Mild and Efficient Hydrolysis of Thioglycosides to Glycosyl Hemiacetals Using N-Iodosaccharin¹

Pintu Kumar Mandal, Anup Kumar Misra*

Medicinal and Process Chemistry Division, Central Drug Research Institute, Chattar Manzil Palace, Lucknow 226001, UP, India Fax +91(522)2223938; E-mail: akmisra69@rediffmail.com

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Abstract: A convenient methodology has been developed for the mild hydrolysis of thioglycosides to the corresponding hemiacetals using N-iodosaccharin without any requirement of co-activator. Most of the functional groups used for the protecting group manipulation of carbohydrates remain unaffected under the reaction conditions.

Key words: carbohydrate, hydrolysis, thioglycoside, hemiacetals, *N*-iodosaccharin

Suitably functionalized glycosyl hemiacetals are useful intermediates for the preparation of various glycosyl donors used in the synthesis of oligosaccharides.^{2–4} They can be converted into more reactive glycosyl donors such as glycosyl fluorides,⁵ trichloroacetimidates⁶ and as such can be used in dehydrative glycosylation reactions.⁷ They have also been applied in Wittig or Horner-Emmons or related reactions for chiral synthesis of various natural products.^{8,9} Glycosyl hemiacetals can be prepared from (a) peracetylated sugars using hazardous hydrazine salts¹⁰ or organic bases like benzyl amine¹¹ or under acidic conditions;¹² (b) acid hydrolysis of alkyl glycosides;¹³ or (c) hydrolysis of thioglycosides. Among them, hydrolysis of thioglycosides is more useful as it provides the preparation of glycosyl hemiacetals having diverse functionalities under mild reaction conditions. As expected, a number of reports appeared in the literature for this purpose, including the use of toxic heavy metal salts,¹⁴ Nbromosuccinimide (NBS)¹⁵ or *N*-iodosuccinimide (NIS) alone¹⁶ or in the presence of an acid,^{17–19} borate salts/*n*-Bu₄NIO₄/HCIO₄,²⁰ V₂O₅/H₂O₂/NH₄Br,²¹ (NH₄)₆Mo₇O₂₄/H₂O₂HCIO₄/NH₄Br,²² and chloramine-T.²³ However, many of these methods suffer from limitations, such as use of expensive reagents, incompatibility with acidic functional groups, relatively low yield and sometime harsh reaction conditions. In search of a generalized reaction condition for the hydrolysis of thioglycosides having acid-labile and base-labile functional groups, we have tested the efficacy of N-iodosaccharin (NISac) for this purpose. Recently, NISac has been used in the glycosylation reaction using armed glycosyl donors²⁴ and iodination of alkenes²⁵ and conversion of alcohols to iodides.²⁶ As discussed earlier,²⁴ due to the lower pK_a value of NISac $(pK_a 1.30)$ than its analogous NIS $(pK_a 9.62)$, it should be

SYNLETT 2007, No. 8, pp 1207–1210 Advanced online publication: 03.04.2007 DOI: 10.1055/s-2007-977412; Art ID: D25606ST © Georg Thieme Verlag Stuttgart · New York a more potent iodinating agent than NIS and it should activate thioglycosides without the requirement of any strong acid as co-activator. Taking cues from the literature reports, we have reasoned that NISac could independently activate thioglycosides in a moist reaction condition resulting in the formation of hemiacetals. Avoiding the addition of acidic co-activator could make this method equally effective for hydrolyzing thioglycosides having both acid-labile as well as base-labile protecting groups. *N*-Iodosaccharin can be easily prepared²⁵ in the laboratory and it is very much cost-effective in comparison to analogous *N*-iodosuccinimide (NIS).





In an initial set of experiments, ethyl 2,3,4,6-tetra-Oacetyl-1-thio- β -D-glucopyranoside was treated with 1.5 equivalents of NISac in wet acetonitrile (MeCN $-H_2O =$ 10:1) at room temperature (Scheme 1). To our satisfaction, a clean formation of glycosyl hemiacetal was achieved in almost quantitative yield in only five minutes. Reducing the quantity of NISac resulted in incomplete hydrolysis of thioglycoside even after 24 hours. Similar reaction conditions were then applied to hydrolyze a diverse set of thioglycosides containing base-labile and acid-labile functional groups, which is presented in Table 1. In Table 1, there are a number of points that need to be highlighted. All reactions were complete in 5-30minutes. The reaction condition is compatible with the acid-labile functional groups (benzylidene, isopropylidene, silyl ether, 4-methoxybenzylidene) as well as baselabile groups (acetyl, benzoyl, chloroacetyl). Interglycosidic linkage remains unaffected under the reaction condition. The rate of activation depends on the alkyl or aryl part linked to the sulfur atom; thus the SPh group takes a longer time to hydrolyze than the SEt or STol glycosides. The rate of hydrolysis for the acyl-protected thioglycosides is slightly slower than the alkyl-protected counterpart, which may be explained by considering the 'armed-disarmed' concept. In most of the cases, an anomeric mixture of glycosyl hemiacetals was formed and the ratio was determined from the ¹H NMR of the crude products. A series of solvents has been tested for the reaction

and MeCN–H₂O (10:1) mixture has been found to be the best option in comparison to other commonly used solvents like dichloromethane and THF. In contrast to the earlier report,²⁴ no trace of formation of glycosyl saccharine derivatives was observed. The formation of the product could be explained by considering the formation of sulfonium ion generated by NISac activation of thioglycoside followed by hydrolysis.

In summary, a relative shortcoming of the hydrolysis of thioglycosides has been overcome by devising a generalized, mild reaction protocol for the hydrolysis of thioglycosides without requirement of any co-activator. Most of the functional groups used for the protecting group manipulation of carbohydrate were found unaffected under the reaction condition. It is important to mention that *N*iodosaccharine (NISac) is very cheap in comparison to its closest analogue, *N*-iodosuccinimide (NIS). Straightforward operation, low-cost activator, very short reaction time, and simple purification of products are the key features of this generalized protocol, which makes it an attractive alternative to the existing literature procedures.

 Table 1
 Hydrolysis of Thioglycosides Using N-Iodosaccharin (NISac) at Room Temperature²⁷

| Entry | Thioglycosides 1 ^a | Products 2 ^a | Time (min) | Yield (%) | α/β | Ref. |
|-------|--------------------------------------|-----------------------------------|------------|-----------|-------|------|
| 1 | Aco Aco SEt | AcO AcO NOH | 5 | 95 | 3:1 | 21 |
| 2 | Aco Aco SPh | AcO AcO ACO ACO ACO | 20 | 92 | 3.5:1 | 21 |
| 3 | ACO ACO STOI | ACO ACO ACO TOH | 5 | 95 | 3:1 | 21 |
| 4 | | ACO OAC ACO ACO OH | 5 | 92 | 2.5:1 | 21 |
| 5 | Aco OAc Aco OAc | AcO AcO AcO OH | 20 | 90 | 1:0 | 21 |
| 6 | BzO OBz BzO SEt | BZO OBZ BZO BZO OH | 5 | 92 | 3:1 | 21 |
| 7 | BnO SEt | BnO OBn BnO BnO OH | 3 | 88 | 2:1 | 23 |
| 8 | CAO OBn CAO OBn SEt | CAO OBn CAO OBn OH | 3 | 85 | 3:1 | 21 |
| 9 | BzO BzO OBz | BZO BZO OBZ OH | 5 | 95 | 2.5:1 | 21 |
| 10 | AcO AcO OAc | Me O O H | 5 | 92 | 4:1 | 12 |
| 11 | SPh Me OBn Bno OBn | Me OT OBn BnO OBn | 3 | 90 | 5:1 | 21 |
| 12 | Ph TO OBZO BZO BZO | Ph O O BzO BzO O H | 30 | 90 | 1:1 | 21 |

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 Table 1
 Hydrolysis of Thioglycosides Using N-Iodosaccharin (NISac) at Room Temperature²⁷ (continued)

| Entry | Thioglycosides 1 ^a | Products 2 ^a | Time (min) | Yield (%) | α/β | Ref. |
|-------|--|--|------------|-----------------|-----|------|
| 13 | Ph TO-O-O-O-O-O-SPh AcO-O-SPh NPhth | Ph TOTO Aco OH NPhth | 20 | 90 | 1:2 | _ |
| 14 | PhOMe O BnO BnO BnO | PhOMe O BnO BnO BnO O H | 20 | 85 | 2:1 | 23 |
| 15 | Me O AcO O O | Me OH Aco O | 5 | 92 | 3:1 | 23 |
| 16 | OBZ OBZ OBZ OBZ | OBZ OBZ OBZ OBZ | 10 | 82 | 5:1 | 21 |
| 17 | AllO OBn MBnO OBn STol | | 5 | 80 | 2:1 | 21 |
| 18 | ACO OAC ACO ACO ACO ACO STOI | ACO OAC ACO ACO ACO ACO OAC | 5 | 95 | 2:1 | 21 |
| 19 | ACO ACO OAC ACO ACO ACO STOI | ACO ACO ACO ACO ACO ACO ACO ACO ACO ACO | 5 | 95 | 2:1 | 21 |
| 20 | AcO QAc AcO CH(SEt) ₂ AcO ÕAc | AcO QAc AcO i CHO AcO OAc | 20 | 80 ^b | - | _ |

^a Tol = tolyl; CA = chloroacetyl; Phth = phthalimido; MBn = 4-Methoxybenzyl. ^b NISac (2.5 equiv) was used.

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- (27) Typical Experimental Procedure: To a solution of thioglycoside (1.0 mmol) in MeCN–H₂O (10:1; 5 mL) was added *N*-iodosaccharin (1.5 mmol) at r.t. and the reaction mixture was allowed to stir for the time mentioned in Table 1. After completion of the reaction, the reaction mixture was concentrated under reduced pressure. The crude mass was purified over SiO₂ using hexane–EtOAc as eluent

to furnish the pure glycosyl hemiacetal as an anomeric mixture. Products of all known compounds gave acceptable ¹H NMR and ¹³C NMR spectra that matched the data reported in the cited references.²⁸

- (28) Spectral data of selected compounds:
 - **3-O-Acetyl-4,6-O-benzylidene-2-deoxy-2-phthalimido-Dglucopyranose (Table 1, Entry 13)**: ¹H NMR (300 MHz, CDCl₃): δ = 7.33–7.91 (m, 9 H, aromatic protons), 5.85 (t, J = 9.0 Hz, 1 H, H-3), 5.69 (d, J = 8.1 Hz, 1 H, H-1), 5.58 (s, 1 H, PhC*H*), 4.48 (dd, J = 4.5, 10.2 Hz, 1 H, H-4), 4.35–4.40 (m, 1 H, H-2), 3.70–3.83 (m, 3 H, H-5, H-6a, H-6b), 1.88 (s, 3 H, OCOMe). ESI-MS: m/z calcd for C₂₃H₂₁NO₈: 439; found: 462 [M + Na]⁺.

2,3,4,5,6-Penta-*O***-acetyl-D-galactose (Table 1, Entry 20)**: ¹H NMR (300 MHz, CDCl₃): $\delta = 2.04-2.12$ (5 × s, 15 H), 3.91 (dd, 1 H), 4.26 (dd, 1 H), 5.26 (d, *J* = 2.0 Hz, 1 H), 5.36 (m, 1 H), 5.49 (d, *J* = 8.0 Hz, 1 H), 5.62 (d, *J* = 8.0 Hz, 1 H), 9.45 (br s, 1 H). ESI-MS: *m*/*z* calcd for C₁₆H₂₂O₁₁: 390; found: 413 [M + Na]⁺. Copyright of Synlett is the property of Georg Thieme Verlag Stuttgart and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.