## Synthesis of Teleocidins A, B and Their Congeners. Part 1.<sup>1</sup> An Efficient Synthesis Method of N-(7-Alkyl-4-indolyl)-N-methyl-L-valine Esters, Essential Intermediates for Teleocidin Synthesis

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Abstract — Details of the synthesis method of N-{7-{3,7-dimethyl-1,6-octadien-3-yl}-4-indolyl}-N-methyl-L-valine esters 19 and 20, essential precursors for the tumor promoters lyngbyatoxin A (= teleocidin A-1) (2) and teleocidin A-2 (3) are presented Key reaction steps are i) introduction of the linally side chain into 12 to form 18 and ii) successive indole cyclization of 21 by way of thoomide derivatives 22

Teleocidin A was first isolated from the mycelia of Streptomyces mediocidicus as a strong skin irritant and a fish poison by Takashima and Sakai in 1960. Two years later, they obtained an extremely toxic substance, teleocidin B having a partial structure similar to the above from another strain of Streptomyces and characterized its chemical behavior by securing crystalline dihydroteleocidin B. In 1966 Hirata and co-workers studied the structure of teleocidin B and proposed a structure 1 for dihydroteleocidin B by chemical and X-ray crystallography <sup>49</sup> Independently in Hawaii, Moore and co-workers studied ingredients of the blue-green alga Lyngbya majuscula Gomont to find the origin of a severe dermatitis known as swimmers' itch, and isolated a highly inflammatory and vesicatory compound, lyngbyatoxin A in 1979 They proposed its structure to be 2 with an unknown absolute configuration at the chiral center C-14.5

Meanwhile Fujiki *et al* found that the above sample of dihydroteleocidin B had a strong tumor promoter activity in the screening system of the tests on irritation of the mouse ear, induction of ornithine decarboxylase in mouse skin, and adhesion of human promyelocytic leukemia cells <sup>6</sup> So the Takashima and Sakai's teleocidins A and B were reinvestigated extensively using high performance liquid chromatography (HPLC) for efficient and complete separation of the natural products and it is now firmly established that the above teleocidin A is actually composed of two compounds teleocidins A-1 (2) and A-2 (3),<sup>7</sup> and teleocidin B has been separated into four stereoisomers, teleocidins B-1 (4), B-2 (5), B-3 (6), and B-4 (7) <sup>8</sup> One of these substances, teleocidin A-1 is identical with lyngbyatoxin A and the previous dihydroteleocidin B is now termed dihydroteleocidin B-4

Recently pendolmycin (8) was isolated from *Nocardiopsis* strain SA 1715, collected in the river near Shanghai.<sup>10</sup> All these teleocidins are proved to be potent tumor promoters <sup>11</sup> <sup>12</sup>

In regard to the synthesis of this class of natural products, only one report on (±)-teleocidins B-3 and B-4 has appeared<sup>13</sup> except our papers concerning teleocidins A-1, A-2, B-3, and B-4 in short communications<sup>114</sup> and pendolmycin <sup>15</sup> In the series of papers starting from this report, we describe the details of our syntheses of all of these natural products as well as their congeners

The idea of the synthesis came from the previous finding<sup>16</sup> that the Grignard reaction of 2-[3-(1,3-dioxolan-2-yl)-1-oxopropyl]-1-[(4-methylphenyl)sulfonyl]pyrrole (9) with the reagent generated *in situ* from magnesium and

vinyl

Me

Me

7 Teleocidin B-4

 $R^4$ 

Me

ı-Pr

Me

1-Pr

(E)-3,7-dimethyl-2,6-octadienyl (geranyl) bromide in tetrahydrofuran afforded regioselectively the allylic reaction product 10 in good yield (Chart 1). This produced readily 7-(3,7-dimethyl-1,6-octadien-3-yl)indole (11) by removal of the nitrogen protecting group, followed by treatment with a catalytic amount of p-toluenesulfonic acid in boiling benzene. So a suitable starting compound leading to the natural products was selected to be either the methyl or the t-butyl ester of N-methyl-N-[4-[1-[(4-methylphenyl)sulfonyl]-2-pyrrolyl]-4-oxobutanoyl]-L-valine (12a, 12b). After introducing necessary terpenoid side chains as above to get 13, we aimed at the indole cyclization reaction to form N-(7-alkyl-4-indolyl)-L-valine derivatives 14. In this report, we describe the initial studies in which we established the synthetic pathway for these essential intermediates 14, which served not only as precursors for the lyngyatoxin A (teleocidin A-1) (2) and teleocidin A-2 (3) but also as useful compounds for the other teleocidin synthesis. With this knowledge, we have now achieved a straightforward synthesis of pendolmycin (8) starting from the key intermediate 12b  $^{15}$ 

Preparation of 4-[1-[(4-methylphenyl)sulfonyl]-2-pyrrolyl]-4-oxobutyric acid (17) from 1-[(4-methylphenyl)sulfonyl]pyrrole<sup>17</sup> by way of the Friedel-Crafts reaction product 16 was described previously <sup>18</sup> It was coupled with methyl<sup>19</sup> or t-butyl<sup>20</sup> N-methyl-L-valinate and methyl or t-butyl L-valinate, <sup>21</sup> using the mixed anhydride method<sup>22</sup> to obtain 12a, 12b, 12c, and 12d in 87%, 90%, 92%, and 90% yields <sup>23</sup> Grignard reaction on 12a – 12d was carried out by stirring with (E)-3,7-dimethyl-2,6-octadienyl (geranyl) bromide and magnesium in tetrahydrofuran at 0°C for 1-3.5 h. The tosyl group was eliminated during the reaction, and 18a, 18b, 18c, and

18d were produced in 80%, 80%, 49%, and 46% yields, respectively.

In search of the cyclization procedure from these amides 18 to the indole derivatives 14, we tried first the conventional Bischler-Napieralski reaction using phosphorus oxychloride<sup>24</sup> on the t-butyl esters 18b and 18d. The results were quite disappointing in that 18b afforded merely an 8% yield of an inseparable mixture of 19b and 20b except for 21b in 30% yield, and 18d gave only a trace of 19d + 20d when refluxed with phosphorus oxychloride in diethyl ether or dichloromethane for long time. Adding pyridine made the yield of 19b + 20b a little better, but it was only 19% yield at best in acctonitrile at 40°C. Other reagents, phosphorus pentoxide, trichloromethyl chloroformate, polyphosphate ester (PPE),<sup>25</sup> triphenylphosphine ditriflate,<sup>26</sup> and trimethylsilyl polyphosphate<sup>27</sup> afforded none of the desired products.

So we turned our attention to the synthesizing of 7-alkyl-4-aminoindole derivatives 19 and 20 by detouring the corresponding thioamides<sup>28</sup> instead of the direct and shortest step from the amides 18 (Chart 3) For that purpose, 18a - 18c were dehydrated by refluxing in benzene in the presence of p-toluenesulfonic acid for a short period to afford 21a, 21b, and 21c in 88%, 91% and 86% yields, respectively. These were then treated with Lawesson's reagent29 in refluxing tetrahydrofuran and thioamide derivatives 22a, 22b, and 22c were obtained in 65%, 65%, and 90% yields with an uncertain geometry around the double bond conjugated with the pyrrole ring The indole formation from 22a - 22c was examined in those conditions using a variety of the alkylating agents (iodomethane, iodoethane, 2-iodopropane, allyl bromide, methyl methanesulfonate, ethyl ptoluenesulfonate) and a metal reagent (silver tetrafluoroborate) in various solvents (dichloromethane, diethyl ether, tetrahydrofuran, t-butanol, acetonitrile, dimethylformamide) Some of the experimental results were summarized in Table 1 The reaction from thioamide 22c was unsatisfactory (Entries 1 – 3) Using iodomethane and allyl bromide as alkylating agents, the major products were alkyl 4-indolyl sulfides 23 (A = methyl and allyl) and the compound 24 Presumably an intermediate 25 arose during the reaction course Either the sulfur or the nitrogen substituent from 25 was liberated to afford the indole derivatives 19 and 20 or the sulfide 23 In the above two cases, the latter path predominated over the desired former step Compound 24 was formed directly from 22 by catalysis of hydrogen iodide, which was generated from 22 to 25 during the reactions

For the next trial, the substrate was changed to thioamide 22b, and the reaction was tested by using a variety of alkylating agents and solvents (Entries 4-10). Iodomethane afforded the best yield of the requisite products 19b + 20b (Entries 4-6) but a longer reaction time was required for the completion in t-butanol and tetrahydrofuran compared to acetonitrile. Other alkyl halides were inferior to iodomethane in demanding elevated reaction temperature and nevertheless providing poor yields of the products (Entries 7 and 8). Using the sulfonate ester and the metal reagent was not encouraging, because the reaction did not occur in clean states and the yields had to be only moderate (Entries 9 and 10). The sulfide 23 was not detected in all cases of 22b

Judging from the result of Entry 4, this combination of the substrate, alkylating agent, and solvent looked acceptable, but disadvantage of the use of 22b consisted in that the reaction product 19b + 20b were hardly separable A small amount of only 19b was obtained in the pure form by chromatography on a Lobar column

Table 1 Reaction of the Thioamide 22 with the Alkylating Agent to Form Indole Derivatives 19 and 20

Entry	Thio- amide	Alkylating agent A-X	Solvent	Temp °C	Time h	Product 19 Yield %	Product 20 Yield %	Sulfide 23 Yield %
1	22c	MeI	MeCN	28	6	19	12	36, <b>24</b> 26
2	22c	allyl bromide	MeCN	18	24	12	7	30, 24 37
3	22c	$AgBF_4$	$CH_2Cl_2$	27	1 5	19	13	
4	22b	MeI	MeCN	26	2 5	62		
5	22b	MeI	t-BuOH	22	7	6	52	
6	22b	MeI	THF	24	20	5	<b>19</b>	
7	22b	EtI	t-BuOH	50-55	8	3	19	
8	22b	Me <sub>2</sub> CHI	THF	reflux	9		8	
9	22b	MsOMe	CICH <sub>2</sub> CH <sub>2</sub> CI	reflux	1 75	4	16	
10	22b	$AgBF_{\mathtt{4}}$	CH <sub>2</sub> Cl <sub>2</sub>	27	3	4	14	
11	22a	MeI	DMF	24	3	37	25	27
12	22a	MeI	MeCN	27	3 5	32	20	4
13	22a	MeI	t-BuOH	25	6	27	18	11

which was not applicable to the preparation of 19b in a large scale.

So we examined the reaction with 22a by selecting iodomethane as the alkylating agent and tried to find a suitable solvent (Entry 11-13) The reaction condition of Entry 11 gave a satisfactory result, although the methyl 4-indolyl sulfide was produced in 27% yield. The resulting 7-alkyl-4-aminoindole derivatives were readily separated by the usual

silica gel chromatography to produce 19a and 20a as crystalline substances in 37% and 25% yields. These crystals contained about 4 – 6% of the double bond isomers having a 3,7-dimethyl-1,7-octadien-3-yl side chain, which were formed during the dehydration process of 18a with p-toluenesulfonic acid 16,30 Repeated recrystallization afforded the pure 19a and 20a in 27% and 19% yields, respectively. The product 20c was correlated with 20a by methylation with iodomethane in the presence of sodium bicarbonate in 78% yield 31. The indole derivative 19b was also correlated with 19a by reducing both with lithium aluminum hydride to give the same alcohol 26 A in 60% and 78% yields. The analogous reduction of 20a afforded 26 B in 75% yield. The configuration of the linallyl side chain of 19a and 20a was uncertain at this stage, and completion of the natural product synthesis revealed the chirality as shown

The compounds 26 A and 26 B are good substrates for examining optical purity. There was a little possibility of partial racemization of the L-valinate chirality at the stage of the Grignard reaction. So the amino alcohols 26 A and 26 B were converted to their esters 27 A and 27 B with (+)- $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetic acid (MTPA) and tested for purity by HPLC. For comparison, enantiomers of 26 A and 26 B (ent-26 A and ent-26 B) were prepared from methyl N-methyl-D-valinate by tracing all of the reaction steps (17  $\rightarrow$  12a  $\rightarrow$  18a  $\rightarrow$  22a  $\rightarrow$  19a and 20a) now established, and transformed them into their (+)-MTPA esters 28 A and 28 B. Good separation of HPLC peaks was secured among all of 27 A, 27 B, 28 A and 28 B, and comparison of their charts revealved that either 27 A or 27 B did not contain any trace of 28 A or 28 B, and vice versa Therefore no racemization had occurred during the processes to synthesize the amino-alcohols 26 A and 26 B. Thus the requisite N-(7-alkyl-4-indolyl)-L-valine esters 19 and 20 were obtained in four steps from the readily available compound 12 Completion of the natural product synthesis is reported in the next paper  $^{32}$ 

## **EXPERIMENTAL**

General Procedures — Melting points (mp) were determined on a Yanagimoto micro-melting point apparatus and are not corrected. Optical rotations were measured on Perkin-Elmer 241and JASCO DIP-370 polarimeters. Mass spectra (MS) were taken on a Hitachi RMS-4 spectrometer. High resolution mass spectra (HRMS) were measured on a JEOL JMS-DX-300 spectrometer. Infrared spectra (IR) were determined on a Hitachi 215 spectrophotometer. Proton magnetic resonance (<sup>1</sup>H NMR) spectra were recorded on a Varian EM 390 (90 MHz) spectrometer in CDCl<sub>3</sub> with TMS as an internal reference unless otherwise specified. Column chromatography was conducted on silica gel Fuji Davison BW 200 and preparative thin-layer chromatography (PTLC) was carried out on glass plates (20 × 20 cm) coated with Merck silica gel 60 PF<sub>254</sub> (1 mm thick). "Usual work-up" refers to washing the organic layers with water or brine, drying over anhydrous sodium sulfate and evaporating the solvents under reduced pressure

Methyl N-Methyl-N-[4-[1-[(4-methylphenyl)sulfonyl]-2-pyrrolyl]-4-oxobutanoyl]-L-valinate (12a), Methyl and t-Butyl N-[4-[1-[(4-Methylphenyl)sulfonyl]-2-pyrrolyl]-4-oxobutanoyl]-L-valinates (12c and 12d) — Preparation of 12a is typical A solution of 17 (128 mg, 0 399 mmol) in THF (5 ml) and Et<sub>3</sub>N (0 l3 ml, 0 934 mmol) was cooled at -20°C, and to this was added 5% v/v ClCOOEt-THF (0 82 ml, 0 43 mmol) After stirring at -20°C for 10 min, methyl N-methyl-L-valinate hydrobromide<sup>19</sup> (108 mg, 0 478 mmol) was added and

the mixture was stirred at that temperature for 10 min and then at room temperature for 45 min. Quenching the reaction with sat NaHCO<sub>2</sub>-H<sub>2</sub>O<sub>3</sub> extraction with CH<sub>2</sub>Cl<sub>2</sub>, usual-work-up and PTLC [hexane-EtOAc (3 2)] afforded 12a (155 mg, 87%), colorless needles, mp 132-133°C (CH<sub>2</sub>Cl<sub>2</sub>-hexane). Anal Calcd for C<sub>2</sub>H<sub>2</sub>N<sub>2</sub>O<sub>6</sub>S C, 58 91, H, 6 29, N, 6 25 Found C, 58 81, H, 6.20; N, 6 17  $[\alpha]_0^{2}$  -69 4° (c 1 01, CH,CL,). MS m/z. 448 (M\*). IR (KBr) cm<sup>-1</sup> 1739, 1672, 1625. <sup>1</sup>H NMR of major and minor rotamers δ 0 77 and 0 84 (3H, d each, J=6 5 Hz), 0 93 and 0 96 (3H, d each, J=6.5 Hz), ca 1 94-2 37 (1H, m), 2 37 (3H, s), 2.95 and 2 80 (3H, s each), 3 64 and 3 68 (3H, s each), 4 82 and 3 99 (1H, d each, J=10 5 Hz), 6 29 (1H, dd, J=3 5, 3.5 Hz), 7 13 (1H, dd, J=3.5, 2 Hz), 7 25 and 7 84 (A,B,, J=8.5 Hz), 7 74 (1H, dd, J=3 5, 2 Hz) Acadification of the NaHCO<sub>4</sub>-H<sub>2</sub>O layer to pH 5 with 30% HOAc-H,O, followed by extraction with CH<sub>2</sub>Cl<sub>2</sub>, usual work-up and PTLC (10% MeOH-CH<sub>2</sub>Cl<sub>2</sub>) recovered 17 (8 mg, 6%) 12c Colorless prisms, mp 94-95°C (Et<sub>2</sub>O) Anal Calcd for  $C_{21}H_{26}N_2O_5S$ . C, 58 05,  $\dot{H}_3$ 6 03, N, 6 45 Found. C, 57 91, H, 5 94, N, 6 36  $[\alpha]_D^{22}$  -1 3° (c 0 91, CH<sub>2</sub>Cl<sub>2</sub>) MS m/z 434 (M\*). IR (CHCl<sub>3</sub>) cm<sup>1</sup> 1738, 1674. <sup>1</sup>H NMR δ 0 83 (3H, d, J=7 Hz), 0.86 (3H, d, J=7 Hz), 2.08 (1H, dqq, J=5, 7, 7 Hz), 2 31-2 79 (2H, m), 2.40 (3H, s), 2.79-3 39 (2H, m), 3 69 (3H, s), 4 46 (1H, dd, J=8 5, 5 Hz), 6 24 (1H, br d, J=8 5 Hz, NH), 6 31 (1H, dd, J=3 5, 3 5 Hz), 7 11 (1H, dd, J=3 5, 2 Hz), 7 28 and 7 88 (A,B,, J=8 5 Hz), 7 78 (1H, dd, J=3 5, 2 Hz) The compound 12d was prepared in the same way as in the previous report 15a 12d Colorless prisms, mp 100-101°C (Et,O). Anal Calcd for C<sub>24</sub>H<sub>4</sub>,N<sub>2</sub>O<sub>6</sub>S C, 60 48, H, 6 77, N, 5 88 Found C, 60 25, H, 6 70, N, 5 75  $[\alpha]_{D}^{22}$  +4 6° (c 0 96, CHCl<sub>4</sub>) MS m/z 476 (M\*) IR (CHCl<sub>4</sub>) cm<sup>1</sup> 1728, 1675 <sup>1</sup>H NMR & 0.84 (6H, d, J=7 Hz), 1 44 (9H, s), 1 79-2 28 (1H, m), ca 2 28-2 78 (2H, m), 2 38 (3H, s), 2 78-3 40 (2H, m), 4 36 (1H, dd, J=8 5, 4 5 Hz), 6 17 (1H, br d, J=8 5 Hz, NH), 6 28 (1H, dd, J=3 5, 3 5 Hz), 7 09 (1H, dd, J=3 5, 1 5 Hz), 7 26 and 7 86  $(A_2B_3, J=8.5 Hz), 7.75 (1H, dd, J=3.5, 1.5 Hz)$ 

Grignard Reaction on 12a-12d — In a similar manner as in the previous report, 15a 18a-18d were prepared by the Grignard reaction of 12a-12d with (E)-3,7-dimethyl-2,6-octadienyl (geranyl) bromide in the presence of Mg at 0°C Methyl N-[5,9-Dimethyl-4-hydroxy-4-(2-pyrrolyl)-5-vinyl-8-decenoyl]-N-methyl-L-valinate (18a) Colorless syrup HRMS Calcd for C<sub>25</sub>H<sub>40</sub>N<sub>2</sub>O<sub>4</sub>. 432 2988 Found 432 2965 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1733, 1622 <sup>1</sup>H NMR δ 0 97 (3H, s), 1 49 (3H, s), 1 60 (3H, s), 2.77 and 2 79 (3H, s each), 3 64 (3H, s), 5 70-5 86 (1H, m), 6 04-6 19 (1H, m), 6 55-6 69 (1H, m), 8 80 (1H, br s, NH) t-Butyl N-[5,9-Dimethyl-4-hydroxy-4-(2pyrrolyl)-5-vinyl-8-decenoyl]-N-methyl-L-valinate (18b) Colorless syrup MS m/z 474 (M\*) IR (CHCl<sub>2</sub>) cm $^{1}$  1729, 1620  $^{1}$ H NMR  $\delta$  0 65, 0 76 and 0 84 (3H, d each, J=7 Hz), 0 69, 0 80 and 0 87 (3H, d each, J=7 Hz), 101 (3H, s), 1.34 and 144 (9H, s each), 152 (3H, s), 163 (3H, s), 280 and 283 (3H, s each), 365, 368, 475 and 4 79 (1H, d each, J=10 5, 10 5, 10 and 10 Hz, respectively), 4 30, 4 36, 4 40 and 4 50 (1H, s each, OH), 5 74-5 90 (1H, m), 6 07-6 23 (1H, m), 6 57-6 73 (1H, m), 8 73 (1H, br s, NH) Methyl N-[5,9-Dimethyl-4-hydroxy-4-(2pyrrolyl)-5-vinyl-8-decenoyl]-L-valinate (18c) Colorless syrup MS m/z 400 (M\*-H,O) IR (CHCl,) cm<sup>-1</sup> 1734, 1652 H NMR 8 1 49 (3H, s), 1 61 (6H, br s), 3 67 (3H, s), 3 91, 4 18 and 4 79 (1H, s each, OH), 4 45 and 4 46 (1H, dd each, J=8 5, 5 Hz), 5 70-5 86 (1H, m), 6 06-6 19 (1H, m), 6 22-6 50 (1H, m, CONH), 6 58-6 71 (1H, m), 8 90 (1H, br s, pyrrole NH) The following by-product was obtained in 13% yield along with 19c Methyl N-[4-oxo-4-(2-pyrrolyl)-butanoyl]-L-valinate Colorless scales, mp 95-96°C (Et,O) Anal Calcd for C<sub>14</sub>H<sub>20</sub>N<sub>2</sub>O<sub>4</sub> C, 59 98, H, 7 19; N, 10 00 Found C, 59 82, H, 7 17, N, 9 99 MS m/z 280 (M\*) IR (KBr) cm<sup>1</sup> 1740, 1635 <sup>1</sup>H NMR  $\delta$  0 87 (3H, d, J=6 5 Hz), 0 89 (3H, d, J=6 5 Hz), 1 84-2 43 (1H, m), 2 47-2 94 (2H, m), 3 15 (2H, dd, J=7, 7 Hz), 4 55 (1H, dd, J=9, 5 5 Hz), 6 23 (1H, ddd, J= 4, 2 5, 2 5 Hz), 6 72 (1H, br d, J=9 Hz, CONH), 6 88-7 08 (2H, m), 9 83 (1H, br s, pyrrole NH) t-Butyl N-[5,9-Dimethyl-4-hydroxy-4-(2-pyrrolyl)-5-vinyl-8-decenoyl]-L-valinate (18d) Colorless syrup MS m/z 460 (M<sup>+</sup>) IR (CHCl<sub>2</sub>) cm<sup>-1</sup> 1722, 1653 <sup>1</sup>H NMR δ 1 43 (9H, s), 1 49 (3H, s), 1 62 (6H, br s), 4 39 (1H, dd, J=8 5, 4 5 Hz), 4 83-5 30 (1H, m), 5 71-5 87 (1H, m), 6 08-6 22 (1H, m), 6 59-6 72 (1H, m), 8 88 (1H, br s, pyrrole NH) The following by-product was obtained in 29% yield along with 19d t-Butyl N-[4-oxo-4-(2-pyrrolyl)-butanoyl]-L-valinate Colorless syrup MS m/z 322 (M<sup>+</sup>) IR (CHCl<sub>2</sub>) cm<sup>-1</sup> 1722, 1668, 1641 <sup>-1</sup>H NMR δ 0 90 (3H, d, J=7 Hz), 0 92 (3H, d, J=7 Hz), 1 48 (9H, s), 1 92-2.37 (1H, m), 2 43-3 04 (2H, m), 3 19 (2H, dd, J=7, 7 Hz), 4 53 (1H, dd, J=9, 4 5 Hz), 6 18-6 34 (1H, m), 6 90-7 11 (2H, m), 7 07 (1H, br d, J=9 Hz, CONH), 10 22 (1H, br s, pyrrole NH)

Indole Cyclization of 18b under Bischler-Napieralski Condition. t-Butyl N-[7-(3,7-Dimethyl-1,6-octadien-3-yl)-4-indolyl]-N-methyl-L-valinate (19b + 20b) — A solution of 18b (29 mg, 0 06 mmol), POCl,

(0 10 ml, 1.08 mmol) and pyridine (0 3 ml) in CH<sub>3</sub>CN (3 ml) was heated at 40°C under N<sub>2</sub> atmosphere for 14 h The reaction was quenched with sat. NaHCO<sub>3</sub>-H<sub>2</sub>O at 0°C and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> Usual work-up and PTLC [hexane-benzene (1.2)] yielded ca. 3:2 mixture of 19b and 20b (5 mg, 19%) as colorless syrup MS m/z. 438 (M\*). IR (CHCl<sub>3</sub>) cm<sup>-1</sup>. 1714, 1630 <sup>1</sup>H NMR (300 MHz) of 19b and 20b  $\delta$  0.96 and 0 94 (3H, d each, J=6.5 Hz), 1 08 and 1 01 (3H, d each, J=6.5 Hz), 1 35 and 1 38 (9H, s each), 1.64 (3H, br s), 2 98 and 3 01 (3H, s each), 3 93 and 3 94 (1H, d each, J=11 Hz), 5 01-5 11 (1H, m), 5 21-5 32 (2H, m), 6 20 and 6 21 (1H, dd each, J=17.5, 11 Hz), 6.58 and 6 60 (1H, d each, J=8 and 7.5 Hz), 6 70 and 6 75 (1H, dd each, J=3, 2 Hz), 6 97 (1H, d, J=8 Hz), 7 03-7.09 (1H, m), 8.58 (1H, br s, NH)

Dehydration of 18a – 18c — In the same manner as in the previous report, <sup>15a</sup> 21a – 21c were prepared from 18a – 18c. Methyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienoyl]-N-methyl-L-valinate (21a) Colorless syrup HRMS Calcd for C<sub>25</sub>H<sub>38</sub>N<sub>2</sub>O<sub>3</sub> 414 2882. Found 414 2873 IR (CHCl<sub>3</sub>) cm <sup>1</sup> 1740, 1632 <sup>1</sup>H NMR δ: 1.15 (3H, s), 1 49 (3H, s), 1 60 (3H, s), 2 84 and 2.88 (3H, s each), 2 96-3 21 (2H, m), 3 67 (3H, s), 5 65 (1H, t, J=7 5 Hz), 6 59-6 74 (1H, m), 9 28 (1H, br s, NH). t-Butyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienoyl]-N-methyl-L-valinate (21b) Colorless syrup MS m/z 456 (M<sup>+</sup>) IR (CHCl<sub>3</sub>) cm <sup>1</sup> 1730, 1628 <sup>1</sup>H NMR of major and minor rotamers δ 0 83 and 0 80 (3H, d each, H=7 Hz), 1 00 and 0 95 (3H, d each, J=7 Hz), 1 18 (3H, s), 1 46 and 1 43 (9H, s each), 1 51 (3H, s), 1 63 (3H, s), 2 91 and 2 87 (3H, s each), 3.06 and 3 13 (2H, d each, J=7 5 Hz), 4 78 and 3 76 (1H, d each, J=10 5 Hz), 6 57-6 75 (1H, m), 9 53 (1H, br s, NH). Methyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienoyl]-L-valinate (21c) Colorless syrup MS m/z 400 (M<sup>+</sup>). IR (CHCl<sub>3</sub>) cm <sup>-1</sup> 1733, 1660 <sup>1</sup>H NMR δ 0 86 (3H, d, J=7 Hz), 0 91 (3H, d, J=7 Hz), 1 17 (3H, s), 1 48 (3H, s), 1 61 (3H, s), 2 16 (1H, dqq, J=5 5, 7, 7 Hz), 2 89 (2H, d, J=8 Hz), 3 72 (3H, s), 4 53 (1H, dd, J=9, 5 5 Hz), ca 4.87-5 19 (1H, m), 5 03 (1H, dd, J=18, 1 5 Hz), 5 07 (1H, dd, J=11, 1 5 Hz), 5 71 (1H, t, J=8 Hz), 5 88-6 33 (4H, m), 6 64-6 77 (1H, m), 9 44 (1H, br s, NH)

Thioamidation of 21a – 21c — In the same way as in the previous report, <sup>15a</sup> 22a – 22c were obtained by the reaction of 21a – 21c with Lawesson's reagent Methyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienthioyl]-N-methyl-L-valinate (22a) Colorless syrup HRMS Calcd for C<sub>25</sub>H<sub>38</sub>N<sub>2</sub>O<sub>2</sub>S 430 2654 Found 430 2654 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1736, 1630 <sup>1</sup>H NMR of major and minor rotamers δ 0 87 and 0 74 (3H, d each, J=6 5 Hz), 1 06 and 0 90 (3H, d each, J=6 5 Hz), 1 18 (3H, s), 1 50 (3H, s), 1 61 (3H, s), 2 29 (1H, dqq, J=10 5, 6 5, 6 5 Hz), 3 01 and 3.27 (3H, s each), 3 69 (3H, s), 6 29 and 4 26 (1H, d each, J=10 5 Hz), ca 4 85-5 11 (1H, m), 5 04 (1H, dd, J=17, 1 5 Hz), 5 07 (1H, dd, J=11, 1 5 Hz), 5 68 and 5 73 (1H, t each, J=7 Hz), 6 57-6 73 (1H, m), 8 85 (1H, br s, NH) t-Butyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienthioyl]-N-methyl-L-valinate (22b): Colorless syrup MS m/z 472 (M\*) IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1731 <sup>1</sup>H NMR of two rotamers δ· 1 16 (3H, s), 1 51 (3H, s), 1 61 (3H, s), 3 03 and 3 30 (3H, s each), 3 47 and 3 57 (2H, d each, J=7 5 Hz), 5 67 and 5 69 (1H, t each, J=7 5 Hz), 6 59-6 75 (1H, m), 8 97 (1H, br s, NH) Methyl N-[5,9-Dimethyl-4-(2-pyrrolyl)-5-vinyl-3,8-decadienthioyl]-L-valinate (22c) Colorless syrup MS m/z 416 (M\*) <sup>1</sup>H NMR δ 0 92 (3H, d, J=7 Hz), 0 97 (3H, d, J=7 Hz), 1 19 (3H, s), 1 52 (3H, s), 1 63 (3H, s), 2 33 (1H, dqq, J=5, 7, 7 Hz), 3 8 (2H, d, J=8 Hz), 3 77 (3H, s), 4 87-5 26 (4H, m), ca 5 64-6 07 (2H, m), 5 94-6 09 (1H, m), 6 07-6 24 (1H, m), 6 70 (1H, ddd, J=2 5, 2 5, 1 5 Hz), 7 69 (1H, br d, J=8 5 Hz, valine NH), 8 92 (1H, br s, pyrrole NH)

Indole Cyclization of 22c to Form 19c, 20c, 23 and 24 — The procedure with MeI is a representative A solution of 22c (23 mg, 0.055 mmol) in CH<sub>3</sub>CN (1 ml) was stirred with MeI (0 50 ml, 8 03 mmol) at room temperature for 6 h. After evaporation to dryness, sat NaHCO<sub>3</sub>-H<sub>2</sub>O was added and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> Usual work-up and PTLC [hexane-EtOAc (9 1)] afforded 23 (6 mg, 36%), 20c (2 5 mg, 12%), 19c (4 mg, 19%) and 24 (6 mg, 26%) in the order of increasing polarity Methyl N-[7-[(R)-3,7-Dimethyl-1,6-octadien-3-yl]-4-indolyl]-L-valinate (19c) Colorless syrup HRMS Calcd for  $C_{24}H_{34}N_2O_2$  382 2620 Found 382 2623 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1734 <sup>1</sup>H NMR  $\delta$  1 04 (3H, d, J=6 5 Hz), 1 08 (, d, J=6 5 Hz), 1 41 (3H, s), 1 45 (3H, s), 1 62 (3H, s), 2 19 (1H, dqq, J=6 5, 6 5, 6 5 Hz), 3 69 (3H, s), 4 03 (1H, d, J=6 5 Hz), 4 12-4 66 (1H, br s, valine NH), ca 4 90-5.21 (1H, m), 5 21 (1H, dd, J=10 5, 1 5 Hz), 5 23 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=3, 3 Hz), 8 57 (1H, br s, indole NH) Methyl N-[7-(S)-[3,7-Dimethyl-1,6-octadien-3-yl]-4-indolyl]-L-valinate (20c) Colorless syrup HRMS Calcd for  $C_{24}H_{34}N_2O_2$  382 2620 Found 382 2632 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1732 <sup>-1</sup>H NMR

δ 1 02 (3H, d, J=6 5 Hz), 1 08 (3H, d, J=6 5 Hz), 1 41 (3H, s), 1 43 (3H, s), 1 61 (3H, s), 2 19 (1H, dqq, J=6 5, 6 5, 6 5 Hz), 3 69 (3H, s), 4 01 (1H, d, J=6 5 Hz), 4 37 (1H, br s, value NH), 4 92-5 19 (1H, m), 5 22 (1H, dd, J=10 5, 1 5 Hz), 5 23 (1H, dd, J=18, 1 5 Hz), 6 19 (1H, d, J=8 Hz), 6 20 (1H, dd, J=18, 1 0 5 Hz), 6 46 (1H, dd, J=3, 2 Hz), 6 93 (1H, d, J=8 Hz), 7.02 (1H, dd, J=3, 3 Hz), 8 57 (1H, br s, indole NH) 7-(3,7-Dimethyl-1,6-octadien-3-yl)-4-(methylthio)indole (23, A=Me) Colorless syrup HRMS Calcd for C<sub>19</sub>H<sub>25</sub>NS: 299 1707 Found 299 1690 IR (CHCl<sub>3</sub>) cm<sup>-1</sup>· 1628 <sup>1</sup>H NMR δ 1 43 (3H, s), 1.45 (3H, s), 1 60 (3H, s), 2 53 (3H, s), 4 88-5 17 (1H, m), 5 24 (1H, dd, J=10 5, 1 5 Hz), 5 25 (1H, dd, J=18, 1 5 Hz), 6.21 (1H, dd, J=18, 10 5 Hz), 6.60 (1H, dd, J=3, 2 5 Hz), 6.95 (1H, d, J=7.5 Hz), 7 08 (1H, d, J=7 5 Hz), 7 11 (1H, dd, J=3, 3 Hz), 8 64 (1H, br s, NH) Methyl N-[5-(3,7-Dimethyl-1,6-octadien-3-yl)-5-(2-pyrrolyl)-2-tetrahydrothiophenylidene]-L-valinate (24) Colorless syrup MS m/z 416 (M\*) IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1733, 1635. <sup>1</sup>H NMR δ 1 11 (3H, s), 1 50 (3H, s), 1 63 (3H, s), 3 69 and 3 72 (3H, s each), 6 56-6.73 (1H, m), 8 34 (1H, br s NH). By use of allyl bromide was obtained 7-(3,7-dimethyl-1,6-octadien-3-yl)-4-(2-propen-1-ylthio)indole (23, A=allyl): Colorless syrup MS m/z 325 (M\*) <sup>1</sup>H NMR δ·1 42 (3H, s), 1 47 (3H, s), 1 62 (3H, s), 3 60 (2H, d, J=7 Hz), 4 87-5 20 (3H, m), 5 12-5 41 (2H, m), 5.93 (1H, ddt, J=17, 10, 7 Hz), 6 23 (1H, dd, J=18, 10 5 Hz), 6 67 (1H, dd, J=3, 2 Hz), 7.04 (1H, d, J=8 Hz), 7 12 (1H, d, J=8 Hz), 7 13 (1H, dd, J=3, 3 Hz), 8 67 (1H, br s, NH)

Indole Cyclization of 22a to Form 19a, 20a and 23 — According to the reported procedure, 15a 22a (740) mg, 1 72 mmol) in DMF (6 ml) was stirred with MeI (3 0 ml, 48 mmol) at 18°C for 4 h After working up as before, column chromatography over silica gel (35 g) [hexane-EtOAc (49 1)] afforded 23 (A=Me) (141 mg, 27%), the crude 19a (252 mg, 37%) and the crude 20a (170 mg, 25%) in the order of increasing polarity The crude 19a was recrystallized from MeOH-H,O to give methyl N-[7-[(R)-3,7-dimethyl-1,6-octadien-3-yl]-4indolyl]-N-methyl-L-valinate (19a) (183 mg, 27%) Colorless needles, mp 58-59 5°C. Anal Calcd for C<sub>2</sub>,H<sub>2</sub>,N<sub>2</sub>O<sub>2</sub>, C, 75 71, H, 9 15, N, 7 07 Found C, 75 69, H, 9 30, N, 6 98 [a]<sub>n</sub><sup>22</sup> -157 7° (c 0 995, CH<sub>2</sub>Cl<sub>2</sub>) MS m/z 396 (M\*) IR (KBr) cm  $^1$  1727, 1627  $^1$ H NMR  $\delta$  0 92 (3H, d, J=6 5 Hz), 1 09 (3H, d, J=6 5 Hz), 1.41 (3H, s), 1 45 (3H, s), 1.62 (3H, s), 2 37 (1H, dqq, J=11, 65, 65 Hz), 2 98 (3H, s), 3 57 (3H, s), 4 04 (1H, d, J=11 Hz), 488-5 17 (1H, m), 5 21 (1H, dd, J=10 5, 1 5 Hz), 5 23 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=18, 10 5 Hz), 654 (1H, d, J=8 Hz), 665 (1H, dd, J=3, 25 Hz), 694 (1H, d, J=8 Hz), 702 (1H, dd, J=3, 3 Hz), 857 (1H, br s, NH) Similarly recrystallization of the crude 20a from MeOH-H<sub>2</sub>O afforded 129 mg (19%) of methyl N-[7-[(S)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-N-methyl-L-valinate (20a) Colorless scales, mp 75-77°C Anal Calcd for  $C_{25}H_{36}N_2O_2$  C, 75 71, H, 9 15, N, 7 07 Found C, 75 71, H, 9 21, N, 6 91  $[\alpha]_D^{22}$  -191 1° (c 1 005,  $CH_2Cl_2$ ) MS m/z 396 (M\*) IR (KBr) cm <sup>1</sup> 1725, 1628 <sup>1</sup>H NMR  $\delta$  0 92 (3H, d, J=6 5 Hz), 1 02 (3H, d, J=6 5 Hz) Hz), 1 43 (3H, s), 1 44 (3H, s), 1 61 (3H, s), 2 37 (1H, dqq, J=11, 65, 65 Hz), 2 99 (3H, s), 3 63 (3H, s), 4 05 (1H, d, J=11 Hz), 4 91-5.19 (1H, m), 5 21 (1H, dd, J=10 5, 1 5 Hz), 5 23 (1H, dd, J=18, 1 5 Hz), 6.20 (1H, dd, J=18, 10 5 Hz), 6 54 (1H, d, J=8 Hz), 6 67 (1H, dd, J=3, 2 5 Hz), 6 94 (1H, d, J=8 Hz), 7 02 (1H, dd, J=3, 3 Hz), 8 57 (1H, br s, NH)

Transformation of 20c to 20a — To a solution of 20c (16 mg, 0.042 mmol) in MeOH (2 ml) were added MeI (2 0 ml, 32 mmol) and NaHCO<sub>3</sub> (35 mg, 0 42 mmol), and the mixture was refluxed for 48 h under Ar atmosphere After evaporation to dryness, H<sub>2</sub>O was added and the whole was extracted with CH<sub>2</sub>Cl<sub>2</sub> Usual work-up and PTLC [hexane-EtOAc (19 1)] afforded 20a (13 mg, 78%)

Reduction of 19b to Form 7-[(R)-3,7-Dimethyl-1,6-octadien-3-yl)-4-[N-[ (S)-1-hydroxy-3-methylbut-2-yl]-N-methyl]aminoindole (26 A) — A mixture of 19b (10 mg, 0 023 mmol) and LiAlH<sub>4</sub> (16 mg, 0.42 mmol) in THF (3 ml) was refluxed for 1 h After cooling, sat Rochelle salt- $H_2O$  was added and the resulting mixture was extracted with Et<sub>2</sub>O Usual work-up and PTLC [hexane-EtOAc (9·1)] gave 26 A (5 mg, 60%), colorless syrup HRMS Calcd for  $C_{24}H_{36}N_2O$  368 2827 Found 368 2832 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1633 <sup>1</sup>H NMR 8· 0 75 (3H, d, J=6 5 Hz), 0 83 (3H, d, J=6 5 Hz), 1 44 (6H, s), 1 61 (3H, s), 2 70-3 09 (1H, m, OH), 2 86 (3H, s), 3 60 (1H, dd, J=12, 12 Hz), 3 66-4 07 (2H, m), 4 93-5 19 (1H, m), 5 22 (1H, dd, J=10 5, 1 5 Hz), 5 25 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=18, 10 5 Hz), 6 57 (1H, d, J=8 Hz), 6 81 (1H, dd, J=3, 2 5 Hz), 6 97 (1H, d, J=8 Hz), 7 01 (1H, dd, J=3, 3 Hz), 8.61 (1H, br s, NH)

Reduction of 19a — Reaction of 19a (18 mg, 0 045 mmol) with LiAlH<sub>4</sub> (10 mg, 0 263 mmol) in THF (3 ml) was carried out at 18°C for 30 min to give 26 A (13 mg, 78%)

7-[(S)-3,7-Dimethyl-1,6-octadien-3-yl]-4-[N-[(S)-1-hydroxy-3-methylbut-2-yl]-N-methyl]amino-indole (26 B) — The ester 20a (10 mg, 0.025 mmol) was reduced with LiAlH<sub>4</sub> (6 mg, 0.158 mmol) as above to give 26 B (7 mg, 75%), colorless syrup HRMS Calcd for  $C_{24}H_{36}N_2O$ : 368 2827. Found 368 2806. IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1627 <sup>-1</sup>H NMR & 0.69 (3H, d, J=6 5 Hz), 0.80 (3H, d, J=6 5 Hz), 1.37 (3H, s), 1.42 (3H, s), 1.60 (3H, s), 2.68-3 16 (1H, m, OH), 2.85 (3H, s), 3.60 (1H, dd, J=12, 12 Hz), ca 3.66-4 11 (2H, m), 4.96-5 23 (1H, m), 5.23 (1H, dd, J=10.5, 1.5 Hz), 5.26 (1H, dd, J=18, 1.5 Hz), 6.23 (1H, dd, J=18, 10.5 Hz), 6.58 (1H, d, J=8 Hz), 6.83 (1H, dd, J=3, 2 Hz), 6.98 (1H, d, J=8 Hz), 6.98-7 13 (1H, m), 8.65 (1H, br s, NH)

The MTPA Ester 27 A — A solution of 26 A (7 mg, 0.019 mmol) in pyridine (0 3 ml) and  $CH_2Cl_2$  (0 3 ml) was stirred with the acid chloride (20 mg, 0 079 mmol) derived from (R)-(+)-MTPA at 0°C – room temperature for 1 h The reaction was quenched with sat NaHCO<sub>3</sub>-H<sub>2</sub>O and the mixture was extracted with Et<sub>2</sub>O Usual work-up and PTLC [hexane-EtOAc (12 1)] afforded 27 A (11 mg, 99%), colorless syrup HRMS Calcd for  $C_{34}H_{43}F_3N_2O_3$  584 3225 Found: 584 3237 IR (CHCl<sub>3</sub>) cm<sup>1</sup> 1747 <sup>1</sup>H NMR  $\delta$  0 93 (3H, d, J=6 5 Hz), 1 04 (3H, d, J=6 5 Hz), 1.44 (6H, s), 1 63 (3H, s), 2 79 (3H, s), 3 36 (3H, s), 3 71-3 99 (1H, m), 4 36 (1H, dd, J=11 5, 2 5 Hz), 4.64 (1H, dd, J=11 5, 6 Hz), 4.96-5 20 (1H, m), 5 12-5 40 (2H, m), 6 20 (1H, dd, J=18, 10 5 Hz), 6 40 (1H, d, J=8 Hz), 6.47 (1H, dd, J=3, 2 Hz), 6 92 (1H, d, J=8 Hz), 6 98 (1H, dd, J=3, 3 Hz), ca 7 28-7 59 (5H, m), 8 58 (1H, br s, NH)

The MTPA Ester 27 B — The same treatment of 26 B (6 mg, 0 016 mmol) as above gave 27 B (9 mg, 95%), colorless syrup HRMS Calcd for  $C_{34}H_{43}F_3N_2O_3$  584 3225 Found 584 3241 IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1743, 1626 <sup>1</sup>H NMR  $\delta$ . 0 94 (3H, d, J=7 Hz), 1 02 (3H, d, J=7 Hz), 1 44 (6H, s), 1 63 (3H, s), 2 80 (3H, s), 3 33 (3H, s), 3 74-4 02 (1H, m), 4 38 (1H, dd, J=11 5, 3 Hz), 4 67 (1H, dd, J=11 5, 6 Hz), 4 93-5 19 (1H, m), 5 22 (1H, dd, J=10 5, 1 5 Hz), 5 24 (1H, dd, J=18, 1 5 Hz), 6 19 (1H, dd, J=18, 10 5 Hz), 6 39 (1H, d, J=8 Hz), 6 47 (1H, dd, J=3, 2 Hz), 6.89 (1H, d, J=8 Hz), 6 97 (1H, dd, J=3, 3 Hz), 7 20-7 59 (5H, m), 8 56 (1H, br s, NH)

Methyl N-Methyl-D-valinate Hydrobromide — This was prepared from D-valine according to the literature <sup>19</sup> mp 133 5-134 5°C (MeOH-Et<sub>2</sub>O), colorless needles Anal Calcd for  $C_7H_{16}BrNO_2$  C, 37 18, H, 7 13, N, 6 20 Found C, 37 13, H, 7 06, N, 6 09  $[\alpha]_D^{23}$  -19 7° (c 2 005, DMF)

Ent-12 — Colorless needles, mp 132-133°C ( $\overline{\text{CH}}_2\text{Cl}_2$ -hexane) Anal Calcd for  $C_{22}\text{H}_{28}\text{N}_2\text{O}_6\text{S}$  C, 58 91, H, 6 29, N, 6 25 Found C, 58 92, H, 6 27, N, 6 22 [ $\alpha$ ]<sub>2</sub><sup>23</sup> +68 3° (c 1 010, CH<sub>2</sub>Cl<sub>2</sub>)

Ent-19a — Colorless needles, mp 62 5-64°C (MeOH- $H_2O$ ) Anal Calcd for  $C_{25}H_{36}N_2O_2$  C, 75 71, H, 9 15, N, 7 07 Found C, 75 82, H, 9 03, N, 7 07 [ $\alpha$ ]<sub>D</sub><sup>23</sup> +155 2° (c 1 000, CH<sub>2</sub>Cl<sub>2</sub>)

Ent-20a — Colorless scales, mp 76-78°C (MeOH-H<sub>2</sub>O) Anal Calcd for  $C_{25}H_{36}N_{2}O_{2}$  C, 75 71, H, 9 15, N, 7 07 Found C, 75 86, H, 9 16, N, 7 15  $[\alpha]_{D}^{23}$  +199 0° (c 1 012, CH<sub>2</sub>Cl<sub>2</sub>)

The MTPA Ester 28 A Derived from Ent-19a — Colorless syrup  $^{\circ}$  MS m/z 584 (M<sup>+</sup>) IR (CHCl<sub>3</sub>) cm  $^{\circ}$  1745, 1628  $^{\circ}$  H NMR  $\delta$  0 96 (3H, d, J=6 5 Hz), 1 03 (3H, d, J=6 5 Hz), 1 44 (6H, s), 1 62 (3H, s), 2 73 (3H, s), 3 39 (3H, s), 3 69-4 01 (1H, m), 4 33 (1H, dd, J=11 5, 3 Hz), 4 65 (1H, dd, J=11 5, 6 Hz), 4 93-5 20 (1H, m), 5 22 (1H, dd, J=10 5, 1 5 Hz), 5 23 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=18, 10 5 Hz), 6 43 (1H, d, J=8 Hz), ca 6 43-6 56 (1H, m), ca 6 86-7 04 (1H, m), 6 94 (1H, d, J=8 Hz), 7 12-7 61 (5H, m), 8 57 (1H, br s, NH)

The MTPA Ester 28 B Derived from Ent-20a — Colorless syrup MS m/z 584 (M<sup>+</sup>) IR (CHCl<sub>3</sub>) cm<sup>-1</sup> 1747, 1630 <sup>1</sup>H NMR δ 0 95 (3H, d, J=6 5 Hz), 1 02 (3H, d, J=6 5 Hz), 1 42 (6H, s), 1 61 (3H, s), 2 75 (3H, s), 3 39 (3H, s), 3 72-4 02 (1H, m), 4 34 (1H, dd, J=11 5, 3 Hz), 4 67 (1H, dd, J=11 5, 6 Hz), 4 91-5 23 (1H, m), 5 23 (1H, dd, J=10 5, 1 5 Hz), 5 24 (1H, dd, J=18, 1 5 Hz), 6 20 (1H, dd, J=18, 10 5 Hz), 6 44 (1H, d, J=8 Hz), *ca* 6 44-6 57 (1H, m), *ca* 6 87-7 03 (1H, m), 6 94 (1H, d, J=8 Hz), 7 20-7 62 (5H, m), 8 58 (1H, br s, NH)

HPLC Analysis of the Four MTPA Esters 27 A, 27 B, 28 A and 28 B — Column TSK Silica 60,  $4.6 \times 250$  mm, mobile phase hexane – benzene – 2-propanol (95  $4.9 \times 0.1$ ), flow rate 0.25 ml/min Retention time of 27 A, 27 B, 28 A and 28 B 30.2 min, 38.2 min, 29.4 min and 35.5 min

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