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Monosaccharide derivatives with low nM lectin affinity and high selectivity based on combined fluorine-amide, phenyl-arginine, sulfur- π , and halogen bond interactions

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Abstract: The design of small and high affinity lectin inhibitors remains a major challenge because lectin natural ligand binding sites often are shallow and have polar character. We report that derivatizing galactose with un-natural structural elements that form multiple non-natural lectin-ligand interactions (orthogonal multipolar fluorine-amide, phenyl-arginine, sulfur- π , and halogen bond) can provide inhibitors with extraordinary affinity (low nM) for the model lectin, galectin-3, which is over 5 orders of magnitude higher than the parent galactose, and, moreover, is selective compared to other galectins.

Lectin binding to glycoconjugates are rate-limiting in many pathophysiological processes^[1] including host-pathogen interaction, inflammation, immunity and cancer. Consequently, the discovery of drug-like inhibitors of such interactions is receiving significant attention.^[2] However, finding small high affinity lectin inhibitors is a major challenge because the lectin carbohydrate-binding sites tend to be polar and shallow. Lectins typically bind natural glycans with multiple hydrogen bonds, sometimes enhanced by CH- π stacking of carbohydrate CH onto aromatic amino acid side chains, and with recently highlighted contributions from conformational entropy.^[3] This usually results in weak-medium-to-affinity (μ M-mM) for a small mono- or disaccharide, although exceptions are known. The challenge then is to find lectin inhibitors with drug like affinities, low nM, and pharmacological properties that are much better than the natural ligands. This has been achieved in some cases

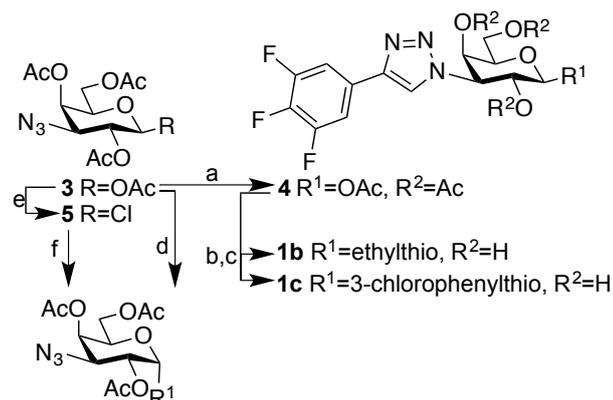
by modifying natural carbohydrate core structures with unnatural chemical groups,^[2a] for example the heparin mimetic Fondaparinux,^[4] nM-affinity inhibitors of the uropathogenic *E. coli* adhesin FimH obtained by optimized interactions with CRD-lining tyrosine side chains,^[5] sodium glucose transporter (SGLT2) inhibitors of the gliofzin family,^[6] influenza neuraminidase inhibitors Zanamivir and Oseltamivir,^[7] selectin inhibitors,^[8] siglec inhibitors based on optimized substituents on a sialic acid core structure,^[9] and galectins,^[10] the topic of this report.

Similar to other lectins, the galectin carbohydrate-binding site is shallow, found along the concave side of the ~130 amino acid β -sandwich carbohydrate recognition domain (CRD).^[11] The carbohydrate binding site contains the galectin-defining galactoside-binding site conferred by a conserved motif of about 7 amino acids. By itself, this subsite has weak binding to galactosides, with K_d in the mM range. Addition of saccharides on either side of the galactose can significantly enhance affinity, but also decrease it. In the most commonly used galectin saccharide inhibitors lactose, N-acetyl-lactosamine, and thiodigalactoside the addition of a monosaccharide on the reducing side of the galactose increases affinity by 10-100 fold, to K_d s in the mid μ M range. Previously we achieved much higher affinities (nM) by derivatizing such disaccharides with artificial moieties that targets additional sites at either end, that is C3-derivatization of N-acetyl-lactosamine and C3,C3'-derivatization of thiodigalactoside with aromatic ester,^[12] amide,^[10a, 10b, 13] or triazole^[10c, 14] moieties. Recently we found that C3 multifluorinated phenyl groups providing orthogonal multipolar fluorine-amide interactions strongly enhanced affinity for galectin-3.^[10c, 14a] This inspired attempts to replace the monosaccharide at the reducing side of galactose with a less polar aglycon, with the aim of still reaching high affinity while keeping the galactose derivatized with a C3 trifluorophenyl group. Indeed, with 4-methylphenylthio as the aglycon **1a**, single digit μ M affinity for galectin-3 was reached.^[14a]

Here we report further optimization of the thioglycosidic aglycon that affords galectin-3 inhibitors with low nM-affinities – unprecedented for a monosaccharide galectin inhibitor. Series of β - (**1b-c**) and α -thio-D-galactopyranosides (**2a-m**) carrying the same C3 4-(3,4,5-trifluorophenyl)-1H-triazole substituent were synthesized (Scheme 1) and affinities compared with **1a** using a competitive fluorescence anisotropy assay (Tables 1-3).^[10c]

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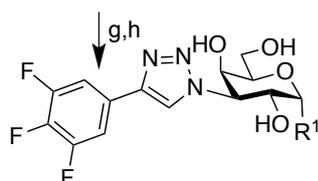


via route e,f

- 6c** R¹=3-chlorophenylthio
6e R¹=phenylthio
6f R¹=3-bromophenylthio
6g R¹=3-iodophenylthio
6h R¹=3,4-dichlorophenylthio
6i R¹=3-chloro-4-cyanophenylthio
6j R¹=2,3-dichlorophenylthio

via route d

- 6a** R¹=4-methylphenylthio
6b R¹=ethylthio
6d R¹=4-chlorophenylthio
6k R¹=3,4-dichlorophenoxy



- 2a** R¹=4-methylphenylthio
2b R¹=ethylthio
2c R¹=3-chlorophenylthio
2d R¹=4-chlorophenylthio
2e R¹=phenylthio
2f R¹=3-bromophenylthio
2g R¹=3-iodophenylthio
2h R¹=3,4-dichlorophenylthio
2i R¹=3-chloro-4-cyanophenylthio
2j R¹=2,3-dichlorophenylthio
2k R¹=3,4-dichlorophenoxy

Scheme 1. Synthesis of compounds **1a-c** and **2a-k**. Reagents and conditions: (a) 3,4,5-trifluorophenylacetylene, CuI, DIPEA, toluene, 40 °C, 49%; (b) R¹-SH, BF₃·OEt₂, mol. sieves, DCM, overnight, 29-50%; (c) NaOMe, MeOH, 2h, 25%-87%; (d) R¹-SH or R¹-OH, BF₃·OEt₂, mol. sieves, DCM or 1,2-dichloroethane, overnight, 0-r.t., 4 days, 21-48%; (e) PCl₅, BF₃·OEt₂, DCM, 20 min., 91%; (f) R¹-SH, CsCO₃ or NaH, 50 °C, DMF 25-66%; (g) 1,2,3-trifluoro-5-[2-(trimethylsilyl)ethynyl]benzen, CuI, DIPEA, toluene, 40 °C; (h) NaOMe, MeOH, 2h, 23-91% over two steps.

Influence of the anomeric configuration. Much to our surprise, α -D-thio-galactopyranoside **2a** had one order of magnitude higher affinity for galectin-3 compared to the reference β -D-thio-galactopyranoside **1a**. For an aliphatic aglycon, the α -anomer **2b** also had enhanced affinity over the corresponding β -anomer **1b**, albeit with a smaller 5-fold difference. Since the methyl α - and β -D-galactopyranosides have similar affinities for galectin-3,^[15] this suggests that the larger α -aglycons finds new interactions with the galectin that are not typically exploited by natural ligands and that have not previously been explored for artificial ligand design.

Table 1. K_d (μ M) values for **1a-b** and **2a-b** for human galectin-3 determined by competitive fluorescence polarization.^[10c]

Compound	Structure	K _d
1a		5.2 ^[14a]
2a		0.33±0.033
1b		5.1±0.53
2b		1.0±0.087

Influence of phenyl aglycon halogen substituents. To explore these potential new interactions of the α -aglycon, we used the phenyl **2e** (similar affinity as **2a**) as a scaffold and examined different halogen substituents. The 3-chloro **2c** enhanced affinity for galectin-3 by another order of magnitude leading to K_d about 50 nM whereas the 4-chloro **2d** did not. 3-Bromo (**2f**) and 3-iodo (**2g**) also lead to enhanced affinity. Introduction of an electron-withdrawing substituent, 4-chloro (**2h**) or 4-cyano (**2i**), next to the 3-chloro enhanced affinity further by a factor of about two, reaching a remarkable K_d of 23 nM for **2i**. In contrast, addition of a 2-chloro to the 3-chloro **2j** decreased affinity by one order of magnitude. The β -anomer **1c** of **2c** had 30-fold lower affinity (K_d 1.60±0.078 μ M) for galectin-3, but was among the best β -anomers (cf. **1a** and **1b** in Table 1).

Influence of the anomeric sulfur. The importance of the α -anomeric sulfur is apparent since the O-glycoside **2k** had about 15-fold lower affinity (K_d 0.49±0.024 μ M) compared to the corresponding S-glycoside **2h**.

Structural analysis. The structure activity relationships (SAR) described above suggests that the strong affinity enhancement of some compounds with α -linked aglycons is due to a subtle combination of different types of sterically precise interactions with galectin-3. To analyze these further, complexes of the galectin-3 CRD (galectin-3C) with one of the best α -glycosides (**2h**) and an analogous β -compound (**1c**) were compared by X-ray crystallography. The crystals diffracted to 1.2 and 1.5 Å, respectively, and clear ligand density was observed for the galactose residue and its 3C substituent. As expected, their positions were essentially identical for the two compounds, including the orthogonal multipolar ligand fluorine-amide interaction with R144, I145, and S237, and the ligand phenyl-R144 side-chain stacking, and similar as observed earlier for disaccharide derivatives^[10c] (Figure 1c and d). The electron density maps were weaker for the aglycons replacing glucose and these showed double conformations for **1c** (Figure 1a), modeled with 0.3 and 0.7 occupancy. Despite this, the key affinity enhancing atoms - the 3-chloro substituent of the phenyl aglycon and the glycosidic sulfur - could be clearly identified, and the phenyl ring modeled.

Table 2. K_d (μM) values for **2c-j** for human galectin-3 determined by competitive fluorescence polarization.^[10c]

Compound		K_d
2c	$R^1=R^3=\text{H}, R^2=\text{Cl}$	0.049 ± 0.0027
2d	$R^1=R^2=\text{H}, R^3=\text{Cl}$	0.38 ± 0.022
2e	$R^1=R^2=R^3=\text{H}$	0.52 ± 0.038
2f	$R^1=R^3=\text{H}, R^2=\text{Br}$	0.031 ± 0.0024
2g	$R^1=R^3=\text{H}, R^2=\text{I}$	0.058 ± 0.0043
2h	$R^1=\text{H}, R^2=R^3=\text{Cl}$	0.037 ± 0.0010
2i	$R^1=\text{H}, R^2=\text{Cl}, R^3=\text{CN}$	0.047 ± 0.0063
2j	$R^1=R^2=\text{Cl}, R^3=\text{H}$	0.85 ± 0.031

For both compounds **1c** and **2h**, the 3-chloro substituent has the direction and position expected to form a halogen bond with the backbone carbonyl oxygen of G182 in the protein, which provides an explanation for the tenfold higher affinity of **2c** and **2h** over **2d** and **2e**. The 4-chloro of **2d** and **2h** points out in solution and cannot form a halogen bond with the protein, and does not enhance affinity by itself (cf. **2d/2e**). Halogen bond strengths can be enhanced by increasing the size of the halide σ -hole.^[16] One way is to replace the chloro substituent with a larger halide; a small affinity enhancement was found here with bromide **2f**, but not with iodide **2g** which may have been too large to fit. Another way to increase the size of the halide σ -hole is to introduce an electron withdrawing group in the vicinity. Adding 4-chloro (**2h**) or a 4-cyano group (**2i**) enhanced affinity compared to the 3-chloro **2c** by about 2-fold. In contrast, adding 2-chloro **2j** decreased affinity by 10-fold possibly due to steric conflict with the protein.

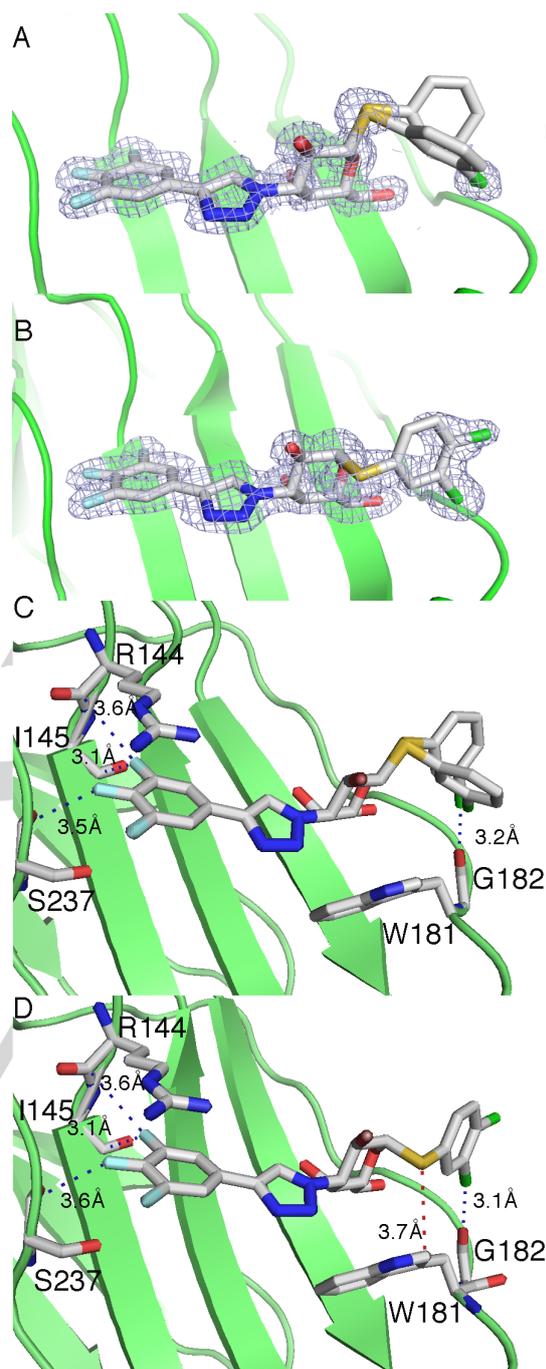


Figure 1. A-B) Electron density map (grey mesh) $2|F_o| - |F_c|$ cc contoured at 1σ for **1c** and **2h** in complex with galectin-3C. Galectin-3C in complex with C) **1c** revealing fluorine-amide carbonyl interactions with residues R144, I145, and S237, and an X-bond interaction with G182 backbone carbonyl oxygen and with D) **2h** revealing fluorine-amide carbonyl interactions with residues R144, I145, and S237, a S- π interaction with W181, and an X-bond interaction with G182 backbone carbonyl oxygen.

The anomeric sulfur of the α -D-galactopyranoside **2h** is positioned near W181 suggesting a beneficial sulfur- π interaction^[17] not shared by the β -galactosides. This could be an important contribution to the affinity differences in the α/β anomeric pairs **1a/2a** (10 fold), **1b/2b** (5 fold), and **1c/2c** (30 fold). In addition, the oxygen analog **2k** has a 15-fold lower affinity than the corresponding thio-galactoside **2h**, which also supports the hypothesis of affinity enhancing effects of the α -anomeric sulfur. The longer C-S bonds and/or smaller C-S-C bond angle of the α -thio-galactosides **2a-2j** may also be important to place the aglycon in a favorable position compared to the equivalent values for O to interact with galectin-3, particularly for 3-chloro to form an optimal halogen bond.

The phenyl aglycon itself did not show evidence for any strong interactions with the protein, which may explain why replacing it with an aliphatic ethyl aglycon in **2b** only led to a 2-fold decrease in affinity (compared to **2a**). Instead it may be more important as a scaffold to position the phenyl 3-chloro substituent to form a halogen bond with G182.

Galectin selectivity. Having arrived at monosaccharide inhibitors with exceptional affinities for galectin-3, the important question of selectivity for different members of the galectin family was addressed (Table 3). In comparison with galectin-3, compound **2h** had >100-fold lower affinity for most of the tested human galectins, and 20 and 4-fold lower affinity for galectin-2 and the C-terminal CRD of galectin-4, respectively. In contrast, methyl α -D-galactopyranoside shows both poor affinity and selectivity for the galectins investigated. Hence, at least some of the specific interactions of the artificial substituents contributing to the high galectin-3 affinity also contribute to the selectivity over other galectins.

Table 3. K_d (μ M) values of **2h** and methyl α -D-galactopyranoside for human galectins determined by competitive fluorescence polarization.^[10c]

Galectin	2h	Me α -gal
1	3.7 \pm 0.15	>10000 ^[15]
2	0.64 \pm 0.11	>20000
3	0.037 \pm 0.0010	2700 ^[15]
4C ^a	0.13 \pm 0.012	>20000
4N ^b	2.9 \pm 0.40	>20000
7	31 \pm 3.7	11000 ^[15]
8C ^a	11 \pm 1.9	>20000
8N ^b	83 \pm 17	6300 ^[15]
9C ^a	2.4 \pm 0.41	6200 \pm 220
9N ^b	2.7 \pm 0.24	2800 ^[15]

[a] C-terminal domain. [b] N-terminal domain.

Conclusion. Derivatizing a low-affinity monosaccharide with functionalities forming a combination of orthogonal multipolar fluorine-amide, phenyl-arginine, sulfur- π , and halogen bond interactions, results in lectin ligands with affinities far surpassing those of common natural ligand fragments (e.g. about 100000 fold more potent than methyl β -D-galactoside^[18] and 5000 fold more potent than methyl β -lactoside^[19]); removal of any of these interactions results in significant loss of affinity. The compounds are the smallest high affinity galectin-3 inhibitors described and thus constitute a new class of promising drug lead structures. We suggest that systematic introduction of interactions, as the ones described here, can be a very useful strategy discovering small ligands that target shallow and polar lectin carbohydrate-binding sites, increasing the drugability for any such target. Polar and sp³-rich monosaccharide scaffolds as drug discovery starting points differ substantially from the small, aromatic, and lipophilic starting scaffolds typically generated by fragment-based lead generation or HTS strategies, and hence may also provide a useful alternative strategy for a broader range of targets.

Experimental Section

Synthetic procedures, crystallization experiments, and data collection and refinement are described in the supplementary information.

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Keywords: lectin • galectin-3 • sulfur- π • halogen bond • fluorine multipolar interaction • inhibitor

- [1] A. Varki, *Glycobiology* **2016**, *27*, 3-49.
- [2] a) B. Ernst, J. L. Magnani, *Nat. Rev. Drug Disc.* **2009**, *8*, 661-677; b) A. Fernández-Tejada, F. J. Cañada, J. Jiménez-Barbero, *Chem. Eur. J.* **2015**, *21*, 10616-10628; c) J. E. Hudak, C. R. Bertozzi, *Chem. Biol.* **2014**, *21*, 16-37.
- [3] a) C. Diehl, O. Engström, T. Delaine, M. Håkansson, S. Genheden, K. Modig, H. Leffler, U. Ryde, U. J. Nilsson, M. Akke, *J. Am. Chem. Soc.* **2010**, *132*, 14577-14589; b) C. MacRaild, A. Daranas, A. Bronowska, S. Homans, *J. Mol. Biol.* **2007**, *368*, 822-832; c) I. V. Nesmelova, E. Ermakova, V. A. Daragan, M. Pang, M. Menéndez, L. Lagartera, D. Solís, L. G. Baum, K. H. Mayo, *J. Mol. Biol.* **2010**, *397*, 1209-1230.
- [4] M. Petitou, C. van Boeckel, *Angew. Chem. Intl ed. Engl.* **2004**, *43*, 3118-3133.
- [5] a) S. Kleeb, X. Jiang, P. Frei, A. Sigl, J. Bezençon, K. Bamberger, O. Schwardt, B. Ernst, *J. Med. Chem.* **2016**, *59*, 3163-3182; b) S. Kleeb, L. Pang, K. Mayer, D. Eris, A. Sigl, R. C. Preston, P. Zihlmann, T. Sharpe, R. P. Jakob, D. Abgottspon, A. S. Hutter, M. Scharenberg, X. Jiang, G.

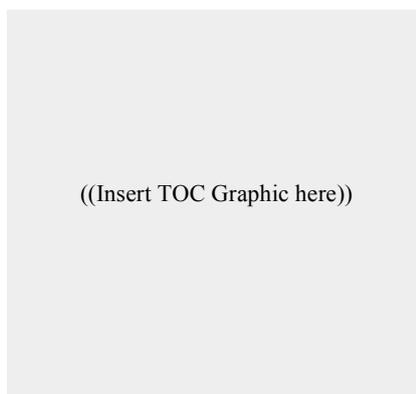
- Navarra, S. Rabbani, M. Smiesko, N. Lüdin, J. Bezençon, O. Schwardt, T. Maier, B. Ernst, *J. Med. Chem.* **2015**, *58*, 2221-2239.
- [6] Y. Zhang, Z. P. Liu, *Curr. Med. Chem.* **2016**, *23*, 832-849.
- [7] E. De Clercq, *Nat. Rev. Drug Discovery* **2006**, *5*, 1015-1025.
- [8] J. Egger, C. Weckerle, B. Cutting, O. Schwardt, S. Rabbani, K. Lemme, B. Ernst, *J. Am. Chem. Soc.* **2013**, *135*, 9820-9828.
- [9] a) S. Kelm, *J. Exp. Med.* **2002**, *195*, 1207-1213; b) H. Prescher, A. Schweizer, E. Kuhfeldt, L. Nitschke, R. Brossmer, *ACS Chem. Biol.* **2014**, *9*, 1444-1450.
- [10] a) I. Cumpstey, E. Salomonsson, A. Sundin, H. Leffler, U. J. Nilsson, *Chem. Eur. J.* **2008**, *14*, 4233-4245; b) I. Cumpstey, A. Sundin, H. Leffler, U. J. Nilsson, *Angew. Chem. Int. Ed.* **2005**, *44*, 5110-5112; c) T. Delaine, C. Collins, A. Mackinnon, G. Sharma, J. Stegmayr, V. K. Rajput, S. Mandal, I. Cumpstey, A. Larumbe, B. A. Salameh, H. von Wachenfeldt, B. Kahl-Knutsson, H. van Hattum, M. van Scherpenzeel, R. J. Pieters, T. Sethi, H. Schambye, H. Leffler, H. Blanchard, U. J. Nilsson, *ChemBioChem* **2016**, *17*, 1759-1770.
- [11] a) S. Di Lella, V. Sundblad, J. Cerliani, C. M. Guardia, D. A. Estrin, G. R. Vasta, G. A. Rabinovich, *Biochemistry* **2011**, *50*, 7842-7857; b) S. P. M. Mendez-Huergo, A. G. Blidner, G. A. Rabinovich, *Curr. Opin. Immun.* **2017**, *45*, 8-15.
- [12] T. Delaine, I. Cumpstey, L. Ingrassia, M. Le Mercier, P. Okechukwu, H. Leffler, R. Kiss, U. J. Nilsson, *J. Med. Chem.* **2008**, *51*, 8109-8114.
- [13] a) P. Sörme, P. Arnoux, B. Kahl-Knutsson, H. Leffler, J. M. Rini, U. J. Nilsson, *J. Am. Chem. Soc.* **2005**, *127*, 1737-1743; b) P. Sörme, Y. Qian, P.-G. Nyholm, H. Leffler, U. J. Nilsson, *ChemBioChem* **2002**, *3*, 183-189.
- [14] a) K. Peterson, R. Kumar, O. Stenström, P. Verma, P. R. Verma, M. Håkansson, B. Kahl-Knutsson, F. Zetterberg, H. Leffler, M. Akke, D. T. Logan, U. J. Nilsson, *submitted 2017*; b) B. A. Salameh, I. Cumpstey, A. Sundin, H. Leffler, U. J. Nilsson, *Bioorg. Med. Chem.* **2010**, *18*, 5367-5378.
- [15] V. K. Rajput, H. Leffler, U. J. Nilsson, B. Mukhopadhyay, *Bioorg. Med. Chem. Lett.* **2014**, *24*, 3516-3520.
- [16] G. Cavallo, P. Metrangolo, R. Milani, T. Pilati, A. Priimagi, G. Resnati, G. Terraneo, *Chem. Rev.* **2016**, *116*, 2478-2601.
- [17] a) B. R. Beno, K.-S. Yeung, M. D. Bartberger, L. D. Pennington, N. A. Meanwell, *J. Med. Chem.* **2015**, *58*, 4383-4438; b) C. R. Forbes, S. K. Sinha, H. K. Ganguly, S. Bai, G. P. A. Yap, S. Patel, N. J. Zondlo, *J. Am. Chem. Soc.* **2017**, *139*, 1842-1855.
- [18] I. Cumpstey, S. Carlsson, H. Leffler, U. J. Nilsson, *Org. Biomol. Chem.* **2005**, *3*, 1922-1932.
- [19] I. Cumpstey, E. Salomonsson, A. Sundin, H. Leffler, U. J. Nilsson, *ChemBioChem* **2007**, *8*, 1389-1398.

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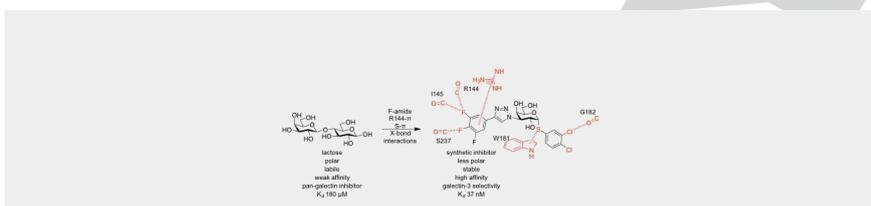
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