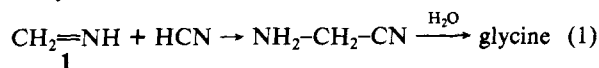
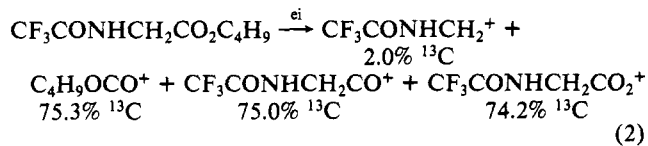


formed by the addition of HCN to **1** (eq 1). For further con-



firmation of this mechanism, a mixture of H^{13}CN and NH_3 was reacted with carbon vapor. Hydrolysis of the product mixture resulted in glycine, which was derivatized and analyzed by GC-MS. The mass of the various fragments gave the ^{13}C distribution shown in eq 2. These results demonstrate that the ^{13}C is in-



corporated almost exclusively into the carboxyl group of glycine as demanded by the mechanism in eq 1 and constitute further proof for the intermediacy of **1**.

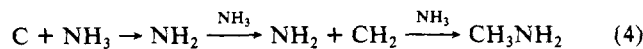
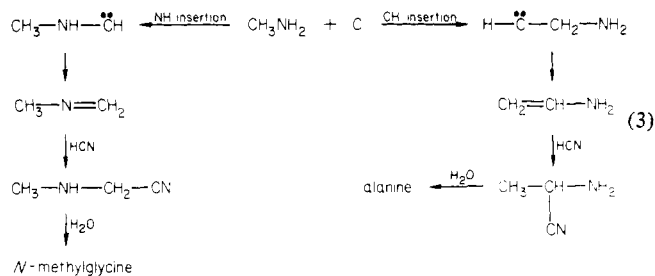
Polymeric HCN as Possible Amino Acid Precursor. The ^{13}C labeling studies also serve to rule out a mechanism for glycine formation involving the hydrolysis of an HCN polymer formed under the reaction conditions. The hydrolysis of HCN polymers and oligomers has been shown to produce amino acids.¹³ However, if HCN were to polymerize under the conditions of the present reaction, hydrolysis of the polymer would generate glycine in which both carbons had incorporated ^{13}C . Equation 2 shows that this is clearly not the case and that no more than 2.6% of the glycine could have arisen from an HCN polymer. When carbon vapor was reacted with HCN alone, only a very small amount of glycine was generated.

In another attempt to evaluate the importance of an HCN polymerization mechanism for the formation of the amino acids, we have reacted ND_3 with carbon vapor. It has been shown that glycine, generated by D_2O hydrolysis of an HCN polymer, incorporates two deuteriums in the methylene group.¹⁴ Hence, amino acids generated by the reaction of carbon with ND_3 and subsequent hydrolysis of the DCN polymer should contain no deuterium. In fact, GC-MS analysis reveals that all the amino acids generated in this experiment were deuterated. Thus we conclude that these amino acids do not have polymeric HCN as their sole precursor.

Formation of Alanine and *N*-Methylglycine. Since amino acid yields are rather variable and appear to depend upon factors that are difficult to control, such as the rate of carbon vaporization, we have normalized the yield of each amino acid to that of glycine in Table II. In this manner, we are able to carry out a number of experiments and obtain a reasonable estimate of experimental errors. Since the mechanism of glycine formation is established, a measure of the change in the ratio of a particular amino acid to glycine in the presence of various additives should provide evidence concerning the mechanism of formation of that amino acid. Absolute yields of amino acids from typical runs are listed in the Experimental Section.

Alanine and *N*-methylglycine contain an additional carbon and are postulated to result from the reaction of carbon with methylamine followed by HCN addition as shown in eq 3.

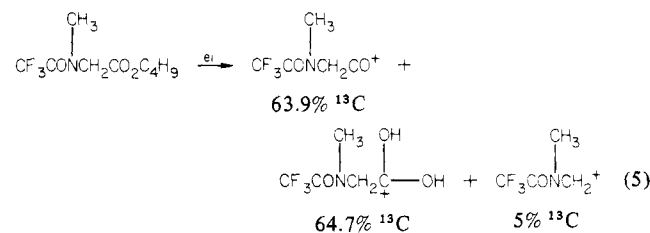
Table I shows that methylamine is formed in substantial yield in our system; it is also reported in the reaction of nucleogenic carbon with ammonia.¹¹ This product is most probably the result of the reaction of CH_2 with NH_3 . The CH_2 , in turn, is formed by a series of hydrogen abstractions by C_1 (eq 4), of ammonia



to generate methylamine has been reported.¹⁶ An alternate mechanism involving a series of hydrogen abstractions by aminomethylene, **2**, appears unlikely for thermodynamic reasons.

Carbon vapor was reacted with a mixture of NH_3 and CH_3NH_2 to further establish the role of methylamine in the formation of *N*-methylglycine and alanine. This reaction resulted in a substantial increase in the ratio of both alanine and *N*-methylglycine to that of glycine (Table II).

When carbon vapor was reacted with a mixture CH_3NH_2 and H^{13}CN , the mass spectrum of *N*-methylglycine derivative showed the majority of the ^{13}C in the carboxy group (eq 5). Although



the mass spectrum of the alanine derivative did not show fragments containing the carboxyl group, the $\text{CF}_3\text{CONHCH}_2^+$ fragment of $m/e = 140$ showed only 7.7% ^{13}C incorporation. These labeling studies support the mechanisms for the formation of alanine and *N*-methylglycine in eq 3.

Since CH_3NH_2 is formed by a series of hydrogen abstractions, the addition of a better hydrogen atom donor than NH_3 ($D_{\text{N-H}} = 109$ kcal/mol) should increase the yield of alanine and *N*-methylglycine relative to that of glycine. For simplicity, we have chosen H_2 ($D_{\text{H-H}} = 104$ kcal/mol) as a hydrogen donor. Table II demonstrates that inclusion of H_2 among the reactants increases the ratio of both alanine and *N*-methylglycine to that of glycine by a factor of 2.

The substitution of D_2 for H_2 results in substantial deuterium incorporation in the alanine and *N*-methylglycine as shown in Table III. The fact that d_1 as well as d_2 amino acids are produced indicates that hydrogen abstraction by carbon proceeds in a stepwise manner through the intermediacy of CH . We have observed a similar effect in the products of the reaction between carbon and a mixture of benzene and benzene- d_6 .¹⁵

It is interesting that the inclusion of D_2 as a reactant results in the formation of glycine and β -alanine with the incorporation of small amounts of d_1 . This deuterium incorporation may result from a competing reaction of CD with NH_3 to generate **1** and a hydrogen atom (eq 6) in a reaction analogous to that of CH with CH_4 that produces ethylene and a hydrogen atom.¹⁷



Precursors to the Remaining Amino Acids. Although mechanistic sequences leading to the precursors of β -alanine, aspartic acid, and serine have not been elucidated, several observations may be relevant. Since the above amino acid precursors are secondary products, it may be that they arise from reactions

(13) (a) Lemmon, R. M. *Chem. Rev.* **1970**, *70*, 95-109. (b) Abelson, P. H. *Proc. Natl. Acad. Sci. U.S.A.* **1966**, *55*, 1365-1372. (c) Oro, J.; Kamat, S. S. *Nature (London)* **1961**, *190*, 442-443. (d) Lowe, C. U.; Rees, M. W.; Markham, R. *Ibid.* **1963**, *199*, 219-222. (e) Matthews, C. N.; Moser, R. E. *Ibid.* **1967**, *215*, 1230-1234. (f) Moser, R. E.; Claggett, A. R.; Matthews, C. N. *Tetrahedron Lett.* **1968**, 1599-1603, 1605-1608. (g) Harada, K. *Nature (London)* **1967**, *214*, 479-480. (h) Ferris, J. P.; Joshi, P. C.; Edelson, E. H.; Lawless, J. G. *J. Mol. Evol.* **1978**, *11*, 293-311.

(14) Matthews, C.; Nelson, J.; Varma, P.; Minard, R. *Science (Washington, D.C.)* **1977**, *198*, 622-625.

(15) Biesiada, K. A.; Koch, C. T.; Shevlin, P. B. *J. Am. Chem. Soc.* **1980**, *102*, 2098-2100.

(16) Chao, K. J.; Lin, C. L.; Hsu, M.; Ho, S. Y. *J. Phys. Chem.* **1979**, *83*, 1241-1243.

(17) Braun, W.; McNesby, J. R.; Bass, A. M. *J. Chem. Phys.* **1967**, *46*, 2071-2080.

alcohol on methanolic NaOH washed firebrick (60/80 mesh) glass column.²³

Reactor. The reactor is modeled after that described by Skell et al.²⁴ Carbon is vaporized by striking an intermittent arc between two graphite rods attached to water-cooled brass electrodes and cocondensed on the walls of the reactor at $-196\text{ }^{\circ}\text{C}$ ($\sim 10^{-4}$ torr) with a reactant. The power source is a Sears Dual Range 295 amp arc welder set at 110 amps. The inlet and outlet tubes of the reactor are attached to a vacuum line.

Reaction of Carbon Vapor with Ammonia. In a typical reaction, ammonia (103.6 mmol) is introduced into the reactor through a vacuum line at a flow rate of ~ 0.19 mmol/s and cocondensed on the walls of the reactor at $-196\text{ }^{\circ}\text{C}$ with carbon vapor (88.0 mmol). If more than one reactant gas is used, for example, ammonia and methylamine, the two gases are allowed to mix for 1 h in the vacuum line before the reaction is run. After all the ammonia is condensed onto the walls of the reactor, the carbon vapor is deposited for an additional 3 min. The reactor is allowed to warm to room temperature for 0.5 h and the volatile products are pumped out through traps at -78 and $-196\text{ }^{\circ}\text{C}$. These volatile products are then analyzed by GC-MS for methylamine (2.5 mmol), by ^1H NMR for acetonitrile (4.6×10^{-1} mmol), and by IR for HCN (0.87 mmol) and $\text{HC}\equiv\text{CH}$ (1.1×10^{-1} mmol).

The residue in the reactor is taken up in 6 N HCl and hydrolyzed at $100\text{ }^{\circ}\text{C}$ for 24 h. The hydrolysate is filtered, evaporated, and derivatized to the *N*-trifluoroacetyl *n*-butyl esters of the amino acids, by the method of Roach and Gehrke,⁸ and analyzed by GC-MS.⁹ The amino acids detected are alanine (2.7×10^{-2} mmol), glycine (1.5×10^{-1} mmol), *N*-methylglycine (3.2×10^{-2} mmol), β -alanine (1.9×10^{-2} mmol), *N*-methylalanine (trace), and aspartic acid (2.2×10^{-3} mmol). Table II shows the average yields of these amino acids from nine runs, normalized to that of glycine, along with their standard deviations.

Reaction of Carbon Vapor with Aqueous Ammonia. The reaction was carried out as described above with the cocondensation of NH_3 (100.7 mmol), H_2O (55.6 mmol), and carbon vapor (64.2 mmol). Analysis of the residue for amino acids revealed the presence of alanine (8.6×10^{-3} mmol), glycine (1.1×10^{-1} mmol), *N*-methylglycine (1.1×10^{-2} mmol), β -alanine (7.5×10^{-3} mmol), *N*-methylalanine (trace), aspartic acid (2.6×10^{-3} mmol), and serine (1.1×10^{-2} mmol).

Reaction of Carbon Vapor with Preformed Ammonia and Hydrogen Cyanide Surfaces. Ammonia (93.2 mmol) was first condensed on the walls of the reactor at $-196\text{ }^{\circ}\text{C}$ in increments of ~ 20.0 mmol. In between the deposition of ammonia, carbon vapor was condensed on the walls for ~ 3 min. After the final aliquot of ammonia had been condensed, the carbon vapor was deposited for an additional 5 min. Total amount of carbon vapor condensed was 64.1 mmol. The reactor was then allowed to warm to room temperature for ~ 0.5 h, and the following amino acids were analyzed as above: alanine (9.1×10^{-3} mmol), glycine (1.2×10^{-1} mmol), *N*-methylglycine (1.3×10^{-2} mmol), β -alanine (1.0×10^{-2} mmol), *N*-methylalanine (trace), and aspartic acid (2.4×10^{-3} mmol).

The same procedure was followed for the reaction of carbon vapor (53.2 mmol) with a preformed hydrogen cyanide (60.0 mmol) surface. The only amino acids detected were glycine (7.66×10^{-3} mmol) and β -alanine (2.58×10^{-3} mmol).

Reaction of Carbon Vapor with Ammonia and Hydrogen (or Deuterium). Ammonia (79.9 mmol) and hydrogen (20.6 mmol) were allowed to mix for 1 h in a vacuum line. The gas mixture was then cocondensed with carbon vapor (133.0 mmol) at $-196\text{ }^{\circ}\text{C}$ at a flow rate of ~ 0.19 mmol/s to keep the pressure low. After the reactor had warmed to room temperature for ~ 0.5 h the volatile products were analyzed for HCN (3.5 mmol) and acetylene (3.5 mmol). The nonvolatile products were hydrolyzed and analyzed as above. The following amino acids were

detected: alanine (3.55×10^{-2} mmol), glycine (3.63×10^{-1} mmol), β -alanine (1.99×10^{-2}), and *N*-methylglycine (8.43×10^{-2} mmol). Average normalized amino acid yields from two runs are listed Table II. When the hydrogen was replaced by deuterium, the amount of deuterium incorporation (Table III) was evaluated from the following mass spectral fragments: glycine ($\text{CF}_3\text{CONHCH}_2^+$), alanine ($\text{CF}_3\text{CONHCHCH}_3^+$), *N*-methylglycine ($\text{CF}_3\text{CONCH}_3\text{CH}_2^+$), and β -alanine ($\text{CF}_3\text{CONHCH}_2\text{CH}_2^+$).

Reaction of Carbon Vapor with Ammonia and HCN (or H^{13}CN). Hydrogen cyanide was prepared by the reaction between sodium or potassium cyanide and concentrated sulfuric acid and checked for purity by IR spectroscopy. Ammonia (15.8 mmol) and the hydrogen cyanide (6.1 mmol) were allowed to mix for 1 h. The gas mixture was then cocondensed with carbon vapor (67.9 mmol) as above. Analysis of the residue for amino acids revealed the presence of glycine (4.02×10^{-1} mmol), alanine (7.85×10^{-2} mmol), *N*-methylglycine (7.36×10^{-2} mmol), β -alanine (8.9×10^{-2} mmol), and aspartic acid (1.18×10^{-2} mmol). Average normalized amino acid yields from four runs are listed in Table II. When H^{13}CN was used, the ^{13}C content of the glycine was determined from the mass spectral fragments in eq 5.

Reaction of Carbon Vapor with Deuterated Ammonia. The ND_3 was prepared by reacting D_2O with Mg_3N_2 on a vacuum line.²⁵ Infrared analysis of the ammonia showed mainly ND_3 with a small amount of ND_2H and no detectable NH_2D or NH_3 .²⁶ The ND_3 (51.6 mmol) was reacted with carbon vapor (89.7 mmol) as above. GC-MS analysis revealed the following amino acids: glycine (1.04×10^{-1} mmol, 59.4% d_2), alanine (1.51×10^{-2} mmol, 38.0% d_4), *N*-methylglycine (3.08×10^{-2} mmol, 30.8% d_5), β -alanine (7.66×10^{-3} mmol, 31.0% d_4), and aspartic acid (6.68×10^{-3} mmol, 5.2% d_2 , 20.1% d_1 , 74.7% d_0).

Reaction of Carbon Vapor with Methylamine and Ammonia. The reaction was carried out in the usual manner by cocondensing carbon (116.7 mmol) with a mixture of NH_3 (81.1 mmol) and CH_3NH_2 (22.4 mmol). The following amino acids were detected: glycine (1.55×10^{-1} mmol), alanine (3.63×10^{-2} mmol), *N*-methylglycine (5.34×10^{-2} mmol), β -alanine (7.86×10^{-3} mmol), aspartic acid (5.62×10^{-3} mmol), and *N*-methylalanine (7.76×10^{-2} mmol).

Reaction of Carbon Vapor with CH_3NH_2 and H^{13}CN . Carbon vapor (132.8 mmol) was cocondensed with CH_3NH_2 (77.9 mmol) and H^{13}CN (16.7 mmol) in the usual manner. Analysis of the residue revealed glycine (6.6×10^{-2} mmol), alanine (3.5×10^{-2} mmol), *N*-methylglycine (1.55×10^{-1} mmol), and *N*-methylalanine (6.7×10^{-2} mmol). Mass spectral analysis indicated that the glycine contained 43.9% ^{13}C on the carboxyl group and 7.8% on the methylene group. The *N*-methylglycine showed 64.3% ^{13}C on the carboxyl group and 5% in the remainder of the molecule. The mass spectrum of the alanine and the *N*-methylalanine did not show fragments containing the carboxyl group. The ^{13}C content in the decarboxylated fragments was 7.7% in the case of alanine and 1.4% in the case of *N*-methylalanine.

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Registry No. 1, 2053-29-4; 2, 35430-17-2; 3, 540-61-4; 6, 17619-22-6; 7, 75-05-8; 8, 4363-36-4; 9, 69245-10-9; methylamine, 74-89-5; carbon, 7440-44-0; ammonia, 7664-41-7; glycine, 56-40-6; DL-alanine, 302-72-7; *N*-methylglycine, 107-97-1; β -alanine, 107-95-9; DL-aspartic acid, 617-45-8; *N*-methyl-DL-alanine, 600-21-5; DL-serine, 302-84-1; hydrogen cyanide, 74-90-8; hydrogen, 1333-74-0; deuterium, 7782-39-0; glycine- d_2 , 84009-43-8; alanine- d_4 , 83998-91-8; *N*-methylglycine- d_5 , 83998-92-9; β -alanine- d_4 , 83998-93-0; aspartic acid- d , 83998-94-1; H^{13}CN , 56162-23-3; ND_3 , 13550-49-7.

(23) Yang, J. Y.; Wolf, A. P. *J. Am. Chem. Soc.* **1960**, *82*, 4488-4492.

(24) Skell, P. S.; Wescott, L. D., Jr.; Golstein, J. P.; Engel, R. R. *J. Am. Chem. Soc.* **1965**, *87*, 2829-2835.

(25) Krannich, L. K.; Thewalt, U.; Cook, W. J.; Jain, S. R.; Sisler, H. H. *Inorg. Chem.* **1973**, *12*, 2304-2313.

(26) Halverson, F. *Rev. Mod. Phys.* **1947**, *19*, 87-131.