Total Synthesis of Mniopetals A, B, C and D

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Dedicated to Prof. Dr. W. Steglich on the occasion of his 80th birthday

Abstract: A total synthesis of the mniopetals A, B, C and D is described. Key steps are a Sharpless asymmetric dihydroxylation and a selective esterification of an equatorial hydroxy group vicinal to an axial hydroxy function.

Key words: asymmetric synthesis, antiviral agents, chemoselectivity, intramolecular Diels–Alder reaction, natural products

In 1994, Anke and Steglich reported on the isolation, structure elucidation and determination of the absolute configuration of six new drimane-type sesquiterpenes from the culture broth of the Canadian fungus Mniopetalum sp. 87256, the mniopetals A-F (**1a–f**; Figure 1).¹



Figure 1 Structures of mniopetals A-F (1a-f)

The mniopetals inhibit the reverse transcriptase of HIV-1, which makes them attractive targets for total synthesis.²

Some years ago, we developed short syntheses of mniopetals E and F.^{2b-d,2g} Esterification of the two secondary alcohol functions with the respective α -hydroxy or α -acetoxy carboxylic acids should lead directly to the mniopetals A–D. Therefore, it was necessary to find a simple

SYNLETT 2013, 24, 1410–1414 Advanced online publication: 10.06.2013 DOI: 10.1055/s-0033-1339172; Art ID: ST-2013-B0271-L © Georg Thieme Verlag Stuttgart · New York strategy for the construction of these acids in enantiomerically pure form (Figure 1) and suitable methods for esterification with mniopetal E. Here, we wish to report our efforts in this direction.

(*R*)-2-Acetoxydecanoic acid (9), MEM-protected (*R*)-2hydroxydecanoic acid (10) and MEM-protected (*R*)-2-hydroxyoctanoic acid (11) were synthesized starting with a Sharpless asymmetric dihydroxylation with AD-MIX- β of the corresponding terminal olefin (Scheme 1).³



Scheme 1 Common synthetic strategy for the carboxylic acid side chains 9, 10 and 11

In order to acetylate the secondary hydroxyl function in 4, the primary one had to be protected temporarily. This could be accomplished in a one-pot procedure by silylation with TBDMSCl–pyridine, acetylation with Ac₂O and finally removal of the TBDMS group with dilute HCl to give 6 in good yield.^{3a} Oxidation of 6 with RuCl₃–NaIO₄^{3a} yielded 9 (side chain for 1a). For the other side chains, the primary hydroxy group of 4 and 5 was silylated, then the secondary hydroxy group was protected as MEM acetal⁴ and the silyl protecting group was removed under standard conditions to give 7 and 8. Oxidation of the primary hydroxy group to the carboxylic acid in two

steps⁴ gave **10** and **11**, respectively. The ee and the absolute configuration of hydroxyacids **10** and **11** were determined according to Mosher's method.⁵ The enantiomeric excess of the synthesized acids exceeds 98%.

With the side chain acids for the mniopetals A–D in hand, esterification of **9** with the mniopetal E precursor 12^{2d} was straightforward, but the final cleavage of the menthyl acetal under standard conditions led to complete decomposition of the product (Scheme 2). Thus, we exchanged the menthyl group with a THP protecting group,^{6,7} starting from 13^{2d} according to Scheme 2. Compound 15 was finally *cis*-dihydroxylated chemo- and stereoselectively^{2,8} to yield 16.



Scheme 2 Cleavage of the menthyl acetal after esterification of 12 with 9 was not possible. Therefore the menthyl group in 11 had to be exchanged with a THP group to give 16.

Mniopetal E precursor **16** contains equatorial and axial hydroxy groups at C2 and C1, respectively. Esterification of the equatorial hydroxy group is possible without protection of the axial hydroxy group (Scheme 3).⁹ For the carboxylic acids **9**, **10** and **11** the Steglich esterification with DCC–DMAP¹⁰ in dichloromethane is the method of choice (Scheme 3).

The THP group in 17 was cleaved with PPTS in acetone– water¹¹ to obtain mniopetal A (1a) in 65% yield. Mniopetals B and C (1b, 1c) were obtained in 40% and 45% yield from 18 and 19, respectively, after deprotection of the THP and MEM acetals in one step with $ZnBr_2$ – CH_2Cl_2 (average yield for cleavage of each protecting group was 63–65%).

For the synthesis of mniopetal D (1d) the equatorial hydroxy group of diol 16 had to be protected selectively, which was achieved with TBDMSOTf-2,6-lutidine in dichloromethane at -78 °C (Scheme 4).¹²



Scheme 3 Syntheses of mniopetals A, B and C (1a, 1b and 1c)



Scheme 4 Synthesis of mniopetal D

After silylation of the hydroxy group at C2, ¹³ leading to **20**, Steglich esterification of the hydroxy group at C1 was not possible, probably due to a shielding effect of the huge silyl group and the innately lower reactivity of an axial hydroxy function. After extensive experimentation, we found that the Yamaguchi esterification with 2,4,6-trichlorobenzoyl chloride, hydroxyacid **10**, triethylamine and DMAP¹⁴ gives **21** in 72% yield. The following desilylation has to be done with Py·HF complex, ^{15,16} since with TBAF migration of the acyl group to the equatorial hydroxyl group occurs. Finally, the acetal protecting groups MEM and THP were both cleaved with ZnBr₂– CH₂Cl₂ leading to mniopetal D (**1d**) in 55% yield (average 74% per protecting group).

All synthesized mniopetals were identical in all respects with the isolated products.

In summary, we have developed short and efficient syntheses of all known mniopetals.¹⁷ Key features of our strategy are a straightforward construction of the side chain carboxylic acids and fine-tuned methods to manipulate an equatorial hydroxy group vicinal to an axial hydroxy group and vice versa.

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- (7) Synthesis of Compound 15: 3,4-Dihydro-2*H*-pyran (565 μ L, 6.18 mmol) was added to a mixture of 14 (324 mg, 1.24 mmol) and pyridinium p-toluenesulfonate (31.0 mg, 0.124 mmol) in CH₂Cl₂ (5.0 mL) and the reaction mixture was stirred at r.t. for 2 h. Sat. aq NaHCO3 solution was added. Extraction with CH₂Cl₂, drying over MgSO₄ and purification by flash chromatography (petroleum ether- Et_2O , 2:1; silica gel, $R_f 0.18$ and 0.24) led to 15 (348 mg, 1.00 mmol, 81.0%). Specifications for one THP-diastereomer (R_f 0.24): ¹H NMR (500 MHz, CDCl₃): $\delta = 9.47$ (s, 1 H, H-12), 7.17 (br d, J = 6.4 Hz, 1 H, H-7), 6.18 (dt, J = 10.2, 1.8 Hz, 1 H, H-1), 5.88 (dt, J = 10.2, 4.0 Hz, 1 H, H-2), 5.53 (s, 1 H, H-11), 5.07 (t, J = 2.8 Hz, 1 H, H-1'), 4.08 (dt, J = 11.1, 2.6Hz, 1 H, H-5 $'_{ax}$), 3.65 (br d, J = 11.5 Hz, 1 H, H-5 $'_{eq}$), 3.20 (br s, 1 H, H-9), 2.55 (dm, J = 19.5 Hz, 1 H, H-6_{ax}), 2.43 $(ddm, J = 19.5, 12.5 Hz, 1 H, H-6_{eq}), 2.00 (dd, J = 4.0, 2.1)$ Hz, 2 H, H-3), 1.82–1.93 (m, 1 H, H-3'_{ax}), 1.80–1.63 (m, 5 H, H-5, H-2', H-4'), 1.57–1.63 (m, 1 H, H-3'_{eq}), 1.24 (s, 3 H, H-14), 1.04 (s, 3 H, H-13). ¹³C NMR (125 MHz, CDCl₃): δ = 192.7 (C-12), 175.3 (C-15), 156.0 (C-7), 138.9 (C-8),

129.5 (C-2), 126.2 (C-1), 99.2 (C-11), 94.7 (C-1'), 61.4 (C-5'), 49.9 (C-9), 48.2 (C-10), 44.9 (C-5), 41.2 (C-3), 31.14 (C-13), 31.05 (C-4), 29.8 (C-2'), 25.2 (C-4'), 24.6 (C-6), 24.2 (C-14), 18.1 (C-3'). $[\alpha]^{20}_{D}$ –22.2 (CHCl₃, *c* = 0.1). HRMS (ESI⁺): *m*/*z* [M + Na]⁺ calcd for C₂₀H₂₆O₅Na: 369.1678; found: 369.1634.

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- (14) **Synthesis of Compound 21**: Compound **10** (21.8 mg, 79.0 μ mol) was diluted in toluene (900 μ L). 2,4,6-Trichlorobenzoyl chloride (10.4 μ L, 67.0 μ mol) and Et₃N (17.0 μ L, 121 μ mol) were added and the solution was stirred for 2 h at r.t. After this a mixture of **20** (30 mg, 61.0 μ mol) and DMAP (9.6 mg, 79 μ mol) in toluene (900 μ L) was added and the solution was stirred for 24 h at r.t.. Sat. aq NaHCO₃ solution was added and extraction with EtOAc, drying over MgSO₄, filtration and evaporation afforded a yellow oil,

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which was purified by flash chromatography (petroleum ether–Et₂O, 1:1; silica gel, R_f 0.13 and 0.09). Yield: 31 mg (41.0 µmol, 67.9%). Specifications for one THPdiastereomer ($R_f 0.09$): ¹H NMR (500 MHz, CDCl₃): $\delta =$ 9.43 (s, 1 H, H-12), 7.11 (br d, *J* = 6.4 Hz, 1 H, H-7), 5.86 (br s, 1 H, H-1), 5.31 (s, 1 H, H-11), 5.05 (br s, 1 H, H-1"), 4.75 (s, 2 H, H-11'), 4.36 (ddd, J = 12.8, 4.1, 2.4 Hz, 1 H, H-2), 4.22 (dd, *J* = 7.5, 5.0 Hz, 1 H, H-2'), 3.81–3.91 (m, 1 H, $H-5''_{ax}$), 3.76 (ddd, $J = 11.0, 6.3, 3.2 Hz, 1 H, H-12'_{a}$), 3.67 $(ddd, J = 11.0, 5.6, 3.2 \text{ Hz}, 1 \text{ H}, \text{H}-12'_{b}), 3.48-3.63 \text{ (m, 3 H},$ H-5"_{eq}), 3.38 (s, 3 H, H-14'), 3.22 (br s, 1 H, H-9), 2.48 (ddd, $J = 19.1, 5.8, 3.0 \text{ Hz}, 1 \text{ H}, \text{H-6}_{ax}), 2.18-2.29 \text{ (m, 1 H, H-6}_{eq}),$ 1.99–2.12 (m, 1 H, H-3"_{ax}), 1.38–1.85 (m, 12 H, H-3, H-5, H-3', H-4', H-2", H-3"_{eq}, H-4"), 1.31 (s, 3 H, H-14), 1.21– 1.31 (m, 10 H, H-5', H-6', H-7', H-8', H-9'), 1.05 (s, 3 H, H-13), 0.86–0.90 (m, 3 H, H-10'), 0.86 (s, 9 H, t-Bu), 0.07 (s, 3 H, SiMe), 0.05 (s, 3 H, SiMe). ¹³C NMR (125 MHz, CDCl₃): $\delta = 192.4$ (C-12), 175.2 (C-15), 170.7 (C-1'), 154.2 (C-7), 137.9 (C-8), 103.8 (C-11), 99.6 (C-1"), 95.0 (C-11'), 75.4 (C-2'), 72.7 (C-1), 71.6 (C-13'), 67.6 (C-12'), 65.9 (C-2), 61.4 (C-5"), 59.0 (C-14'), 52.2 (C-10), 47.4 (C-9), 43.0 (C-3), 40.7 (C-5), 33.6 (C-13), 33.5 (C-4), 33.2 (C-3'), 31.9 (C-8'), 29.5 (C-2"), 29.33 (C-5'), 29.31 (C-6'), 29.25 (C-7'), 25.8 (t-Bu), 25.07 (C-4'), 25.05 (C-4''), 24.7 (C-6), 23.5 (C-14), 22.6 (C-9'), 18.3 (t-Bu), 17.6 (C-3"), 14.1 (C-10'), -5.1 (SiMe), -5.3 (SiMe). $[\alpha]^{20}_{D}$ -25.3 (CHCl₃, c = 0.3). HRMS (ESI⁺): m/z [M + Na]⁺ calcd. for C₄₀H₆₈O₁₁SiNa: 775.4423; found: 775.4400.

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- (16) Cleavage of the Silvl Protection Group in 21: Compound 21 (30.0 mg, 40.0 µmol) was diluted in HF pyridine (1.14 mL, 1.59 mmol) in THF-pyridine prepared according to a procedure of Trost's (see ref. 15a). The solution was stirred at r.t. for 24 h. Sat. aq NaHCO₃ solution was added. Extraction with EtOAc, drying over MgSO₄, filtration and evaporation afforded a yellow oil, which was purified by flash chromatography (petroleum ether-acetone, 5:1; silica gel, R_f 0.12). Yield: 20.0 mg (31.0 µmol, 79.0%). Specifications for one THP-diastereomer: ¹H NMR (500 MHz, CDCl₃): $\delta = 9.47$ (s, 1 H, H-12), 7.11 (d, J = 6.5 Hz, 1 H, H-7), 5.76 (s, 1 H, H-1), 5.40 (s, 1 H, H-11), 4.96 (br s, 1 H, H-1"), 4.67-4.77 (m, 2 H, H-11'), 4.29-4.39 (m, 1 H, H-2), 4.21–4.28 (m, 1 H, H-2'), 3.93 (dd, *J* = 11.4, 2.5 Hz, 1 H, H-5"_{ax}), 3.69–3.77 (m, 1 H, H-12'_a), 3.56–3.62 (m, 1 H, H-12′_b), 3.53–3.56 (m, 1 H, H-5″_{eq}), 3.45–3.50 (m, 2 H, H-13′), 3.32 (s, 3 H, H-14'), 3.30 (br s, 1 H, H-9), 2.44 (dm, J=19.3 Hz, 1 H, H- 6_{ax}), 2.14–2.26 (m, 1 H, H- 6_{eq}), 1.82 (d, J = 3.3Hz, 1 H, OH), 1.66–1.77 (m, 4 H, H-3', H-2"), 1.47–1.65 (m, 7 H, H-3, H-5, H-3", H-4"), 1.33–1.44 (m, 2 H, H-4'), 1.24 (s, 3 H, H-14), 1.16-1.27 (m, 10 H, H-5', H-6', H-7', H-8', H-9'), 1.00 (s, 3 H, H-13), 0.81 (t, J = 7.0 Hz, 3 H, H-10'). ¹³C NMR (125 MHz, CDCl₃): δ = 192.2 (C-12), 175.0 (C-15), 172.2 (C-1'), 153.8 (C-7), 137.8 (C-8), 99.9 (C-11), 94.9 (C-11'), 94.4 (C-1"), 75.4 (C-2'), 72.9 (C-1), 71.7 (C-13'), 67.6 (C-12'), 65.7 (C-2), 61.2 (C-5"), 59.0 (C-14'), 52.3 (C-10), 45.9 (C-9), 41.6 (C-5), 41.1 (C-3), 33.6 (C-13), 33.5 (C-4), 33.0 (C-3'), 31.9 (C-8'), 29.8 (C-2"), 29.4 (C-5'), 29.3 (C-6'), 29.2 (C-7'), 25.1 (C-4'), 25.0 (C-4''), 24.7 (C-6), 23.4 (C-14), 22.6 (C-9'), 17.5 (C-3''), 14.1 (C-10'). $[\alpha]^{20}_{D}$ -32.1 (CHCl₃, c = 1.2). HRMS (ESI⁺): m/z [M + Na]⁺ calcd for C₃₄H₅₄O₁₁Na: 661.3564; found: 661.3549.
- (17) Mniopetal A (**1a**): ¹H NMR (500 MHz, CDCl₃): $\delta = 9.44$ (s, 1 H, H-12), 7.12 (br d, J = 6.7 Hz, 1 H, H-7), 5.48 (br s, 1 H,

H-11), 5.37 (ddd, J = 12.7, 4.0, 2.3 Hz, 1 H, H-2), 4.87 (dd, J = 6.5, 6.5 Hz, 1 H, H-2'), 4.59 (br s, 1 H, H-1), 3.89 (br s, 1 H, H-9), 2.46 (dm, J = 19.5 Hz, 1 H, H-6_{ax}), 2.24 (dm, J =19.5 Hz, 1 H, H- 6_{eq}), 2.14 (s, 3 H, H-12'), 2.08 (dd, J = 12.6, 12.6 Hz, 1 H, H-3_{ax}), 1.81–1.85 (m, 2 H, H-3'), 1.77 (dd, J= 12.7, 3.4 Hz, 1 H, H-5), 1.50 (dd, J = 12.2, 3.6 Hz, 1 H, H-3_{eq}), 1.37–1.44 (m, 2 H, H-4'), 1.33 (s, 3 H, H-14), 1.20–1.30 (m, 10 H, H-5', H-6', H-7', H-8', H-9'), 1.05 (s, 3 H, H-13), 0.88 (t, J = 7.1 Hz, 3 H, H-10'). ¹³C NMR (125 MHz, CDCl₃): δ = 193.3 (C-12), 175.9 (C-15), 171.7 (C-11'), 169.6 (C-1'), 154.8 (C-7), 138.5 (C-8), 99.6 (C-11), 73.0 (C-2'), 71.1 (C-2), 68.1 (C-1), 53.4 (C-10), 46.2 (C-9), 39.7 (C-5), 37.5 (C-3), 33.6 (C-4), 33.3 (C-13), 31.8 (C-8'), 30.9 (C-3'), 29.3 (C-5'), 29.14 (C-6'), 29.11 (C-7'), 25.1 (C-4'), 24.7 (C-6), 23.3 (C-14), 22.6 (C-9'), 20.7 (C-12'), 14.1 (C-10'). [α]²⁰_D -64.0 (CHCl₃, c = 0.4); Lit.^{1b} $[\alpha]^{20}_{D} - 63$ (CHCl₃, c = 0.05). HRMS (ESI⁺): m/z [M + Na]⁺ calcd for C₂₇H₄₀O₉Na: 531.2570; found: 531.2563. Mniopetal B (1b): ¹H NMR (500 MHz, CDCl₃): δ = 9.46 (s, 1 H, H-12), 7.18 (br d, J = 6.5 Hz, 1 H, H-7), 5.52 (br s, 1 H, H-11), 5.40 (ddd, J = 12.6, 4.0, 2.4 Hz, 1 H, H-2), 4.56 (br s, 1 H, H-1), 4.49 (br s, 1 H, OH), 4.18 (br s, 1 H, H-2'), 3.85 (br s, 1 H, H-9), 2.99 (br s, 1 H, OH), 2.92 (br s, 1 H, OH), 2.49 (dddm, J = 19.5, 6.7, 3.9 Hz, 1 H, H-6_{ax}), 2.26 (ddm, J = 19.5, 12.9 Hz, 1 H, H- 6_{eq}), 2.02 (ddd, J = 12.6, 12.6, 6.7 Hz, 1 H, H- 3_{ax}), 1.77 (dd, J = 12.6, 3.3 Hz, 1 H, H-5), 1.68– 1.74 (m, 1 H, H-3'_a), 1.62–1.67 (m, 1 H, H-3'_b), 1.55–1.59 (m, 1 H, H-3_{eq}), 1.37-1.43 (m, 2 H, H-4'), 1.35 (s, 3 H, H-14), 1.20–1.30 (m, 10 H, H-5', H-6', H-7', H-8', H-9'), 1.07 (s, 3 H, H-13), 0.88 (t, J = 6.8 Hz, 3 H, H-10'). ¹³C NMR (125 MHz, CDCl₃): δ = 193.2 (C-12), 175.7 (C-15), 174.4 (C-1'), 154.9 (C-7), 138.1 (C-8), 99.4 (C-11), 71.4 (C-2), 70.6 (C-2'), 68.8 (C-1), 53.4 (C-10), 46.2 (C-9), 39.6 (C-5), 37.4 (C-3), 34.5 (C-3'), 33.7 (C-4), 33.3 (C-13), 31.8 (C-8'), 29.4 (C-5'), 29.3 (C-6'), 29.2 (C-7'), 24.8 (C-4'), 24.7 (C-6), 23.2 (C-14), 22.7 (C-9'), 14.1 (C-10'). $[\alpha]^{20} - 47.3$ (CHCl₃, c = 0.1); Lit.^{1b} $[\alpha]^{20}_{D}$ -46 (CHCl₃, c = 0.05). HRMS (ESI⁺): m/z [M + Na]⁺ calcd for C₂₅H₃₈O₈Na: 489.2465; found: 489.2475. Mniopetal C (1c): ¹H NMR (500 MHz, CDCl₃): $\delta = 9.44$ (s, 1 H, H-12), 7.14 (br d, J = 6.8 Hz, 1 H, H-7), 5.51 (br s, 1 H, H-11), 5.38 (ddd, J = 12.7, 4.0, 2.3 Hz, 1 H, H-2), 4.56 (br s, 1 H, H-1), 4.18 (dd, J = 7.5, 4.1 Hz, 1 H, H-2'), 3.83 (br s, 1 H, H-9), 2.48 (ddd, J = 18.6, 6.0, 3.0 Hz, 1 H, H-6_{ax}), 2.25 $(ddm, J = 19.0, 12.5 Hz, 1 H, H-6_{eq}), 2.04 (dd, J = 12.6, 12.6)$ Hz, 1 H, H-3_{ax}), 1.76 (dd, *J* = 12.2, 3.9 Hz, 1 H, H-5), 1.69– $1.75 \text{ (m, 1 H, H-3'_a)}, 1.63-1.68 \text{ (m, 1 H, H-3'_b)}, 1.55 \text{ (dd, } J =$ 12.3, 4.2 Hz, 1 H, H-3_{eq}), 1.38–1.44 (m, 2 H, H-4'), 1.34 (s, 3 H, H-14), 1.24–1.33 (m, 6 H, H-5', H-6', H-7'), 1.06 (s, 3 H, H-13), 0.88 (t, J = 6.8 Hz, 3 H, H-8'). ¹³C NMR (125 MHz, CDCl₃): $\delta = 194.3$ (C-12), 175.9 (C-15), 174.4 (C-1'), 155.1 (C-7), 135.2 (C-8), 99.5 (C-11), 71.3 (C-2), 70.7 (C-2'), 68.6 (C-1), 53.5 (C-10), 46.2 (C-9), 39.6 (C-5), 37.4 (C-3), 34.4 (C-3'), 33.7 (C-4), 33.2 (C-13), 31.6 (C-6'), 28.9 (C-5'), 24.8 (C-4'), 24.7 (C-6), 23.2 (C-14), 22.5 (C-7'), 14.0 (C-8'). $[\alpha]^{20}_{D}$ -47.3 (CHCl₃, c = 0.1); Lit.^{1b} $[\alpha]^{20}_{D}$ -45 (CHCl₃, c= 0.05). HRMS (ESI⁺): m/z [M + Na]⁺ calcd for C₂₃H₃₄O₈Na: 461.2152; found: 461.2150. Mniopetal D (1d): ¹H NMR (500 MHz, CDCl₃): $\delta = 9.46$ (s, 1 H, H-12), 7.16 (br d, J = 6.6 Hz, 1 H, H-7), 5.84 (br s, 1 H, H-1), 5.50 (s, 1 H, H-11), 4.49 (s, 1 H, OH), 4.39-4.44 (m, 1 H, H-2), 4.29-4.33 (m, 1 H, H-2'), 3.27 (br s, 1 H, H-9), 2.75 (d, J = 5.7 Hz, 1 H, OH), 2.52 (ddd, J = 19.6, 6.7, 3.3 Hz, 1 H)H, H- 6_{ax}), 2.27 (ddd, J = 19.7, 12.8, 2.5 Hz, 1 H, H- 6_{eq}), 1.95 $(d, J = 2.8 \text{ Hz}, 1 \text{ H}, \text{OH}), 1.80-1.88 (m, 1 \text{ H}, \text{H-3}'_{a}), 1.57-$ 1.73 (m, 4 H, H-3, H-5, H-3'_b), 1.44–1.54 (m, 2 H, H-4'),

1.32 (s, 3 H, H-14), 1.23–1.30 (m, 10 H, H-5', H-6', H-7', H-8', H-9'), 1.08 (s, 3 H, H-13), 0.87 (t, J = 7.1 Hz, 3 H, H-10'). ¹³C NMR (125 MHz, CDCl₃): δ = 192.6 (C-12), 176.1 (C-15), 174.4 (C-1'), 154.6 (C-7), 137.5 (C-8), 99.3 (C-11), 74.5 (C-1), 70.6 (C-2'), 65.2 (C-2), 52.3 (C-10), 47.0 (C-9), 41.3 (C-3), 40.8 (C-5), 34.6 (C-3'), 33.53 (C-4), 33.49 (C-13), 31.8 (C-8'), 29.4 (C-5'), 29.3 (C-6'), 29.1 (C-7'), 24.7 (C-6), 24.5 (C-4'), 23.4 (C-14), 22.6 (C-9'), 14.1 (C-10'). $[\alpha]^{20}_{\rm D}$ -36.6 (CHCl₃, *c* = 0.1); Lit.^{1b} $[\alpha]^{20}_{\rm D}$ -40 (CHCl₃, *c* = 0.05). HRMS (ESI⁺): *m/z* [M + Na]⁺ calcd for C₂₅H₃₈O₈Na: 489.2465; found: 489.2434. Copyright of Synlett is the property of Georg Thieme Verlag Stuttgart and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.