Stereoselective Synthesis of Tetrahydropyrans through Tandem and Organocatalytic Oxa-Michael Reactions: Synthesis of the Tetrahydropyran Cores of *ent*-(+)-Sorangicin A

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Tandem and organocatalytic oxa-Michael reactions of α , β unsaturated aldehydes were explored for the stereoselective synthesis of structurally complex tetrahydropyrans. The stereoselective synthesis of 2,6-*trans*-tetrahydropyrans, which are thermodynamically unfavorable, was accomplished through a reagent-controlled, organocatalytic

Introduction

Substituted tetrahydropyrans are important structural motifs found in a wide range of biologically interesting natural products.^[1,2] Among substituted tetrahydropyrans, 3methyl-2,6-disubstituted tetrahydropyrans are one of the most abundant classes in natural products and have attracted considerable interest (Figure 1).^[1a,1c,1c,1f] Although an increasing amount of interest has focused on the generation of these structures, it is surprising that the oxa-Michael reaction of α,β -unsaturated aldehydes has rarely been used for the stereoselective synthesis of 3-methyl-2,6-disubstituted tetrahydropyrans.^[3,4] In addition, the stereoselective synthesis of 2,6-*trans*-tetrahydropyrans has been a challenge in organic synthesis because of their poor thermodynamic stability.

Recently, we reported the stereoselective synthesis of 2,6*cis*-tetrahydropyrans through the tandem oxidation/oxa-Michael reaction of α , β -unsaturated aldehydes promoted by the *gem*-disubstituent effect.^[5] The reaction required no activation of either the oxygen nucleophiles or the aldehydes. The reaction was applicable to a broad range of substrates and proceeded with excellent stereoselectivity (>20:1 *dr*). We used the oxa-Michael reaction in conjunction with a dithiane coupling reaction to perform the highly stereoselective synthesis of 2,3,6-trisubstituted tetrahydropyrans.^[6] We also demonstrated the utility and efficiency of the combination of the oxa-Michael reaction and the dioxa-Michael reaction. A temperature-dependent configurational switch allowed the preparation of both 2,3-*trans*-2,6-*trans*- and 2,3-*cis*-2,6-*cis*-tetrahydropyrans from a common substrate. This switch was then used to synthesize the precursors of the C21–C29 and C30–C37 fragments of *ent*-(+)-sorangicin A.

thiane coupling reaction in the stereoselective synthesis of neopeltolide,^[5] cyanolide A,^[6a] leucascandrolide A,^[6b] psymberin,^[6c] and SCH 351448.^[7]

Intrigued by the excellent efficiency and stereoselectivity of the tandem oxa-Michael reaction of α , β -unsaturated aldehydes in the synthesis of 2,6-*cis*-tetrahydropyrans, we decided to extend this method to the synthesis of 3-methyl-2,6-disubstituted tetrahydropyrans. Herein, we describe our investigation of tandem and organocatalytic oxa-Michael reactions of α , β -unsaturated aldehydes for the stereoselective synthesis of structurally complex tetrahydropyrans and their application to the efficient synthesis of the precursors to the C21–C29 and C30–C37 fragments of *ent*-(+)-sorangicin A.

Results and Discussion

The tandem oxidation/oxa-Michael reaction required no activation of the substrates and proceeded in a substratecontrolled manner under neutral and mild conditions. Therefore, we proposed that it would be possible to predict the stereochemical outcome of the oxa-Michael reaction through conformational analysis of the transition states (Figure 2).

On the basis that the bulky C2, C3, and C6 substituents of 2,3-*trans*-2,6-*cis*-tetrahydropyran 12 occupy equatorial positions, we expected that 12 could be stereoselectively prepared through the most favorable transition state 11B in the tandem oxidation/oxa-Michael reaction. We also anticipated that aldehyde (*E*)-13 should adopt the more favorable chair-like transition state (*E*)-13B to stereoselectively provide 2,3-*cis*-2,6-*cis*-tetrahydropyran 14 owing to the severe 1,3-diaxial interaction between the C4 dithiane group and the C6 alkyl group in competing transition state (*E*)-13A.

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Figure 1. Examples of natural products containing 3-methyl-2,6-disubstituted tetrahydropyrans.



Figure 2. Conformational analysis of the oxa-Michael reactions.

However, the stereoselective synthesis of 2,3-*trans*-2,6-*trans*-tetrahydropyran **15** by using the tandem oxidation/oxa-Michael reaction was expected to be more challenging. A stereochemical mismatch between the C3 methyl group and the C6 alkyl group in (Z)-**13A** and (Z)-**13B** was expected to hamper the formation of a well-defined transition state in the oxa-Michael step leading to **15**, thus having an impact on the stereoselectivity and reactivity.

To test the feasibility of the oxa-Michael reaction for the stereoselective synthesis of 2,3-*trans*-2,6-*trans*-tetrahydropyrans, we prepared allyl alcohol (Z)-**18a**, by coupling (Z)-**17**^[6b] and (S)-glycidyl benzyl ether (**16a**), and subjected it to the tandem oxidation/oxa-Michael reaction (Scheme 1). As predicted, the tandem reaction of (Z)-**18a** provided aldehyde (Z)-**19a** as the major product (65–70%) because of repulsive interactions in the transition states. Both starting material and product also decomposed during the prolonged reaction time (72 h).



Scheme 1. Preparation of allyl alcohol (*Z*)-**18a** and attempts towards the synthesis of 2,3-*trans*-2,6-*trans*-tetrahydropyrans through the tandem oxa-Michael reaction.

To overcome the stereochemical mismatch in the transition states and to promote the oxa-Michael step by increasing the reactivity of (Z)-19a, we converted (Z)-19a into the corresponding iminium ion by reacting it with a range of amines and acids (Table 1). The iminium activation of (Z)-19a by treatment with pyrrolidine and BzOH at 25 °C dramatically promoted the oxa-Michael reaction, but afforded undesired 2,3-cis-2,6-cis-tetrahydropyran 21a as a single diastereomer (Table 1, Entry 1).^[8] Surprisingly, when the reaction was attempted at low temperature (-40 or -78 °C), the stereoselectivity was reversed to provide 2,3-trans-2,6trans-tetrahydropyran 20a as the major diastereomer (3.5-4.1:1 dr; Table 1, Entries 2 and 3).^[8] Use of piperidine as an alternative amine source further improved the stereoselectivity (7.4:1 dr; Table 1, Entry 6). Encouraged by these results, we decided to test chiral organocatalysts to further improve the stereoselectivity of the oxa-Michael reaction.^[9,10] When (Z)-19a was treated with (S)-22^[11] at -40 °C, the organocatalytic oxa-Michael reaction proceeded smoothly to provide **20a** with excellent stereoselectivity and yield (12:1 dr, 96%; Table 1, Entry 9). When (R)-22 was employed, the organocatalytic oxa-Michael reaction provided 20a with no stereoselectivity (Table 1, Entry 10).



Table 1. The organocatalytic oxa-Michael reaction of α , β -unsaturated aldehydes.



[a] Combined yield of isolated **20a** and **21a**. [b] The diastereomeric ratio (**20a/21a**) was determined by integration of relevant ¹H NMR spectroscopic signals of the crude product.

Our proposed rationales (Rationale 1 and Rationale 2, Figure 3) for the stereochemical outcome, as a function of reaction temperature observed in the organocatalytic oxa-Michael reaction, are illustrated in Figure 3. Rationale 1 shows that at low temperature (-40 or -78 °C), the iminium ion of (Z)-19a would adopt conformation 23B to avoid the severe 1,3-diaxial interaction between the axially oriented C3 methyl group and the C2 iminium diene group to afford 2,3-trans-2,6-trans-tetrahydropyran 20a. At 25 °C, (Z)-iminium ions 23A and 23B readily undergo isomerization to give the more stable (E)-iminium ion 23C, which subsequently cyclizes to give the kinetically and thermodynamically more favorable product 2,3-cis-2,6-cis-tetrahydropyran 21a. An alternative explanation (Rationale 2) is that the iminium ion of (Z)-19a initially forms the kinetically more favorable product 20a at 25 °C. Compound 20a could then be converted into the corresponding (E)-enal through a retro-Michael reaction to form 21a. The low reaction temperature would minimize the isomerization of (Z)-iminium ions and/or the retro-Michael/isomerization/oxa-Michael reaction of 20a. When 20a was subjected to the oxa-Michael reaction conditions (pyrrolidine, BzOH and CH₂Cl₂ at 25 °C for 2.5 h), 21a (96%, >20:1 dr) was formed exclusively, suggesting that Rationale 2 is most likely. Because no equilibrium in the retro-Michael reaction was observed for the tandem oxidation/oxa-Michael reaction,^[5] the equilibrium between 20a and 21a observed in the organocatalytic oxa-Michael reaction was attributed to activation by the iminium ion formation and/or the stereochemical mismatch between the C3 methyl group and the C6 alkyl chain in 20a.

To the best of our knowledge, this is the first report of a temperature-dependent configurational switch for the synthesis of both 2,3-*trans*-2,6-*trans*- and 2,3-*cis*-2,6-*cis*-tetra-hydropyrans from a common substrate by using oxa-Michael reactions.



Figure 3. Proposed rationales for the stereochemical outcome of the organocatalytic oxa-Michael reaction.

To investigate the substrate scope and stereochemical outcome of the organocatalytic oxa-Michael reaction, we prepared α,β -unsaturated aldehydes (*Z*)-**19a**–**e** with a variety of substituents at the C6 position by coupling (*Z*)-**17** with commercially or readily available chiral epoxides **16a**–**e** (Scheme 2).

Under the standard reaction conditions [(S)-22 and BzOH in CH₂Cl₂ at -40 °C], the organocatalytic oxa-Michael reaction of (*Z*)-19a–e proceeded smoothly to provide the corresponding 2,3-*trans*-2,6-*trans*-tetrahydropyran aldehydes **20a–e** with good to excellent stereoselectivities (11–20:1 *dr*; Table 2).



Scheme 2. Preparation of α,β -unsaturated aldehydes (*Z*)-**19a**–**e** for the organocatalytic oxa-Michael reaction.

Table 2. Substrate scope of the organocatalytic oxa-Michael reaction.



1	19a	14	96	12:1	
2	19b	7	97	20:1	
3	19c	8	98	11:1	
4	19d	13	98	>20:1	
5	19e	12	95	13:1	

[a] Combined yield of the isolated 2,3-*trans*-2,6-*trans*- and 2,3-*cis*-2,6-*cis*-tetrahydropyrans. [b] The diastereomeric ratio (2,3-*trans*-2,6-*trans*-tetrahydropyran/2,3-*cis*-2,6-*cis*-tetrahydropyran) was determined by integration of the ¹H NMR spectroscopic signals of the crude product.

As illustrated in Figure 2, conformational analysis suggested that the stereoselective synthesis of 2,3-*trans*-2,6-*cis*-tetrahydropyrans and 2,3-*cis*-2,6-*cis*-tetrahydropyrans through the tandem oxa-Michael reaction should be straightforward. To explore the utility of the tandem oxa-Michael reaction in the stereoselective synthesis of these compounds, we prepared allyl alcohols (*E*)-**25a**-**d**, (*Z*)-**25a**-**d**, and (*E*)-**18a**-**d** by coupling (*E*)-**24**, (*Z*)-**24**, and (*E*)-**17**, respectively, with chiral epoxides **16a**-**d** (Scheme 3).

As expected, the tandem oxa-Michael reaction (MnO₂ in CH₂Cl₂ at 25 °C) of (*E*)- and (*Z*)-**25a**-**d** afforded 2,3-*trans*-2,6-*cis*-tetrahydropyrans **26a**-**d** with excellent stereoselectivities (Table 3, Entries 1–8).^[8,12] Under the same reaction conditions, (*E*)-**18a**-**d** gave rise to 2,3-*cis*-2,6-*cis*-tetrahydropyrans **21a**-**d** as single diastereomers (Table 3, Entries 9–12).^[8]



Scheme 3. Preparation of allyl alcohols.

Table 3. Synthesis of 2,3-*trans*-2,6-*cis*- and 2,3-*cis*-2,6-*cis*-tetrahydropyrans **26a**-**d** and **21a**-**d** through tandem oxa-Michael reactions.



 \mathbf{c} , R = Ph; \mathbf{d} , R = C(CH₃)₂CH₂OBn

Entry	Substrate	Time [h]	Yield [%]	$dr^{[a]}$
1	(<i>E</i>)-25a	12	83	>20:1
2	(E)-25b	12	90	>20:1
3	(E)-25c	12	83	>20:1
4	(E)-25d	12	82	>20:1
5	(Z)-25a	10	84	>20:1
6	(Z)-25b	10	85	>20:1
7	(Z)-25c	10	81	>20:1
8	(Z)-25d	12	88	>20:1
9	(E)-18a	10	87	>20:1
10	(E)-18b	10	84	>20:1
11	(E)-18c	10	85	>20:1
12	(<i>E</i>)-18d	12	81	>20:1

[a] The diastereomeric ratio was determined by integration of the ¹H NMR spectroscopic signals of the crude product.

The tandem and organocatalytic oxa-Michael reactions of α,β -unsaturated aldehydes can be broadly applicable to the stereoselective synthesis of structurally complex tetra-

hydropyrans and natural products. To demonstrate the efficiency of the temperature-dependent configurational switch from a common substrate, we synthesized the precursors to the C21-C29 and C30-C37 fragments of ent-(+)-sorangicin A (ent-1, Figure 4). The marine macrolide (+)-sorangicin A (1, Figure 1) was isolated from the myxobacterium Sorangium cellulosum by Jansen and co-workers.^[13] (+)-Sorangicin A is active against both Gram-positive (MIC = 0.01–0.1 μ g/mL) and Gram-negative (MIC = 3–30 μ g/mL) bacteria.^[14] Reichenbach and co-workers determined that the antibacterial activity of 1 arises from its inhibition of RNA polymerase.^[14] Owing to its potent antibiotic activity and architectural complexity, the synthesis of 1 has attracted considerable interest from a number of groups,^[15–17] with the first total synthesis reported by Smith and coworkers.^[15] We envisioned that both 2,3-trans-2,6-transtetrahydropyran and 2,3-cis-2,6-cis-tetrahydropyran units embedded in ent-1 could be constructed from a common substrate through the temperature-dependent configurational switch of the organocatalytic oxa-Michael reaction.



Figure 4. Structure of ent-(+)-sorangicin A (ent-1).

The synthesis of the precursors to the C21–C29 and C30–C37 fragments of *ent*-1 started with the preparation of chiral epoxide **31** (Scheme 4). Bn protection of commercially available (*R*)-(+)-glycidol (**27**), opening of epoxide **28** by trimethylsulfonium iodide, and Sharpless asymmetric epoxidation of allyl alcohol **29** provided known epoxide **30**.^[18] Protection of **30** by using *p*-methoxybenzyl chloride completed the synthesis of chiral epoxide **31**.



Scheme 4. Preparation of chiral epoxide 31.

Dithiane coupling of (Z)-17 with 31 followed by MnO₂ oxidation of the corresponding allyl alcohol (Z)-32 set the stage for the organocatalytic oxa-Michael reactions (Scheme 5). The organocatalytic oxa-Michael reaction of (Z)-33 at 25 °C in the presence of pyrrolidine proceeded smoothly to provide 2,3-*cis*-2,6-*cis*-tetrahydropyran 34 as a single diastereomer (>20:1 dr).^[12] When (S)-22 was used for the oxa-Michael reaction of (Z)-33 at -40 °C, 2,3-*trans*-2,6-*trans*-tetrahydropyran 35 was obtained with excellent stereoselectivity (17:1 dr). The temperature-dependent con-



Scheme 5. Synthesis of the precursors to the C21–C29 and C30–C37 fragments of *ent*-1 through the organocatalytic oxa-Michael reaction.



Scheme 6. Synthesis of the precursor to the C21–C29 fragment of ent-(+)-sorangicin A through a tandem oxidation/oxa-Michael reaction.

figurational switch was successfully used to prepare diastereomeric tetrahydropyrans 34 and 35 from common substrate (Z)-33.

In addition, the tandem oxidation/oxa-Michael reaction was effective in the stereoselective synthesis of 2,3-*cis*-2,6*cis*-tetrahydropyran **34** as a single diastereomer (Scheme 6). Coupling of (*E*)-**17** and **31** provided allyl alcohol (*E*)-**32** in 76% yield. The tandem oxidation/oxa-Michael reaction of (*E*)-**32** (MnO₂ in CH₂Cl₂ at 25 °C for 12 h) proceeded smoothly to provide **34** with excellent stereoselectivity and yield (>20:1 *dr*, 85%). Compounds **34** and **35** can be further elaborated into C21–C29 fragment **36** and C30–C37 fragment **37** of *ent*-**1**.

Conclusion

In summary, tandem and organocatalytic oxa-Michael reactions have been used to stereoselectively synthesize structurally complex tetrahydropyrans. In particular, the synthesis of thermodynamically unfavorable 2,6-*trans*-tetra-hydropyrans was achieved through a reagent-controlled, organocatalytic oxa-Michael reaction. A temperature-dependent configurational switch allowed the preparation of both 2,3-*trans*-2,6-*trans*- and 2,3-*cis*-2,6-*cis*-tetrahydropyrans from a common substrate, which was applied in the synthesis of the precursors to the C21–C29 and C30–C37 fragments of *ent*-(+)-sorangicin A. We expect that tandem and organocatalytic oxa-Michael reactions will be used for the stereoselective synthesis of a diverse set of tetrahydropyrans and be applied to the synthesis of complex natural products with interesting biological activities.

Experimental Section

General Methods: All reactions were conducted in oven-dried glassware under nitrogen. All commercial chemical reagents were used as supplied. Anhydrous tetrahydrofuran (THF) was distilled from sodium/benzophenone. Analytical thin layer chromatography (TLC) was performed on SiO₂ (60 Å) with florescent indication (Whatman). Visualization was accomplished by UV irradiation at 254 nm and/or by staining with para-anisaldehyde solution. Flash column chromatography was performed by using silica gel 60 (particle size 4063 $\mu m,~230400$ mesh). 1H NMR, ^{13}C NMR, and 2D NMR (COSY, NOESY) spectra were recorded with a Varian 400 (400 MHz) and a Bruker 500 (500 MHz) spectometer in CDCl₃ by using the signal of residual CHCl₃ as an internal standard. All NMR δ values are given in ppm, and all J values are in Hz. Electrospray ionization (ESI) mass spectra (MS) were recorded with an Agilent 1100 series (LC/MSD trap) spectrometer and were performed to obtain the molecular masses of the compounds. Infrared (IR) absorption spectra were determined with a Thermo-Fisher (Nicolet 6700) spectrometer. Optical rotation values were measured with a Rudolph Research Analytical (A21102, API/1W) polarimeter.

Typical Procedure for the Organocatalytic Oxa-Michael Reaction: To a cooled (-40 °C) solution of aldehyde (Z)-**19a** (28.8 mg, 0.079 mmol) in CH₂Cl₂ (0.039 M, 2.0 mL) was added dropwise a mixture of (S)-(-)- α , α -diphenyl-2-pyrrolidinemethanol trimethylsilyl ether (5.1 mg, 0.016 mmol) and BzOH (2.0 mg, 0.016 mmol) in CH₂Cl₂ (0.5 mL). After stirring at -40 °C for 14 h, the reaction mixture was diluted with hexanes (25.0 mL), filtered through a short pad of silica gel (hexanes/EtOAc, 3:1), and concentrated in vacuo. The residue was purified by column chromatography (silica gel; hexanes/EtOAc, 2:1) to afford 2,3-trans-2,6-trans-tetrahydropyran 20a (25.9 mg, 90%) and 2,3-cis-2,6-cis-tetrahydropyran 21a (2.1 mg, 7%) as colorless oils. Data for **20a**: $[a]_{D}^{25} = +16.0$ (c = 0.92, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ = 9.75 (dd, J = 2.0, 2.0 Hz, 1 H), 7.26–7.37 (m, 5 H), 4.54 (s, 2 H), 4.25 (ddd, J = 7.0, 7.0, 7.0 Hz, 1 H), 4.15 (dddd, J = 5.5, 5.5, 5.5, 5.5 Hz, 1 H), 3.80 (dd, J = 6.0, 1.5 Hz, 2 H), 3.08 (ddd, J = 14.5, 11.5, 3.0 Hz, 1 H),2.96 (ddd, J = 14.5, 11.5, 3.0 Hz, 1 H), 2.65–2.75 (m, 5 H), 2.26 (dd, J = 14.5, 5.5 Hz, 1 H), 2.00-2.07 (m, 1 H), 1.94 (dddd, J =7.0, 7.0, 7.0, 7.0 Hz, 1 H), 1.79–1.89 (m, 1 H), 1.21 (d, J = 7.0 Hz, 3 H) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 201.4, 138.1, 128.3, 127.6, 73.3, 70.55, 70.37, 69.6, 52.3, 47.5, 43.2, 36.5, 26.1, 25.7, 25.2, 13.9 ppm. IR (neat): $\tilde{v} = 1722$, 1452, 1277, 1097, 906, 738, 698 cm⁻¹. HRMS (ESI): calcd. for $C_{19}H_{26}O_3S_2$ [M + H]⁺ 367.1396; found 367.1396. Data for **21a**: $[a]_{D}^{25} = -26.9$ (c = 1.07, CHCl₃). ¹H NMR (500 MHz, CDCl₃): $\delta = 9.77$ (dd, J = 2.0, 2.0 Hz, 1 H), 7.24– 7.36 (m, 5 H), 4.84 (ddd, J = 9.5, 4.5, 2.0 Hz, 1 H), 4.53 (AB, Δv = 16.5, J_{AB} = 11.5 Hz, 2 H), 4.06–4.12 (m, 1 H), 3.48 (dd, J = 10.5, 5.5 Hz, 1 H), 3.42 (dd, J = 10.5, 4.5 Hz, 1 H), 2.66–2.90 (m, 5 H), 2.34 (ddd, J = 17.0, 4.5, 2.0 Hz, 1 H), 2.09 (ddd, J = 7.5, 7.5, 7.5 Hz, 1 H), 1.90–2.02 (m, 3 H), 1.85 (dd, J = 13.5, 11.5 Hz, 1 H), 1.11 (d, J = 7.5 Hz, 3 H) ppm. ¹³C NMR (125 MHz, CDCl₃): $\delta =$ 201.0, 138.1, 128.3, 127.6, 73.3, 72.59, 72.56, 70.1, 53.3, 47.4, 38.3, 34.6, 26.0, 25.4, 25.2, 8.9 ppm. IR (neat): $\tilde{v} = 1722$, 1453, 1376, 1108, 1026, 738, 698 cm⁻¹. HRMS (ESI): calcd. for C₁₉H₂₆O₃S₂ [M + H]⁺ 367.1393; found 367.1396.

Typical Procedure for the Tandem Oxidation/Oxa-Michael Reaction: To a solution of diol (E)-25a (15.5 mg, 0.042 mmol) in CH₂Cl₂ (0.021 M, 2.0 mL) was added MnO₂ (18.3 mg, 0.21 mmol). The resulting mixture was stirred for 1 h at 25 °C. An addition of MnO₂ (18.3 mg, 0.21 mmol) was repeated three times every 1 h. After stirring for an additional 6 h, the reaction mixture was filtered through a pad of Celite and concentrated in vacuo. The residue was purified by column chromatography (silica gel; hexanes/EtOAc, 2:1) to afford 2,3-trans-2,6-cis-tetrahydropyran 26a (12.8 mg, 83%) as a colorless oil. $[a]_{D}^{25} = +16.4$ (c = 0.17, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ = 9.78 (dd, J = 3.5, 1.5 Hz, 1 H), 7.26–7.40 (m, 5 H), 4.56 (s, 2 H), 4.09–4.15 (m, 2 H), 3.52 (dd, J = 10.0, 5.0 Hz, 1 H), 3.47 (dd, J = 10.5, 5.0 Hz, 1 H), 3.14 (ddd, J = 14.5, 12.5, 2.5 Hz, 1 H), 2.91 (ddd, J = 14.5, 12.0, 2.5 Hz, 1 H), 2.76 (dd, J = 14.0, 1.5 Hz, 1 H), 2.62–2.69 (m, 2 H), 2.58 (ddd, J = 16.0, 4.0, 2.0 Hz, 1 H), 2.44 (ddd, J = 15.5, 9.0, 3.5 Hz, 1 H), 2.05–2.12 (m, 1 H), 1.77-1.88 (m, 2 H), 1.69-1.76 (m, 1 H), 1.16 (d, J = 7.0 Hz, 3 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 201.6, 138.2, 128.4, 127.7, 73.4, 73.1, 72.8, 72.5, 54.3, 47.4, 45.5, 39.9, 25.68, 25.55, 25.0, 12.2 ppm. IR (neat): $\tilde{v} = 1722$, 1452, 1380, 1054, 908, 736, 698 cm⁻¹. HRMS (ESI): calcd. for $C_{19}H_{26}O_3S_2$ [M + H]⁺ 367.1392; found 367.1396.

Supporting Information (see footnote on the first page of this article): Complete characterization data and copies of the ¹H and ¹³C NMR spectra.

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