S-Glucosylated Hydantoins as New Antiviral Agents

Ahmed A. El-Barbary,[†] Ahmed I. Khodair,[†] Erik B. Pedersen,^{*} and Claus Nielsen[‡]

Department of Chemistry, Odense University, DK-5230 Odense M, Denmark, and Retrovirus Laboratory, Department of Virology, Statens Seruminstitut, Artillerivej 5, DK-2300 Copenhagen, Denmark

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S-Glycosylation took place on reaction of 5-alkylidene- and 5-arylidene-3-aryl-2-thiohydantoins with glycosyl halides under alkaline conditions. Bisglucosylation also took place when N-3 unsubstituted hydantoins were reacted. The bisglucosylated hydantoins produced N-3 glucosylated hydantoins on treatment with ammonia in methanol. In antiviral studies the most active compounds against both HSV-1 and HSV-2 were 5-(2-thienylmethylene)-3-phenyl-2-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoin and 5-(2-thienylmethylene)-3-(4-chlorophenyl)-2-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoin.

Introduction

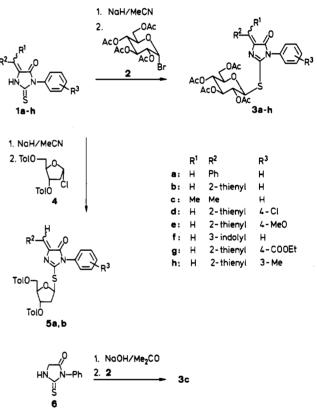
Some acyclic nucleoside analogues have achieved considerable success as antiviral agents¹ because of their low toxicity for normal cells while having an inhibitory activity against herpes simplex virus (HSV). One of the most potent compounds is 9-[(2-hydroxyethoxy)methyl]guanine (acyclovir).^{2,3} Recently, novel phosphonate isosteres of acyclovir and ganciclovir monophosphates have also been found selective antiherpes virus agents.⁴ Among pyrimidine nucleosides,⁵ 5-iodo-2'-deoxyuridine (IdUrd) has been in clinical use as a drug for years. The most active congeners among the 5-substituted 2'-deoxyuridine derivatives are (E)-5-(2-halogenovinyl)-2'-deoxyuridines.⁶ These compounds are particularly active against herpes simplex virus type 1 (HSV-1) and varicella-zoster virus. The structure activity relationship of 5-substituted 2'deoxyuridine analogues has been studied.^{7,8} Many examples of new nucleoside analogues have been described that show broad spectrum antiviral activity.⁹ On the contrary, non-nucleosides are expected to interact with very specific targets, hence a narrow spectrum and a good chance of a high therapeutic index could be the result.⁹ As a part of our program to find new antiviral agents, we report our results on the synthesis and evaluation of the antiherpes virus activity of the 5-arylidene-3-phenyl-2- $[3,5-bis-O-(4-methylbenzoyl)-\beta-D-erythro-pentofuranosyl]-$ 2-thiohydantoins and 5-arylidene-2-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoins. Our interest in 2-glucosylated hydantoin derivatives has been outlined in an earlier publication.¹⁰

Chemistry

For the synthesis of hydantoins **5a,b** (Scheme 1), compounds **1a,b** were treated with 1.1 equiv of NaH in anhydrous acetonitrile followed by 1.1 equiv of the 2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranosyl chloride, 4, prepared from 2-deoxy-D-ribose according to the published methods.¹¹⁻¹³ For the synthesis of glucosyl hydantoins **3a-h**, compounds **1a-h** were likewise treated with 1.1 equiv of NaH in anhydrous acetonitrile followed by 1.1 equiv of the 2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl bromide 2. Alternatively, the isopropylidene derivative **3c** could be synthesized by treatment of 3-phenyl-2-thiohydantoin 6

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with aqueous NaOH in acctone followed by 2. In all cases the yields were in the range of 72-95%.

When the N-3 unsubstituted compounds 7a-c were reacted with 2 in aqueous acetone, both mono- and bisglucosylation reactions took place with formation of the compounds 8 and 9, respectively (Scheme 2). Upon deprotection of 9a,b with ammonia in methanol, the glucosylthio group was most likely replaced by a methoxide group in a nucleophilic substitution reaction. Subsequent demethylation afforded the N-3 glucosyl hydantoin derivatives 10a,b. However, formation of the 2-oxo derivative directly from moisture cannot be excluded. This type of cleavage also explains why we were not successful in the deprotection of the compounds 3, 5, or 8 in saturated methanolic ammonia.

The ¹H-NMR (Me₂SO- d_{6}) spectrum of 5-(phenylmethylene)-3-methylhydantoin has been reported to show N¹-H at 10.72 ppm whereas 5-(phenylmethylene)-1-methylhy-

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[†]On leave from Chemistry Department, Faculties of Science and Education, Tanta University, Tanta, Egypt. [‡]Statens Seruminatitut.

Scheme 2

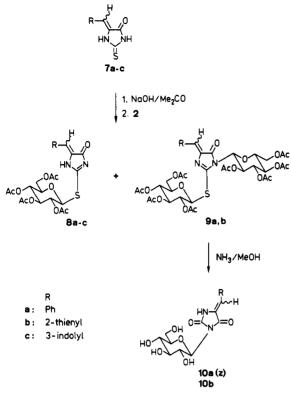


Table 1. Antiviral Activity of

3-Aryl-2-glucopyranosyl-2-thiohydantoin Derivatives 3a-h against HSV-1 in Vero Cells

	ED ₅₀ , ^α μM			SI¢	
compd	HSV-1	HSV-2	$\mathrm{CD}_{50}{}^b\mu\mathrm{M}$	HSV-1	HSV-2
3a	>100	100	>100		****
3b	0.4	0.7	24	60	34
3c	119	100	>200	>1.6	>2
3d	0.6	1	20	33	20
3e	27	27	35	1.3	1.3
3f	46	40	>100	>2.2	>2.5
3g	32	46	46	1.4	1
3b	10	19	25	2.5	1.3
BVDU	0.3	6.8	>100	>333	>15
Acyclovir	0.5	0.5	>50	>100	>100

^a Effective dose of compound, achieving 50% protection of Vero cells against either HSV-1 or HSV-2. ^b Cytotoxic dose of compound, required to reduce the viability of normal uninfected Vero cells by 50%. ^c Selectivity index: ratio CD_{50}/ED_{50} . ED_{50} and CD_{50} are expressed as the mean values of three independent determinations.

dantoin showed N³-H at 11.38 ppm.¹⁴ The ¹H-NMR (DMSO- d_6) spectra of 10a,b showed NH at 10.91 and 10.70 ppm, respectively, corresponding to N¹-H, proving N-3 glycosylation. N-3 glucosylation was further confirmed by nuclear Overhauser effect (NOE) experiments on compound 10a. On irradiation of the NH resonance 10.91 ppm a large NOE enhancement was found for the ortho protons of the phenyl group (9%) and on irradiation of the ortho protons a small NOE enhancement was found for NH (2%). Actually, this also proves Z configuration of the exocyclic double bond in compound 10a.

Antiviral Activity

The glucopyranosylthiohydantoin derivatives 3a-h were tested for their activity against herpes simplex virus, type 1 (HSV-1) and type 2 (HSV-2) in Vero cells. As shown in Table 1, only 3a was devoid of any activity against HSV-1 at 100 μ M whereas 3b-h were active; inhibitory concentrations (ED₅₀) ranged from 0.4 to 119 μ M. Most compounds, however, had an *in vitro* therapeutic index of only 1-3. Compounds **3b** and **3d** were found to be interesting, having activities against HSV-1 comparable to those of acyclovir and BVDU. However, their selectivity index (SI) was in the range of 33-60 and was lower than the one of acyclovir. Since **3b** and **3d** belong to a new class of active compounds and also were active against HSV-2, further investigations are needed in order to determine the mechanism of their action against herpes virus.

None of the 3-unsubstituted 2-glucopyranosyl-2-thiohydantoin derivatives 8a-c, the 3-phenyl-2-(2-deoxy-Dpentofuranosyl)hydantoin derivatives 5a,b, or the bisglucosyl derivative 9b showed any selectivity and/or antiviral activity against HSV-1 or HIV-1. Nor were any of the 3-aryl-2-glucopyranosyl-2-thiohydantoins 3a-h endowed with substantial anti-HIV activity in MT-4 cells.

Experimental Section

The ¹H-NMR spectra were recorded on a Bruker A 250 FT NMR spectrometer with tetramethylsilane as an internal standard. Chemical shifts are reported in parts per million (δ), and signals are expressed as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), or br (broad). Mass spectra (MS) were recorded using electron ionization (EI) on a Varian Mat 311A spectrometer or using fast atom bombardment (FAB) on a Kratos MS 50 spectrometer. The silica gel (0.040–0.063 mm) used for column chromatography was purchased from Merck. IR spectra were recorded with a Perkin-Elmer 1720 spectrometer.

3-Aryl-2-thichydantoin Derivatives. These compounds were prepared according to the published method¹⁵ for the preparation of 3-phenyl-2-thichydantoin.

3-[4-(Ethoxycarbonyl)phenyl]-2-thiohydantoin. Glycine (0.75 g, 10 mmol) was dissolved in a mixture of water (25 mL) and pyridine (25 mL), and the pH was adjusted to 9 by addition of 1 M NaOH. The solution was heated at 40 °C during addition of 4-(ethoxycarbonyl)phenyl isothiocyanate (3.9g) under vigorous stirring. NaOH (1 M) was added portionwise to keep the pH at 9. The reaction was complete when the alkali consumption ceased after 30 min. Pyridine and excess of 4-(ethoxycarbonyl)phenyl isothiocyanate were then removed by repeated extractions with equal volumes of benzene. A volume of 1 M HCl equivalent to the total volume of 1 M NaOH was added. The solvent was removed under reduced pressure, and the residue was suspended in 30 mL of a saturated solution of HCl in EtOH and refluxed for 4 h. The reaction mixture was concentrated to dryness in vacuo. The residue was triturated with water, and the solid product was filtered off and recrystallized from methanol to give 0.89 g (30%) of 3-[4-(ethoxycarbonyl)phenyl]-2-thiohydantoin: mp 164 °C; MS (EI) m/z 264 (M⁺); ¹H NMR (DMSO-d₆) δ 1.32 $(3H, t, J = 6.8 Hz, CH_3), 3.86 (2H, q, J = 6.4 Hz, CH_2), 7.46 (2H, q, J = 6.4 Hz, CH_2), 7$ d, J = 8.0 Hz, aryl), 8.05 (2H, d, J = 8.1 Hz, aryl), 10.42 (1H, s, NH); ¹³C NMR (DMSO-d₆) δ 13.99 (CH₃), 49.11 (C-5), 60.79 (CH₂), 128.99, 129.29, 129.67, 137.45 (aryl), 165.00 (C=O), 171.72 (C-4), 182.51 (C-2).

5-Arylidene-3-aryl-2-thiohydantoins (1a-h) and 5-Arylidene-2-thiohydantoins 7a-c. The standard procedure was used.¹⁶

3-(4-Chlorophenyl)-5-(2-thienylmethylene)-2-thiohydantoin (1d): yield 2.28 g (71%); mp 275 °C; MS (EI) m/z 320 (M⁺); ¹H NMR (DMSO- d_{e}) δ 6.84 (1H, s, =CH), 7.23–7.95 (7H, m, aryl); ¹³C NMR (DMSO- d_{e}) δ 105.79 (=CH), 124.27, 128.78, 129.17, 130.59, 130.79, 131.03, 132.09, 133.32, 135.31 (C-5, aryl), 163.35 (C-4), 177.57 (C-2).

3-(4-Methoxyphenyl)-5-(2-thienylmethylene)-2-thiohydantoin (1e): yield 2.34 g (74%); mp 274 °C; MS (EI) m/z 316 (M⁺); ¹H NMR (DMSO- d_6) δ 3.81 (3H, s, CH₃), 7.02–7.80 (8H, m, —CH, aryl), 12.47 (1H, s, NH); ¹³C NMR (DMSO- d_6) δ 55.27 (CH₃), 112.04 (—CH), 113.88, 125.07, 125.60, 127.71, 129.85, 131.86, 135.02, 135.56, 159.16 (C-5, aryl), 161.31 (C-4), 174.81 (C-2).

5-(3-Indolylmethylene)-3-phenyl-2-thiohydantoin (1f): yield 2.5 g (78%); mp 300 °C; MS (EI) m/z 319 (M⁺); ¹H NMR (DMSO-d₆) δ 7.10-9.1 (11H, m, aryl), 12.00 (2H, s, 2 NH); ¹³C NMR (DMSO-d₆) δ 108.96, 112.40, 117.35, 120.80, 122.57, 123.21, 127.44, 128.48, 128.64, 128.83, 130.58, 133.60, 135.90 (-CH, C-5, aryl), 161.50 (C-4), 172.14 (C-2).

3-[4-(Ethoxycarbonyl)phenyl]-5-(2-thienylmethylene)-2thiohydantoin (1g). To a mixture of 3-[4-(ethoxycarbonyl)phenyl]-2-thiohydantoin (0.89 g, 3 mmol) and piperidine (0.26 g, 3 mmol) in absolute ethanol (20 mL), was added 2-thiophenecarboxaldehyde (0.34 g, 3 mmol). The reaction mixture was stirred at room temperature for 6 h until the starting material was consumed (TLC). The reaction mixture was poured into water, and the yellow solid obtained was filtered off. Recrystallization from ethanol gave 0.9 g (84%) of 1g: mp 200 °C dec; MS (EI) m/z 358 (M⁺); ¹H NMR (DMSO-d₆) δ 1.35 (3H, t, J =6.9 Hz, CH₃), 4.36 (2H, q, J = 6.8 Hz, CH₂), 7.06 (1H, s, =CH), 7.11-8.11 (8H, m, aryl), 12.50 (1H, s, NH); ¹³C NMR (Me₂SO-d₆) δ 14.05 (CH₃), 60.91 (CH₂), 105.95 (=CH), 124.25, 128.87, 129.06, 129.17, 129.48, 130.85, 131.12, 135.31, 137.30 (C-5, aryl), 163.25 (C=O), 165.02 (C-4), 177.27 (C-2).

3-(3-Methylphenyl)-5-(2-thienylmethylene)-2-thiohydantoin (1h): yield 1.2 g (80%); mp 152 °C; MS (EI) m/z 300 (M⁺); ¹H NMR (DMSO- d_6) δ 2.35 (3H, s, CH₃, 6.79–7.90 (8H, m, =-CH, aryl), 12.35 (1H, s, NH); ¹³C NMR (DMSO- d_6) δ 20.67 (CH₃), 105.37 (=-CH), 125.48, 125.72, 128.44, 128.97, 129.16, 130.44, 130.75, 133.36, 135.69, 138.11 (C-5, aryl), 163.90 (C-4), 178.01 (C-2).

5-Arylidene- and 5-Isopropylidene-3-phenyl-2-(2,3,4,6tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoin (3a-h). General Procedure. The hydantoin 1 (5 mmol) was suspended in anhydrous MeCN (25 mL) at room temperature. To this suspension was added NaH (50%, 0.26g, 5 mmol), and the mixture was stirred at room temperature for 30 min. The mixture became clear after 15 min. 2,3,4,6-Tetra-O-acetyl- α -D-glucopyranosyl bromide (2, 2.26 g, 5.5 mmol) was added, and the mixture was stirred at room temperature for 12 h until the starting material was consumed (TLC) and then filtered. The residue from evaporation of the filtrate under reduced pressure was purified by silica gel column chromatography with Et₂O/petroleum ether (1:1, v/v) to give the products **3a-h**.

5-Benzylidene-3-phenyl-2-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (3a): yield 3.1 g (91%); mp 182 °C (lit.¹⁰ mp 180 °C).

3-Phenyl-5-(2-thienylmethylene)-2-(2,3,4,6-tetra-O-acetylβ-D-glucopyranosyl)-2-thiohydantoin (3b): yield 2.75 g (89%); mp 189–190 °C; MS (EI) m/z 616 (M⁺), 286 (base), 331 (glycon + 1); IR (KBr) 1752 (C=O); ¹H NMR (CDCl₃) δ 1.87 (3H, s, CH₃), 2.02 (6H, s, 2CH₃), 2.07 (s, 3H, CH₃), 4.04 (1H, m, 5'-H), 4.23 (m, 2H, 6'-H), 5.17 (t, J = 9.7 Hz, 1H, 4'-H), 5.27 (t, J = 9.9Hz, 1H, 2'-H), 5.41 (t, J = 9.3 Hz, 1H, 3'-H), 5.91 (d, J = 10.5 Hz, 1H, 1'-H), 7.12–7.70 (m, 8H, aryl); ¹³C NMR (CDCl₃) δ 20.22, 20.33 (4 Ac), 61.51 (C-6'), 67.88 (C-2'), 68.86 (C-3'), 73.88 (C-4'), 76.84 (C-5'), 81.27 (C-1'), 119.52 (=CH), 126.81, 127.34, 129.12, 129.45, 131.76, 133.54, 134.08, 135.33, 137.65 (C-5, aryl), 159.29 (C-2), 167.46 (C-4), 169.11, 169.17, 170.38 (4 C=O). Anal. (C₂₈H₂₈N₂O₁₀S₂) C, H, N.

5-Isopropylidene-3-phenyl-2-(2,3,4,6,tetra-*O*-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (3c): yield using the general procedure 2.4 g (85%); mp 167–168 °C; MS (EI) m/z 562 (M⁺), 232 (base), 331 (glycon + 1); IR (KBr) 1757, 1718 (C=O); ¹H NMR (DMSO-d₆) δ 1.94 (s, 3H, CH₃), 197 (s, 6H, 2 CH₃), 199 (s, 3H, CH₃), 2.29, 2.34 (2 s, 6H, (CH₃)₂CH=), 4.04 (m, 1H, 6'-H), 4.16 (m, 2H, 5'-H, 6'-H), 4.97 (1H, t, J = 9.2 Hz, 4'-H), 5.10 (1H, t, J = 9.8 Hz, 2'-H), 5.50 (t, J = 9.8 Hz, 1H, 3'-H), 5.90 (d, J = 10.4 Hz, 1H, 1'-H), 7.27–7.55 (m, 5H, aryl); ¹³C NMR (DMSO-d₆) δ 18.51, 21.74 (2 Me), 20.11, 20.17, 20.23 (4 Ac), 61.50 (C-6'), 67.78 (C-2'), 68.93 (C-3'), 72.80 (C-4'), 74.99 (C-5'), 80.16 (C-1'), 153.34 (C-2), 165.64 (C-4), 169.06, 169.30, 169.75 (4 C=O). Anal. (C₂₈H₈₀N₂O₁₀S) C, H, N.

Synthesis of 3c from 6. 3-Phenyl-2-thiohydantoin (6, 0.96 g, 5 mmol) was suspended in aqueous NaOH (4.4%, 5 mL) at room temperature. To this suspension was added acetone (25 mL), and the mixture became clear after stirring at room temperature for 5 min. 2,3,4,6-Tetra-O-acetyl- α -D-glucopyranosyl bromide (2, 2.26 g, 5.5 mmol) was added, and the mixture was stirred overnight at room temperature. After filtration and

evaporation of the filtrate under reduced pressure, the residue was chromatographed on a silicagel column with ether/petroleum ether (1:1, v/v) to obtain 1.3 g (67%) of 3c.

3-(4-Chlorophenyl)-5-(2-thienylmethylene)-2-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (3d): yield 3.0 g (92%); mp 183–185 °C; MS (FAB) m/z 651 (M + H⁺), 331 (glycon + 1), 321 (base + 1); IR (KBr) 1752 (C==O); ¹H NMR (DMSO-d₆) δ 1.81 (s, 3H, Ac), 1.96 (s, 3H, Ac), 1.99 (s, 3H, Ac), 2.02 (s, 3H, Ac), 4.07 (d, J = 11.1 Hz, 1H, 6'-H), 4.22 (m, 2H, 5'-H, 6'-H), 5.15 (t, J = 9.5 Hz, 1H, 4'-H), 5.37 (t, J = 9.7 Hz, 1H, 2'-H), 5.55 (t, J = 9.3 Hz, 1H, 3'-H), 5.87 (d, J = 10.2 Hz, 1H, 1'-H), 7.19–7.94 (m, 7H, aryl); ¹³C NMR (DMSO-d₆) δ 20.18, 20.21, 20.29 (4 Ac), 61.49 (C-6'), 67.67 (C-2'), 68.60 (C-3'), 72.85 (C-4'), 75.57 (C-5'), 80.63 (C-1'), 119.34 (=CH), 127.73, 129.39, 129.58, 130.78, 133.99, 134.75, 135.10, 137.41 (C-5, aryl), 158.31 (C-2), 166.59 (C-4), 169.23, 169.26, 169.46, 169.85 (4 C==O).

3-(4-Methoxyphenyl)-5-(2-thienylmethylene)-2-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (3e): yield 2.95 g (91%); mp 190–192 °C; MS (EI) m/z 646 (M⁺), 331 (glycon + 1), 316 (base); IR (KBr) ν (C=O) 1752; ¹H NMR (DMSO-d₆) δ 1.82 (s, 3H, Ac), 1.96 (s, 3H, Ac), 1.99 (s, 3H, Ac), 2.03 (s, 3H, Ac), 3.82 (s, 3H, OMe), 4.07 (d, J = 10.1 Hz, 1H, 6'-H), 4.22 (m, 2H, 5'-H, 6'-H), 5.13 (t, J = 9.7 Hz, 1H, 4'-H), 5.53 (d, J = 10.3 Hz, 1H, 2'-H), 5.53 (t, J = 9.3 Hz, 1H, 3'-H), 5.85 (d, J = 10.3 Hz, 1H, 1'-H), 7.05–7.93 (=CH, m, 7H, Aryl); ¹³C NMR (DMSO-d₆) δ 20.13, 20.16, 20.24 (4 Ac), 55.35 (OMe), 61.44 (C-6'), 67.66 (C-2'), 68.59 (C-3'), 72.86 (C-4'), 75.53 (C-5'), 80.47 (C-1'), 114.65 (=CH), 118.79, 124.20, 127.63, 128.98, 134.46, 134.79, 134.99, 137.44, 159.50 (C-5, aryl), 159.71 (C-2), 166.96 (C-4), 169.14, 169.19, 169.38, 169.78 (4 C=O). Anal. (C₂₉H₃₀N₂O₁₁S₂) C, H, N.

5-(3-Indolylmethylene)-3-phenyl-2-(2,3,4,6-tetra-O-acetylβ-**D-glucopyranosyl)-2-thiohydantoin (3f)**: yield 5.07 g (78%); mp 215 °C; MS 651 (M⁺); ¹H NMR (DMSO-d₆) δ 1.71 (3H, s, Ac), 1.98 (3H, s, Ac), 2.01 (3H, s, Ac), 2.04 (3H, s, Ac), 4.06 (1H, d, J = 10.7 Hz, 6'-H), 4.18 (1H, dd, J = 5.6, 12.5 Hz, 6'-H), 4.40 (1H, m, 5'-H), 5.02 (1H, t, J = 9.7 Hz, 4'-H), 5.11 (1H, t, J = 9.8 Hz, 2'-H), 5.72 (1H, t, J = 9.3 Hz, 3'-H), 6.27 (1H, d, J = 10.5 Hz, 1'-H), 7.19–8.78 (11H, m, =CH, aryl), 12.16 (1H, d, J = 10.5 Hz, NH); ¹³C NMR (DMSO-d₆) δ 20.00, 20.23, 20.35 (4 Ac), 61.66 (C-6'), 67.96 (C-2'), 69.09 (C-3'), 72.81 (C-4'), 75.34 (C-5'), 80.15 (C-1'), 110.82, 112.21, 119.00, 119.84, 120.92, 122.52, 126.90, 127.43, 129.04, 129.47, 132.42, 132.52, 133.96, 136.32 (=CH, C-5, aryl), 154.65 (C-2), 167.02 (C-4), 169.26, 169.52, 169.85 (4 C=O).

3-[4-(Ethoxycarbonyl)phenyl]-5-(2-thienylmethylene)-2-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoin (3g): yield 0.4 g (72%); mp 187 °C; MS (EI) m/z 688 (M⁺), 358 (base), 331 (glycon); IR (KBr) 1757, 1725 (C=O); ¹H NMR (DMSO- d_6) δ 1.34 (3H, t, J = 7.1 Hz, CH₃), 1.82, 1.96, 2.00, 2.03 (12H, 4 s, 4 Ac), 4.10 (1H, d, J = 10.9 Hz, 6'-H), 4.16-4.29 (2H, m, 5'-H, 6'-H), 4.37 (2H, q, J = 7.1 Hz, CH₂), 5.14 (1H, t, J = 9.5Hz, 4'-H), 5.36 (1H, t, J = 9.7 Hz, 2'-H), 5.55 (1H, t, J = 9.3 Hz, 3'-H), 5.90 (1H, d, J = 10.2 Hz, 1'-H), 7.21-8.13 (8H, m, =CH, aryl); ¹³C NMR (DMSO- d_6) δ 14.00 (CH₃), 20.17, 20.28 (4 Ac), 61.00 (CH₂), 61.48 (C-6'), 67.68 (C-2'), 68.58 (C-3'), 72.83 (C-4'), 75.55 (C-5'), 80.67 (C-1'), 119.56 (=CH), 127.55, 127.76, 130.27, 130.37, 134.62, 134.89, 135.23, 135.94, 137.35 (C-5, aryl), 157.76 (C-2), 164.79 (COOEt), 166.35 (C-4), 169.22, 169.42, 169.81 (4 C=O).

3-(3-Methylphenyl)-5-(2-thienylmethylene)-2-(2,3,4,6-tetra-O-acetyl-\beta-D-glucopyranosyl)-2-thiohydantoin (3h): yield 1.0 g (95%); mp 177 °C; MS (EI) m/2 630 (M⁺), 300 (base), 331 (glycon); IR (KBr) 1757 (C=O); ¹H NMR (DMSO-d_6) \delta 1.82, 1.96, 2.00, 2.03 (12H, 4s, 4Ac), 2.36 (3H, s, CH₃), 4.10 (1H, d, J = 10.2 Hz, 6'-H), 4.26 (2H, m, 5'-H, 6'-H), 5.14 (1H, t, J = 9.6 Hz, 4'-H), 5.37 (1H, t, J = 9.7 Hz, 2'-H), 5.54 (1H, t, J = 9.3 Hz, 3'-H), 5.86 (1H, d, J = 10.3 Hz, 1'-H), 7.14-7.94 (8H, m, =CH, aryl); ¹³C NMR (DMSO-d_6) \delta 20.16, 20.27, 20.61 (4 Ac + CH₃), 61.47 (C-6'), 67.67 (C-2'), 68.59 (C-3'), 72.84 (C-4'), 75.54 (C-5'), 80.55 (C-1'), 119.04 (=CH), 124.50, 127.68, 127.86, 129.26, 129.99, 131.76, 134.61, 134.89, 134.95, 137.42, 139.16 (C-5, aryl), 158.86 (C-2), 166.70 (C-4), 169.14, 169.21, 169.40, 169.79 (4 C=O).

5-Benzylidene-3-phenyl-2-(3,5-di-O-4-toluoyl- β -D-erythro-pentofuranosyl)-2-thiohydantoin (5a). Typical Procedure. 5-Benzylidene-3-phenyl-2-thiohydantoin (1a, 2.80 g, 10 mmol) was suspended in anhydrous MeCN (50 mL) at room temperature. To this suspension was added NaH (50%, 0.53 g,

11 mmol), and the mixture was stirred at room temperature for 30 min. The mixture became clear after 15 min. 2-Deoxy-3,5di-O-4-toluoyl- α -D-erythro-pentofuranosyl chloride (4, 4.27 g, 11 mmol) was added, and the mixture was stirred at room temperature for 4 h until the starting material was consumed (TLC). Evaporation of the filtrate gave a dry residue which was purified by silica gel column chromatography with Et₂O/petroleum ether (1:1, v/v) to give 4.5 g (71%) of 5a: mp 163-165 °C; MS (FAB, MeOH + 3-nitrobenzyl alcohol) m/z 633 (M + H⁺), 281 (BH⁺), 353 (glycon + H⁺); IR (KBr) 1744, 1719 (C=O); ¹H NMR (CDCl₃) δ 2.38 (s, 3H, CH₃), 2.42 (s, 3H, CH₃), 2.77 (m, 1H, 2'-H), 2.93 (m, 1H, 2'-H), 4.56 (m, 2H, 5'-H), 4.64 (m, 1H, 4'-H), 5.65 (m, 1H, 3'-H), 6.70 (t, J = 5.5 Hz, 1H, 1'-H), 7.08 (s, 1H, ==CH), 7.12-8.49 (m, 18H, aryl); ¹⁸C NMR (CDCl₃) δ 21.48 (CH₃), 21.55 (CH₃), 39.56 (C-2'), 63.74 (C-5'), 74.95 (C-3'), 83.44 (C-4'), 83.79 (C-1'), 125. 32 (=CH), 126.39, 126.54, 126.96, 127.97, 128.45, 128.93, 129.09, 129.33, 129.58, 129.83, 131.86, 132.02, 134.12, 137.60, 143.68, 144.24 (C-5, aryl), 162.37 (C-2), 165.75 (C=O), 165.96 (C=O), 168.44 (C-4). Anal. $(C_{37}H_{32}N_2O_6S \cdot 0.5H_2O)$ C, H, N.

3-Phenyl-5-(2-thienylmethylene)-2-(3,5-di-*O***-4-toluoyl**-β-D-erythro-pentofuranosyl)-2-thiohydantoin (5b): yield 1.4 g (88%); mp 168–170 °C; MS (FAB, DMSO + 1% CH₃COOH + 3-nitrobenzyl alcohol) m/z 639 (M + H⁺), 353 (glycon + H⁺), 287 (BH⁺); IR (KBr) 1719 (C=O); ¹H NMR (CDCl₃) δ 2.38 (s, 3H, CH₃), 2.43 (s, 3H, CH₃), 2.78 (td, J = 7.0, 14.1 Hz, 1H, 2'-H), 3.11 (ddd, J = 3.1, 6.1, 14.4 Hz, 1H, 2-H), 4.51 (d, J = 4.3 Hz, 2H, 5'-H), 4.63 (m, 1H, 4'-H), 5.67 (m, 1H, 3'-H), 6.66 (t, J = 6.6 Hz, 1H, 1'-H), 7.12–8.00 (m, 17H, =CH, aryl); ¹³C NMR (CDCl₃) δ 21.48 (CH₃), 21.55 (CH₃), 40.04 (C-2'), 63.78 (C-5'), 75.11 (C-3'), 83.41 (C-4'), 84.40 (C-1'), 118.86 (=CH), 126.60, 126.65, 126.83, 127.19, 128.87, 128.94, 129.09, 129.33, 129.61, 129.67, 132.14, 132.91, 133.62, 135.58, 138.02, 143.66, 144.19 (C-5, aryl), 160.96 (C-2), 165.66 (C=O), 165.95 (C=O). Anal. (C₃₅H₃₀N₂O₆S₂·

5-(3-Indolylmethylene)-2-thiohydantoin (7c): yield 1.97 g (81%); mp 312 °C; MS (EI) m/z 243 (M⁺); ¹H NMR (DMSO-d₆) δ 6.91 (=CH), 7.15–8.93 (5H, m, aryl), 12.05 (3H, s, 3 NH); ¹³C NMR (DMSO-d₆) δ 105.52, 108.32, 112.10, 118.04, 120.69, 122.63, 123.30, 127.13, 128.90, 135.92 (=CH, C-5, aryl), 165.50 (C-4), 176.76 (C-2).

5-Benzylidene-2-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (8a) and 5-Benzylidene-3-(2,3,4,6-tetra- $O\text{-}acetyl-\beta\text{-}D\text{-}glucopyranosyl)\text{-}2\text{-}(2,3,4,6\text{-}tetra\text{-}O\text{-}acetyl-\beta\text{-}D\text{-}detyl-\beta\text{-}D\text{-}detyl-\beta\text{-}D\text{-}detyl-\beta\text{-}D\text{-}detyl-\beta\text{-}D\text{-}detyl-\beta\text{-}D\text{-}detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-}detyl-\beta\text{-}Detyl-}detyl-detyl-}detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-}detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-detyl-detyl-detyl-}detyl-detyl-detyl-detyl-detyl-detyl-detyl-detyl-detyl-detyl-detyl-}detyl-detyl$ glucopyranosyl)-2-thiohydantoin (9a). 5-Benzylidene-2thiohydantoin (7a, 1.02 g, 5 mmol) was suspended in aqueous NaOH (4.4%, 5 mL) at room temperature. To this suspension was added acetone (25 mL), and the mixture became clear after stirring at room temperature for 5 min. 2,3,4,6-Tetra-O-acetyl- α -D-glucopyranosyl bromide (2, 2.26 g, 5.5 mmol) was added, and the mixture was stirred overnight at room temperature. After filtration and evaporation of the filtrate under reduced pressure, the residue was chromatographed on a silica gel column with ether/petroleum ether (6:4, v/v) to obtain 1.07 g (40%) of 8a (mp 171-172 °C) and 0.86 g (20%) 9a (mp 111-112 °C). 8a: MS (EI) m/z 534 (M⁺, 331 (glycon + 1), 204 (base); IR (KBr) 3436 (NH), 1757 (C=O); ¹H NMR (CDCl₃) δ 1.93 (s, 3H, Ac), 2.04 (s, 3H, 2Ac), 2.08 (s, 6H, 2Ac), 4.00 (m, 1H, 5'-H), 4.22 (d, J = 3.4 Hz, 2H, 6'-H), 5.15 (t, J = 9.6 Hz, 1H, 4'-H), 5.27 (t, J = 9.7 Hz, 1H, 3'-H), 5.41 (t, J = 9.2 Hz, 1H, 2'-H), 5.68 (d, J = 10.3 Hz, 1H, 1'-H), 7.03 (1H, s, =-CH), 7.42-8.14 (m, 5H, aryl), 9.98 (s, 1H, NH); ¹³C NMR (CDCl₃) δ 20.37, 20.42 (4Ac), 61.83 (C-6'), 67.97 (C-2'), 69.15 (C-3'), 73.43 (C-4'), 76.60 (C-5'), 81.04 (C-1'), 126.25 -CH), 128.60, 130.33, 132.03, 133.74, 138.10 (C-5, aryl), 158.56 (C-2), 169.28 (C-4), 169.41, 169.99, 170.58, 171.14 (4 C=O). Anal. $(C_{24}H_{28}N_2O_{10}S \cdot 0.25H_2O)$ C, H, N. 9a: MS (FAB, MeOH + 3-nitrobenzyl alcohol) m/z 865 (M + H⁺); IR (KBr) 1757 (C=O); ¹H NMR (CDCl₈) δ 1.85, 1.91, 2.02, 2.04, 2.05, 2.07, 2.12 (7s, 24H, 8Ac), 3.86-3.91 (m, 2H, 5'-H, 5"-H), 4.11-4.31 (m, 4H, 6'-H, 6"-H), 5.15-5.25 (m, 2H, 4'-H, 4"-H), 5.30-5.42 (m, 4H, 3'-H, 3"-H, 2''-H, 1''-H), 5.64 (t, J = 9.3 Hz, 1H, 2'-H), 5.84 (d, J = 10.1 Hz, 1H, 1'-H), 7.04 (s, 1H, =CH), 7.43-8.14 (m, 5H, aryl); ¹³C NMR (CDCl₃) & 19.97, 20.20, 20.30, 20.35, 20.41 (8Ac), 61.31 (C-6'), 61.73 (C-6"), 67.40 (C-2"), 68.09 (C-2"), 69.01 (C-3', C-3"), 72.78 (C-4'), 73.83 (C-4''), 75.97 (C-5'), 76.40 (C-5''), 79.60 (C-1'), 81.56 (C-1"), 126.76 (=CH), 128.62, 130.48, 131.99, 133.62, 136.47 (C-

5, aryl), 158.30 (C-2), 167.80 (C-4), 168.61, 169.08, 169.11, 169.20, 169.80, 169.97, 170.42, 170.49 (8 C=O). Anal. (C₃₈H₄₃N₂O₁₉S) C, H, N.

 $5-(2-Thienylmethylene)-2-(2,3,4,6-tetra-O-acetyl-\beta-D-glu$ copyranosyl)-2-thiohydantoin (8b) and 5-(2-Thienylmethylene)-3-(2,3,4,6-tetra-O-acetyl-D-glucopyranosyl)-2-(2,3,4,6tetra-O-acetyl- β -D-glucopyranosyl)-2-thiohydantoin (9b). The reaction of 7b with 2 under the same conditions as for preparation of 8a: yield 1.55 g (57%) of 8b, mp 180-182 °C, and 0.80 g (18%) of 9b, mp 120-122 °C. 8b: MS (FAB, DMSO + 1% $CH_{3}COOH + 3$ -nitrobenzyl alcohol) m/z 541 (M + H⁺), 331 $(glycon + 1); {}^{1}H NMR (DMSO) - d_{8} \delta 1.81 (3H, s, Ac), 1.98 (s, 3H, s)$ Ac), 2.03 (s, 6H, 2 Ac), 4.07 (m, 1H, 6'-H), 4.18 (m, 2H, 5'-H, 6'-H), 5.13 (t, J = 9.5 Hz, 1H, 4'-H), 5.34 (t, J = 10.2 Hz, 1H, 3'-H), 5.52 (t, J = 9.4 Hz, 1H, 2'-H), 5.85 (d, J = 10.2 Hz, 1H, 1'-H), 7.17 (dd, J = 3.9, 4.9 Hz, 1H, 4"-H), 7.21 (s, 1H, =CH), 7.69 (d, J = 3.5 Hz, 1H, 3"-H), 7.85 (d, J = 5.0 Hz, 1H, 5"-H), 11.78 (1H, s, NH); 18C NMR (DMSO-d₆) & 20.10, 20.17, 20.21, 20.27 (4 Ac), 61.47 (C-6'), 67.75 (C-2'), 68.76 (C-3'), 72.87 (C-4'), 75.49 (C-5'), 80.16 (C-1'), 117.04 (=CH), 127.51, 133.81, 134.13, 136.34, 137.58 (C-5 and aryl), 158.59 (C-2), 169.16, 169.20, 169.41, 169.78 (C-4 and 4 C=O). 9b: MS (EI) m/z 870 (M⁺); IR (KBr) 1763, 1742 (2 C=O); ¹H NMR (CDCl₃) δ 1.89, 1.91, 2.02, 2.03, 2.04, 2.05, 2.07, 2.11 (s, 24H, 8 Ac), 3.86-3.91 (m, 2H, 5'-H, 5"-H), 4.11-4.28 (m, 4H, $2 \times 6'$ -H, $2 \times 6''$ -H), 5.20-5.42 (m, 4H, 3'-H, 3''-H, 2''-H, 1''-H), 5.64 (t, J = 9.3 Hz, 1H, H-2'), 5.84 (d, J = 10.1Hz, 1H, H-1'), 7.11-7.73 (m, 4H, =CH, aryl); ¹³C NMR (CDCl₈) δ 20.09, 20.31, 20.40, 20.47 (8 × Ac), 61.31 (C-6'), 61.67 (C-6''), 67.45 (C-2'), 68.09 (C-2''), 68.95, 69.01 (C-3', C-3"), 72.92 (C-4'), 74.05 (C-4"), 75.01 (C-5'), 76.91 (C-5"), 79.64 (C-1'), 81.78 (C-1"), 120.30 (=CH), 127.43, 134.23, 134.39, 137.45 (aryl, C-5), 156.85 (C-2), 166.99 (C-4), 168.61, 169.11, 169.20, 169.26, 169.86, 170.10, 170.55 (8 C=O).

5-(3-Indolylmethylene)-2-(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)-2-thiohydantoin (8c). The reaction of 7c with 2 under the same conditions as for preparation of 8a gave 3.95 g (69%) of 8c: mp 150 °C; MS (EI) m/z 573 (M⁺), 243 (base), 331 (glycon); ¹H NMR (DMSO-d₆) δ 1.74 (3H, s, Ac), 1.99 (3H, s, Ac), 2.05 (6H, s, 2Ac), 4.06 (1H, d, J = 10.7 Hz, 6'-H), 4.14-4.33 (2H, m, 5'-H and 6'-H), 5.05 (1H, t, J = 9.7 Hz, 4'-H), 5.14 (1H, t, J = 9.8 Hz, 3'-H), 5.67 (1H, t, J = 9.3 Hz, 2'-H), 6.18 (1H, d, J = 10.4 Hz, 1'-H), 7.16-8.61 (6H, m, Aryl), 11.56 (1H, s, NMR (DMSO-d₆) δ 20.03, 20.25, 20.27, 20.35 (4 Ac), 61.66 (C-6'), 68.02 (C-2'), 69.15 (C-3'), 72.77 (C-4'), 75.27 (C-5'), 79.80 (C-1'), 110.69 (=CH), 112.10, 117.56, 118.94, 120.64, 122.32, 126.80, 133.11, 134.16, 136.25 (C-5, aryl), 154.59 (C-2), 169.25 (C-4), 169.32, 169.49, 169.53, 169.83 (4 C=O).

(Z)-5-Benzylidene-3-D-glucopyranosylhydantoin (10a). The protected nucleoside 9a (0.5 g, 0.58 mmol) was stirred in saturated NH₃/MeOH (25 mL) at room temperature for 1 day. The solvent was removed in vacuo, and the residue was chromatographed on silica gel with a gradient of 5-10% MeOH in CHCl₃ to afford 100 mg (49%) of 10a as a white foam. 10: MS (EI) m/z 350 (M⁺), 188 (base), 162 (glycon); IR (KBr) 3408 (OH, NH), 1770 (C=O); ¹H NMR (DMSO-d₆) § 3.12 (m, 1H, 4'-H), 3.26 (m, 1H, 3'-H), 3.43 (m, 1H, 5'-H, 6'-H), 3.71 (dd, 3.72 (dd, J = 4.0, 11.4 Hz, 1H, 6'-H), 4.20 (m, 1H, 2'-H), 4.52 (m, 1H, 2'-H)6'-OH), 4.85 (d, J = 9.0 Hz, 1H, 1'-H), 5.00 (1H, d, J = 5.0 Hz, 4'-OH), 5.06 (d, J = 3.7 Hz, 1H, 3'-OH), 5.28 (d, J = 4.8 Hz, 1H, 2'-OH), 6.55 (s, 1H, =CH), 7.35-7.66 (m, 5H, aryl), 10.91 (s, 1H, NH); ¹³C NMR (CD₃OD-d₆) δ 62.71 (C-6'), 69.79 (C-4'), 71.20 (C-2'), 79.04 (C-5'), 80.92 (C-3'), 82.20 (C-1'), 112.57 (=CH), 127.04 (C-5), 130.03, 130.33, 134.24 (aryl), 156.50 (C-2), 165.85 (C-4).

3-D-Glucopyranosyl-5-(2-thienylmethylene)hydantoin (10b). The protected nucleoside 9b (0.5 g, 0.92 mmol) was treated similarly as described for the preparation of the deprotected nucleosides 10a. Yield 95 mg (29%) of 10b as a yellow solid: mp 215 °C; MS (EI) m/z 356 (M⁺), 194 (base), 162 (glycon); ¹H NMR (DMSO- d_{e}) δ 3.09 (m, 1H, 4'-H), 3.20 (m, 1H, 3'-H), 3.38 (m, 2H, 5'-H, 6'-H), 3.71 (dd, J = 5.8, 11.0 Hz, 1H, 6'-H), 4.20 (m, 1H, 2'-H), 4.58 (t, J = 5.7 Hz, 1H, 6'-OH), 4.83 (d, J = 9.4 Hz, 1H, 1'-H), 5.00 (d, J = 5.2 Hz, 1H, 4'OH), 5.06 (d, J = 4.7 Hz, 1H, J'-GH), 5.28 (d, J = 5.0 Hz, 2'-OH), 6.72 (s, 1H, ==CH), 7.19 (dd, J = 3.9, 4.9 Hz, 1H, 4''-H), 7.64 (d, J = 3.6 Hz, 1H, 3''-H), 7.74 (d, J = 4.8 Hz, 1H, 5''-H), 10.70 (s, 1H, NH); ¹³C NMR (DMSO-

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 d_{6}) δ 61.01 (C-6'), 67.82 (C-4'), 69.93 (C-2'), 77.37 (C-5'), 79.96 (C-3'), 80.72 (C-1'), 102.95 (=CH), 128.52 (C-5), 128.69, 129.02, 129.50, 135.60 (thienyl), 153.61 (C-2), 166.83 (C-4).

Virus and Cells. The HIV-1 strain HTLV-IIIB¹⁷ was propagated in H9 cells¹⁸ at 37 °C using RPMI 1640 with 10% heat-inactivated Fetal Calf Serum (FCS) and antibiotics (growth medium). Culture supernatant was filtered (0.45 nm), aliquotted, and stored at -80 °C until use. HSV-1 strain McIntyre (American Type Culture Collection, VR-539) and HSV-2 strain MS (American Type Culture Collection, VR-540) were propagated in the African green monkey kidney cell line Vero (American Type Culture Collection, CRL-1587) using Eagle medium with 10%FCS and antibiotics (growth medium). Culture supernatants were filtered (0.45 nm), aliquotted, and stored at -80 °C until use.

Drugs. Acyclovir was a gift from The Wellcome Research Laboratories, Beckenham, Kent, and (E)-5-(2-bromovinyl)-2'deoxyuridine (BVDU) was purchased from Sigma Chemical Co., St. Louis, MO. Stock solutions of these drugs were prepared in H₂O and stored at -80 °C until use.

Inhibition of HSV-1 and HSV-2 Replication. Nucleoside analogues were examined for possible antiviral activity against HSV-1 and HSV-2 using Vero cells as target cells. For screening studies Vero cells were incubated with virus for 2.5 h, washed, and thereafter added in a proportion of 1:50 to uninfected Vero cells, which had been preincubated in growth medium containing the test compound for 2.5 h. Cultures were maintained with the test compound for 5 days in parallel with virus-infected control cultures without compound added. Expression of HSV-1 and HSV-2 in the culture medium was quantiated by HSV-1 and HSV-2 antigen detection ELISA.¹⁹ Compounds mediating less than 30% reduction of antigen expression were considered without biological activity. Compounds mediating a reduction of 30% or more were examined for cytotoxic potential using concentration-dependent inhibition of Vero cell proliferation as measure of cytoxicity. A 30% inhibition of cell growth relative to control cultures was considered significant.

Inhibition of HIV-1 Replication. Nucleoside analogues were examined for possible antiviral activity against HIV-1 using MT-4 cells as target cells. For screening studies MT-4 cells were incubated with virus for 2 h, washed, and thereafter added in a proportion of 1:10 to uninfected cells, which had been preincubated in growth medium containing the test compound for 2 h. Cultures were maintained with the test compound for 7 days in parallel with virus-infected control cultures without compound added. Expression of HIV in the culture medium was quantiated by HIV antigen detection ELISA.²⁰ Compounds mediating less than 30% reduction of antigen expression were considered without biological activity. Compounds mediating a reduction of 30% or more were examined for cytotoxic potential using concentration-dependent inhibition of MT-4 cell proliferation as measure of cytoxicity. A 30% inhibition of cell growth relative to control cultures was considered significant.

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