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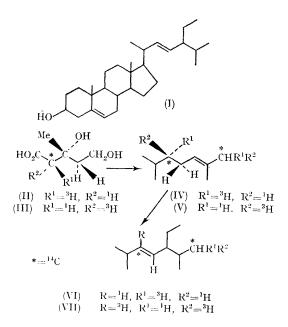
## The Stereochemistry of Hydrogen Elimination at C(7) and C(22) in Phytosterol Biosynthesis by Ochromonas malhamensis

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A NUMBER of phytosterols are characterised by a  $\Delta^{22}$  bond in the side-chain.<sup>1</sup> Little is known about the mechanism of formation of the  $\Delta^{22}$  bond but, as would be expected, a hydrogen atom is eliminated from C(23) during introduction of this double bond into ergosterol.<sup>2</sup> We now report the use of 3R-[2-<sup>14</sup>C-(2R)-2-<sup>3</sup>H<sub>1</sub>]mevalonic acid [MVA, (III)] and 3R-[2-<sup>14</sup>C-(2S)-2-<sup>3</sup>H<sub>1</sub>]MVA (III) to study the stereospecific hydrogen eliminations at C(7) and C(22) during the biosynthesis of poriferasterol (I) by Ochromonas malhamensis.

Two cultures of O. malhamensis were grown as previously described<sup>3</sup> but with the addition to the growth medium of either  $3R-[2-^{14}C-(2R)-2-^{3}H_1]$ MVA (II) (2  $\mu$ c of <sup>14</sup>C; <sup>3</sup>H : <sup>14</sup>C = 8.58) or 3*R*-[2- $^{14}C-(2S)-2-^{3}H_{1}MVA$  (III) (2.0  $\mu c$  of  $^{14}C$ ;  $^{3}H$ :  $^{14}C =$ 8.59). The cells from the two experiments were harvested after six days and treated in an identical manner. The non-saponifiable lipid was chromatographed on silica gel thin layers and the sterol band eluted. Poriferasterol (100 mg.) was added to a portion of the radioactive sterol and crystallised to constant specific activity. Acetylation, followed by oxidation<sup>4</sup> gave  $3\beta$ -acetoxy-24Sethylcholesta-5,22-dien-7-one, m.p. 188-191°, purified by chromatography and crystallisation. A second portion of the radioactive poriferasterol, after addition of carrier sterol, was converted into



24-ethyl-cholesta-4,22-dien-3-one by Oppenauer oxidation.<sup>5</sup> Ozonolysis then gave 3-oxobisnor-4-cholen-22-al, m.p. 154-156° which was finally converted into 3-oxobisnor-4-cholenic acid, m.p.

266-269° (decomp.) by treatment with chromic acid.<sup>6</sup> The results are given in the Table.

The decrease in the <sup>3</sup>H ; <sup>14</sup>C ratio observed when

 $[(V) \rightarrow (VII)]$ . Oxidation of the 3-oxobisnor-4cholen-22-al obtained by ozonolysis of the poriferasterol from the  $3R-[2^{-14}C-(2R)-2^{-3}H_1]MVA$ 

Incorporation o 3R-[2-14C-(2R)-2-3H1]MVA and 3R-[2-14C-(2S)-2-3H1]MVA into poriferasterol in Ochromonas malhamensis

	$3R - [2^{-14}C - (2R) - 2^{-3}H_1]MVA$		3R-[2- <sup>14</sup> C-(2S)-2- <sup>3</sup> H <sub>1</sub> ]MVA	
	Specific activity d.p.m. of <sup>14</sup> C/mg.	<sup>3</sup> H : <sup>14</sup> C ratio	Specific activity d.p.m. of <sup>14</sup> C/mg.	<sup>3</sup> H : <sup>14</sup> C ratio
Poriferasterol $3\beta$ -Acetoxy-24S-ethylcholesta-5,22-dien-7-one $24$ -Ethylcholesta-4,22-dien-3-one $3$ -Oxobisnor-4-cholen-22-al $3$ -Oxobisnor-4-cholenic acid	1728 1510 1715 1685	4.65 3.90 4.68 4.45 4.00	1840 1670 1790 1885	3.80 3.66 3.88 3.63 2.21

the poriferasterol biosynthesised from 3R-[2-14C-(2R)-2-<sup>3</sup>H<sub>1</sub>]MVA (II) was converted into 3 $\beta$ acetoxy 24S-ethylcholesta-5,22-dien-7-one demonstrates the biosynthetic retention of the C(7)hydrogen derived from the 2-proR hydrogen<sup>7</sup> of MVA. By contrast, the much smaller fall in the <sup>3</sup>H:<sup>14</sup>C ratio upon formation of the 7-ketone, when  $3R-[2-^{14}C-(2S)-2-^{3}H_1]MVA$  is the poriferasterol precursor, shows the biosynthetic loss of the  $7\beta$ hydrogen derived from the 2-proS hydrogen of MVA. This is in complete accord with results obtained for cholesterol biosynthesis in liver.8

Incorporation of (II; 2R) or (III; 2S) into the side-chain of the poriferasterol precursor sterol will give tritium at C(22) in the configuration shown in (IV; 22R) and (V; 22S) respectively. The large drop in the <sup>3</sup>H : <sup>14</sup>C ratio upon oxidation of the 3-oxobisnor-4-cholen-22-al obtained from the  $3R-[2-^{14}C-(2S)-2-^{3}H_1]MVA$  (III) culture proves that a tritium atom was located at C(22) and therefore that the C(22) proton of poriferasterol was derived from the 2-proS hydrogen of MVA

(III) incubation resulted in only a relatively small decrease in the <sup>3</sup>H:<sup>14</sup>C ratio, thus showing the presence of only a small amount of tritium at C(22)of the poriferasterol. The two results demonstrate that the C(22) hydrogen originating from the 2proR hydrogen of MVA [*i.e.* the C(22) proR hydrogen] is lost in the introduction of the  $\Delta^{22}$  bond into poriferasterol  $[(IV) \rightarrow (VI)]$  by O. malhamensis. The retention of some tritium at C(22) from the  $[2R-2-{}^{3}H_{1}]MVA$  and at C7 from the  $[2S-2-{}^{3}H_{1}]$ -MVA is probably due to loss of stereospecificity caused by the reversibility of the isopentenyl pyrosphosphate-dimethyl allyl pyrophosphate enzymic isomerisation.9 This equilibration would also explain the unexpectedly large drop in the <sup>3</sup>H:<sup>14</sup>C ratios observed upon conversion of the doubly labelled mevalonates into sterol. These findings will be discussed in more detail elsewhere.

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<sup>1</sup> L. J. Goad, in "Terpenoids in Plants", ed., J. B. Pridham, Academic Press, London, 1967, p. 159.

<sup>2</sup> M. Akhtar, M. A. Parvez, and P. F. Hunt, Biochem. J., 1968, 106, 623.

<sup>3</sup> Poriferasterol (24S-ethyl-cholesta-5-22-dien- $3\beta$ -ol) is the C(24) epimer of stigmasterol. The identification of the O. malhamensis sterol is based upon mass spectral, n.m.r., g.l.c., and i.r. evidence. It is distinguished from stig-masterol at present solely by the m.p. of the free sterol  $(154^\circ)$  and the steryl acetate  $(146^\circ)$  (A. R. H. Smith, L. J. Goad, T. W. Goodwin, and E. Lederer, Biochem. J., 1967, 104, 15c; M. C. Gershengorn, A. R. H. Smith, G. Goulston, L. J. Goad, T. W. Goodwin, and T. H. Haines, Biochemistry, 1968, 7, 1698.

- <sup>7</sup> K. R. Hanson, J. Amer. Chem. Soc., 1966, 88, 2731.
  <sup>8</sup> G. F. Gibbons, L. J. Goad, and T. W. Goodwin, 1968, unpublished results.
- <sup>9</sup> P. W. Holloway and G. Popjak, Biochem. J., 1968, 106, 835.

<sup>&</sup>lt;sup>4</sup> L. F. Fieser, M. Fieser, and R. N. Chakraverti, *J. Amer. Chem. Soc.*, 1949, **71**, 2226. <sup>5</sup> D. A. Shepherd, R. A. Donia, J. A. Campbell, B. A. Johnson, R. P. Holysz, G. Stomp, S. E. Stafford, R. L. Pederson, and A. C. Ott, J. Amer. Chem. Soc., 1955, 77, 1212. M. E. Herr and F. W. Heyl, J. Amer. Chem. Soc., 1952, 74, 3627.