was prepared by acid-catalyzed dehydration of cyclohexanedione and cyclopropanemethanol in benzene at 140 °C for 6 h. Ester 3 was synthesized in two steps by first converting cyclohexanone to the corresponding

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Scheme 1. Synthesis of compounds **2** and **3**: (i) *p*-TsOH, benzene, reflux; (ii) Et₃N, trimethylsilyl chloride, DMF, 120° C; (iii) potassium *t*-butoxide, cyclopropanecarbonyl chloride, THF, -20° C.



Scheme 2. Preparation of compounds 17-21.

trimethylsilyl enol ether,⁷ followed by treatment with cyclopropanecarbonyl chloride in the presence of potassium *t*-butoxide in THF at -20 °C with 63% overall yield. Compounds **4–7** were easily prepared in one step by a standard coupling reaction of cyclopropanecarbonyl chloride in the presence of triethylamine as a base in methylene chloride with 5,5-dimethylcyclohexane-1,3-dione, cyclopentane-1,3-dione, resorcinol, and 4-hydroxy-coumarin, respectively. Preparation of compounds **8–13**, however, called for first preparing the appropriate cyclohexane-1,3-dione derivatives **17–21**



Figure 1. Various compounds synthesized for SAR studies.

shown in Scheme 2 according to the literature procedures,⁸ followed by the same coupling reaction mentioned above. Base-catalyzed self-condensation of the commercially available diethyl acetonylsuccinate afforded 5-ethoxycarbonyl-cyclohexane-1,3-dione,⁹ which upon treatment with cyclopropanecarbonyl chloride under basic conditions gave rise to compound 14 with 57% combined yield. The synthesis of compounds 14 and 15 was started by first introducing an aldehyde functional group onto 1,3,5-trimethoxybenzene via the Vilsmeier¹⁰ reaction as illustrated in Scheme 3. The resulting aldehydes were then oxidized under Baeyer-Villiger¹¹ conditions to produce phenols 23a and 23b. After coupling with cyclopropanecarbonyl chloride as described above, the esters 24a and 24b were exposed to silver oxide and aqueous 6N nitric acid in acetone to afford the final benzoquinones 15 and 16 with 48 and 35% overall yield, respectively.

All compounds described herein were tested for their ability to inhibit the activity of 4-hydroxyphenylpyruvate dioxygenase using the spectrophotometric enol borate method described by Lindstedt and Rundgren.¹² Results of the inhibition reactions of these compounds with pig liver 4-HPPD are summarized in Table 1. The importance of the position of the two carbonyl groups of 1 was investigated by comparing analogues 2 and 3. When the ring carbonyl group of 1 was removed to form 3, the inhibition potency decreased 23-fold relative to 1 indicating that the ring carbonyl group is essential for activity. In addition, replacement of the cyclic α , β unsaturated keto moiety of 1 with a phenol derivative 7 also resulted in significant losses in potency. In the case in which the ester carbonyl group of 1 was removed to form 2, no enzyme inhibition was observed up to the concentration of 0.2 mM. This result suggests the carbonyl oxygen atom on the ester functionality is crucial for binding, presumably by chelating with ferric ion in the enzyme active site.¹³ The role played by the substitution on the six-membered ring of 1 toward 4-HPPD inhibition was explored by investigation of the inhibition activity of compounds 4-16. Contraction of the sixmembered ring to the five-membered ring of 1 to 5, fusion a benzene ring to the 4- and 5-position of the ring system to a 4-hydroxycoumarin derivative 6, as well as the dimethyl and ethoxycarbonyl substitutions at 5position of the ring system of 1 to compounds 4 and 14

Table 1. Inhibition constants for reactions of 1–16 with 4-HPPD from pig liver by the enol borate method

Compound	$IC_{50} \ (\mu M)^a$	Compound	IC ₅₀ (µM)
1	0.03 (7.52) ^b	9	5.6 (-0.75)
2	NAc	10	135(-2.13)
3	0.7 (6.15)	11	20 (-1.30)
4	0.33 (6.48)	12	0.015 (7.82)
5	0.07 (7.15)	13	0.028 (7.55)
6	0.11 (6.96)	14	0.1 (7.00)
7	NA	15	25 (-1.40)
8	92 (-1.96)	16	30 (-1.48)

^aAssay detail is described in Experimental. Values are the means of at least three independent determinations.

^bpIC₅₀ unit is given in parentheses.

^cNA, not active; IC_{50}> 200 $\mu M.$



Scheme 3. Preparation of compounds 15 and 16: (i) POCl₃, DMF; (ii) H_2O_2 , H_2SO_4 , MeOH; (iii) cyclopropanecarbonyl chloride, Et_3N , CH_2Cl_2 ; (iv) AgO, 6 N HNO₃, acetone.

all have moderate effects on inhibition potency, but modifications at the 2-position of the ring system gave more variable results. When the 2-substituents were a chlorine or bromine atom, the resulting 12 or 13 were as potent as 1, whereas bulkier and less electronegative substituents like methyl (9) or aryl groups (10 and 11) led to a major reduction of activity. These results implied that both the steric and electrostatic effects of these substituents at the 2-position of the ring system might promote a conformational change to give a lower binding affinity for the 4-HPPD enzyme. Furthermore, the observation of 3-order of magnitude reduction of inhibition potency when the ester bond of 1 was replaced with the amide functionality might be attributed to the resulting conformational-constrained structure 8 that prevents tight binding with the enzyme active site.

Analogue 15 explored the effect of an additional oxo group at C-4 position when the six-membered ring was replaced by a benzoquinone moiety. In this case, inhibition potency of 15 decreased 833-fold (IC₅₀ = $25 \,\mu$ M) as compared to 1, and 227-fold as compared to 6. The results indicate that incorporation of a small but strongly electronegative atom like oxygen at 4-position of the ring system of 1 is highly detrimental to the inhibition activity, whereas incorporation of a large, planar, but less electronegative aromatic moiety like benzene at 4- and 5-positions of the ring (6) is not. We reasoned that the presence of a C-4 oxo group in the ring system might interfere with either of the two carbonyl groups in the molecule to chelate with the active site ferric ion and thus result in a weak affinity with the enzyme 4-HPPD, although more investigation is needed to confirm this assumption. Finally, further incorporation of a methoxy substituent on the 6-position of the benzoquinone moiety (16) resulted in little improvement on inhibition potency.

Conclusion

As a result of this investigation, we now have a general understanding of the SAR for this class of 4-hydroxyphenylpyruvate dioxygenase inhibitors, as depicted in Figure 2. The two carbonyl groups as well as the cyclo-



Figure 2. General SAR for 3-cyclopropanecarbonyloxy-2-cyclohexen-1-one inhibitors.

propyl group of 3-cyclopropanecarbonyloxy-2-cyclohexen-1-one were required for potent 4-HPPD inhibition. Substitution at the 2-position of ring system has a significant effect on inhibitor potency, while the 5position can undergo substantial variations and retain inhibitor potency. The ability to vary the 5-position of compound **1** should become important as this class of 4-HPPD inhibitors undergo further development, as it may be possible to vary the in vivo profile of these inhibitors without affecting inhibitor potency. The results of our studies will provide an alternate treatment for the fatal disease tyrosinemia type I.

Experimental

General

Melting points were determined on a Mel-Temp melting point apparatus in open capillaries and are uncorrected. Ultraviolet-visible spectroscopy was performed on a Hewlett-Packard 8453 spectrophotometer. High resolution FAB-MS was measured with a JEOL JMS-SX/SX 102A spectrometer. ¹H NMR spectra were recorded at 300 MHz on a Varian VXR300 spectrometer. Chemical shifts were reported in ppm on the δ scale relative to internal standard (tetramethylsilane, or appropriate solvent peaks) with coupling constants given in hertz. ¹H NMR multiplicity data were denoted by s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and b (broad). Flash chromatography was performed in columns of various diameters with Merck silica gel (230-400 mesh ASTM 9385 kieselgel 60H) by elution with the solvents reported. Analytical thin-layer chromatography (TLC) was carried out on Merck silica gel 60F-254 plates (0.2 mm) and developed with the solvents mentioned. All new compounds exhibited satisfactory spectroscopic and analytical data.

Enzyme purification and assay

4-HPPD was purified from pig liver by the method of Hamilton¹⁴ with a specific activity of 0.1 μ mol min⁻¹ mg⁻¹. The protein concentration was determined by BCA method. Routine assay of the enzyme utilized the spectrophotometric enol borate method of Lindstedt and Rundgren.¹² The typical assay mixtures contained 0.85 mL potassium phosphate/borate buffer (prepared by adjusting the pH of 0.42 M H₃BO₃ to 6.2 with a 0.17 M Na₃PO₄ solution), 0.06 mL 4-hydroxyphenyl-

pyruvic acid (1.8 mM, in 0.2 M Na₃PO₄ buffer), 0.03 mL dichlorophenolindophenol (reduced form, prepared by mixing 1 mL of 3.3 mM sodium dichlorophenolindophenol in H₂O and 0.16 M glutathione in 0.2 M sodium phosphate buffer), 0.01 mL phenylpyruvate tautomerase (10 U/mL, Sigma). The above solution was equilibrated for about 15 min (monitored at 308 nm), then 4-HPPD to be tested was added (0.05 mL solution). For calculation of dioxygenase activity the change in absorbance between the 8th and 10th min was used. The inhibition reaction of 1-16 with the enzyme 4-HPPD was evaluated by measuring the decrease in absorbance at 308 nm over a 15 min period following coadministration of varying concentration of the inhibitors and 4-HPPD. The IC₅₀ values were determined by fitting the data to the equation $v_i = v_o/v_i$ $(1 + [inhibitor]/IC_{50})$, where v_i is the rate of absorbance change at a given inhibitor concentration and v_o the rate of absorbance change in the absence of inhibitor, and converted into pIC_{50} units for modeling SAR.

3-Cvclopropvlmethoxv-2-cvclohexen-1-one (2). To a solution of 1,3-cyclohexanedione (166 mg, 1.48 mmol) in benzene (30 mL) was added cyclopropanemethanol (107 mg, 1.48 mmol) and catalytic amounts of *p*-toluenesulfonic acid. The resulting mixture was heated to 140 °C. The by-product water was removed by the Dean–Stark trap. The reaction was completed within 6h, then quenched with water. The vinyl ether was extracted with ethyl acetate, dried in MgSO₄, and concentrated in vacuo to obtain the crude product, which was further purified by column chromatography to give a colorless liquid with a 90% yield. $R_f = 0.54$ (40% EtOAc/hexanes). ¹H NMR $(CDCl_3)$ δ 5.17 (s, 1H, 2-H), 3.54 (d, J=6.9 Hz, 2H, OCH₂), 2.32–2.29 (m, 2H, 6-H), 2.23–2.20 (m, 2H, 4-H), 1.87-1.83 (m, 2H, 5-H), 1.10-1.02 (m, 1H, OCH₂CH), 0.52–0.47 (m, 2H, CH₂), 0.20–0.17 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₄O₂ 166.0994, found 166.0989.

2-Cyclopropanecarbonyloxy-1-cyclohexene (3). A solution of potassium t-butoxide (560 mg, 5 mmol) in THF (5 mL) was added to a stirred solution of 1-cyclohexenyloxy-trimethylsilane7 (850 mg, 5 mmol) in THF (10 mL) at $-78 \,^{\circ}$ C under nitrogen and the mixture was stirred at this temperature for 1 h. After this, the temperature was increased to -20 °C, then a solution of cyclopropanecarbonyl chloride (523 mg, 5 mmol) in THF (5 mL) was added to it. After 15 min, the mixture was quenched with saturated aqueous NaHCO3 and extracted with Et₂O. The extract was dried and evaporated under reduced pressure and the crude product was purified by flash chromatography on silica gel to give a colorless liquid with a 70% yield. $R_f = 0.25$ (100% hexanes). ¹H NMR (CDCl₃) δ 6.59 (t, J = 1.9 Hz, 1H, 2-H), 2.57-2.49 (m, 4H, 3-H, 6-H), 2.09-2.03 (m, 2H, 4-H), 1.80-1.75 (m, 1H, CH), 1.65-1.50 (m, 2H, 5-H), 1.14-1.11 (m, 2H, CH₂), 1.00–0.97 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₄O₂ 166.0994, found 166.0985.

General procedure for preparation of compounds 4–16, 24a, and 24b

The cyclopropanecarbonyl chloride (523 mg, 5 mmol) was added to a solution of 5,5-dimethylcyclohexane-1,3-

dione (700 mg, 5 mmol) with triethylamine (1.4 mL, 10 mmol) in dry methylene chloride (30 mL). The mixture was stirred at room temperature for 1 h, then washed with water, diluted hydrochloric acid, saturated aqueous sodium hydrogen carbonate, and water. The organic phase was dried with MgSO₄ and evaporated to dryness in vacuo to give the crude enol ester, which was further purified by flash chromatography to give the desired product.

3-Cyclopropanecarbonyloxy-5,5-dimethyl-2-cyclohexen-1 - **one (4).** This compound was obtained as a colorless liquid with a 87% yield. R_f =0.30 (25% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 5.91 (s, 1H, 2-H), 2.42 (s, 2H, 6-H), 2.27 (s, 2H, 4-H), 1.75–1.68 (m, 1H, CH), 1.13–1.10 (m, 2H, CH₂), 1,11 (s, 6H, 5-Me), 1.04–1.00 (m, 2H, CH₂). HRMS: calcd for C₁₂H₁₆O₃ 208.1100, found 208.1092.

3-Cyclopropanecarbonyloxy-2-cyclopenten-1-one (5)

This compound was obtained as a colorless liquid with a 70% yield. R_f =0.17 (50% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.20 (s, 1H, 2-H), 2.79–2.75 (m, 2H, 5-H), 2.47–2.43 (m, 2H, 4-H), 1.81–1.76 (m, 1H, CH), 1.19–1.16 (m, 2H, CH₂), 1.11–1.06 (m, 2H, CH₂). HRMS: calcd for C₉H₁₀O₃ 166.0630, found 166.0635.

4-Cyclopropanecarbonyloxy-coumarin (6). This compound was obtained as a brown liquid with a 75% yield. $R_f = 0.39$ (50% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 7.66 (d, J = 7.8 Hz, 1H, Ar H), 7.59 (t, J = 8.2 Hz, 1H, Ar H), 7.38–7.28 (m, 2H, Ar H's), 6.51 (s, 1H, 3-H), 1.99–1.59 (m, 1H, CH), 1.29–1.19 (m, 2H, CH₂), 1.18–1.15 (m, 2H, CH₂). HRMS: calcd for C₁₃H₁₀O₄ 230.0579, found 230.0570.

3-Cyclopropanecarbonyloxy-phenol (7). This compound was obtained as a light blue liquid with a 92% yield. $R_f = 0.44$ (25% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 7.18 (t, J = 8.4 Hz, 1H, Ar H), 6.68–6.62 (m, 2H, Ar H's), 6.57 (s, 1H, Ar H), 5.90 (bs, 1H, OH), 1.85–1.80 (m, 1H, CH), 1.17–1.14 (m, 2H, CH₂), 1.05–1.00 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₀O₃ 178.0630, found 178.0636.

N-(3-Oxo-cyclohex-1-enyl)-cyclopropanecarboxamide (8). This compound was obtained from 3-amino-2-cyclohexen-1-one¹⁵ **17** as a brown solid with a 65% yield. Mp 177–178 °C. R_f =0.40 (100% EtOAc). ¹H NMR (CDCl₃) δ 7.18 (bs, 1H, NH), 6.48 (s, 1H, 2-H), 2.62 (t, J=6.0 Hz, 2H, 6-H), 2.38 (t, J=6.3 Hz, 2H, 4-H), 2.21– 2.10 (m, 2H, 5-H), 1.49–1.41 (m, 2H, CH), 1.10–1.07 (m, 2H, CH₂), 0.91–0.87 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₃NO₂ 179.0947, found 179.0957.

3-Cyclopropanecarbonyloxy-2-methyl-2-cyclohexen-1one (9). This compound was obtained from 2-methylcyclohexane-1,3-dione¹⁶ **18** as a colorless liquid with a 87% yield. R_f =0.36 (25% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 2.53–2.51 (m, 2H, 6-H), 2.46–2.42 (m, 2H, 4-H), 2.04–1.99 (m, 2H, 5-H), 1.79–1.75 (m, 1H, CH), 1.67 (s, 3H, 2-CH₃), 1.34–1.11 (m, 2H, CH₂), 1.03–1.00 (m, 2H, CH₂). HRMS: calcd for $C_{11}H_{14}O_3$ 194.0943, found 194.0940.

2-*p*-Nitrophenyl-3-cyclopropanecarbonyloxy-2-cyclohexen-1-one (10). This compound was obtained from 2*p*-nitrophenyl-cyclohexane-1,3-dione¹⁷ 19 as a yellow solid with a 75% yield. Mp 92–93 °C. R_f =0.33 (40% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 8.21 (d, J=9.0 Hz, 2H, Ar H's), 7.31 (d, J=9.0 Hz, 2H, Ar H's), 2.75 (t, J=6.0 Hz, 2H, 6-H), 2.62 (t, J=6.0 Hz, 2H, 4-H), 2.18 (quintet, J=6.0 Hz, 2H, 5-H), 1.55–1.49 (m, 1H, CH), 0.90–0.85 (m, 4H, CH₂CH₂). HRMS: calcd for C₁₆H₁₅NO₅ 301.0951, found 301.0946.

2-*o*-Nitrophenyl-3-cyclopropanecarbonyloxy-2-cyclohexen-1-one (11). This compound was obtained from 2*o*-nitrophenyl-cyclohexane-1,3-dione¹⁷ as a yellow solid with a 85% yield. Mp 82–83 °C. R_f =0.42 (40% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 8.11 (dd, J=8.1, 1.5 Hz, 1H, Ar H), 7.61 (dt, J=7.5, 1.5 Hz, 1H, Ar H), 7.50 (dt, J=8.4, 1.8 Hz, 1H, Ar H), 7.20 (dd, J=7.5, 1.8 Hz, 1H, Ar H), 2.88–2.58 (m, 4H, 4-H, 6-H), 2.24–2.13 (m, 2H, 5-H), 1.53–1.45 (m, 1H, CH), 0.89–0.75 (m, 4H, CH₂CH₂). HRMS: calcd for C₁₆H₁₅NO₅ 301.0951, found 301.0935.

2-Chloro-3-cyclopropanecarbonyloxy-2-cyclohexen-1-one (12). This compound was obtained from 2-chloro-cyclohexane-1,3-dione¹⁸ **20** as a yellow liquid with a 75% yield. R_f =0.23 (25% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 2.71–2.62 (m, 2H, 6-H), 2.60–2.58 (m, 2H, 4-H), 2.11–2.03 (m, 2H, 5-H), 1.84–1.79 (m, 1H, CH), 1.26–1.18 (m, 2H, CH₂), 1.16–1.03 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₁ClO₃ 214.0397, found 214.0403.

2-Bromo-3-cyclopropanecarbonyloxy-2-cyclohexen-1-one (13). This compound was obtained from 2-bromo-cyclohexane-1,3-dione¹⁹ **21** as a yellow liquid with a 50% yield. R_f =0.26 (25% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 2.71–2.61 (m, 4H, 4-H, 6-H), 2.12–2.04 (m, 2H, 5-H), 1.84–1.76 (m, 1H, CH), 1.23–1.20 (m, 2H, CH₂), 1.09–1.05 (m, 2H, CH₂). HRMS: calcd for C₁₀H₁₁BrO₃ 257.9891, found 257.9886.

3-Cyclopropanecarbonyloxy-5-ethoxycarbonyl-2-cyclohexen-1-one (14). This compound was obtained from 5-ethoxycarbonyl-cyclohexane-1,3-dione⁹ as a yellow liquid with a 67% yield. R_f =0.35 (35% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 5.96 (s, 1H, 2-H), 4.18 (q, J=7.2 Hz, 2H, OCH₂CH₃), 3.15–3.05 (m, 1H, 5-H), 2.93–2.55 (m, 4H, 4-H, 6-H), 1.77–1.71 (m, 1H, CH), 1.27 (t, J=7.2 Hz, 3H, OCH₂CH₃), 1.14–1.12 (m, 2H, CH₂), 1.06–1.01 (m, 2H, CH₂). HRMS: calcd for C₁₃H₁₆O₅ 252.0998, found 252.1002.

2, 4, 5-Trimethoxybenzaldehyde (22b). To a solution of 1, 2, 4-trimethoxybenzene (336 mg, 2.00 mmol) in DMF (10 mL) was added dropwise phosphorus oxychloride (307 mg, 2.00 mmol) at room temperature. The resulting mixture was stirred at that temperature for 1 h. After completion of the reaction, monitored by TLC, 100 mL of water and 10% NaOH solution (20 mL) was added to the solution. The precipitate was then filtered and the

product was purified by column chromatography to give a gray solid with a yield of 81%. Mp 114–115 °C. R_f =0.48 (40% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 10.32 (s, 1H, CHO), 7.33 (s, 1H, Ar H), 6.50 (s, 1H, Ar H), 3.98 (s, 3H, OMe), 3.93 (s, 3H, OMe), 3.88 (s, 3H, OMe). HRMS: calcd for C₁₀H₁₂O₄ 196.0736, found 196.0740.

2,5-Dimethoxyphenol (23a). To a solution of commercially available 2,5-dimethoxybenzaldehyde (196 mg, 1.00 mmol) was added dropwise 30% hydrogen peroxide (0.23 mL, 2.00 mmol) and sulfuric acid (0.05 mL). The resulting mixture was then stirred at room temperature for 2h. After completion of the reaction, water was added to the mixture and the solution was neutralized to pH 7. The product was extracted with ethyl acetate twice. The combined organic extracts were dried over MgSO₄, filtered, and concentrated. The resulting syrup was purified by column chromatography to give a yellow liquid with a yield of 65%. $R_f = 0.45$ (40%) EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.78 (d, J = 8.7 Hz, 1 H, Ar H), 6.56 (d, J = 3.0 Hz, 1 H, Ar H),6.38 (dd, J=8.7, 3.0 Hz, 1H, Ar H), 5.67 (bs, 1H, OH), 3.84 (s, 3H, OMe), 3.75 (s, 3H, OMe). HRMS: calcd for C₈H₁₀O₃ 154.0630, found 154.0634.

2, 4, 5-Trimethoxyphenol (23b). This compound was obtained as a white solid with a yield of 69% following the procedure of 23a. Mp 60–61 °C. R_f =0.40 (50% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.61 (s, 1H, Ar H), 6.58 (s, 1H, Ar H), 5.28 (bs, 1H, OH), 3.85 (s, 3H, OMe), 3.84 (s, 3H, OMe), 3.82 (s, 3H, OMe). HRMS: calcd for C₉H₁₂O₄ 184.0736, found 184.0739.

1 - Cyclopropanecarbonyloxy - **2**,**5** - dimethoxybenzene (24a). This compound was obtained as a yellow liquid with a yield of 92%. R_f =0.60 (40% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.89 (d, J=8.7 Hz, 1H, Ar H), 6.72 (dd, J=8.7, 3.0 Hz, 1H, Ar H), 6.65 (d, J=3.0 Hz, 1H, Ar H), 3.38 (s, 3H, OMe), 3.75 (s, 3H, OMe), 1.91–1.83 (m, 1H, CH), 1.21–1.16 (m, 2H, CH₂), 1.05–0.98 (m, 2H, CH₂). HRMS: calcd for C₁₂H₁₄O₄ 222.0892, found 222.0880.

1 - Cyclopropanecarbonyloxy - 2,4,5 - trimethoxybenzene (24b). This compound was obtained as a colorless liquid with a yield of 75%. R_f =0.47 (50% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.64 (s, 1H, Ar H), 6.60 (s, 1H, Ar H), 3.87 (s, 3H, OMe), 3.85 (s, 3H, OMe), 3.82 (s, 3H, OMe), 1.92–1.84 (m, 1H, CH), 1.19–1.03 (m, 2H, CH₂), 1.02–0.99 (m, 2H, CH₂). HRMS: calcd for C₁₃H₁₆O₅ 252.0998, found 252.0993.

2-Cyclopropanecarbonyloxy-1,4-benzoquinone (15). To a solution of **24a** (127 mg, 0.64 mmol) in acetone was added silver (II) oxide (372 mg, 3.00 mmol), and 6 N nitric acid (1 mL) dropwise at room temperature. The reaction was finished within 1 h. After acetone was removed under reduced pressure, the residue was dissolved in water. The aqueous layer was neutralized with sodium bicarbonate and extracted with ether. The combined organic extracts were dried over MgSO₄, filtered, concentrated, and the residue was purified by column

chromatography to give a yellow solid with a yield of 80%. Mp 173–175 °C. R_f =0.55 (40% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 7.26 (s, 1H, Ar H), 6.87 (s, 1H, Ar H), 6.70 (s, 1H, Ar H), 1.91–1.84 (m, 1H, CH), 1.27–1.20 (m, 2H, CH₂), 1.15–1.09 (m, 2H, CH₂). HRMS: calcd for C₁₀H₈O₄ 192.0422, found 192.0428.

2-Cyclopropanecarbonyloxy-5-methoxy-1,4-benzoquinone (16). This compound was obtained as a yellow solid with a yield of 83% following the procedure of 15. Mp 85–87 °C. R_f =0.43 (50% EtOAc/hexanes). ¹H NMR (CDCl₃) δ 6.54 (s, 1H, Ar H), 5.95 (s, 1H, Ar H), 3.86 (s, 3H, OMe), 1.90–1.84 (m, 1H, CH), 1.26–1.20 (m, 2H, CH₂), 1.13–1.08 (m, 2H, CH₂). HRMS: calcd for C₁₁H₁₀O₅ 222.0528, found 222.0533.

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References and Notes

(a) Scheparts, B.; Gurin, S. J. Biol. Chem. 1949, 180, 663.
(b) Vitol, M. J.; Vilks, S. R.; Zabarovska, I. M.; Maurinia, K. A. Kokl. Akad. Nauk. SSSR 1970, 192, 908.

- 2. Jefford, C. W.; Cadby, P. Experimentia 1981, 37, 1134.
- 3. Lindblad, B.; Lindstedt, S.; Steen, G. Proc. Natl. Acad. Sci. U.S.A. 1977, 74, 4641.
- 4. Puntoni, R.; Merlo, F.; Fini, A.; Meazza, L.; Santi, L. Lancet 1986, *ii*, 525.
- 5. Lindstedt, S.; Holme, E.; Lock, E.; Hialmarson, O.; Strandvik, B. *Lancet* **1992**, *340*, 813.
- 6. Lin, S.-W.; Lin, Y.-L.; Lin, T.-C.; Yang, D.-Y. Bioorg. Med. Chem. Lett. 2000, 10, 1297.
- 7. Suginome, H.; Kondoh, T.; Gogonea, C.; Singh, V.; Goto,
- H.; Osawa, E. J. Chem. Soc., Perkin Trans. 1, 1995, 69.
- 8. Montes, I. F.; Burger, U. Tetrahedron Lett. 1996, 37, 1007.
- 9. Wu, B.; Bai, D. J. Org. Chem. 1997, 62, 5978.
- 10. Vilsmeier, A.; Haack, A. Chem. Ber. 1927, 60, 119.
- 11. Baeyer, A.; Villiger, V. Chem. Ber. 1902, 35, 3013.
- 12. Lindstedt, S.; Rundgren, M. Biochim. Biophys. Acta 1982, 704, 66.
- 13. Crouch, N. P.; Adlington, R. M.; Baldwin, J. E.; Lee, M.-
- H.; MacKinnon, C. H. *Tetrahedron* **1997**, *53*, 6993.
- 14. Buckthal, D. J.; Roche, P. A.; Moorehead, T. J.; Forbes, B. J. R.; Hamilton, G. A. *Methods in Enzymol.* **1987**, *142*, 132.
- 15. Huang, Y.; Hartmann, R. W. Synth. Commun. 1998, 28, 1197.
- 16. Barrack, S. A.; Okamura, W. H. J. Org. Chem. 1986, 51, 3201.
- 17. Sole, D.; Bosch, J.; Bonjoch, J. Tetrahedron 1996, 52, 4013.
- 18. Shepherd, R. G.; White, A. C. J. Chem. Soc., Perkin Trans. 1, 1987, 2153.
- 19. Arakawa, C. Pharm. Bull. 1957, 5, 528.