

# CHEMISTRY

## A European Journal

A Journal of



### Accepted Article

**Title:** A multicolor large Stokes shift fluorogen-activating RNA aptamer with cationic chromophores

**Authors:** Christian Steinmetzger, Navaneethan Palanisamy, Kiran R. Gore, and Claudia Höbartner

This manuscript has been accepted after peer review and appears as an Accepted Article online prior to editing, proofing, and formal publication of the final Version of Record (VoR). This work is currently citable by using the Digital Object Identifier (DOI) given below. The VoR will be published online in Early View as soon as possible and may be different to this Accepted Article as a result of editing. Readers should obtain the VoR from the journal website shown below when it is published to ensure accuracy of information. The authors are responsible for the content of this Accepted Article.

**To be cited as:** *Chem. Eur. J.* 10.1002/chem.201805882

**Link to VoR:** <http://dx.doi.org/10.1002/chem.201805882>

Supported by  
**ACES**

WILEY-VCH

## COMMUNICATION

# A multicolor large Stokes shift fluorogen-activating RNA aptamer with cationic chromophores

Christian Steinmetzger,<sup>[a]</sup> Navaneethan Palanisamy,<sup>[b]#</sup> Kiran R. Gore<sup>[c]§</sup> and Claudia Höbartner<sup>\*[a,b,c]</sup>

**Abstract:** Large Stokes shift (LSS) fluorescent proteins (FPs) exploit excited state proton transfer pathways to enable fluorescence emission from the phenolate intermediate of their internal 4-hydroxybenzylidene imidazolone (HBI) chromophore. An RNA aptamer named Chili mimics LSS FPs by inducing highly Stokes-shifted emission from several new green and red HBI analogs that are non-fluorescent when free in solution. The ligands are bound by the RNA in their protonated phenol form and feature a cationic aromatic side chain for increased RNA affinity and reduced magnesium dependence. In combination with oxidative functionalization at the C2 position of the imidazolone, this strategy yielded DMHBO<sup>+</sup>, which binds to the Chili aptamer with a low-nanomolar  $K_D$ . Because of its highly red-shifted fluorescence emission at 592 nm, the Chili-DMHBO<sup>+</sup> complex is an ideal fluorescence donor for Förster resonance energy transfer (FRET) to the rhodamine dye Atto 590 and will therefore find applications in FRET-based analytical RNA systems.

Fluorogen-activating RNA aptamers (FLAPs) have emerged as powerful tools for tagging and visualizing RNA *in vitro* and *in vivo*, and, among various other applications in biochemistry and synthetic biology, they are used as components of metabolite and protein sensors.<sup>[1]</sup> These artificial functional RNAs form specific, non-covalent complexes with conditionally fluorescent chromophores that are non-emissive free in solution, but show strongly enhanced fluorescence emission in the bound state. Early examples, including the Malachite green aptamer<sup>[2]</sup> and the sulforhodamine aptamer,<sup>[3]</sup> are currently receiving renewed attention for further optimization and reselection,<sup>[4,5]</sup> and new chromophore-binding aptamers, such as cyanine dye-activating Dir2s<sup>[6]</sup> and Mango RNA aptamers,<sup>[7,8]</sup> continue to be developed. A prominent class of fluorogenic aptamers known as mimics of fluorescent proteins, named Spinach,<sup>[9]</sup> Broccoli,<sup>[10]</sup> and Corn,<sup>[11]</sup> were evolved to bind analogs of the GFP chromophore 4-hydroxybenzylidene imidazolone (HBI). Synthetic HBI derivatives are non-fluorescent in aqueous solution, but non-covalent binding

to an RNA aptamer prevents non-radiative deactivation of the excited state, thereby restoring the intrinsic fluorescence.

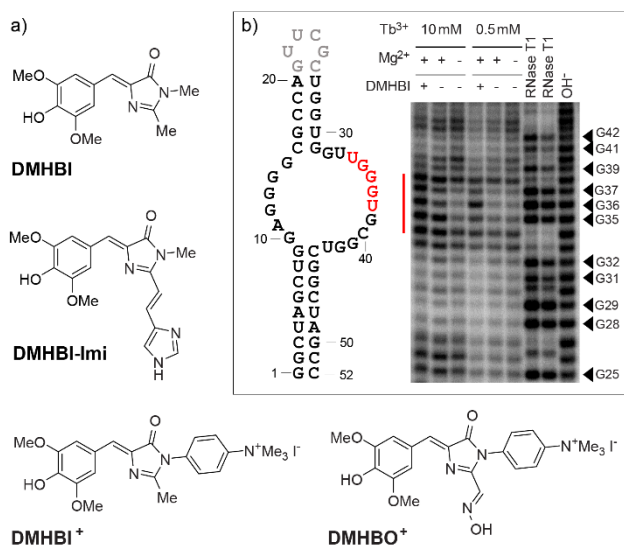
The majority of known fluorogenic aptamers is characterized by a small or moderate difference between their excitation and emission maxima. However, fluoromodules with large Stokes shifts display significant advantages, such as reduced reabsorption of the emitted light,<sup>[12]</sup> convenient use as FRET donors for far-red fluorescence reporters,<sup>[13]</sup> and the ability to produce multiple emission colors by simultaneous excitation of different fluorophores at a single wavelength.<sup>[14]</sup> Large energy differences between absorption and emission peaks can result from intramolecular charge transfer (ICT), or from inter- or intramolecular excited state proton transfer (ESPT), among other mechanisms.<sup>[12]</sup> Fluorescent proteins that exhibit large apparent Stokes shifts (termed LSS FPs) have been engineered to exploit ESPT from the aromatic OH group of the HBI chromophore to a neighboring amino acid of the  $\beta$ -barrel scaffold. Prominent members of this family are the orange/red emitting proteins LSSmOrange<sup>[14]</sup> and LSSmKate,<sup>[15]</sup> which are excited at 437 and 460 nm and emit at 572 and 605 nm, respectively. Similarly large Stokes shifts and fluorescence emission in the 600 nm region have not been reported for fluorogenic RNA aptamers. In this study, we aim to mimic LSS green and red fluorescent proteins by capitalizing on fluorogen-activating RNA aptamers that bind the protonated form of HBI analogs and enable ESPT and emission from an excited deprotonated intermediate.

Spinach, Broccoli and Corn are members of the HBI-binding aptamer family that activate derivatives of 3,5-difluoro-HBI (DFHBI), which are almost completely deprotonated at physiological pH.<sup>[9]</sup> Here, we capitalized on 3,5-dimethoxy-substituted HBI (DMHBI) derivatives that have a higher  $pK_a$  and can be excited at the absorption maximum of the phenol form. We first examined the 13-2 aptamer, which was originally reported to activate the fluorescence of DMHBI, showing an emission at 529 nm upon excitation at 398 nm.<sup>[9]</sup> Based on the analysis of the predicted secondary structure,<sup>[16]</sup> we introduced an extra-stable UUCG tetraloop into 13-2min, with the intention to stabilize an apical hairpin and to assist folding of the ligand binding pocket. This modified 52-nt RNA aptamer (Figure 1) showed approx. 2-fold enhanced fluorescence of bound DMHBI compared to the original 60-nt RNA, and the fraction of RNA that folded into a ligand binding-competent conformation was significantly enhanced (Figure S1). The 52-nt RNA resulting from the optimization of 13-2min was used for all further experiments discussed in this manuscript, and – in line with the “vegetable nomenclature” – was named Chili because of its special properties to activate green, yellow and red chromophores.

- [a] C. Steinmetzger, Prof. Dr. C. Höbartner  
Institute of Organic Chemistry, University of Würzburg  
Am Hubland, 97074 Würzburg (Germany)  
E-mail: claudia.hoebartner@uni-wuerzburg.de
- [b] N. Palanisamy, Prof. Dr. C. Höbartner  
International Max Planck Research School Molecular Biology,  
University of Göttingen (Germany)
- # present address: BIOS Center for Biological Signaling Studies,  
University of Freiburg (Germany)
- [c] Dr. K.R. Gore, Prof. Dr. C. Höbartner  
Center for Nanoscale Microscopy and Molecular Physiology of the  
Brain (CNMPB), Göttingen, (Germany)
- § present address: Department of Chemistry, University of Mumbai  
(India)

Supporting information for this article is given via a link at the end of the document.

## COMMUNICATION



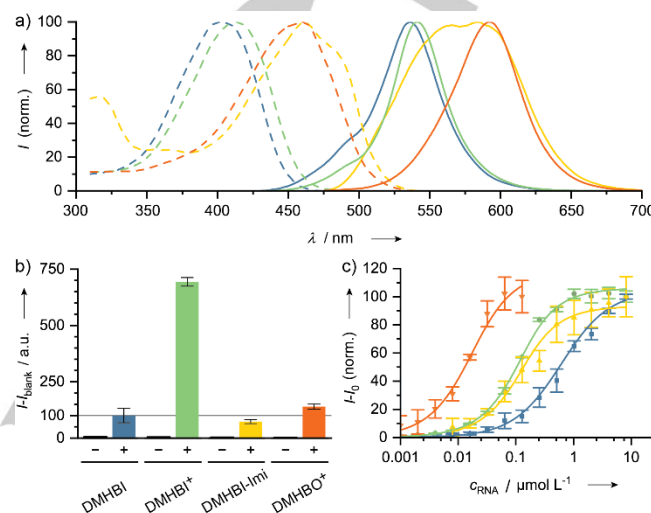
**Figure 1.** a) Structures and names of the HBI derivatives investigated in this work. b) Sequence of 52-nt Chili RNA and gel image of Tb<sup>3+</sup>-footprinting, comparing unfolded and folded RNA in the absence and presence of DMHBI. Full gel image in Figure S3. The flexible loop region is indicated in red.

The high guanine content of the Chili RNA sequence suggests that it contains guanine quartets in the ligand binding site. This hypothesis is supported by the finding that DMHBI fluorescence is strongly dependent on the  $K^+$  concentration with an apparent  $K_D$  of 12 mM and a Hill coefficient of 2.5 (Figure S2), suggesting the involvement of a two-tiered G quadruplex.<sup>[17,18]</sup> This feature is reminiscent of other HBI-binding aptamers, including Spinach and Corn, for which the crystal structures show highly unusual quadruplex topologies,<sup>[18]</sup> as well as Mango aptamers, which also contain quadruplexes as ligand-binding structural elements.<sup>[19]</sup> Despite the overall similarity of the 2D secondary structure schemes of Chili and Spinach/Broccoli (Figure S1), their sequences are significantly different. In the absence of a 3D structural model, it remains unknown which guanines of Chili are involved in putative G quartets. However, it is interesting to note that, compared to Spinach, Chili has additional nucleotides that are expected to form larger loops extruding from a putative quadruplex core. This is supported by structural probing data that clearly identify the region of nt 34–38 in the folded and ligand-bound state as more susceptible to backbone hydrolysis compared to the unfolded state, i.e., in the absence of ligand and  $Mg^{2+}$  (Figure 1b and Figure S3).

The presence of the flexible bulge in Chili suggested an opportunity to modulate the fluorescence properties of the RNA-chromophore complexes by ligand engineering. Candidate sites for modification are the positions N1 and C2 of the imidazolone, similar to variations observed in fluorescent proteins, originating from the primary protein sequences. In analogy, the DFHBI ligand was earlier modified at these two sites, resulting in DFHBI-1T as a brighter Spinach ligand<sup>[20]</sup> and DFHO as the ligand responsible for orange emission in complex with the Corn aptamer.<sup>[11]</sup>

Here, we designed and synthesized three new DMHBI ligands (Figure 1) and investigated their fluorescence properties in

complex with the Chili RNA. The DMHBI<sup>+</sup> derivative carries a trimethylammonium-substituted phenyl ring at N3, which may provide additional stacking interactions with nucleobases, and the cationic substituent may assist electrostatic interactions with the negatively charged RNA backbone. These features should enforce a rigid fluorophore conformation and stabilize the position of the fluorophore at the binding site. The other two new DMHBI analogs were extended at the C2 position: DMHBI–Imi mimics



**Figure 2.** a) Normalized excitation and emission spectra (dashed and solid lines, resp.) of DMHBI (blue), DMHBI\* (green), DMHBI-Imi (yellow), and DMHBO\* (orange) bound to Chl*a* RNA. b) Integrated fluorescence intensity of each dye in the presence (+) or absence (–) of the Chl*a* RNA at pH 7.5. c) Titration with Chl*a* RNA reveals higher affinity for all three new chromophores compared to DMHBI ( $K_D$  listed in Table 1, experimental details given in supporting information).

the chromophore of the protein Kaede, which contains a histidine-derived arylvinyl substituent.<sup>[21]</sup> Other red fluorescent proteins, such as DsRed and its relatives, carry acylimine substituents instead of arylvinyl groups at C2.<sup>[22]</sup> In combination with the trimethylammoniumphenyl side chain described above, the oxime in DMHBO<sup>+</sup> was chosen as a hydrolytically stable acylimine mimic.

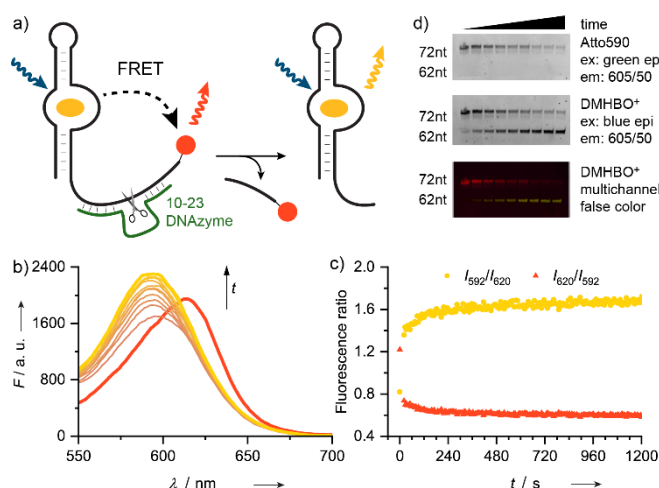
The three new chromophores were synthesized starting from 4-hydroxy-3,5-dimethoxybenzaldehyde (Scheme S1). DMHBI<sup>+</sup> was obtained in three steps, starting with the condensation with 4-(dimethylamino)aniline. The imine was incorporated into the HBI scaffold in a 1,3-dipolar cycloaddition reaction with an imidate ylide, which was prepared from ethyl acetimidate and methyl glycinate.<sup>[23]</sup> The aromatic dimethylamino group was alkylated with methyl iodide to obtain DMHBI<sup>+</sup>. To generate DMHBO<sup>+</sup>, the cycloaddition product was oxidized with selenium dioxide and the resulting aldehyde was reacted with hydroxylamine, followed by alkylation with methyl iodide. The Kaede analog DMHBI-Imi was generated from DMHBI via a scandium triflate-catalyzed Aldol condensation with imidazole-4-carboxaldehyde.<sup>[24]</sup>

The fluorescence properties of the four dyes and their binding and activation by Chili were investigated. Chili–DMHBI and Chili–DMHBI<sup>+</sup> showed excitation maxima near 400 nm and emission maxima near 540 nm (Figure 2a), demonstrating that the



## COMMUNICATION

rhodamine dye at its 3'-end.<sup>[27]</sup> Upon binding of DMHBO<sup>+</sup> and excitation at 456 nm, fluorescence emission from Atto 590 was observed, which can be rationalized by the strong overlap of the DMHBO<sup>+</sup> emission and Atto 590 absorption bands (Figure S14). The large Stokes shift of Chili-DMHBO<sup>+</sup> rules out direct excitation of the acceptor, as shown in the absence of DMHBO<sup>+</sup> (Figure S14). Furthermore, the FRET effect allowed monitoring of DNA-catalyzed site-specific cleavage of the Atto 590-labeled extension of the aptamer. Upon addition of a 10-23 deoxyribozyme with appropriate binding arms,<sup>[28]</sup> the DMHBO<sup>+</sup> emission intensity increased, while Atto 590 intensity decreased concomitantly (Figure 4b). The time-dependent change in the ratio of fluorescence signals ( $I_{590}/I_{620}$ ) (Figure 4c) correlates well with the gel-based analysis (Figure 4d). The polyacrylamide gel was stained with DMHBO<sup>+</sup> and imaged with green and blue epi illumination (i.e., green: excitation of Atto 590; blue: excitation of DMHBO<sup>+</sup>). The false-color overlay confirms the presence of both chromophores in the full-length construct and identifies the single-labeled cleavage products (Figure 4d, S15). This proof-of-principle experiment suggests that the Chili-DMHBO<sup>+</sup>-Atto 590 FRET pair can be used for ratiometric RNA sensors and multicolor fluorescence experiments.



**Figure 4.** a) A Chili aptamer-based FRET sensor monitors DNA-catalyzed RNA cleavage. Donor: DMHBO<sup>+</sup> (orange), acceptor: Atto 590 (red). b) Fluorescence emission spectra of Atto 590-labeled Chili-DMHBO<sup>+</sup> before addition of DNA enzyme (red curve). Upon addition of 10-23 DNA enzyme, spectra were taken every 6 sec (every 20<sup>th</sup> spectrum shown up to 20 min). c) Kinetics of RNA cleavage monitored by a decrease in acceptor emission (red) and an increase in donor emission (orange). d) Denaturing PAGE imaged with green and blue epi illumination after staining with a 1  $\mu$ M solution of DMHBO<sup>+</sup>. Time points: 0.25, 0.5, 1, 2, 5, 10, 20, 40, 60 min. Experimental conditions for c) and d): 0.5  $\mu$ M RNA, 5  $\mu$ M DNA enzyme, 40 mM HEPES pH 7.5, 125 mM KCl, 5 mM MgCl<sub>2</sub>.

In summary, the Chili RNA aptamer in complex with DMHBI<sup>+</sup> and DMHBO<sup>+</sup> mimics green and red LSS fluorescent proteins. Rational design resulted in the first variants of HBI aptamer ligands with permanently cationic sidechains. The Chili-DMHBO<sup>+</sup> complex is the longest wavelength-emitting (592 nm) and tightest binding (12 nM  $K_D$ ) RNA fluoromodule in the growing family of HBI-binding aptamers. In the future, it will be exciting to elucidate

the three-dimensional structure of the binding site to comprehend the structural basis of the proton relay in the RNA responsible for the large apparent Stokes shift, and to enable engineering of even brighter LSS aptamer variants for applications *in vitro* and in cells.

## Acknowledgements

This work was supported by the European Research Council (ERC consolidator grant No. 682586), by the Cluster of Excellence and DFG Research Center Nanoscale Microscopy and Molecular Physiology of the Brain, University of Göttingen, and by the University of Würzburg.

**Table 1.** Photophysical data of Chili aptamer complexes<sup>[a]</sup> in comparison to selected fluorescent proteins.

Fluorophore	Ex/Em <sup>[b]</sup> (nm)	Stokes shift (nm)	Quantum yield <sup>[c]</sup>	$K_D$ ( $\mu$ M)
DMHBI	400/537	137	0.08	0.57
DMHBI <sup>+</sup>	413/542	129	0.40	0.063
DMHBI-Imi	463/545, 594 <sup>[d]</sup>	82, 131	0.08	0.071
DMHBO <sup>+</sup>	456/592	136	0.10	0.012
GFP <sup>[22]</sup>	475/508	33	0.79	-
Kaede <sup>[21]</sup>	572/582	10	0.33	-
LSSmOrange <sup>[14]</sup>	437/572	135	0.45	-
LSSmKate2 <sup>[15]</sup>	460/605	145	0.17	-

[a] in 40 mM HEPES pH 7.5, 125 mM KCl, 5 mM MgCl<sub>2</sub> [b] Excitation and emission maxima. [c] relative quantum yield using Coumarin 153 as standard. [d] Two maxima of the bimodal emission spectrum.

**Keywords:** RNA aptamer • fluorescence • large Stokes shift • fluorescent protein • fluorescence resonance energy transfer

- [1] a) S. Hennig, S. Neubacher, *Angew. Chem. Int. Ed.* **2018**, doi:10.1002/anie.201806482; *Angew. Chem.* **2018**, doi: 10.1002/ange.201806482; b) S. R. Jaffrey, *Adv. Pharmacol.* **2018**, 82, 187; c) K. Rau, A. Rentmeister, *ACS Cent. Sci.* **2017**, 3, 701; d) S. C. Alexander, N. K. Devaraj, *Biochemistry* **2017**, 56, 5185.
- [2] J. R. Babendure, S. R. Adams, R. Y. Tsien, *J. Am. Chem. Soc.* **2003**, 125, 14716.
- [3] L. A. Holeman, S. L. Robinson, J. W. Szostak, C. Wilson, *Fold. Des.* **1998**, 3, 423.
- [4] a) M. Sunbul, A. Jäschke, *Angew. Chem. Int. Ed.* **2013**, 52, 13401; *Angew. Chem.* **2013**, 125, 13643; b) M. Sunbul, A. Jäschke, *Nucleic Acids Res.* **2018**, doi: 10.1093/nar/gky543.
- [5] M. Gotrik, G. Sekhon, S. Saurabh, M. Nakamoto, M. Eisenstein, H. T. Soh, *J. Am. Chem. Soc.* **2018**, 140, 3583.
- [6] X. Tan, T. P. Constantin, K. L. Sloane, A. S. Waggoner, M. P. Bruchez, B. A. Armitage, *J. Am. Chem. Soc.* **2017**, 139, 9001.
- [7] A. Autour, S. C. Y. Jeng, A. D. Cawte, A. Abdolazadeh, A. Galli, S. S. S. Panchapakesan, D. Rueda, M. Ryckelynck, P. J. Unrau, *Nat. Commun.* **2018**, 9, 656.

## COMMUNICATION

- [8] E. V. Dolgosheina, S. C. Y. Jeng, S. S. S. Panchapakesan, R. Cojocar, P. S. K. Chen, P. D. Wilson, N. Hawkins, P. A. Wiggins, P. J. Unrau, *ACS. Chem. Biol.* **2014**, *9*, 2412.
- [9] J. S. Paige, K. Y. Wu, S. R. Jaffrey, *Science* **2011**, *333*, 642.
- [10] G. S. Filonov, J. D. Moon, N. Svensen, S. R. Jaffrey, *J. Am. Chem. Soc.* **2014**, *136*, 16299.
- [11] W. Song, G. S. Filonov, H. Kim, M. Hirsch, X. Li, J. D. Moon, S. R. Jaffrey, *Nat. Chem. Biol.* **2017**, *13*, 1187.
- [12] F. Vollmer, W. Rettig, E. Birckner, *J. Fluoresc.* **1994**, *4*, 65.
- [13] a) O. Zapata-Hommer, O. Griesbeck, *BMC biotechnol.* **2003**, *3*, 5; b) H.-W. Ai, K. L. Hazelwood, M. W. Davidson, R. E. Campbell, *Nat. Methods* **2008**, *5*, 401.
- [14] D. M. Shcherbakova, M. A. Hink, L. Joosen, T. W. J. Gadella, V. V. Verkhusha, *J. Am. Chem. Soc.* **2012**, *134*, 7913.
- [15] K. D. Piatkevich, V. N. Malashkevich, S. C. Almo, V. V. Verkhusha, *J. Am. Chem. Soc.* **2010**, *132*, 10762.
- [16] Secondary structure was predicted using the Vienna RNA webservices: <http://rna.tbi.univie.ac.at/> and compared to the predicted secondary structure reported in Ref. 9.
- [17] M. A. Mullen, S. M. Assmann, P. C. Bevilacqua, *J. Am. Chem. Soc.* **2012**, *134*, 812.
- [18] a) K. D. Warner, L. Sjekloča, W. Song, G. S. Filonov, S. R. Jaffrey, A. R. Ferré-D'Amaré, *Nat. Chem. Biol.* **2017**, *13*, 1195; b) H. Huang, N. B. Suslov, N.-S. Li, S. A. Shelke, M. E. Evans, Y. Koldobskaya, P. A. Rice, J. A. Piccirilli, *Nat. Chem. Biol.* **2014**, *10*, 686; c) P. Fernandez-Millan, A. Autour, E. Ennifar, E. Westhof, M. Ryckelynck, *RNA* **2017**, *23*, 1788; d) K. D. Warner, M. C. Chen, W. Song, R. L. Strack, A. Thorn, S. R. Jaffrey, A. R. Ferré-D'Amaré, *Nat. Struct. Mol. Biol.* **2014**, *21*, 658.
- [19] a) R. Trachman, A. Abdolazadeh, A. Andreoni, R. Cojocar, J. R. Knutson, M. Ryckelynck, P. J. Unrau, A. Ferré-D'Amaré, *Biochem.* **2018**; b) R. J. Trachman, N. A. Demešhkina, M. W. L. Lau, S. S. S. Panchapakesan, S. C. Y. Jeng, P. J. Unrau, A. R. Ferré-D'Amaré, *Nat. Chem. Biol.* **2017**, *13*, 807.
- [20] W. Song, R. L. Strack, N. Svensen, S. R. Jaffrey, *J. Am. Chem. Soc.* **2014**, *136*, 1198.
- [21] R. Ando, H. Hama, M. Yamamoto-Hino, H. Mizuno, A. Miyawaki, *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 12651.
- [22] a) R. N. Day, M. W. Davidson, *Chem. Soc. Rev.* **2009**, *38*, 2887; b) R. M. Wachter, J. L. Watkins, H. Kim, *Biochemistry*, **2010**, *49*, 7417.
- [23] a) J. M. Lerestif, J. Perrocheau, F. Tonnard, J. P. Bazureau, J. Hamelin, *Tetrahedron* **1995**, *51*, 6757; b) A. Baldrige, J. Kowalik, L. Tolbert, *Synthesis* **2010**, 2424.
- [24] Scandium triflate was the preferred catalyst compared to more commonly used  $ZnCl_2$  in previous reports of analogous reactions: W.-T. Chuang, B.-S. Chen, K.-Y. Chen, C.-C. Hsieh, P.-T. Chou, *Chem. Commun.* **2009**, 6982.
- [25] G. Feng, C. Luo, H. Yi, L. Yuan, B. Lin, X. Luo, X. Hu, H. Wang, C. Lei, Z. Nie et al., *Nucleic Acids Res.* **2017**, *45*, 10380.
- [26] S. Bose, S. Chakrabarty, D. Ghosh, *J. Phys. Chem. B* **2017**, *121*, 4790.
- [27] A 20-nt 3'-amino-modified RNA was labeled with Atto 590-NHS ester and ligated to the Chili RNA with T4 RNA ligase. Experimental details are given in the Supporting information.
- [28] a) S. W. Santoro, G.F. Joyce, *Proc. Natl. Acad. Sci. USA*, **1997**, *94*, 4262; b) S.K. Silverman, *Nucleic Acids Res.* **2005**, *33*, 6151.

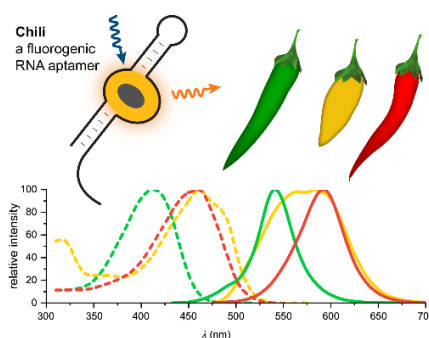
## COMMUNICATION

## Entry for the Table of Contents

## COMMUNICATION

"My shadow side so amplified." (Red Hot Chili Peppers)

A large Stokes shift fluorogen-activating RNA aptamer (LSS FLAP) named "Chili" strongly amplifies green or red fluorescence emission of otherwise non-fluorescent, tightly bound cationic ligands upon excitation with blue light. Chili allows visualization of RNA and can be used in FRET sensors.



C. Steinmetzger, N. Palanisamy,  
K.R. Gore, C. Höbartner\*

Page No. – Page No.

**A multicolor large Stokes shift  
fluorogen-activating RNA aptamer  
with cationic chromophores**