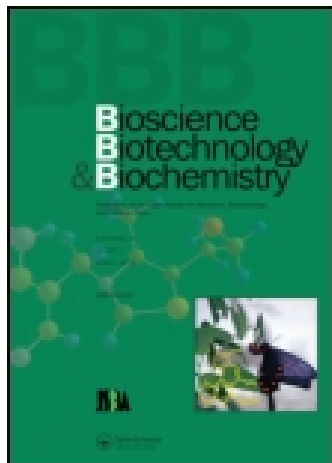


This article was downloaded by: [University of Hong Kong Libraries]

On: 15 November 2014, At: 20:04

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Bioscience, Biotechnology, and Biochemistry

Publication details, including instructions for authors and subscription information:  
<http://www.tandfonline.com/loi/tbbb20>

### Lipase-Catalyzed Kinetic Resolution of ( $\pm$ )-cis-Flavan-4-ol and Its Acetate: Synthesis of Chiral 3-Hydroxyflavanones

Tamotsu TODOROKI<sup>a</sup>, Akiko SAITO<sup>b</sup> & Akira TANAKA<sup>a</sup>

<sup>a</sup> College of Technology, Toyama Prefectural University Kosugi, Toyama 939-0398, Japan

<sup>b</sup> Japan Science and Technology Corporation, Domestic Research Fellow Kosugi, Toyama 939-0398, Japan

Published online: 22 May 2014.

To cite this article: Tamotsu TODOROKI, Akiko SAITO & Akira TANAKA (2014) Lipase-Catalyzed Kinetic Resolution of ( $\pm$ )-cis-Flavan-4-ol and Its Acetate: Synthesis of Chiral 3-Hydroxyflavanones, *Bioscience, Biotechnology, and Biochemistry*, 66:8, 1772-1774, DOI: [10.1271/bbb.66.1772](https://doi.org/10.1271/bbb.66.1772)

To link to this article: <http://dx.doi.org/10.1271/bbb.66.1772>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

## Note

## Lipase-Catalyzed Kinetic Resolution of ( $\pm$ )-*cis*-Flavan-4-ol and Its Acetate: Synthesis of Chiral 3-Hydroxyflavanones

Tamotsu TODOROKI,<sup>1</sup> Akiko SAITO,<sup>2</sup> and Akira TANAKA<sup>1,†</sup>

<sup>1</sup>College of Technology, Toyama Prefectural University, Kosugi, Toyama 939-0398, Japan

<sup>2</sup>Japan Science and Technology Corporation, Domestic Research Fellow, Kosugi, Toyama 939-0398, Japan

Received February 6, 2002; Accepted March 25, 2002

**Lipase-catalyzed kinetic resolution of ( $\pm$ )-*cis*-flavan-4-ol and its acetate led to enantiomerically enriched flavan-4-ol and its acetate. These chiral compounds were converted to (2*R*, 3*R*)- and (2*S*, 3*S*)-3-hydroxyflavanones.**

**Key words:** kinetic resolution; lipase; 3-hydroxyflavanone

3-Hydroxyflavanones are widely distributed in the plant kingdom<sup>1)</sup> and some of them show interesting biological activities involving a hepatoprotective effect. In association with the synthetic work on condensed tannins,<sup>1,2)</sup> we needed chiral 3-hydroxyflavanones which could be promising monomeric sources for the synthesis of condensed tannins having the oligomeric structure of flavan-3-ols. Three methods for accessing chiral 3-hydroxyflavanones have been reported to date: 1) enzymatic resolution of racemic 3-hydroxyflavanone,<sup>3)</sup> 2) conversion of the epoxy ketones obtained by asymmetric epoxidation of chalcone derivatives,<sup>4,5)</sup> 3) synthesis *via* asymmetric dihydroxylation of cinnamic esters.<sup>6)</sup> These methods, however, involve the disadvantages of low enantiomeric purity of the products and multi-step reaction sequences. We have recently developed a novel approach to accessing enantiomerically enriched 3-hydroxyflavanones.

We have observed earlier that treatment of the enolate derived from racemic flavanone **1** with 2-(phenylsulfonyl)-3-phenyloxaziridine<sup>7)</sup> below  $-80^{\circ}\text{C}$  gave 3-hydroxyflavanone **2** in a good yield (Fig. 3). In this case, temperature control was crucial, because elevation of the reaction temperature caused easy opening of the pyranone ring. This useful experimental technique that could afford 3-hydroxyflavanones from flavanones in high yields required the development of a more efficient method for the preparation of enantiomerically enriched flavanones. In this paper, we report the synthesis of enantiomerically enriched 3-hydroxyflavanones *via* lipase-catalyzed kinetic resolution of ( $\pm$ )-*cis*-flavan-4-ol (**3**) and its acetate **4**.

In 1992, Izumi *et al.*<sup>8)</sup> reported that the kinetic resolution of ( $\pm$ )-*cis*-flavan-4-ol acetate (**4**) with lipase PS (Amano) resulted in the formation of (2*R*, 4*R*)-**3** (93% ee) and (2*S*, 4*S*)-**4** (95% ee). On the other hand, the transesterification of ( $\pm$ )-**3** with the same enzyme, using vinyl acetate as the acyl donor afforded acetate **4**, but with no optical rotation. In order to further examine the potent resolving ability of lipases, we conducted a kinetic resolution experiment on some lipases (lipase PS, lipase M, lipase A, lipase R and lipase AY). Among them, lipase AY (Amano) was the most efficient enzyme for both kinetic hydrolysis and transesterification.

When ( $\pm$ )-**3** was treated with lipase AY and vinyl acetate at  $25^{\circ}\text{C}$  for 6 d, (2*R*, 4*R*)-**4** (44% yield, 93% ee) and unreacted (2*S*, 4*S*)-**3** (55% yield, 61% ee) were obtained (Fig. 1). (2*S*, 4*S*)-**3** was oxidized to (*S*)-flavanone (*S*)-**1** with PCC/ $\text{CH}_2\text{Cl}_2$  to determine the enantiomeric purity by HPLC. Similarly, (2*R*, 4*R*)-**4** was converted to (*R*)-**1** *via* hydrolysis ( $\text{K}_2\text{CO}_3/\text{CH}_3\text{OH}$ ) and subsequent oxidation.

The kinetic hydrolysis of ( $\pm$ )-*cis*-flavan-4-ol acetate **4** in a phosphate buffer (pH 6.8) with the same enzyme gave (2*R*, 4*R*)-**3** (40% yield, 96% ee)

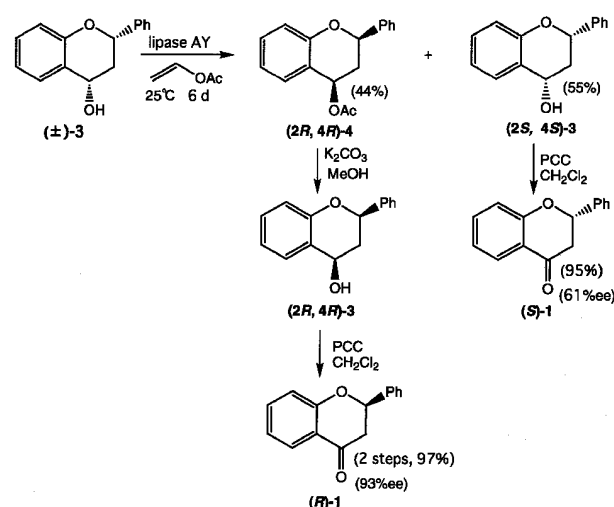
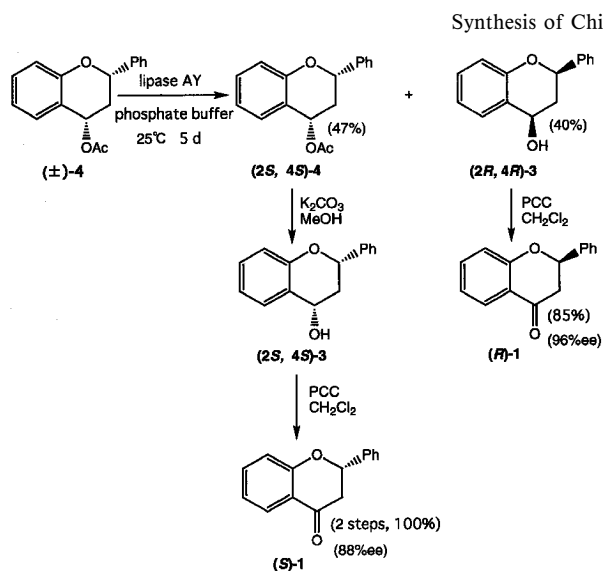


Fig. 1.

<sup>†</sup> To whom correspondence should be addressed. Fax: +81-766-56-0396; E-mail: atanaka@pu-toyama.ac.jp



and unreacted (2*S*, 4*S*)-4 (47% yield, 88% ee) (Fig. 2). The ee values of these compounds were also determined by an HPLC analysis of (*R*)- and (*S*)-flavanones (*R*)-1 and (*S*)-1 obtained by the same process as that just described.

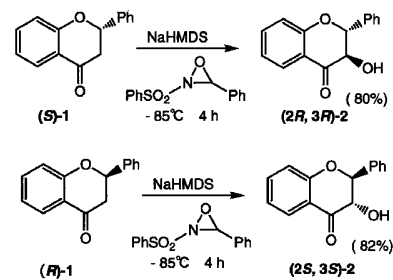
Finally, the enantiomerically enriched (*S*)- and (*R*)-flavanones (1) were transformed into (2*R*, 3*R*)- and (2*S*, 3*S*)-3-hydroxyflavanones (2), respectively (Fig. 3). Thus, the reaction between the enolate derived from (*S*)- or (*R*)-1 and the oxaziridine reagent at  $-85^{\circ}\text{C}$  with subsequent hydrolytic work-up led to (2*R*, 3*R*)- or (2*S*, 3*S*)-2, each in a good yield.

## Experimental

All melting point (mp) data are uncorrected. Optical rotation was measured with a HORIBA SEPA-300 spectrometer, and IR spectra were taken with a JASCO IR-810 spectrometer.  $^1\text{H-NMR}$  spectra were measured with a JEOL JNM-LA 400 spectrometer. HPLC analyses were conducted with SHIMADZU LC 10AD apparatus equipped with a UV/VIS detector and CHIRALCEL OD (DAICEL) column.

**Kinetic Resolution of Racemic *cis*-Flavan-4-ol ( $\pm$ )-3 with Lipase AY.** A suspension of ( $\pm$ )-*cis*-flavan-4-ol<sup>9)</sup> ( $\pm$ )-3 (60 mg, 0.265 mmol), vinyl acetate (4 ml) and lipase AY (Amano) (600 mg) was stirred at  $25^{\circ}\text{C}$  for 6 d. Filtration through a Celite® pad and evaporation of the solvent left a mixture of products which was separated by preparative TLC (hexane:AcOEt = 3:1) to give (2*R*, 4*R*)-4 (29 mg, 44%) and unreacted (2*S*, 4*S*)-3 (33 mg, 55%).

The enantiomeric purity of these compounds was determined to be 93% ee for (2*R*, 4*R*)-4 and 61% ee for (2*S*, 4*S*)-3 by their conversion to (*R*)- and (*S*)-flavanones (*R*)-1 and (*S*)-1 and by an HPLC analysis.



Data for (2*R*, 4*R*)-4: mp  $95\text{--}96^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} - 33.4$  ( $c$  1.68,  $\text{CHCl}_3$ ); IR (nujol)  $\nu_{\text{max}} \text{cm}^{-1}$ : 1740, 1610, 1580, 1240, 1060, 1030, 750, 700;  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.11 (3H, s), 2.21 (1H, ddd,  $J = 10, 11, 13$  Hz), 2.64 (1H, ddd,  $J = 2, 6, 13$  Hz), 5.22 (1H, dd,  $J = 2, 11$  Hz), 6.20 (1H, dd,  $J = 6, 10$  Hz), 6.92–6.99 (2H, m), 7.20–7.25 (2H, m), 7.28–7.58 (5H, m). Data for (2*S*, 4*S*)-3: mp  $125\text{--}126^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} + 7.9$  ( $c$  1.0,  $\text{CHCl}_3$ ) {lit.,<sup>8)</sup> mp  $115\text{--}116^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} + 11.2$  ( $\text{CHCl}_3$ )}; IR (nujol)  $\nu_{\text{max}} \text{cm}^{-1}$ : 3300, 3200, 1605, 1580, 1230, 900, 750, 700;  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 1.75 (1H, d,  $J = 9$  Hz), 2.14 (1H, ddd,  $J = 10, 11, 13$  Hz), 2.54 (1H, ddd,  $J = 2, 8, 13$  Hz), 5.12 (1H, dd,  $J = 9, 16$  Hz), 5.19 (1H, dd,  $J = 2, 12$  Hz), 6.90 (1H, dd,  $J = 1, 8$  Hz), 6.99 (1H, dt,  $J = 1, 7$  Hz), 7.21 (1H, m), 7.33–7.47 (5H, m), 7.52 (1H, d,  $J = 8$  Hz).

**Kinetic Resolution of Racemic *cis*-Flavan-4-ol Acetate 4 with Lipase AY.** A solution of ( $\pm$ )-*cis*-flavan-4-ol acetate<sup>9)</sup> (4; 30 mg, 0.133 mmol) in toluene (0.5 ml) was stirred with a mixture of lipase AY (150 mg), a phosphate buffer (pH 6.8, 2 ml) and  $\text{H}_2\text{O}$  (5 ml) at  $25^{\circ}\text{C}$  for 5 d. The reaction mixture was filtered through a Celite® pad and extracted with  $\text{CHCl}_3$ . Purification of the crude mixture by preparative TLC (hexane:AcOEt = 3:1) afforded (2*R*, 4*R*)-3 (12 mg, 40%) and unreacted (2*S*, 4*S*)-4 (14 mg, 47%).

The enantiomeric purity [(2*R*, 4*R*)-3]: 96% ee; (2*S*, 4*S*)-4]: 88% ee] of these compounds was determined by the same procedure as that just described. Data for (2*R*, 4*R*)-3: mp  $118\text{--}119^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} - 13.7$  ( $c$  0.98,  $\text{CHCl}_3$ ) {lit.,<sup>8)</sup> mp  $115\text{--}116^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} - 14.4$  ( $\text{CHCl}_3$ )}. Data for (2*S*, 4*S*)-4: mp  $96\text{--}97^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} + 34.6$  ( $c$  1.26,  $\text{CHCl}_3$ ) {lit.,<sup>8)</sup> mp  $94\text{--}95^{\circ}\text{C}$ ;  $[\alpha]_{\text{D}}^{20} + 23.9$  ( $\text{CHCl}_3$ )}.

**Conversion of (2*S*, 4*S*)- and (2*R*, 4*R*)-Alcohols, (2*S*, 4*S*)- and (2*R*, 4*R*)-3, to (*S*)- and (*R*)-Flavanones, (*S*)- and (*R*)-1.**

a) **Synthesis of (*S*)-1 from (2*S*, 4*S*)-3.** (2*S*, 4*S*)-alcohol 3 (12 mg, 0.053 mmol) was treated with PCC (34 mg) and molecular sieves 4A (50 mg) in  $\text{CH}_2\text{Cl}_2$  (2 ml) at room temperature for 15 min. Ether was added, and the solution was filtered through a

Celite® pad. Evaporation of the solvent gave a residue which was passed through a short SiO<sub>2</sub> column using ether. The crude sample was purified by preparative TLC to give pure (*S*)-**1** (10 mg, 85%), which was then submitted to an HPLC analysis to determine the enantiomeric purity.

Data for (*S*)-**1**: mp 75–76°C;  $[\alpha]_{\text{D}}^{20} - 39.0$  (*c* 0.49, CHCl<sub>3</sub>) {lit.,<sup>8</sup> mp 75–77°C;  $[\alpha]_{\text{D}}^{20} - 54.1$  (CHCl<sub>3</sub>)}

b) *Synthesis of (R)-1 from (2R, 4R)-3*. This reaction was carried out by the same procedure as that just described for (*S*)-**1**.

Data for (*R*)-**1**: mp 76–77°C;  $[\alpha]_{\text{D}}^{20} + 61$  (*c* 0.86, CHCl<sub>3</sub>) {lit.,<sup>8</sup> mp 76–77°C;  $[\alpha]_{\text{D}}^{20} + 66.5$  (CHCl<sub>3</sub>)}

*Conversion of (2R, 4R)- and (2S, 4S)-Acetates 4 to (R)- and (S)-Flavanones 1.*

a) *Synthesis of (R)-1 from (2R, 4R)-4*. (*2R, 4R*)-acetate **4** (26 mg, 0.097 mmol) was then treated with K<sub>2</sub>CO<sub>3</sub> (19 mg) in MeOH (0.5 ml) at room temperature for 15 min. A sat. NaCl solution was added and the solution was extracted with CHCl<sub>3</sub>. Evaporation of the solvent gave a crude alcohol which was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2 ml) and treated with PCC (62 mg) and molecular sieves 4A (70 mg) as just described. Purification of the crude product by preparative TLC furnished pure (*R*)-**1** (21 mg, 97% from acetate). The enantiomeric purity of this sample was determined by the HPLC analysis already described.

Data for (*R*)-**1**: mp 78–79°C;  $[\alpha]_{\text{D}}^{20} + 57$  (*c* 0.38, CHCl<sub>3</sub>) {lit.,<sup>8</sup> mp 76–77°C;  $[\alpha]_{\text{D}}^{20} + 66.5$  (CHCl<sub>3</sub>)}

b) *Synthesis of (S)-1 from (2S, 4S)-4*. This reaction was carried out by the same procedure as that just described for (*R*)-**1**.

Data for (*S*)-**1**: mp 76–77°C;  $[\alpha]_{\text{D}}^{20} - 55.0$  (*c* 0.20, CHCl<sub>3</sub>) {lit.,<sup>8</sup> mp 75–77°C;  $[\alpha]_{\text{D}}^{20} - 54.1$  (CHCl<sub>3</sub>)}

*(2R, 3R)- and (2S, 3S)-3-Hydroxyflavanones (2R, 3R)- and (2S, 3S)-2.*

a) *Synthesis of (2R, 3R)-2 from (S)-1*. To a stirred solution of (*S*)-**1** (63 mg, 0.281 mmol) in THF (3 ml) was added sodium bis(trimethylsilyl)amide (NAHMDS) (0.39 ml, 1 M in THF, 0.393 mmol) below –80°C and, after 5 min, a solution of 2-(phenylsulfonyl)-3-phenyloxaziridine<sup>10</sup> (102 mg, 0.393 mmol) in THF (1 ml) was added. Stirring was continued for 4 h at –85°C and the reaction was quenched with sat. NH<sub>4</sub>Cl. After stirring at room temperature for 2 h, the aq. solution was extracted with CHCl<sub>3</sub>, and the organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>). Evaporation and preparative TLC (hexane:AcOEt = 2:1) gave pure (*2R, 3R*)-**2** (54 mg,

80%), mp 187–188°C;  $[\alpha]_{\text{D}}^{20} - 18$  (*c* 0.34, CH<sub>2</sub>Cl<sub>2</sub>) {lit.,<sup>2</sup> mp 189–190°C for 59%ee; lit.,<sup>6</sup>  $[\alpha]_{\text{D}}^{20} - 25.0$  (CH<sub>2</sub>Cl<sub>2</sub>)}; IR (nujol)  $\nu_{\text{max}}$  cm<sup>-1</sup>: 3450, 1680, 1610, 1580, 1310, 1230, 1150, 1040, 760, 700; <sup>1</sup>H-NMR (CDCl<sub>3</sub>)  $\delta$ : 3.67 (1H, d, *J* = 2 Hz, OH), 4.65 (1H, dd, *J* = 2, 12 Hz), 5.15 (1H, d, *J* = 12 Hz), 7.06 (1H, d, *J* = 12 Hz), 7.12 (1H, t, *J* = 7 Hz), 7.42–7.50 (3H, m), 7.54–7.60 (3H, m), 7.94 (1H, dd, *J* = 2, 8 Hz).

b) *Synthesis of (2S, 3S)-2 from (R)-1*. (*2S, 3S*)-3-Hydroxyflavanone **2** was obtained from (*R*)-**1** in an 82% yield by the same procedure as that just described for (*2R, 3R*)-**2**.

Data for (*2S, 3S*)-**2**: mp 186–187°C;  $[\alpha]_{\text{D}}^{20} + 22.8$  (*c* 0.6, CH<sub>2</sub>Cl<sub>2</sub>).

## References

- 1) Harborne, J. B., and Baxter, H., *The Handbook of Natural Flavonoids*, John Wiley & Sons, New York (1999).
- 2) Harborne, J. B., *The Flavonoids: Advances in Research from 1986*, Chapman and Hall, London (1993).
- 3) Izumi, T., and Murakami, S., Enzymatic resolution of *trans*-2,3-dihydro-3-hydroxy-2-phenyl-4*H*-1-benzopyran-4-one(*trans*-flavanon-3-ol) by lipase. *J. Heterocycl. Chem.*, **32**, 1125–1127 (1995).
- 4) Takahashi, H., Kubota, Y., Miyazaki, H., and Onda, M., Heterocycles. XV. Enantioselective synthesis of chiral flavononols and flavan-3,4-diol. *Chem. Pharm. Bull.*, **32**, 4852–4857 (1984).
- 5) van Rensburg, H., van Heerden, P. S., Bezuidenhout, B. C. B., and Ferreira, D., Stereoselective synthesis of flavonoids. Part 4. *Trans- and cis-dihydroflavonoids*. *Tetrahedron*, **53**, 14141–14152 (1997).
- 6) Jew, S.-S., Kim, H.-A., Bae, S.-Y., Kim, J.-H., and Park, H.-G., Enantioselective synthetic method for 3-hydroxyflavanones: an approach to (*2R, 3R*)-3',4'-*O*-dimethyltaxifolin. *Tetrahedron Lett.*, **41**, 7925–7928 (2000).
- 7) For a review, see Davis, F. A., and Sheppard, A. C., Applications of oxaziridines in organic synthesis. *Tetrahedron*, **45**, 5703–5742 (1989).
- 8) Izumi, T., Hino, T., and Kasahara, A., Enzymatic kinetic resolution of flavanone and *cis*-4-acetoxylavan. *J. Chem. Soc. Perkin Trans. 1*, **1992**, 1265–1267.
- 9) Suzuki, M., Amano, J., Morioka, M., Mizuno, H., and Matsuura, R., Photochemical reactions of 4-flavanols in the presence of ketone sensitizers. *Bull. Chem. Soc. Jpn.*, **50**, 1169–1172 (1977).
- 10) Vishwakarma, L. C., Stringer, O. D., and Davis, F. A., ( $\pm$ )-*trans*-2-(Phenylsulfonyl)-3-phenyloxaziridine. *Org. Synth., Coll. Vol. VIII*, 546–550 (1993).