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Epoc group: transformable protecting group with gold(III)-catalyzed fluorene formation†

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This study presents the novel concept of a transformable protecting group, which changes its properties through structural transformation. Based on this concept, we developed a 2-(2-ethynylphenyl)-2-(5-methylfuran-2-yl)-ethoxycarbonyl (Epoc) group. The Epoc group was transformed into an Fmoc-like structure with gold(III)-catalyzed fluorene formation and was removable under Fmoc-like mild basic conditions post-transformation even though it was originally stable under strongly basic conditions. As an application for organic synthesis, the Epoc group provides the novel orthogonality of gold(III)-labile protecting groups in solid-phase peptide synthesis. In addition, the high turnover number of fluorene formation in aqueous media is suggestive of the applicability of the Epoc group to biological systems.

Introduction

The mild deprotection of the Fmoc protecting group is advantageous to avoid side reactions and to obtain free functional groups efficiently.¹ Thus, the Fmoc group has been applied to the synthesis of a wide variety of compounds including natural products, glycans, and peptides.^{2–5} This lability is also applicable to the controlled release of drugs *in vivo*; the modification of peptide drugs with an Fmoc structure prolongs their lifetime in circulation.^{6,7} In addition, some Fmoc-protected amino acids, peptides, and sugars have been used for low-molecular-weight gelators and applied as biomaterials.^{8–13} These compounds form fibrils with the π - π or CH- π stacking of fluorene rings, which results in the formation of hydrogels.

Although this base lability is useful, the Fmoc group is not tolerant to reactions using strong basic reagents. A protecting group that achieves both Fmoc-like mild deprotection and high stability would solve this problem. However, it is difficult to develop such a protecting group because the stability of the protecting group is generally inseparable from its harsh deprotection conditions.

The base lability of the Fmoc group originates from the aromaticity of fluorenes, which stabilizes its anion and accelerates E1cB elimination. The pK_a value of fluorene is 10 less than that of diphenylmethane (Fig. 1a).^{14,15} Thus, the

substitution of diarylmethane for fluorene in the Fmoc group increases stability. Based on this, we proposed the novel

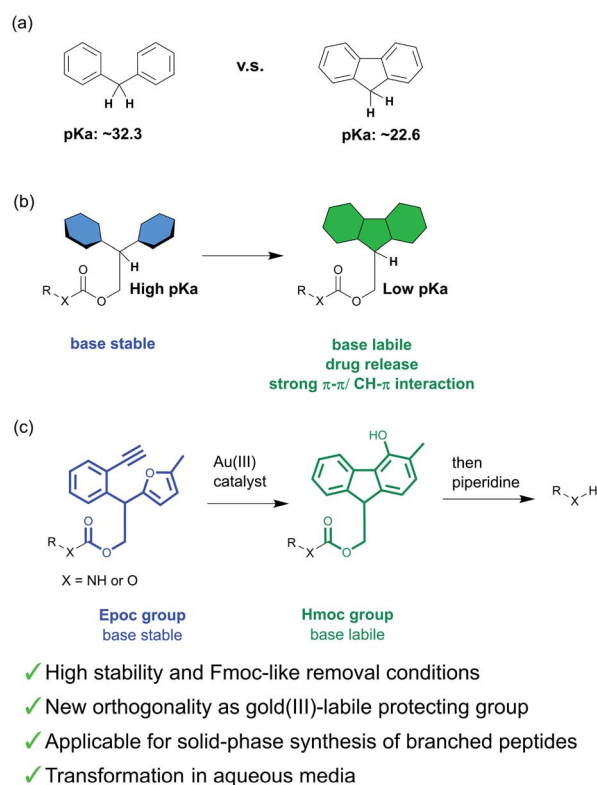


Fig. 1 (a) The difference in pK_a between diphenylmethane and fluorene. (b) Our strategy for the development of a transformable protecting group. (c) The novel protecting group developed in this work. The Epoc group is removed with a unique two-step procedure: gold(III)-catalyzed fluorene formation and base-induced E1cB elimination.

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concept of a transformable protecting group, which changes its properties through structural transformation (Fig. 1b). For example, if the protecting group with diarylmethane is transformable into an Fmoc-like structure, its lability will be changed by the transformation, and this protecting group will achieve both high stability (before the transformation) and Fmoc-like lability (after the transformation). This system can also be applied to biological systems. Recently, metal catalysts have been used for prodrug activation and fluorescence labeling in biological systems.^{16–20} If these catalysts perform the transformation *in vivo*, this could create potential new application of the Fmoc group in biological systems such as triggering drug release and hydrogelations.

For the fluorene formation of this design, we focused on the gold(III)-catalyzed^{21,22} phenol synthesis reported by Hashmi *et al.*^{23–27} They have reported that the furan-yne system tethered with an aromatic linker is transformed into 4-OH-fluorenol with gold(III) catalysts within 10 minutes.²⁸ Thus, we envisioned that the protecting group containing this sub-structure could meet our criteria for the design of a transformable protecting group.

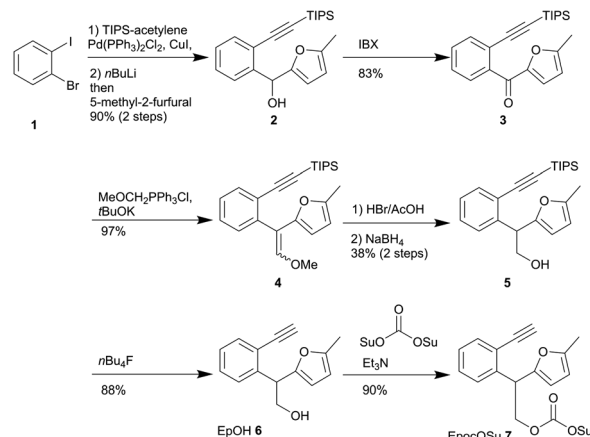
In this study, we developed a novel protecting group containing the precursor structure of fluorene, 2-(2-ethynylphenyl)-2-(5-methylfuran-2-yl)-ethoxycarbonyl (Epoc) (Fig. 1c), which is the first example of a transformable protecting group. The Epoc group is stable under strongly basic conditions. On the other hand, a catalytic amount of gold(III) salt transforms the Epoc group into a (4-hydroxy-3-methyl-9H-fluoren-9-yl)-methoxycarbonyl (Hmoc) group, and shows Fmoc-like lability. The gold(III)-labile protecting group has not been reported to the best of our knowledge, and thus this group provided the novel orthogonality of the protecting group strategy. The Epoc group was successfully applied for the synthesis of branched peptides on resins. In addition, the fluorene formation proceeded even in aqueous media, which is suggestive of the application of the Epoc group for controlling the function of the Fmoc group *in vivo*.

Results and discussions

Synthesis of reagents for Epoc protecting groups

To develop the Epoc protecting group explained above, we first established the synthetic route of the reagent for introducing the Epoc group to various functional groups (Scheme 1). The TIPS-ethynyl group was introduced to 2-bromo-1-iodobenzene **1** with Sonogashira coupling, followed by lithiation and nucleophilic attack of 5-methyl-2-furfural to obtain alcohol **2**. Ketone **3** was obtained *via* the oxidation of alcohol **2** with 2-iodoxybenzoic acid and the subsequent Wittig reaction gave enol ether **4**. After hydrolysis of enol ether **4** under acidic conditions to obtain the aldehyde, this aldehyde was subsequently reduced with sodium borohydride to yield alcohol **5** as a racemic mixture. The TIPS group of alcohol **5** was removed with tetrabutylammonium fluoride to obtain EpOH **6**.

EpOH **6** can be used for the introduction of the 2-(2-ethynylphenyl)-2-(5-methylfuran-2-yl)-ethyl (Ep) protecting group to carboxylic acid, which is transformed to a structure similar to the fluorenylmethyl (Fm) group by gold catalysis. EpocOSu **7**



Scheme 1 Synthesis of reagents for introducing Epoc group.

was prepared by the reaction of EpOH **6** with di(*N*-succinimidyl) carbonate, which can be used to introduce the Epoc group to amines.

Introduction and deprotection conditions for protecting group

Before investigating the applicability of the Epoc group for protecting groups, we sought suitable conditions for gold-catalyzed fluorene formation. The Epoc-protected aniline **8** (Table 1) was obtained in the reaction with EpOH **6** and phenyl isocyanate and used as the substrate for screening reaction conditions. A catalytic amount of commercially available

Table 1 Condition screening for fluorene formation

Entry	Solvent	NaAuCl ₄ ·2H ₂ O (mol%)	Yield ^a (%)
1	CH ₂ Cl ₂	10	>99
2	CH ₂ Cl ₂	1	>99
3	1,4-Dioxane	10	94
4	1,4-Dioxane	1	15
5	THF	10	0
6	Acetone	10	0
7	DMF	10	0
8	CH ₃ CN	10	0
9	DMSO	10	0

^a Yields determined by HPLC. The reactions were performed with 300 nmol of compound **8** in 200 μL of solvents (1.5 mM), at room temperature for 1 hour. The reactions were run in triplicate and the average yields were shown.



gold(III) salt, $\text{NaAuCl}_4 \cdot 2\text{H}_2\text{O}$, transformed the Epoc group into the Hmoc group. This reaction proceeded the most efficiently in CH_2Cl_2 , where 1 mol% of gold(III) catalyst smoothly transformed the Epoc group into the Hmoc group. Thus, we decided to use this solvent as the standard deprotection condition. Although the reaction proceeded in 1,4-dioxane, a larger amount of gold catalyst is needed for its completion. On the other hand, in other polar solvents such as THF, acetone, DMF, CH_3CN , and DMSO, this reaction did not proceed at all.

Next, we tested the reaction conditions for introducing the Epoc group (Scheme 2a). The introduction of the Epoc group into the amino group in compound **10** was successful with EpocOSu **7**, which is a general condition for introducing carbamate protecting groups into amino groups. Carboxylic acids can also be protected with the Ep group. The Ep group was introduced to the carboxylic acid of compound **12** by esterification with EpOH **6**. This is similar to the fluorenylmethyl (Fm) group, which is used for the protecting group of carboxylic acid. For the protection of alcohols with the Epoc group, the *in situ* formation of Epoc chloride with EpOH **6** and triphosgene and the subsequent addition of substrate **14** gave the best yield. The Epoc- and Ep- protected amino acids and sugars were obtained

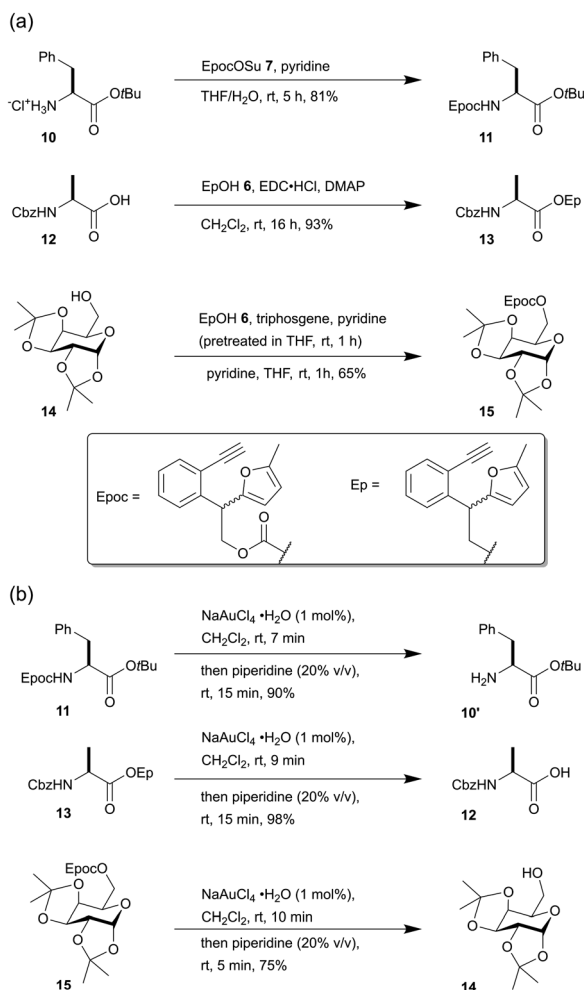
as the mixtures of diastereomers because the reagents for introducing the Epoc groups were obtained as racemic mixtures using the current technique.

The fluorene formation of the Epoc and Ep group in substrates **11**, **13**, **15** proceeded smoothly (within 10 minutes) with 1 mol% of gold(III) catalyst in CH_2Cl_2 (Scheme 2b). The transformed protecting groups were easily removed with the subsequent addition of piperidine in a one-pot procedure after the completion of the fluorene formation. Although the yield for the deprotection of substrate **15** was relatively low (75%) compared to substrate **11** and **13**, the fluorene formation was complete within 10 minutes. Production of a small amount of byproducts and loss of the product **14** during its purification decreased the overall yield. These results indicate that the Epoc and Ep groups are removed under mild conditions in almost the same manner as the Fmoc group.

Orthogonality of Epoc group

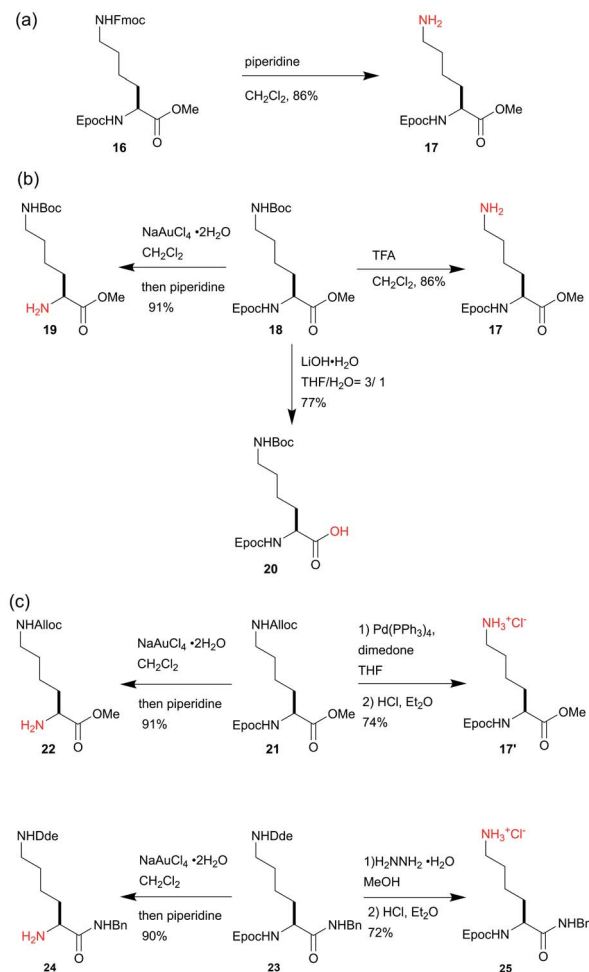
To demonstrate the stability and the mild removal conditions of the Epoc group, we tested the orthogonality of the Epoc group against other commonly used protecting groups. When compound **16**, which is protected with both the Epoc and the Fmoc groups, was treated with piperidine, the Fmoc group was chemoselectively removed (Scheme 3a). This indicates that the Epoc group is tolerant to the conditions of Fmoc deprotection, and thus can be applied to the orthogonal protecting group for Fmoc solid-phase peptide synthesis (SPPS) and automated glycan assembly. The selective deprotection experiments for compound **18**, possessing the Epoc, Boc, and ester groups, indicate that the Epoc group is also tolerant to acidic conditions for the Boc deprotection, and the conditions of the Epoc deprotection do not affect either the Boc or the ester groups (Scheme 3b). As expected, in the hydrolysis of methyl ester in compound **18** under strongly basic conditions, the Epoc group was stable and compound **20** was selectively obtained, where the Fmoc group is known to be labile. This indicates that the Epoc group achieves the criteria of our initial design for a transformable protecting group: high stability against bases and Fmoc-like lability post-transformation. Thus, the Epoc group will be useful for the protection of the amino group of compounds that are planned to undergo reactions under strongly basic conditions and require mild conditions for removal at the end of the synthetic route, where the Fmoc group is not applicable.

As well as the acid-labile and base-labile protecting groups, the Epoc group is orthogonal to the Pd-labile Alloc protecting group²⁹ and the hydrazine-labile Dde protecting group (Scheme 3c),³⁰ which are commonly used as the orthogonal protecting groups for amines. When PhSiH_3 was used as a nucleophilic scavenger, the Alloc deprotection of compound **21** was unsuccessful because of the undesired reduction of alkyne in the Epoc group. On the other hand, the use of dimedone instead improved this problem. The Alloc group was chemoselectively removed with a palladium catalyst and compound **17'** was obtained. The treatment of an excess amount of hydrazine to compound **23** gave compound **25**, where the Dde group was



Scheme 2 (a) Introduction and (b) removal of the Epoc and Ep groups.





Scheme 3 Orthogonality of the Epoc group. (a) Fmoc-selective deprotection. (b) Orthogonality of the Epoc group to acid- and base-labile protecting groups. (c) Orthogonality of Epoc group to Pd- and hydrazine-labile protecting groups.

selectively removed. Thus, the Epoc group was also stable under the conditions required for Dde removal. The Epoc group was selectively removed from compounds **21** and **23** to obtain compounds **22** and **24** respectively, indicating that the removal of the Epoc group requires relatively mild conditions to keep other protecting groups unaffected. These results indicate the orthogonality of the Epoc group against the Alloc and Dde groups. The Alloc and Dde groups have been used for the solid-phase synthesis of cyclic and branched peptides. Thus, the addition of the Epoc group to a series of these orthogonal protecting groups enables the synthesis of more complex cyclic and branched peptides on resins.

Application to solid-phase peptide synthesis

We tested whether the transformation from Epoc to Hmoc and its subsequent deprotection proceeded on resins using *N*-Epoc-valylphenylalanine bound to Trt(2-Cl) resin **26** (Fig. 2). The transformation from the Epoc to the Hmoc group proceeded smoothly even on resins with the treatment of 10 mol% of NaAuCl₄·2H₂O for 10 minutes. The resulting peptide on resin

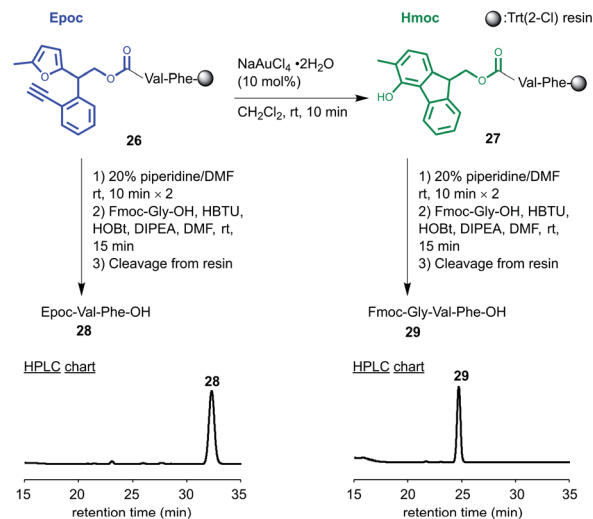


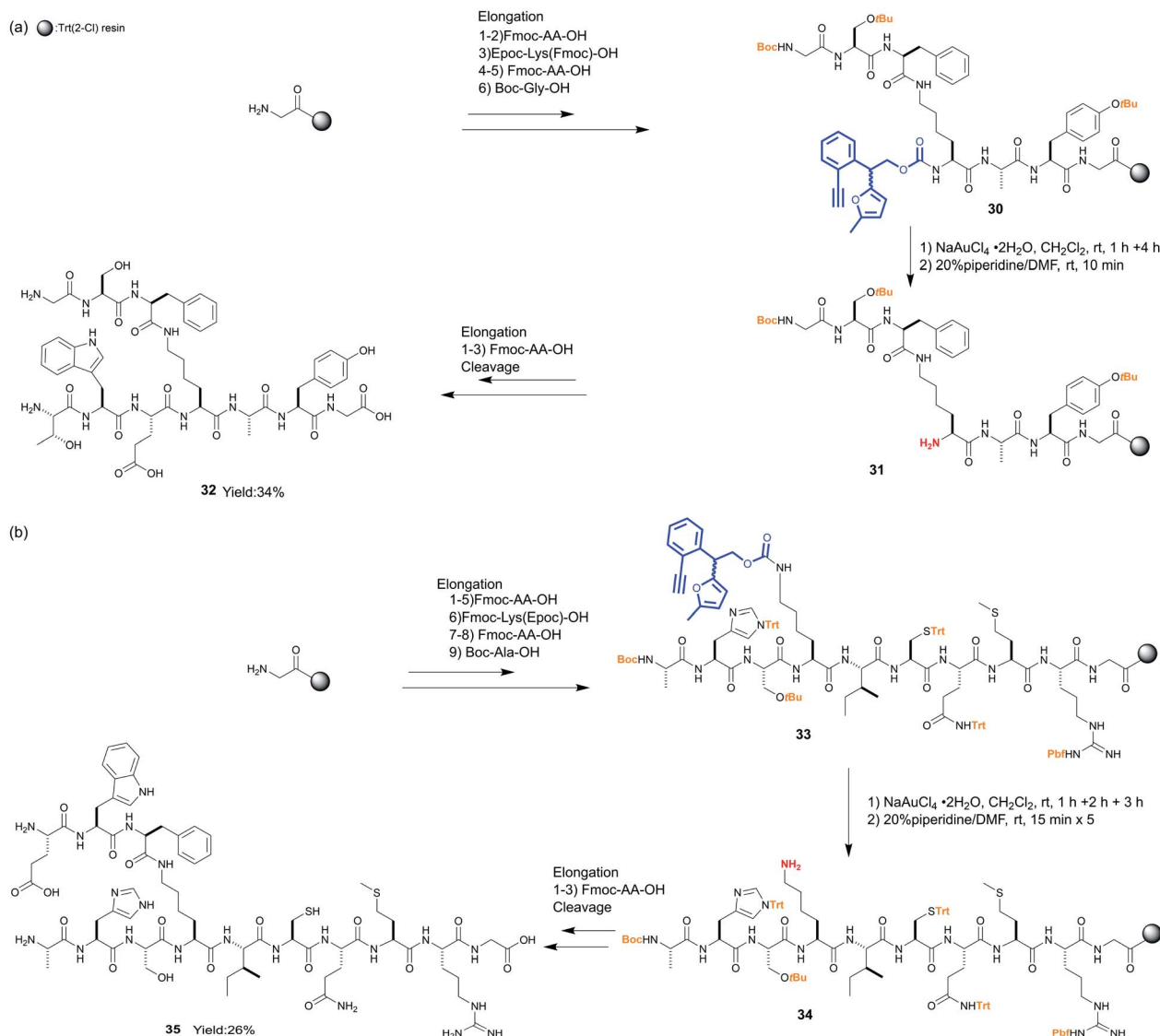
Fig. 2 Transformation and removal of the Epoc group on resin. HPLC charts of the cleaved peptide are shown. HPLC methods: Table S2.

27 was then treated with piperidine to remove the Hmoc group. After the coupling with Fmoc-Gly-OH and cleavage, Fmoc-Gly-Val-Phe-OH **29** was successfully obtained. This indicates that the deprotection of the Epoc group proceeds properly even on resins and the Epoc group works as a protecting group for solid-phase synthesis.

As a negative control, we treated a 20% piperidine/DMF solution and the coupling reagents with the same resin **26** without the transformation from the Epoc to the Hmoc group. The Epoc group did not react with piperidine or the coupling reagents without transformation into the Hmoc group, and Epoc-Val-Phe-OH **28** was selectively obtained after cleavage. In this experiment, the peptides other than Epoc-Val-Phe-OH **28**, such as the Fmoc-Gly-Val-Phe-OH **29** and other peptides derived from side reactions on the Epoc group were not obtained based on the HPLC chart (Fig. S5†). Therefore, the Epoc group is tolerant of the conditions for Fmoc removal and coupling on resins, and applicable to the orthogonal protecting group for the on-resin synthesis of peptides.

The use of the Epoc group for SPPS enabled the elongation of the branched peptides in two different directions with different amino acid sequences. In the first example, Epoc-Lys(Fmoc)-OH was incorporated into the elongation of the linear peptide with 7 amino acids on resin **30**, whose N-terminus was protected with the Boc group (Scheme 4a). Although the Epoc group in the middle part of peptides on the resin was less reactive compared to the case of *N*-Epoc-valylphenylalanine, the transformation into the Hmoc group proceeded with a longer reaction time (5 hours total). After the removal of the Hmoc group with piperidine to obtain the resin-bound peptide **31**, several amino acids were coupled from *N*α at lysine. After the cleavage from resin and HPLC purification, the branched peptide **32** was obtained in a 34% yield. Fmoc-Lys(Epoc)-OH was applied in the second example, and peptides with 10 amino acids on resin **33** were synthesized (Scheme 4b). The transformation and removal of the Epoc group proceeded well also in this peptide. After the





Scheme 4 Application of the Epc group as an orthogonal protecting group for branched peptide synthesis.

elongation from *N*ε at the lysine, peptides were cleaved and the branched peptide **35** was obtained at 26% yield after HPLC purification. In these syntheses, we confirmed that acid-labile protecting groups such as Trt, *t*Bu, Boc, and Pbf, which are commonly used as permanent protecting groups in SPPS, were not affected by treatment with NaAuCl₄·2H₂O. These experiments demonstrate that the Epc group is tolerant to multiple cycles of Fmoc deprotection and couplings. In addition, the transformation into the Hmoc group and its removal were shown to proceed even in the sterically hindered middle parts of peptides. Thus, the Epc group is applicable for the synthesis of complex branched or cyclic peptides on resins. Similar to the Fmoc group, the absorbance of the Hmoc-piperidine adduct cleaved from resins is detectable using UV spectroscopy, which helps monitor the completion of the deprotections. At this point, the Epc protecting group is more useful than other orthogonal protecting groups previously applied for the solid-phase synthesis of peptides and glycans.

Transformation of the Epc group under aqueous conditions

To demonstrate the applicability of the Epc group to biological systems, we performed the fluorene formation of the Epc group under aqueous conditions (Table 2). The fluorene formation of compound **8** proceeded in PBS buffer at 37 °C for 24 hours and gave 50% yield with 1 mol% of NaAuCl₄·2H₂O. The reaction itself proceeded even with a very small amount of gold(III) catalyst (0.1 mol%) with a turnover number of approximately 340.

These results indicate that the fluorene formation of the Epc group proceeds under conditions similar to biological settings, where a small concentration of gold catalysts exists in an aqueous solution. Although the gold catalysts, the substrate structure, pH of the solvent *etc.* have to be further optimized before its applications to biological systems, we were able to demonstrate that the transformation of the Epc group could be applied to trigger drug release and hydrogelations.

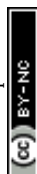
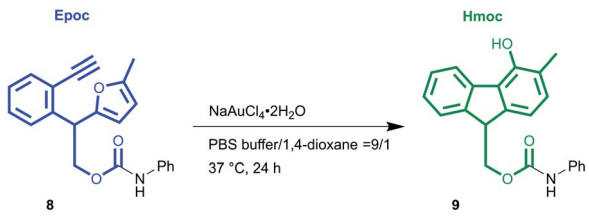


Table 2 Fluorene formation under aqueous conditions



Entry	NaAuCl ₄ ·2H ₂ O (mol%)	Yield ^a (%)
1	0.1	34
2	1	50

^a Yields were determined by HPLC. The reactions were performed with 300 nmol of compound **8** in 100 μ L of PBS buffer/1,4-dioxane = 9/1 (3.0 mM), at 37 °C for 24 hours. The reactions were run in triplicate and the average yields were shown.

Conclusions

In this study, we designed the novel concept of a transformable protecting group, which includes the transformation of the protecting group, changing its properties. To the best of our knowledge, a protecting group with this type of unique two-step deprotection procedure has not previously been developed. Based on this concept, we developed the Epoc and Ep groups, which transform from a stable to Fmoc-like labile structure with gold(III) catalysts. Because of the requirement for synthesizing complex molecules and constructing efficient synthetic strategies, the novel orthogonal protecting groups have still been developed recently such as reduction-labile, tetrazine-labile and Cu(I)-labile protecting groups.^{31–34} This work provided the novel orthogonality in the series of protecting groups as a gold(III)-labile protecting group with the extremely mild conditions for its removal. Its applicability to SPPS enables the efficient synthesis of branched and cyclic peptides including complex natural products, which require multiple orthogonal protecting groups.³⁵ This study also demonstrated that the transformation of the Epoc group proceeds in aqueous media. Thus, the Epoc group is expected to be useful for future applications in biological systems for prodrugs and hydrogelators.

Data availability

The experimental procedure, characterization data, and NMR spectra are provided in the ESI.

Author contributions

Conceptualization: T. Y. and K. T.; funding acquisition: K. T.; investigation: T. Y. and T.-C. C.; writing-original draft preparation: T. Y.; writing-review and editing T. Y., T.-C. C., and K. T.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 L. A. Carpino and G. Y. Han, *J. Org. Chem.*, 1972, **37**, 3404–3409.
- 2 C.-D. Chang and J. Meienhofer, *Int. J. Pept. Protein Res.*, 1978, **11**, 246–249.
- 3 E. Atherton, H. Fox, D. Harkiss, C. J. Logan, R. C. Sheppard and B. J. Williams, *J. Chem. Soc., Chem. Commun.*, 1978, 537–539.
- 4 L. Kröck, D. Esposito, B. Castagner, C.-C. Wang, P. Bindschädler and P. H. Seeberger, *Chem. Sci.*, 2012, **3**, 1617–1622.
- 5 P. H. Seeberger, *Acc. Chem. Res.*, 2015, **48**, 1450–1463.
- 6 E. Gershonov, I. Goldwasser, M. Fridkin and Y. Shechter, *J. Med. Chem.*, 2000, **43**, 2530–2537.
- 7 Y. Shechter, L. Preciado-Patt, G. Schreiber and M. Fridkin, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 1212–1217.
- 8 Z. Yang, H. Gu, D. Fu, P. Gao, J. K. Lam and B. Xu, *Adv. Mater.*, 2004, **16**, 1440–1444.
- 9 A. Mahler, M. Reches, M. Rechter, S. Cohen and E. Gazit, *Adv. Mater.*, 2006, **18**, 1365–1370.
- 10 V. Jayawarna, M. Ali, T. A. Jowitt, A. F. Miller, A. Saiani, J. E. Gough and R. V. Ulijn, *Adv. Mater.*, 2006, **18**, 611–614.
- 11 R. A. Pires, Y. M. Abul-Haija, D. S. Costa, R. Novoa-Carballal, R. L. Reis, R. V. Ulijn and I. Pashkuleva, *J. Am. Chem. Soc.*, 2015, **137**, 576–579.
- 12 C. Diaferia, G. Morelli and A. Accardo, *J. Mater. Chem. B*, 2019, **7**, 5142–5155.
- 13 P. Chakraborty, Y. Tang, T. Yamamoto, Y. Yao, T. Guterman, S. Zilberzwige-Tal, N. Adadi, W. Ji, T. Dvir, A. Ramamoorthy, G. Wei and E. Gazit, *Adv. Mater.*, 2020, **32**, 1906043.
- 14 F. G. Bordwell, W. S. Matthews and N. R. Vanier, *J. Am. Chem. Soc.*, 1975, **97**, 442–443.
- 15 W. S. Matthews, J. E. Bares, J. E. Bartmess, F. G. Bordwell, F. J. Cornforth, G. E. Drucker, Z. Margolin, R. J. McCallum, G. J. McCollum and N. R. Vanier, *J. Am. Chem. Soc.*, 1975, **97**, 7006–7014.
- 16 P. Destito, C. Vidal, F. López and J. L. Mascareñas, *Chem.–Eur. J.*, 2021, **27**, 4789–4816.



- 17 J. Clavadetscher, E. Indrigo, S. V. Chankeshwara, A. Lilienkamp and M. Bradley, *Angew. Chem., Int. Ed.*, 2017, **56**, 6864–6868.
- 18 K. Tsubokura, K. K. H. Vong, A. R. Pradipta, A. Ogura, S. Urano, T. Tahara, S. Nozaki, H. Onoe, Y. Nakao, R. Sibgatullina, A. Kurbangaliev, Y. Watanabe and K. Tanaka, *Angew. Chem., Int. Ed.*, 2017, **56**, 3579–3584.
- 19 M. A. Miller, B. Askevold, H. Mikula, R. H. Kohler, D. Pirovich and R. Weissleder, *Nat. Commun.*, 2017, **8**, 15906.
- 20 K. Vong, T. Tahara, S. Urano, I. Nasibullin, K. Tsubokura, Y. Nakao, A. Kurbangaliev, H. Onoe, Y. Watanabe and K. Tanaka, *Sci. Adv.*, 2021, **7**, eabg4038.
- 21 A. S. K. Hashmi and G. J. Hutchings, *Angew. Chem., Int. Ed.*, 2006, **45**, 7896–7936.
- 22 D. Pflästerer and A. S. K. Hashmi, *Chem. Soc. Rev.*, 2016, **45**, 1331–1367.
- 23 A. S. K. Hashmi, T. M. Frost and J. W. Bats, *J. Am. Chem. Soc.*, 2000, **122**, 11553–11554.
- 24 A. S. K. Hashmi, J. P. Weyrauch, M. Rudolph and E. Kurpejović, *Angew. Chem., Int. Ed.*, 2004, **43**, 6545–6547.
- 25 A. S. K. Hashmi, M. Rudolph, J. P. Weyrauch, M. Wölfl, W. Frey and J. W. Bats, *Angew. Chem., Int. Ed.*, 2005, **44**, 2798–2801.
- 26 A. S. K. Hashmi, R. Salathé and W. Frey, *Chem.–Eur. J.*, 2006, **12**, 6991–6996.
- 27 A. S. K. Hashmi, M. Rudolph, H.-U. Siehl, M. Tanaka, J. W. Bats and W. Frey, *Chem.–Eur. J.*, 2008, **14**, 3703–3708.
- 28 A. S. K. Hashmi, J. Hofmann, S. Shi, A. Schütz, M. Rudolph, C. Lothschütz, M. Wietek, M. Bührle, M. Wölfl and F. Rominger, *Chem.–Eur. J.*, 2013, **19**, 382–389.
- 29 F. Guibé, *Tetrahedron*, 1998, **54**, 2967–3042.
- 30 B. W. Bycroft, W. C. Chan, S. R. Chhabra and N. D. Hone, *J. Chem. Soc., Chem. Commun.*, 1993, 778–779.
- 31 M. Staderini, A. Gambardella, A. Lilienkamp and M. Bradley, *Org. Lett.*, 2018, **20**, 3170–3173.
- 32 S. Chithanna, S. Vyasamudri and D.-Y. Yang, *Org. Lett.*, 2020, **22**, 2391–2395.
- 33 H. Liu, S.-Y. Zhou, G.-E. Wen, X.-X. Liu, D.-Y. Liu, Q.-J. Zhang, R. R. Schmidt and J.-S. Sun, *Org. Lett.*, 2019, **21**, 8049–8052.
- 34 A. Kumar, V. Gannedi, S. A. Rather, R. A. Vishwakarma and Q. N. Ahmed, *J. Org. Chem.*, 2019, **84**, 4131–4148.
- 35 H. Itoh and M. Inoue, *Org. Biomol. Chem.*, 2019, **17**, 6519–6527.

