Accepted Manuscript

Synthesis, Antileishmanial Activity and Structure-Activity Relationship of 1-*N*-X-phenyl-3-*N*,-Y-phenyl-benzamidines

Cl² udio Eduardo Rodrigues-Santos, Leonor L. Leon, Adailton J. Bortoluzzi, Marilene Marcuzzo Canto-Cavalheiro, G^o rzia C. Machado, Aurea Echevarria

PII: S0223-5234(13)00408-X

DOI: 10.1016/j.ejmech.2013.06.040

Reference: EJMECH 6268

To appear in: European Journal of Medicinal Chemistry

Received Date: 21 March 2013

Revised Date: 21 May 2013

Accepted Date: 18 June 2013

Please cite this article as: C.E. Rodrigues-Santos, L.L. Leon, A.J. Bortoluzzi, M.M. Canto-Cavalheiro, G.C. Machado, A. Echevarria, Synthesis, Antileishmanial Activity and Structure-Activity Relationship of 1-*N*-X-phenyl-3-*N*,-Y-phenyl-benzamidines, *European Journal of Medicinal Chemistry* (2013), doi: 10.1016/j.ejmech.2013.06.040.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.





Synthesis, Antileishmanial Activity and Structure-Activity Relationship of 1-N-Xphenyl-3-N'-Y-phenyl-benzamidines

Cláudio Eduardo Rodrigues-Santos,^a Leonor L. Leon,^b Adailton J. Bortoluzzi,^c Marilene Marcuzzo Canto-Cavalheiro,^b Gérzia C. Machado,^b and Aurea Echevarria^{a,*}

^a Departamento de Química, Instituto de Ciências Exatas, Universidade Federal Rural do Rio de Janeiro, 23890-900, Seropédica, RJ, Brazil

^b Departamento de Imunologia, Instituto Oswaldo Cruz, FIOCRUZ, 21042-900, Rio de Janeiro, RJ, Brazil

^c Departamento de Química, Universidade Federal de Santa Catarina, 88040-900, Florianópolis, SC, Brazil

Corresponding author: Phone: +55 21 26822807, Fax: +55 21 26822807, E-mail: echevarr@ufrrj.br

Abstract

Two series of N_{N} -diphenyl-benzamidines were synthesized as part of a study to search potential new drugs with antileishmanial activity. These compounds were obtained by anilides in PCl₅ halogenation reaction with generation in situ of the corresponding benzimidoyl chlorides, and subsequently treatment with adequate anilines. The series I showed expressive results of antileishmanial activity, highlighted the compounds 9a with $IC_{50} = 81.28 \ \mu M \ (\log IC_{50} = 1.91 \ \mu M) \ against L. chagasi, 8e \ with \ IC_{50} = 26.30 \ (\log IC_{50} = 1.91 \ \mu M)$ 1.52 µM) against L. braziliensis. From the results obtained from SAR study (series I), the series II was planned from Craig 2-dimensional map, in which was possible the discovery of the potent compounds, 9v and 9j with $IC_{50} = 12.60 \ \mu M (\log IC_{50} = 1.10 \ \mu M)$ and 13.00 μ M (log IC₅₀ = 1.11 μ M), respectively, against *L. amazonensis*. The results obtained from the SAR and QSAR studies indicated the best results when electron-donors groups in the ring attached to amidinic carbon, unlike when electron-withdrawing groups at the phenyl-Nring showing inhibitory activity increased. Furthermore, the QSAR model obtained indicated the hydrophobicity as a fundamental property for antileishmanial activity presented by these series.

Keywords: *N*,*N*'-diphenyl-benzamidines; *Leishmania amazonenses; Leishmania chagasi; Leishmania braziliensis;* QSAR; Hansch model.

1. Introduction

Leishmania parasites are small and intriguing organisms that cause leishmaniasis, and the symptoms of these diseases are skin sores that erupt weeks to months after the person affected is bitten by sandflies. Other consequences include fever, damage to the spleen and liver [1-2]. This disease is endemic in some geographical areas of world, where constitutes a serious public health problem [3].

Leishmania amazonensis has been isolated from patients with all the different clinical forms of the disease. The extracellular promastigote stage of this protozoan parasite is introduced into subcutaneous tissue in the human host during the bite of an infected sandfly vector. It is phagocytosed by a mononuclear phagocyte, after which it converts into the obligate intracellular amastigote form [3].

The World Health Organization considers leishmaniasis one of the most serious diseases worldwide caused by protozoan parasites. However, the control of this disease remains a problem; the available antileishmanial drugs still rely on the highly toxic pentavalent antimonials (meglumine antimoniate, Glucantime and sodium stibogluconate, Pentostam), which cause serious side effects and require long-term treatment [4].

Second-line drugs include pentamidine (1) and amphotericin B, but these drugs have not experienced widespread use because of toxicity and cost. Recently, the oral drug miltefosine (2), an alkylphosphocoline was approved for the treatment of human visceral *Leishmania* infections, but it present high cost and a long half-life (100 - 200 h) in humans and low therapeutic ratio, the characteristics that could encourage the development of resistance. In addition, the miltefosine is not suitable for use during pregnancy because of teratogenecity and also cause mild to severe gastrointestinal side effects (Figure 1) [5].

Please, insert the Figure 1

Since the chemotherapy against leishmaniasis is still inefficient, there is an urgent need for the development of new, efficient, and safe drugs for the treatment of this disease.

Amidines are much used in synthesis. In some cases for the preparation of acyclic compounds, but mostly for the synthesis of heterocyclic compounds such as aziridines, pyrroles, oxazoles, oxadiazoles, pyridines, pyrimidines, imidazoles and triazines [6]. They combine the properties of an azomethine-like and C-N single bond having some partial double bond character [7].

The amidines are important medical and biochemical agents. Their anti-inflammatory, antiviral, antifungal, antibacterial, antibiotic, antihypertensive and anesthetic activities have been reported [8]. We have previously demonstrated that *N*,*N*'-diphenyl-*p*-methoxy-benzamidine (**3**) was effective against *Leishmania amazonensis* promastigotas and axenic amastigotes and *Trypanosoma evansi* trypomastigotes. Besides it was the most effective derivative in the parasite-macrophage interaction [9-11].

Then, considering the previous results of compound **3** against trypanosomatid parasites and the importance of the discovery of new compounds for leishmaniasis treatment, our research group describes the synthesis and the *L. amazonensis* inhibitory activity of two new series of amidine derivatives. The series I was based in the methoxy moiety importance considering the compound **3**, the most active in the *N*,*N*^{*}-diphenyl-*p*-Xbenzamidine series [9-11] led us to prepare 8 derivatives with mono, di and tri-methoxy substituents on *para* and *meta* position in the *N*-phenyl groups. The results of antileishmania assays supplied a Structure-Activity Relationship (SAR) study and the Craig Graphic [12] allowed the synthesis of series II with 14 derivatives (Figure 2).

Please, insert the Figure 2

2. Chemistry

The route adapted [8] for the synthesis of the two series is outlined in Scheme 1. Initially the benzanilides (6) were prepared from of the corresponding aniline (4) and the appropriately substituted acid chlorides (5) supplying excellent yields (85-90%). The benzanilides (6) were purified by ethanol recrystallization.

After, the benzanilides were converted to benzimidoyl chlorides, *in situ*, by treatment with halogenating reagent (PCl₅) under reflux in dry toluene for 8h, that subsequently reacts with aniline dissolved in dry toluene to furnish the compounds **8a-v** in good yields (60-85%). The reactions were monitored by thin layer chromatography (TLC) and the benzamidinic chlorides (**8a-v**) were purified from acetone. Following, **8a-v** were treated with aqueous solution of NaHCO₃ (5%, m/v) submitted to CHCl₃ extraction affording the target compounds **9a-h** (series I) and **9i-v** (series II).

The formation of the benzimidoyl chloride *in situ* using dry toluene [13-14] demonstrated to be a more effective way than solvent-free [8, 15] to obtain N,N'-diphenylbenzamidine derivatives, because it generated in high purity (it was verified by TLC). This small change was significant to affording the target compounds with high degree of purity (\geq 95%, CG-MS). The compounds that were obtained with less than 95% of purity were recrystallized from methanol afforded needle crystals.

Please, insert the Scheme

The infrared spectra show the disappearance of the benzanilide (6) ν (C=O) band at 1646-1655 cm⁻¹ and the new ν (C=N) band at 1587-1632 cm⁻¹ resulted from benzamidine evidence in agreement with the literature [15].

The values of ¹H and ¹³C NMR spectra (supplementary data) permitted the full characterization of all compounds. The ¹³C NMR chemical shifts indicated peak at 154-158 ppm corresponding to C=N. As the expected the central carbon atom in benzamidines resonates at higher field than in the corresponding benzanilides (164-165 ppm) and benzamidinium cations (161-163 ppm).

The structures of compounds **9b** and **9o** were determined by single crystal X-ray diffraction. Asymmetric unit of **9o** contains two independent molecules, which are different rotamers (**A** and **B**) with different methoxy group position (Figure 3).

The solid-state structure reveals a noncentrosymmetric molecule, with an *E* configuration around the C=N double bond. The crystal structures **9b**, **9o**-rotamer **A** and **9o** -rotamer **B**, respectively, shows lengthened C=N (1.282, 1.283 and 1.281) and shortened C-N (1.371, 1.364 and 1.372) bonds, a feature of $n-\pi$ conjugation [15], an important characteristic of this moiety. In the crystal structures, molecules are linked *via* N-H....O interactions. It is already known that the difference between the C-N and C=N distances is related to the degree of delocalization in the N-C=N skeleton. An interesting fact that can be observed in these results was the electronic delocalization also depends of conformation, the compound **9o**-rotamer A, 0.081Å and **9o**-rotamer B, 0.091 Å.

Please, insert the Figure 3.

3. Results and Discussion

3.1. Antileishmanial Assays

The 50 % growth inhibitory activity value, IC_{50} , of each compound (**9a-v**) was determined using *L. amazonensis* in the evolutive form of promastigotes. For compounds **9a, 9e** and **9f**, and its salt forms (**8a, 8e** and **8f**), the *L. braziliensis* and *L. chagasi* inhibitory activities were evaluated. The remaining parasites were counted in a Naubauer chamber, and the IC_{50} /24h values were determined by linear regression, relating percentage and log of drug concentration in μ g/mL and μ M, as shown in Tables 1, 2 and 3.

The compound with one methoxy moiety (9a) and two methoxy moieties (9e) in phenyl group attached at amidinic carbon, and with two methoxy moieties (9f) in phenyl groups linked to *N* and *N'* atoms were evaluated against promastigotas of *L. amazonenesis*, *L. braziliensis* and *L. chagasi*, and also against macrophages in order to evaluate the toxicity (Table 1). The results for 9a, in neutral form, $IC_{50} = 1.91 \mu M$ presented the best antileishmanial activity against *L. chagasi*, in similar potency of the pentamidine ($IC_{50} = 1.76 \mu M$), reference drug, already assayed against *L. braziliensis* the compound 9e, both neutral ($IC_{50} = 1.52 \mu M$) and salt form 8e ($IC_{50} = 1.42 \mu M$) presented comparable values with the pentamidine ($IC_{50} = 1.36 \mu M$). However, the results against *L. amazonensis* to 8f ($IC_{50} = 1.62 \mu M$) presented the best antileishmanial activity presented the best antileishmanial activity (pentamidine, $IC_{50} = 1.05 \mu M$). Interesting, the macrophage toxicity presented of 9a and 9f was the 15% and 0%, respectively, while the pentamidine presented 100%, at 320 µg/mL of concentration. This SAR study made possible to verify that the substitutions on two *N*-phenyl rings do not

increase the antileishmanial activity against *L. amazonensis* and furthermore cause toxicity increase (Table 1). Then faced this data, this series I (**9a-h**) was proposed and evaluated against *L. amazonensis*. The results obtained demonstrated that the **9a** and **9h** highlighted among the compounds substituted with methoxy groups presenting values of $IC_{50} = 1.85$ μ M and $IC_{50} = 1.60 \mu$ M, respectively (Table 2).

Please insert the Tables 1 and 2

From the results obtained of SAR study, the new series II was planned, with methoxy group linked in the *para* position of the phenyl ring attached to amidinic carbon, because as it can be seen, the compound **9a** presented low toxicity, as well as the compound **9f**. Thus, it was selected the appropriate substituent groups with a wide range of both lipophilicity (π) and Hammett electronic (σ) parameters from 2-dimensional map proposed by Craig [12].

The compounds of series II were assayed against *L. amazonensis*, the results are showed in the Table 3. The most active compounds were the **9j** ($R_3 = H$; $R_5 = Cl$; $IC_{50} = 1.11 \mu$ M); **9m** ($R_3 = Br$; $R_5 = H$; $IC_{50} = 1.34 \mu$ M); **9o** ($R_3 = H$; $R_5 = NO_2$; $IC_{50} = 1.22 \mu$ M) and highlighted the compound **9v** ($R_3 = Cl$; $R_5 = NO_2$; $IC_{50} = 1.10 \mu$ M) that presented inhibitory activity close to reference drug, pentamidine ($IC_{50} = 1.05 \mu$ M). The results obtained from series II indicated that unlike the series I, when the best results were observed to electron donors moieties attached to benzamidine ring (methoxy group), the inhibitory activity increased when electron-withdrawing groups at the phenyl-*N* ring.

Please, insert the Table 3.

3.2. Hansch Model

To try to understand the intrinsic multivariate nature of the results of the antileishmanial activity presented by compounds, it decided to calculate the electronic, steric and lipophilicity properties represented for polarizability (POLZ), superficial tension (ST), volume molar (VM), molar refractivity (MR) and log P descriptors (Table 4). All these parameters were calculated from the ACDLabs software package (version 12.0), because Spessard [17] demonstrated that this is a good software to simulate these parameters. From the results was established the Hansh model [18-19].

Please, insert the Table 4.

The models of 2D-QSAR were obtained from of multiple linear regression (MLR) utilized BuildQSAR software [20].

After to analyze several models, it was possible to indicate a model with statistics parameters expressive, so much in the quality of the adjustment of the data in the model ($r^2 = 0.85$; F = 24.90) as in the predictability of this ($q^2 = 0.74$), according to the works of Erikssons *et al* [21] (Equation 1).

Equation 01.

 $\log 1/\text{IC}_{50} = 1.04 \ (\pm 0.29)\log P^2 - 10.80 \ (\pm 3.23)\log P - 0.04 \ (\pm 0.02)\text{MR} + 29.04 \ (\pm 9.98)$

$$n = 18$$
; $r^2 = 0.85$; $s = 0.26$; $F = 24.90$; $q^2 = 0.74$

The 2D-QSAR model (Equation 01) demonstrates a parabolic relationship with log *P*, and the linear dependence of MR. The lipophilicity presents larger importance that the refractivity molar due to your largest coefficient. The model indicates that the N-phenyl-N'-phenyl-benzamidines reaches good results of antileishmanial activity with molecules

with small volume and it lowers polarity, since the refractivity molar is an ambiguous descriptor, that combines as much volume effect as of polarizability, according to Verma & Hansch [22]. The proposed model presented 4 outliers, the compounds **9a**, **9k**, **9n** and **9o**.

4. Conclusions

The new series of *N*,*N*'-diphenyl-4-methoxybenzamidine derivatives showed important antileishmanial activity, especially when compared with the licensed drug, pentamidine, highlighted the compounds **9a** with $IC_{50} = 81.28 \mu M$ against *L. chagasi*, **8e** with $IC_{50} = 26.30 \mu M$ against *L. braziliensis*, and **9v** and **9j** with $IC_{50} = 12.60$ and 13.00 μM respectively, against *L. amazonensis*. Also, it was observed that the inhibitory activity increase when electron-withdrawing groups are attached on the *N*-phenyl ring, furthermore, the QSAR model obtained were satisfactory showing that the hydrophobicity is the fundamental property for antileishmanial activity. The results of the antileishmanial activity presented by *N*,*N*'-diphenyl-benzamidines indicated a promising new class of leishmanicidal drugs.

5. Experimental Section

5.1. Chemistry

5.1.1. Reagents and Instruments

Melting points were determined on a Kofler hot plate apparatus and are uncorrected. Infrared spectra were recorded on a Perkin-Elmer 1600 FTIR spectrophotometer using potassium bromide tablets. ¹H and ¹³C NMR spectra were obtained in a Brucker Avance-II

400 or Avance-III 500 spectrometers, with tetramethysilane as the internal reference, in DMSO-d₆ as solvent; the chemical shifts were reported in ppm. Mass spectrums were recorded using Saturn GC-MS - CP-SIL8CB ($30m \times 25mm \times 25mm$). Reactions were monitored on Merck silica gel 60 F254 aluminum sheets. TLC spots were visualized by inspection of plates under UV light (254 and 365 mm). All commercial reagents were obtained from Aldrich or Across Co. and used without any further purification, only aniline was distillated.

5.1.2. N- R_2 -phenyl-R, R_1 -benzamides (**6a-d**)

These compounds were prepared and characterized according to the literature [23-27].

5.1.3. General Procedure for the Preparation of the N- R_2 -phenyl-N'- R_3 , R_4 -phenyl-R, R_1 -benzamidine (**9a-v**).

A mixture of *N*-R₂-phenyl-R,R₁-benzamide (**6a-d**) (2.25 mmol), and phosphorous pentachloride (PCl₅) (2.25 mmol) was refluxed in dry toluene for 8 h under N₂ atmosphere furnished, *in situ*, the benzimidoyl chlorides. Subsequently, was added drop wise to mixture the corresponding aniline (**4**) (2.25 mmol) in dry toluene reflux. After, the mixture was stirred for 3 h; was cooled and the precipitate was filtered afforded the corresponding benzamidinic chlorides (**8a-v**). The **8a-v** were sequentially washed with acetone (5 x 5 mL), and neutralized with NaHCO₃ 5% (m/v) under stirred at room temperature for 4 h. Finally, the mixture was extracted with CHCl₃, dried and concentrated to afford the target compounds (**9a-v**) purified by recrystallization from ethanol.

5.1.3.1. N,N'-Diphenyl-4-methoxybenzamidine (**9***a*). Yield 87%; mp 122-124 °C (118-120 °C [8]); IR (KBr): v 3312, 2927, 2835, 1627, 1591, 1533, 1252, 1030, 789 cm⁻¹; ¹H NMR

11

(DMSO- d_6): δ 9.10 (s, 1H), 7.84 (d, J = 8.0 Hz, 2H), 7.26 (d, J = 8.0 Hz, 1H), 7.21 (d, J = 8.0 Hz, 2H), 7.04 (t, J = 8.0 and 4.0 Hz, 2H), 6.95 (t, J = 8.0 and 4.0 Hz, 1H), 6.84 (t, J = 8.0 Hz, 2H), 6.75 (t, J = 8.0 and 4.0 Hz, 1H), 6.57 (t, J = 4.0 Hz, 2H), 3.71 (s, 3H); ¹³C NMR (DMSO- d_6): δ 159.5, 154.4, 150.8, 141.4, 130.5, 128.3, 126.9, 122.1, 121.7, 119.5, 113.5, 55.1. MS, m/z (%): 302 (M+⁻, 13), 286 (1), 210 (100), 77 (15), 51 (9).

5.3.1.2. *N*,*N*'-*Diphenyl-3-methoxybenzamidine* (**9b**). Yield 65%; mp 210-213 °C; IR (KBr): v 3349, 2839, 1625, 1590, 1528, 1325, 1269, 1022 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.21 (s, 1H), 7.88 (sl, 2H), 7.21 (dd, *J* = 6.0 Hz, 3H), 7.06 (d, *J* = 8.0 Hz, 3H), 6.88 (t, *J* = 8.0 Hz, 4H), 6.61 (d, *J* = 8.0 Hz, 2H), 3.65 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 158.7, 154.3, 136.0, 129.1, 128.3, 121.8, 121.1, 119.6, 114.7, 114.5, 55.0. MS, m/z (%): 302 (M+, 100), 287 (1), 210 (55), 75 (5), 51 (3). Anal. Calcd. for C₂₀H₁₈N₂O: C, 79.44, H, 6.00, N, 9.26. Found: C, 80.02, H, 5.90, N, 9.30.

5.3.1.3. *N*-(4-methoxyphenyl)-*N*'-phenyl-benzamidine (**9**c). Yield 80%; mp 110-111 °C (114 °C [27]); IR (KBr): v 2935, 2839, 1632, 1513, 1242, 1026, 730, 639 cm⁻¹; ¹H NMR (DMSO-*d*-₆): δ 9.09 (s, 0.5H), 9.03 (s, 0.5 H), 7.76 (d, *J* = 8.0 Hz, 2H), 7.28 (d, *J* = 8.0 Hz, 3H), 7.01 (t, *J* = 8.0 and 4.0 Hz, 1H), 6.85 (d, *J* = 4.0 Hz, 1H), 7.2 (d, *J* = 4.0 Hz, 1H), 6.60 (s, 1H), 6.55 (s, 1H), 6.49 (s, 1H), 3.71 (s, 1.7H), 3.60 (s, 1.3H); ¹³C NMR (DMSO-*d*₆): δ 161.9, 158.1, 136.2, 133.0, 132.3, 130.5, 128.6, 126.8, 125.0, 122.2, 114.2, 55.4. MS, m/z (%): 302 (M+, 50), 286 (1), 180 (75), 77 (32), 51 (16).

5.3.1.4. *N*-(4-methoxyphenyl)-*N*'-phenyl-3-methoxybenzamidine (**9d**). Yield 70%; mp 127-128 °C; IR (KBr): v 3349, 2948, 2834, 1634, 1593, 1541, 1335, 1240, 1034 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.05 (s, 1H), 7.87 (d, *J* = 8.0 Hz, 1H), 7.77 (d, *J* = 6.0 Hz, 1H), 7.20 (m, *J* =8.0 Hz, 2H), 7.03 (m, *J* = 8.0 Hz, 2H), 6.84 (m, *J* = 8.0 Hz, 5H), 6.58 (m, *J* = 4.0 Hz, 2H), 3.71 (s, 3H), 3.64 (sl, 3H); ¹³C NMR (DMSO-*d*₆): δ 158.7, 154.4, 136.2, 129.2, 128.3, 122.9, 122.2 121.2, 119.2, 114.6, 55.0. MS, m/z (%): 332 (M+, 63), 240 (100), 91 (1), 77 (28), 51 (15). Anal. Calcd. for C₂₁H₂₀N₂O₂: C, 75.88, H, 6.06, N, 8.43. Found: C, 76.05, H, 5.91, N, 8.60.

5.3.1.5. *N*-(4-methoxyphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9**e). Yield 65%; mp 121-122 °C (124°C [28]) ; IR (KBr): v 3334, 3037, 2950, 2833, 1626, 1592, 1533, 1505, 1244, 1030, 832 cm⁻¹, ¹H NMR (DMSO- d_6): δ 8.98 (s, 0.56H), 8.93 (s, 0.44H), 7.84 (d, *J* = 10.0 Hz, 1H), 7.75 (d, *J* = 5.0 Hz, 1H), 7.24 (d, *J* = 5.0 Hz, 1H), 7.19 (d, *J* = 10.0 Hz, 2H), 7.02 (t, *J* = 10.0 and 5.0 Hz, 1H), 6.93 (t, *J* = 10.0 and 5.0 Hz, 0.5H), 6.84 (sl, 3H), 6.73 (sl, 0.5H), 6.61 (d, *J* = 10.0 Hz, 1H), 6.55 (d, *J* = 10.0 Hz, 1H), 6.49 (d, *J* = 10.0 Hz, 1H), 3.72 (s, 5H), 3.62 (s, 1H); ¹³C NMR (DMSO- d_6): δ 159.4, 154.2, 134.0, 130.7 128.2, 127.1, 122.8, 122.2, 121.0, 120.5, 119.3, 113.6 113.4, 113.3, 55.12. MS, m/z (%): 332 (M+, 47), 240 (100), 211 (90), 77 (21), 51 (11).

5.3.1.6. *N*-(4-methoxyphenyl)-*N*'-(4-methoxyphenyl)-benzamidine (**9***f*). Yield 60%; mp 119-120 °C (125 °C [28]); IR (KBr): v 3352, 3055, 2933, 2834, 1625, 1505, 1241, 1032, 831 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 8.92 (s, 1H), 7.78 (d, *J* = 10.0 Hz, 2H), 7.30 (t, *J* = 5.0 Hz, 3H), 7.25 (dd, *J* = 5.0 Hz, 2H), 6.84 (d, *J* = 5.0 Hz, 2H), 6.59 (d, *J* = 10.0 Hz, 2H), 6.45 (d, J = 5.0 Hz, 2H), 3.71 (s, 3H), 3.59 (s, 3H); ¹³C NMR (DMSO- d_6): 162.4, 157.8, 136.0, 132.3, 130.3, 128.6, 126.8, 121.1, 113.9, 55.4. MS, m/z (%): 332 (M+, 33), 210 (100), 77 (10), 51 (5).

5.3.1.7. N-(4-methoxypheny)-N'-(4-methoxyphenyl)-4-methoxybenzamidine (9g). Yield
70%; mp 110-113 °C (105 °C [28]); IR (KBr): v 3414, 3065, 2837, 1606, 1551, 1512, 1458, 1253, 1027 cm⁻¹; ¹H NMR (DMSO-d₆): 7.62 (d, J = 8.0 Hz, 2H), 7.15 (d, J = 8.8 Hz, 4H), 6.82 (d, J = 8.6 Hz, 4H), 6.70 (sl, 2H); 3.69 (sl, 6H), 3.63 (sl, 3H); ¹³C NMR (DMSO-d₆): δ 161.9, 157.7, 132.7, 128.3, 126.6, 119.8, 114.1, 55.7, 55.4. MS, m/z (%): 362 (M+; 100), 241 (100), 226 (30), 197 (10), 123 (15), 77 (27).

5.3.1.8. *N*-(4-methoxyphenyl)-*N*'-phenyl-3,4-methylenedioxybenzamidine (**9**h). Yield 80%; mp 146-148 °C; IR (KBr): v 3349, 2964, 2840, 1633, 1592, 1533, 1348, 1245, 1037 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 8.95 (sl, 1H), 7.78 (sl, 1H), 7.15 (sl, 3H), 6.82 (t, *J* = 8.0 Hz, 6H), 6.57 (sl, 2H), 5.99 (s, 2H), 3.67 (sl, 3H); ¹³C NMR (DMSO-*d*₆): δ 155.1, 147.5, 146.7, 128.7, 128,3, 123,2, 107.9, 113,5, 101.5, 55.0. MS, m/z (%): 346 (M+, 55), 254 (93), 224 (100), 91 (1), 77 (22), 51 (13). Anal. Calcd. for C₂₁H₁₈N₂O₃: C, 72.82, H, 5.24, N, 8.09. Found: C, 72.30, H, 5.54, N, 8.30.

5.3.1.9. *N*-(4-chorophenyl)-N'-phenyl-4-methoxybenzamidine (**9***i*). Yield 55%; mp 168-170 °C; IR (KBr): v 3124, 2963, 2839, 1620, 1591, 1534, 1340, 1252, 1030 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.24 (d, 1H), 7.86 (dd, *J* = 6.0 Hz, 2H), 7.22 (m, *J* = 8.0 Hz, 3H), 7.07 (d, *J* = 10.0 Hz, 2H), 6.89 (d, *J* = 6.0 Hz, 4H), 6.57 (d, *J* = 8.0 Hz, 2H), 3.72 (s, 3H); ¹³C NMR

14

(DMSO-*d*₆): δ 159.7, 152.11, 149.9, 136.9, 130.5, 128.3, 128.2, 1256.6, 123.8, 122.0, 119.6, 120.9, 113.4, 55.1. MS, m/z (%): 337 (M+, 100), 301 (2), 210 (92), 92 (3), 77 (6), 51 (5). Anal. Calcd. for C₂₀H₁₇ClN₂O: C, 71.32, H, 5.09, N, 8.32. Found: C, 70.80, H, 5.25, N, 8.48.

5.3.1.10. *N*-(*3*-chorophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***j*). Yield 67%; mp 215-218 °C; IR (KBr): v 3422, 2841, 1586, 1512, 1327, 1257, 1025 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.29 (d, 1H), 7.66 (m, *J* = 6.0 Hz, 2H), 7.24 (m, *J* = 6.0 Hz, 4H), 7.03 (m, *J* = 6.0 Hz, 3H), 6.87 (m, *J* = 10.0 Hz, 3H), 6.82 (m, *J* = 4.0 Hz, 1H), 3.72 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 162.1, 158.1, 152.1, 149.3, 135.0, 131.9, 130.7, 129.6, 128,0, 123.8, 122.9, 121.3, 114.5, 55.7. MS, m/z (%): 337 (M+, 100), 301 (2), 210 (85), 92 (3), 77 (6), 51(5). Anal. Calcd. for C₂₀H₁₇ClN₂O: C, 71.32, H, 5.09, N, 8.32. Found: C, 71.40, H, 5.05, N, 8.55.

5.3.1.11. *N*-(4-fluorophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9k**). Yield 89%; mp 149-150 °C; IR (KBr): v 3300, 2932, 2843, 1627, 1596, 1535, 1335, 1254, 1031 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.15 (s, 1H), 7.80 (m, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 4H), 7.07 (d, *J* = 10.0 Hz, 2H), 6.88 (m, *J* = 8.0 Hz, 3H), 6.57 (d, *J* = 8.0 Hz, 2H), 3.72 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 159.6, 154.8, 150.8, 147.4, 141.4, 130.5, 128.3, 123.4, 122.2, 121.9, 120.9, 115.0, 114.6, 113.4, 55.1. MS, m/z (%): 320 (M+⁺, 20), 305 (5), 210 (85), 91 (1), 77 (20), 51 (9). Anal. Calcd. for C₂₀H₁₇FN₂O: C, 74.98, H, 5.35, N, 8.74. Found: C, 74.71, H, 5.23, N, 8.33. 5.3.1.12. *N*-(*3-fluorophenyl*)-*N*'-*phenyl*-4-*methoxybenzamidine* (**9***l*). Yield 85%; mp 95-97 °C; IR (KBr): v 3329, 2960, 2839, 1628, 1597, 1531, 1336, 1251, 1031 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.27 (d, 1H), 7.87 (m, *J* = 6.0 Hz, 2H), 7.54 (d, *J* = 8.0 Hz, 1H), 7.25 (dd, *J* = 6.0 Hz, 3H), 7.06 (m, *J* = 6.0 Hz, 2H), 6.83 (d, *J* = 8.0 Hz, 3H), 6.59 (d, *J* = 8.0 Hz, 1H), 6.39 (d, *J* = 8.0 Hz, 1H), 3.72 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 164.6, 159.8, 154.6, 153.4, 150.5, 141.2, 130.9, 129.8, 128.5, 126.7, 122.9, 119.6, 115.4, 112.5, 107.8, 55.3. MS, m/z (%): 320 (M+, 20), 304 (1), 210 (68), 91 (2), 77 (20), 51 (8). Anal. Calcd. for C₂₀H₁₇FN₂O: C, 74.98, H, 5.35, N, 8.74. Found: C, 75.80, H, 5.44, N, 8.28.

5.3.1.13. N-(4-bromophenyl)-N'-phenyl-4-methoxybenzamidine (**9m**). Yield 65%; mp 175-178 °C; IR (KBr): v 3120, 2963, 2838, 1618, 1591, 1534, 1341, 1251, 1030 cm⁻¹; ¹H NMR (DMSO- d_6): δ 9.23 (d, 1H), 7.87 (d, J = 8.0 Hz, 2H), 7.43 (d, J = 8.0 Hz, 1H), 7.22 (m, J = 8.0 Hz, 4H), 7.05 (d, J = 8.0 Hz, 2H), 6.88 (d, J = 8.0 Hz, 2H), 6.54 (t, J = 8.0 Hz, 2H), 3.73 (s, 3H); ¹³C NMR (DMSO- d_6): δ 159.7, 154.9, 150.3, 141.2, 140.8, 131.1, 130.5, 128.3, 126.6, 124.4, 122.6, 122.0, 121.3, 119.8, 113.3, 55.2. MS, m/z (%): 381 (M+, 50), 350 (1), 210 (100), 91 (5), 77 (15), 51 (5). Anal. Calcd. for C₂₀H₁₇BrN₂O: C, 63.00, H, 4.49, N, 7.35. Found: C, 62.85, H, 4.55, N, 7.77.

5.3.1.14. *N*-(3-methoxyphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***n*). Yield 65%; oil; IR (KBr): v 3384, 2932, 2842, 1603, 1266, 1023 cm ⁻¹; ¹H NMR (DMSO-*d*₆): δ 7.75 (d, J =8.0 Hz, 2H), 7.28 (m, J = 10.0 Hz, 8H), 6.77 (d, J = 8.0 Hz, 3H), 3.88 (s, 3H), 3.78 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 161.7, 138.1, 135.9, 133.1, 131, 130.1, 126.8, 125.2, 121.9, 115.3, 114.1, 111.0, 55.9, 55.24. MS, m/z (%): 332 (M+⁻, 20), 317 (1), 210 (80), 91 (3), 77 (27), 51 (10). Anal. Calcd. for C₂₁H₂₀N₂O₂: C, 75.88, H, 6.06, N, 8.43. Found: C, 74.11, H, 5.94, N, 8.50.

5.3.1.15. *N*-(4-nitrophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***o*). Yield 60%; mp 150-153 °C; IR (KBr): v 3375, 2930, 2838, 1625, 1582, 1527, 1322, 1251, 1028 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.72 (d, 1H), 8.19 (d, *J* = 8.0 Hz, 1H), 8.07 (d, *J* = 8.0 Hz, 1H), 7.95 (d, *J* = 8.0 Hz, 1H), 7.75 (d, *J* = 8.0 Hz, 1H), 7.28 (m, *J* = 4.0 Hz, 4H), 6.90 (d, *J* = 8.0 Hz, 2H), 7.05 (m, *J* = 8.0 Hz, 1H), 6.73 (d, *J* = 8.0 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 1H), 3.73 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 159.9, 158.0, 149.9, 140.6, 130.8, 128.5, 124.6, 122.8, 122.6, 122.0, 120.2, 113.7, 113.7, 55.27. MS, m/z (%): 347 (M+; 10), 300 (1), 210 (30), 91 (1), 77 (6), 51 (6). Anal. Calcd. for C₂₀H₁₇N₃O₃: C, 69.15, H, 4.93, N, 12.10. Found: C, 68.77, H, 4.85, N, 11.86.

5.3.1.16. *N*-(3-nitrophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***p*). Yield 76%; oil; IR (KBr): v 3410, 2923, 2850, 1602, 1527, 1349, 1260, 1031 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.52 (d, 1H), 7.82 (d, *J* = 8.0 Hz, 1H), 7.60 (d, *J* = 8.0 Hz, 1H), 7.27 (m, *J* = 8.0 Hz, 4H), 7.03 (m, *J* = 8.0 Hz, 3H), 6.90 (m, *J* = 8.0 Hz, 3H), 6.63 (d, *J* = 6.0 Hz, 1H), 3.72 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 160.0, 156.0, 152.4, 140.8, 147.9, 150.1, 130.8, 129.5, 128.4, 122.5, 122.2, 120.0, 115.5, 113.6, 55.2. MS, m/z (%): 303 (30), 210 (100), 91 (1), 77 (20), ,51 (10). Anal. Calcd. for C₂₀H₁₇N₃O₃: C, 69.15, H, 4.93, N, 12.10. Found: C, 68.85, H, 4.87, N, 11.82.

5.3.1.17. *N*-(4-hydroxyphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***q*). Yield 80%; mp 160-162 °C; IR (KBr): v 3202, 2923, 2852, 1626, 1605, 1536, 1361, 1236, 1028 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 7.66 (d, *J* = 2.0 Hz, 2H), 7.10 (m, *J* = 6.0 Hz, 10H), 6.80 (d, *J* = 8.0 Hz, 2H), 3.83 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 154.6, 152.3, 130.7, 128.4, 127.1, 122.2, 121.2, 120.9, 115.0, 55.1. MS, m/z (%): 318 (M+, 30), 226 (100), 210 (100), 91 (2), 77 (26), 51 (20). Anal. Calcd. for C₂₀H₁₈N₂O₂: C, 75.45, H, 5.70, N, 8.80. Found: C, 75.05, H, 5.58, N, 8.76.

5.3.1.18. *N*-(3-hydroxyphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9r**). Yield 75%; mp 190-192 °C; IR (KBr): v 3118, 2918, 1604, 1536, 1347, 1266, 1025 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 7.34 (dd, *J* = 8.0 Hz, 2H), 7.17 (t, *J* = 8.0 Hz, 3H), 7.01 (d, *J* = 8.0 Hz, 2H), 6.88 (m, *J* = 8.0 Hz, 5H), 6.35 (dd, *J* = 8.0 Hz, 1H), 3.77 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 157.6, 154.7, 154.6, 151.6, 131.8, 130.1, 129.5, 122.8, 115.1, 114.3, 113.1, 111.1, 55.7. MS, m/z (%): 318 (M+', 20), 226 (100), 210 (90), 91 (3), 77 (18), 51 (10). Anal. Calcd. for C₂₀H₁₈N₂O₂: C, 75.45, H, 5.70, N, 8.80. Found: C, 74.80, H, 5.62, N, 8.23.

5.3.1.19. *N*-(4-methylphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9**s). Yield 60%; mp 140-142 °C; IR (KBr): v 3290, 2926, 2857, 1624, 1591, 1535, 1336, 1250, 1031 cm⁻¹; ¹H NMR (DMSO- d_6): δ 9.09 (sl, 1H), 7.77 (dd, *J* = 8.0 Hz, 2H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.05 (d, *J* = 10.0 Hz, 2H), 6.85 (d, *J* = 12.0 Hz, 5H), 6.65 (d, *J* = 6.0 Hz, 2H), 371 (s, 3H), 2.18 (d, 3H); ¹³C NMR (DMSO- d_6): δ 159.5, 154.3, 151.0, 141.5, 139.0, 130.5, 128.7, 128.3, 127.0, 122.1, 119.6, 113.3, 55.1. MS, m/z (%): 316 (M+, 20), 224, (100), 210 (85), 91 (5), 77 (20), 51 (6). Anal. Calcd. for C₂₁H₂₀N₂O: C, 79.72, H, 6.37, N, 8.85. Found: C, 78.67, H, 6.33, N, 8.54.

5.3.1.20. *N*-(3-methylphenyl)-*N*'-phenyl-4-methoxybenzamidine (**9**t). Yield 86%; mp 234-236 °C; IR (KBr): v 3391, 2851, 1603, 1547, 1334, 1263, 1024 cm⁻¹; ¹H NMR (DMSO- d_6): δ 9.02 (sl, 1H), 7.83 (d, *J* = 12.0 Hz, 1H), 7.63 (d, *J* = 10.0 Hz, 1H), 7.22 (m, *J* = 8.0 Hz, 3H), 7.10 (m, *J* = 8.0 Hz, 1H), 6.85 (m, *J* = 6.0 Hz, 4H), 6.56 (d, *J* = 6.0 Hz, 2H), 6.32 (m, *J* = 8.0 Hz, 1H), 3.71 (s, 3H), 2.18 (sl, 3H); ¹³C NMR (DMSO- d_6): 159.6, 154.2, 150.7, 141.4, 137.3, 128.3, 128.1, 122.9, 122.2, 121.6, 122.5, 119.1, 116.8, 113.32, 55.15, 30.7. MS, m/z (%): 316 (M+, 25), 224, (100), 210 (80), 91 (12), 77 (15), 51 (8). Anal. Calcd. for C₂₁H₂₀N₂O: C, 79.72, H, 6.37, N, 8.85. Found: C, 78.87, H, 6.24, N, 8.77.

5.3.1.21. *N*-(4-sulfonylaminophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9***u*). Yield 60%; mp 159-161 °C; IR (KBr): v 3199, 2913, 1602, 1552, 1337, 1262, 1014 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.67 (s, 0.2H), 9.43 (s, 0.6H), 8.73 (s, 0.3H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.25 (d, *J* = 8.0 Hz, 2H), 7.21 (d, *J* = 4.0 Hz, 1H), 7.05 (t, *J* = 8.0 Hz, 1H), 6.97 (d, *J* = 8.0 Hz, 1H). 6.90 (d, *J* = 10.0 Hz, 2H), 6.83 (d, *J* = 8.0 Hz, 1H), 6.78 (dd, *J* = 8.0 and 4.0 Hz, 1H), 6.58 (d, *J* = 8.0 Hz, 2H), 371 (s, 2.2H), 3.68 (0.7H); ¹³C NMR (DMSO-*d*₆): δ 162.24, 151.2, 143.5, 129.5, 132, 127.6, 123.9, 122.6, 114.6, 55.2. MS, m/z (%): 301, (13), 210 (100), 91 (1), 77 (18), 51 (10). Anal. Calcd. for C₂₀H₁₉N₃O₃S: C, 62.98, H, 5.02, N, 11.02. Found: C, 61.76, H, 4.89, N, 11.20.

5.3.1.22. *N*-(4-choro-3-nitrophenyl)-*N*'-phenyl-4-methoxybenzamidine (**9**ν). Yield 50%; mp 128-130 °C; IR (KBr): v 3418, 2920, 2849, 1603, 1537, 1336, 1265, 1026 cm⁻¹; ¹H NMR (DMSO-*d*₆): δ 9.45 (s, 1H), 8.00 (d, *J* = 6.0 Hz, 2H), 7.39 (m, 7H), 6.86 (d, *J* = 6.0 Hz, 2H), 6.65 (s, 1H), 3.75 (s, 3H); ¹³C NMR (DMSO-*d*₆): δ 160.1, 154.4, 151.3, 147.1, 140.7, 131.4, 130.8, 128.4, 127.6, 125.8, 122.6, 120.1, 115.7, 113.7, 55.2. MS, m/z (%): 381 (M+⁺, 15), 289, (100), 210 (37), 91 (1), 77 (13), 51 (6). Anal. Calcd. for C₂₁H₁₆ClN₃O₃: C, 62.91, H, 4.22, N, 11.01. Found: C, 61.89, H, 4.15, N, 10.92.

5.2. Biology.

5.2.1. Parasite Culture. *L. amazonensis* promastigotes MHOM/BR/77/LTB0016 strain were grown at 25 °C in LIT medium supplemented with 10% (v/v) heat-inactivated fetal calf serum (FCS). Cells were harvested in the late log phase, resuspended in fresh medium, counted in Neubauer's chamber, and adjusted to a final concentration of 4 x 10^6 /mL. This strain has been characterized by molecular and immunological techniques [9].

5.2.2. Antileishmanial Assays. The assays were carried out in 96-well plates in volume of 180 μ L/well. The drugs were added to a parasite culture in a concentration range from 160 to 5 μ g/mL, solubilized in DMSO (the highest percentage used was 1.6%, v/v, which was not hazardous to the parasites). After 24 h incubation, the remaining parasites were counted and the percentage of inhibition was calculated, comparing to the controls (DMSO without the drugs and with the parasites alone). The IC₅₀ values were determined by linear regression from these percentages of inhibition using statistical error limits up to 10%. All

tests were done in triplicate and pentamidine isethionate (May & Baker Lab., England) was used as reference drug.

5.3. Molecular Descriptors. The molecular structures were drawn by ACD/ChemSketch software (ACDLabs software package, version 12.0) and $\log P$, polarizability, superficial tension, volume molar and molar refractivity and $\log P$ descriptors were calculated for each compound.

5.4. QSAR Analysis. QSAR models were derived by multiple regression analyses that were performed using the BuildQSAR program to determine the coefficients of the correlation equations. In all equations in this paper, the numbers in parentheses represent the 95% confidence intervals of the coefficients, n is number of data points, r is the correlation coefficient, s is the standard deviation, q^2 is the cross-validated and F is Fisher value; measures for the statistical significance.

5.5. X-ray diffraction Analyses. A crystal of each sample was selected from representative crystalline mass. The intensity data for compounds **9b** and **9o** were collected with an Enraf-Nonius CAD4 diffractometer, at room temperature, with graphite-monochromated Mo Kα radiation. The unit cell parameters were determined on the setting angles of 25 centered reflections. All data were corrected for Lorentz and polarization effects. The structure was solved by direct methods and refined by full-matrix least-squares methods using SIR97 [29] and SHELXL97 [30] programs, respectively. All non-hydrogen atoms were refined anisotropically. H atoms attached to C atoms were placed at their idealized positions, with

C-H distances and U_{eq} values taken from the default settings of the refinement program. H atoms of the secondary amine groups were located from Fourier difference maps and treated as free atoms. ORTEP plots and cif validate procedure were performed by using PLATON software [31]. Further data of crystallographic analysis for compounds **9b** and **9o** are summarized in Table S1 (supplementary data). Crystallographic data (without structure factors) have been deposited at the Cambridge Crystallographic Data Centre (CCDC) as supplementary publication no. CCDC-837978 and CCDC-837979. Copies of the data may be obtained free of charge from the CCDC (12 Union Road, Cambridge CB2 1EZ, UK; Tel.: +44-1223-336408; Fax: +44-1223-336003; e-mail: deposit@ccdc.cam.ac.uk; http://www.ccdc.cam.ac.uk).

Acknowledgment. The authors thank CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico) and CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) for financial support and fellowships received. The authors also thank Prof. Dr. Anderson Coser Gaudio for the freeware use of the BuildQSAR program.

Appendix. Supplementary data

Supplementary data related to this article can be found at http...

References

- M.C. Vendrametto, A.O. Santos, C.V. Nakamura, B.P.D. Filho, D.A.G. Cortez, T.U. Nakamura, Parasitol Int. 59 (2010) 154-158.
- [2] R.J. Soares-Bezerra, E.F. Da Silva, A. Echevarria, L. Gomes-Silva, L. Cysne-Finkelstein, F.P. Monteiro, L.L. Leon, M. Genestra, J. Enzyme Inhib. Med. Chem. 23 (2008) 328-333.
- [3] (a) L. Paloque, P. Verhaghe, M. Casanova, C. Castera-Ducros, A. Dumètre, L. Mbatchi, S. Hetter, M. Kraiem-M'Rabet, M. Laget, V. Remusat, S. Rault, P. Rathelot, N. Azas, P. Vanelle, Eur. J. Med. Chem. 54 (2012) 75-86. (b) M. Genestra, R.J. Soares-Bezerra, L. Gomes-Silva, D.L. Fabrino, T. Bellato-Santos, D.B. Castro-Pinto, M.M. Canto-Cavalheiro, L.L. Leon, Cell Biochem Funct. 26 (2008) 709-717.
- [4] (a) R.F. Rodrigues, D. Castro-Pinto, A. Echevarria, C.M. Reis, C.N. Del Cistia, C.M.R. Sant'Anna, F. Teixeira, H. Castro, M. Canto-Cavalheiro, L.L. Leon, A. Tomás, Bioorg. Med. Chem, 20 (2012) 1760-1766. (b) D. Plano, Y. Baquedano, D. Moreno-Mateos, M. Font, A. Jiménez-Ruiz, J.A. Palop, C. Sanmartín, Eur. J. Med. Chem. 46 (2011) 3315-3323. (c) R.F. Rodrigues, K.S. Charret, E.F. Silva, A. Echevarria, V.F. Amaral, L.L. Leon, M.M. Canto-Cavalheiro, Antimicrob. Agents Chemother. 53 (2009) 839-842.
- [5] (a) M.M. Alam, E.H. Joh, Y. Kim, Y.I. Oh, J. Hong, B. Kim, D.H, Kim, Y.S. Lee, Eur.
 J. Med. Chem. 47 (2012) 485-492. (b) S. Porwal, S.S. Chauhan, P.M.S. Chauhan, N.
 Shakya, A. Verma, S. Gupta, J. Med. Chem. 52 (2009) 5793-5802.
- [6] M.S. Santos, A.M.R. Bernardino, M.C. Souza, Quim. Nova. 29 (2006) 1301-1306.
- [7] J. Suwinski, W. Szczepankiesicz, E.A. Basso, C.F. Tormena, M.P. Freitas, R. Rittner, Spectrochim. Acta A. 59 (2003) 3139-3145.

- [8] (a) S. Surbhi, N. Kumar, P. Roy, S. M. Sondhy, Eur. J. Med. Chem. 59 (2013) 7-14.
 (b) A. Echevarria, L.H. Santos, J. Miller, N. Mahmood, Bioorg. Med. Chem. Lett. 6 (1996) 1901-1904.
- [9] R.M. Temporal, L.C. Finkelstein, A. Echevarria, M.A.S. Souza, M. Sertã, A.J. Silva-Gonçalves, C. Pirmez, L.L. Leon, Arzneim.-Forsch./Drug Res. 6 (2002) 489-493.
- [10] (a) M. Genestra, A. Echevarria, L. Cysne-Finkelstein, L. Vignólio-Alves, L.L. Leon, Nitric Oxide. 8 (2003) 1-6. (b) R.M. Temporal, L. Cysne-Finkelstein, A. Echevarria, A.J. Silva-Gonçalves, L.L. Leon, M.S. Genestra, J. Enzyme Inhib. Med. Chem. 20 (2005) 13-18.
- [11] D.B. Castro-Pinto, A. Echevarria, M.S. Genestra, L. Cysne-Finkelstein, L.L. Leon, J. Enzyme Inhib. Med. Chem. 19 (2004) 57-63.
- [12] P.N. Craig, J. Med. Chem. 14 (1971) 680-684.
- [13] W.M. Fathalla, M. Cajan, J. Marek, P. Pazdera, Molecules. 6 (2001) 574-587.
- [14] W.M. Fathalla, P. Pazdera, Molecules. 7 (2002) 96-103.
- [15] A.J. Bortoluzzi, A. Echevarria, C.E. Rodrigues-Santos, Acta Cryst. E. 60 (2004) 1837-1839.
- [16] A.K. Pal, G.S. Hanan, Acta Cryst E. 65 (2009) 2777-2779.
- [17] G.O. Spessard, J. Chem. Inf. Comput. Sci.38 (1998) 1250-1253.
- [18] H. Kubinyi, Research Focus. 2 (1997) 457-467.
- [19] C. Hansch, A. Leo, R.W. Taft, Chem. Rev. 91 (1991) 165-195.
- [20] D.D. Oliveira, A.C. Gaudio, Quant. Sctruct.-Act. Relat. 19 (2000) 599-601.
- [21] L. Eriksson, J. Jaworska, A.P. Worth, M.T. Cronin, R.M. McDowell, P. Gramatica, Environ. Health Perspect. 111 (2003) 1361-1375.
- [22] R.P. Verma, C. Hansh, J. Pharm. Sci. 97 (2008) 88-110.

- [23] E. Bouron, G. Goussard, C. Marchad, M. Bonin, X. Pannecoucke, J.C. Quirion, H.P. Husson, Tetrahedron Lett. 40 (1999) 7227-7230.
- [24] Z. Hu, W. Ye, H. Zou, Y. Yu, Synth. Commun. 40 (2010) 222-228.
- [25] F. Shi, J. Li, C. Li, X. Jia, Tetrahedron Lett. 51 (2010) 6049-6051.
- [26] C. Quian, X. Zhang, J. Li, F. Xu, Y. Zhang, Q. Shen, Organometallics. 28 (2009) 3856-3862.
- [27] K.E. Fairfull-Smith, I.D. Jenkins, W.A. Loughlin, Org. Biomol. Chem. 2 (2004) 1979-1986.
- [28] N.J. Sintov, J.S. Rodia, J.A. Tursich, H.L. Davis, G.L. Webster, J. Am. Chem. Soc. 71 (1949) 3990-3992.
- [29] A. Altomare, M.C. Burla, M. Camalli, G.L. Cascarano, C. Giacovazzo, A. Guagliardi, A.G.G. Moliterni, G. Polidori, R. Spagna, J. Appl. Cryst. 32 (1999) 115-119.
- [30] G.M. Sheldrick, Acta Cryst. A64 (2008) 112-122.
- [31] A.L. Spek, J. Appl. Cryst. 36 (2003) 7-13.

Table 1. IC₅₀ values for compounds 9a, 9e and 9f neutral and salts forms (8a, 8e and 8f) assayed against *L. amazonesis*, *L. brazilensis and L. chagasi*

Promastigotes and macrophage toxicity.



Compound	Sı	ubstituen	its	L. amaze	onensis	L. brazi	liensis	L. chaz	gasi	Toxicity (%)
	R	R_2	R ₃	IC ₅₀ (µg/mL)	$IC_{50}(\mu M)$	IC ₅₀ (µg/mL)	$IC_{50}(\mu M)$	IC ₅₀ (µg/mL)	IC ₅₀ (µM)	
9a	OCH ₃	Н	Н	1.34 ± 0.18	1.85 ± 0.25	1.48 ± 0.04	1.90 ± 0.14	1.41 ± 0.05	1.91 ± 0.66	No toxic
9e	OCH ₃	OCH ₃	Н	2.33 ± 0.15	2.80 ± 0.19	1.05 ± 0.21	1.52 ± 0.31	1.48 ± 0.09	1.95 ± 0.12	15
9f	Н	OCH ₃	OCH ₃	1.77 ± 0.11	2.24 ± 0.14	1.77 ± 0.66	2.25 ± 0.14	1.75 ±0.067	2.25 ± 0.25	100
8a ^a	OCH ₃	Н	Н	1.43 ± 0.07	1.90 ± 0.10	1.52 ± 0.11	2.05 ± 0.06	2.25 ± 0.17	2.76 0.21	-
8e ^a	OCH ₃	OCH ₃	-	1.39 ± 0.27	1.82 ± 0.35	0.99 ± 0.36	1.42 ± 0.52	1.61± 0.12	2.04 ± 0.16	-
8f ^a	Н	OCH ₃	OCH ₃	1.19 ± 0.31	1.62 ± 0.42	1.70 ± 0.16	2.13 ± 0.20	> 2.5	>3.0	-
Pentamidine ^b	-	-	-	0.82 ± 0.19	1.05 ± 0.25	1.12 ± 0.11	1.36 ± 0.14	1.53 ± 0.11	1.76 ± 0.12	100

^a benzamidine hydrochorides ; ^b Reference drug

Y

 Table 2. IC₅₀ values for compounds 9a-h (series I) assayed against L. amazonensis

 promastigotes.

		R ₄ R ₁	NH 9a-g	R ₃		OCH ₃
Comp		Subs	tituent		L. amo	nzonensis
-	\mathbf{R}_1	R ₂	R ₃	\mathbf{R}_4	IC ₅₀ (μg/L)	IC ₅₀ (µM)
9a	OCH ₃	Н	Н	Н	1.34 ± 0.18	1.85 ± 0.25
9b	Н	OCH ₃	Н	н	2.0 ± 0.11	2.56 ± 0.14
9c	Н	Н	OCH ₃	Н	2.28 ± 0.21	2.80 ± 0.26
9d	Н	OCH ₃	OCH ₃	Н	2.10 ± 0.01	2.57 ± 0.020
9e	OCH ₃	Н	OCH ₃	н	2.33 ± 0.16	2.80 ± 0.19
9f	Н	Н	OCH ₃	OCH ₃	1.77 ± 0.11	2.24 ± 0.14
9g	OCH ₃	Н	OCH ₃	OCH ₃	> 2.50	> 3.0
9h	OCH ₂ O	OCH ₂ O	OCH ₃	Н	1.13 ± 0.08	1.60 ± 0.11
Pent. ^a	-		-	-	-	1.05 ± 0.25

^aPentamidine: reference drug

Table 3. IC_{50} values for compounds 9i-v (series II) assayed against L. amazonensispromastigotes.



Compound	Substi	tuents	L. amazon	ensis
	R ₃	R ₅	IC ₅₀ (µg/ml)	IC ₅₀ (μM)
9i	Cl	Н	1.43 ± 0.43	1.90 ± 0.57
9j	Н	Cl	0.64 ± 0.0	1.11 ± 0.0
9k	F	Н	0.94 ± 0.0	1.43 ± 0.0
91	Н	F	1.13 ± 0,0	1.62 ± 0.0
9m	Br	Н	0.92 ± 0.0	1.34 ± 0.0
9n	Н	OCH ₃	1.20 ± 0.12	1.68 ± 0.17
90	NO ₂	Н	0.76 ± 0.11	1.22 ± 0.18
9p	н	NO ₂	2.13 ± 0.17	2.59 ± 0.2
9q	OH	Н	2.01 ± 0.27	2.51 ± 0.34
9r	Н	ОН	2.15 ± 0.076	2.65 ± 0.09
9s	CH ₃	Н	1.92 ± 0.21	2.42 ± 0.27
9t	Н	CH ₃	1.92 ± 0.32	2.42 ± 0.41
9u	SO_2NH_2	Н	1.37 ± 0.11	1.79 ± 0.14
9v	Cl	NO_2	0.69 ± 0.0	1.10 ± 0.0
Pentamidine ^a	-	-	0.82 ± 0.19	1.05 ± 0.25

^aPentamidine: reference drug.

Compound	$\log P^{a}$	MR ^a	\mathbf{MV}^{a}	POLZ ^a	ST ^a
9a	5.28	94.38	284.0	37.41	39.7
9b	5.28	94.38	284	37.41	39.7
9c	5.07	94.38	284.0	37.41	39.7
9d	5.23	100.19	305.7	39.72	38.9
9e	5.23	100.19	305.7	39.72	38.9
9f	4.89	100.19	305.7	39.72	38.9
9g	5.05	106.41	327.3	42.02	38.1
9h	4.48	93.28	265.7	36.97	44.12
9i	6.27	98.98	293.3	39.24	40.9
9j	6.31	98.98	293.3	39.24	40.9
9k	5.73	94.25	286.9	37.36	38.2
91	5.77	94.25	286.9	37.36	382
9m	6.45	101.94	296.5	40.41	42.4
9n	5.44	100.19	305.7	39.72	38.9
90	5.74	100.04	289.3	39.66	47.3
9р	5.67	100.04	289.3	39.66	47.3
9q	4.54	95.23	281.2	37.75	42.6
9r	4.93	95.23	281.2	37.75	42.6
9s	5.74	98.8	299.2	37.17	38.3
9t	5.74	98.8	299.2	39.17	38.3
9u	4.09	106.05	301.0	42.04	50.2
9v	6.45	106.64	298.6	41.48	48.4

Table 04. Table of descriptors; lipophilicity, log *P*, molecular refractivity (MR), molecular volume (MV), polarizability (POLZ) and superficial tension (ST).

^aACDLabs software package version 12.0



Figure 1. Drugs 1 and 2 and compound 3 with antileishmanial activity.









Figure 3. Molecular structure 9b and 9o-rotamer A and 9o-rotamer B.



Series I

Series II

9a = 3	$R_1 = OCH_3, R_2, R_3, R_4 = H$	9i*	$R_3 = Cl; R_5 = H$	9p*	$R_5 = H; R_5 = NO_2$
9b*	$R_2 = OCH_3; R_1, R_3, R_4 = H$	9j*	$R_3 = H; R_5 = Cl$	9q*	$R_5 = OH; R_5 = H$
9c	$R_3 = OCH_3; R_1, R_2, R_4 = H$	9k*	$R_3 = F; R_5 = H$	9r*	$R_5 = H; R_5 = OH$
9d*	$R_2, R_3 = OCH_3; R_1, R_4 = H$	91 *	$R_3 = H; R_5 = F$	9s*	$R_5 = CH_3; R_5 = H$
9e	$R_1, R_3 = OCH_3; R_2, R_4 = H$	9m*	$R_3 = Br; R_5 = H$	9t*	$R_5 = H; R_5 = CH_3$
9f	$R_3, R_4 = OCH_3; R_1, R_2 = H$	9n*	$R_3 = H; R_5 = OCH_3$	9u*	$R_5 = SO_2NH_2; R_5 = H$
9g	$R_1, R_3, R_4 = OCH_3; R_1 = H$	90*	$R_3 = NO_2; R_5 = H$	9v*	$R_5 = Cl; R_5 = NO_2$
9h*	$\mathbf{R}_1, \mathbf{R}_2 = \mathbf{OCH}_2\mathbf{O}; \mathbf{R}_3 = \mathbf{OCH}_3 \mathbf{R}_4 = \mathbf{H}$				
*New	compounds				

^aReagents and conditions: (i) NaOH 10 % (m/v), 45 min; (ii) PCl₅; dry toluene, 8 h.; (iii) *p*-R₃.aniline in dry toluene, reflux, 6 h; (iv) 5% aq. NaHCO₃, 4 h, CH₂Cl₃ extraction.

Highlights

- 1. Antileishmania activity evaluation of new N,N'-substituted benzamidines series
- 2. QSAR study involving the hydrophobicity and electronic parameters
- 3. Synthesis and characterization of N, N'-substituted benzamidines series

Supplementary data

Synthesis, Antileishmanial Activity and Structure-Activity Relationship of 1-*N*-X-phenyl-3-*N*'-Y-phenyl-benzamidines

Cláudio Eduardo Rodrigues-Santos,^a Leonor L. Leon,^b Adailton J. Bortoluzzi,^c Marilene

Marcuzzo Canto-Cavalheiro,^b Gérzia C. Machado,^b and Aurea Echevarria^{a,*}

^a Departamento de Química, Instituto de Ciências Exatas, Universidade Federal Rural

do Rio de Janeiro, 23890-900, Seropédica, RJ, Brazil

^b Departamento de Imunologia, Instituto Oswaldo Cruz, FIOCRUZ, 21042-900, Rio de

Janeiro, RJ, Brazil

^c Departamento de Química, Universidade Federal de Santa Catarina, 88040-900, Florianópolis, SC, Brazil

Corresponding author: Phone: +55 21 26822807, Fax: +55 21 26822807, E-mail: echevarr@ufrrj.br

Table of Contents

1	Molecular structure 9b and 9o-rotamer A and 9o-rotamer B	S 3
2	Crystal data for compounds 9b and 90	S 4
3	¹ H NMR spectrum of 9a	S 5
4	¹ H NMR spectrum of 9b	S 6
5	¹ H NMR spectrum of 9c	S 7
6	¹ H NMR spectrum of 9d	S 8
7	¹ H NMR spectrum of 9e	S 9
8	¹ H NMR spectrum of 9f	S 10
9	¹ H NMR spectrum of 9 g	S 11
10	¹ H NMR spectrum of 9h	S 12
11	¹ H NMR spectrum of 9i	S 13
12	¹ H NMR spectrum of 9j	S14
13	¹ H NMR spectrum of 9k	S15

14	¹ H NMR spectrum of 91	S16
15	¹ H NMR spectrum of 9m	S17
16	¹ H NMR spectrum of 9n	S 18
17	¹ H NMR spectrum of 90	S19
18	¹ H NMR spectrum of 9p	S20
19	¹ H NMR spectrum of 9q	S21
20	¹ H NMR spectrum of 9r	S22
21	¹ H NMR spectrum of 9s	S23
22	¹ H NMR spectrum of 9t	S24
23	¹ H NMR spectrum of 9u	S25
24	¹ H NMR spectrum of 9v	S26
25	¹³ C NMR spectrum of 9a	S27
26	¹³ C NMR spectrum of 9b	S28
27	¹³ C NMR spectrum of 9c	S29
28	¹³ C NMR spectrum of 9d	S 30
29	¹³ C NMR spectrum of 9e	S 31
30	¹³ C NMR spectrum of 9f	S 32
31	¹³ C NMR spectrum of 9g	S 33
32	¹³ C NMR spectrum of 9h	S 34
33	¹³ C NMR spectrum of 9i	S35
34	¹³ C NMR spectrum of 9j	S 36
35	¹³ C NMR spectrum of 9k	S 37
36	¹ H NMR spectrum of 91	S 38
37	¹³ C NMR spectrum of 9m	S39
38	¹³ C NMR spectrum of 9n	S40
39	¹³ C NMR spectrum of 90	S41
40	¹³ C NMR spectrum of 9p	S42
41	¹³ C NMR spectrum of 9q	S43
42	¹³ C NMR spectrum of 9r	S44
43	¹ H NMR spectrum of 9s	S45
44	¹³ C NMR spectrum of 9t	S46
45	¹³ C NMR spectrum of 9u	S47
46	¹³ C NMR spectrum of 9v	S48

X-ray crystal data



(Compound 9b	Compound 90
Empirical formula	C ₂₀ H ₁₈ N ₂ O	C ₂₀ H ₁₇ N ₃ O ₃
Formula weight	302.36	347.37
Temperature (K)	293(2)	293(2)
Wavelength (Å)	0.71069	0.71069
Crystal system	Monoclinic	Triclinic
Space group	P21/c	Pī
Unit cell dimensions		
A (Å)	9.5968(19)	10.4554(15)
b (Å)	14.2860(9)	11.4787(16)
c (Å)	12.0218(7)	16.6234(18)
α (°)		78.403(9)
β (°)	103.998(9)	73.919(9)
γ (°)		70 702(9)
Volume $(Å^3)$	1599 2(4)	1796 0(4)
7	4	4
Density (cal.) (Mg/m^3)	1 256	1 285
(mm^{-1})	0.078	0.088
μ (mm) E(000)	640	728
$\Gamma(000)$	$0.50 \times 0.36 \times 0.36$	$0.46 \times 0.30 \times 0.26$
Thoto rongo (°)	0.50 X 0.50 X 0.50	1.28 to 25.07
Perfections collected	2.19 10 23.07	6502
Independent reflections	2980 2841 [B(int) = 0.0125]	6356 [D (int) = 0.0222]
Refinament method	$E_{\text{VIII}} = 0.0123$	$\frac{1}{10000000000000000000000000000000000$
Deta / restrainta / response	Full-mail ix least-squales on F	Full-Inatian least-squares on F
Data / restraints / parameters	2841/0/214	1.045
Einel D indiana II: 2- (D)	1.040	1.043
Final R indices $[1>2\sigma(1)]$	R1 = 0.0377, WR2 = 0.0958	R1 = 0.0534, WR2 = 0.1382
R indices (all data)	R1 = 0.0668, WR2 = 0.1065	R1 = 0.1481, WR2 = 0.1677
Extinction coefficient	0.0213(18)	0.007
Largest diff. peak and hole (e.A) 0.161 and -0.159	0.296 and -0.170

Table S1. Crystal data and structure refinement for compounds 9b and 9o.

Acquisition Time (sec)	7.9299	Date	25 Oct 2007	7 12:27:12		File Name	E:\75\pdata	\1_075001r	
Frequency (MHz)	200.13	Nucleus	1H	Number of Transients	16	Original Points Count	32768	Points Count	32768
Pulse Sequence	zg30	Solvent	DMSO-D6	Sweep Width (Hz)	4132.23	Temperature (degree C	27.000		



Acquisition Time (sec)	7.9299	Date	30 Oct 200	7 17:07:40		File Name	F:\81\pdata\	1_081001r	
Frequency (MHz)	200.13	Nucleus	1H	Number of Transients	16	Original Points Count	32768	Points Count	32768
Pulse Sequence	zg30	Solvent	DMSO-D6	Sweep Width (Hz)	4132.23	Temperature (degree C)	27.000		



Acquisition Time (sec)	7.9299	Date	25 Oct 2007	7 09:50:14		File Name	E:\73\pdata	1_073001r	
Frequency (MHz)	200.13	Nucleus	1H	Number of Transients	16	Original Points Count	32768	Points Count	32768
Pulse Sequence	zg30	Solvent	DMSO-D6	Sweep Width (Hz)	4132.23	Temperature (degree C	27.000		





S8

Acquisition Time (sec)	7.9299	Date	24 Oct 2007	13:38:26		File Name E:	\69\pdata\	1_069001r	
Frequency (MHz)	200.13	Nucleus	1H	Number of Transients	16	Original Points Count 32	768	Points Count	32768
Pulse Sequence	zg30	Solvent	DMSO-D6	Sweep Width (Hz)	4132.23	Temperature (degree C) 27	.000		
								NH N	Br
956 	1999 1995 1996 1999 1996 1999 1996	1986 1986	-7.47 -7.18,22	6, 5, 5, 5, 5, 5, 5, 5, 5, 5, 5, 5, 5, 5,	Minery anges	п ₃ со			
9.0	8.5	2.13 1 	.16 4.86 2	7.0 6.5				-3.73	DMSO-d6
song denic - operance a mane managempi ang muccupat an	-9.26 -9.21	98.7 1997 -	20-1-4-1-1-2-4-4-4-4-4-4-4-4-4-4-4-4-4-4-4	225-722 27.03 17.18 27.05.90 6.82 6.86 6.89 6.59 6.54 6.54	włędowerty kang z czył – skoływany	non a la againgtait se nasarait an dhaingtain ag bha the ghan an dha dhaingtai	ánh vetere) ad sa braha	contemporation and a second	
10.0	0.52	2.1					45	3.00	·····







22 Feb 2013

Acquisition Time (sec)	1.3631	Comment A	MOSTRA: CE1	101-d (13C_CPD) - 400MH	Z SOLVENTE: DMS	SO	
Date	21 Feb 2013 12:5	12:56:32		File Name D:\claudio_029001r		lr	
Frequency (MHz)	100.61	Nucleus 13	С	Number of Transients	1024	Original Points Count	32768
Points Count	32768	Pulse Sequence zg	pg30	Solvent	DMSO-D6	Sweep Width (Hz)	24038.46
Temperature (degree C,	27.000 27.000 40 40 40 40 40 40 40 40 40	2 4 100 000 000 2 2 9 2 9 2 9 2 9 2 9 2 9 2 9 2	130 47 130 47 130 130 47 130 25	98.727 98.727 98.727 125 120 115		CH ₃ O	NH NH N-OCH ₃
		97 22 69 1	740 130.47 1	282727 282727 282727 282727 292727 292727 292727 292727 292727 292727 292727 292727 292727 292727 2010 100 Chemical Shift	90 80 (ppm)	70 60 50	40 30 20 10 0 -10 -20
170 16	5 160 155	150 145 140 135 1	30 125 12	20 115 110 105 Chemical St	100 95 90 nift (ppm)	85 80 75 70	0 65 60 55 50 45 40 35



0.04 (±0.02)MR + 29.04 (±9.98)

Chilling Mark