An Efficient Synthesis of (-)-Deacetylanisomycin Starting from D-Tyrosine¹

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Abstract: The antibiotic (–)-deacetylanisomycin was synthesized starting from D-tyrosine using Sharpless asymmetric dihydroxylation as a key reaction.

Key words: anisomycin, (–)-deacetylanisomycin, D-tyrosine, Sharpless asymmetric dihydroxylation

Anisomycin (1) is an antibiotic that has been isolated from fermentation broth filtrates of various species of *Streptomyces*.² X-ray crystallographic analysis³ and chemical studies⁴ reveal the structure and relative stereochemistry of anisomycin to be that depicted in structure 1 (Figure 1). The absolute stereochemistry was established as 2R,3S,4Sby chemical correlation studies.⁵ Anisomycin possesses strong and selective activity against pathogenic protozoa and fungi and has been proved successful clinically in the treatment of amoebic dysentry and trichomonas vaginitis.⁶ It has been shown to block ribosomal peptide synthesis.⁶

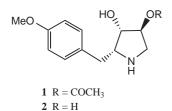
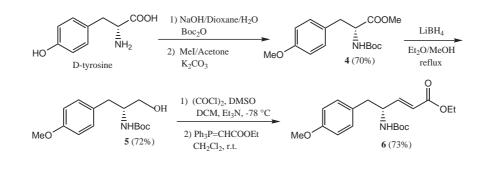


Figure 1 The structure of Anisomycin (1)

Although a few total synthesis of (–)-anisomycin have been reported⁷ previously, the molecule still continues to attract the attention of synthetic organic chemists owing to its high biological profile. Our continued interest in the development of new and efficient synthetic routes to clinically significant amino compounds prompted us to investigate the total synthesis of (–)-deacetylanisomycin (2).⁸ We wish to report herein the synthesis of 2 from D-tyrosine, which also constitutes a formal total synthesis of 1. It appeared to us that an expedient approach to 2 from Dtyrosine would involve a reaction sequence outlined as below (Scheme 1).

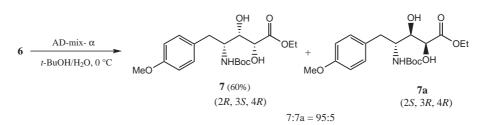
Compound **4** was obtained in 70% yield from D-tyrosine by protecting the amino functionality using di-*tert*-butyl dicarbonate, followed by methylation of the acid and phenolic hydroxyl group with MeI/K₂CO₃. Reduction of ester **4** with lithium borohydride⁹ furnished the alcohol **5** (72%). A Swern oxidation¹⁰ followed by Wittig reaction with (ethoxycarbonylmethylene)triphenylphosphorane in CH₂Cl₂ yielded the *trans* α , β -unsaturated ester **6** (73%). In order to avoid or minimize any possible racemization at the chiral centre, the aldehyde was used immediately without purification.

Our strategy required that the olefination product **6** contained the desired *trans* geometry at the double bond for the subsequent creation of chiral diol via Sharpless asymmetric dihydroxylation. Upon reacting the *N*-Boc-protected γ -amino- α , β -unsaturated ester **6** with the modified Sharpless asymmetric dihydroxylation conditions using AD-mix- α ,¹¹ the expected (2*R*,3*S*,4*R*) configured ester **7** (60%) was obtained with high stereoselectivity (Scheme 2). This result has precedence.^{12,13}



Scheme 1

Synthesis 2002, No. 13, Print: 20 09 2002. Art Id.1437-210X,E;2002,0,13,1867,1870,ftx,en;Z06502SS.pdf. © Georg Thieme Verlag Stuttgart · New York ISSN 0039-7881



Scheme 2

Then diol **7** was subjected to acetonation with 2,2dimethoxypropane (2,2-DMP) in the presence of a catalytic amount of camphorsulfonic acid (CSA) in anhydrous CH_2Cl_2 to afford the acetonide **8** (72%). Then **8** was reduced to the corresponding alcohol **9** (68%) with LiBH₄ (Scheme 3).

The alcohol **9** was converted to its corresponding bromide **10** (65%) via the tosylate (TsCl/Py) using LiBr/DMF. The above obtained Boc-amine **10** was subjected to acidolysis using TFA and the resulting amine was in situ treated with triethylamine, thereby facilitating cyclisation to the desired (–)-deacetylanisomycin **2** (70%). The sample had identical spectral data, to that previously reported in the literature¹⁴ (Scheme 3).

The present work offers an alternative route to the antibiotic in enantiomerically pure form. It is to be noted that a completely different approach to the synthesis of (–)deacetylanisomycin has been achieved starting from the chiral pool D-tyrosine. Further work on the synthesis of anisomycin analogues utilizing the above method is in progress and will be reported in due course.

Crude products were purified by column chromatography on silica gel (60–120 mesh). ¹H NMR were obtained in CDCl₃ at 200 MHz. Chemical shifts are given in ppm, with respect to internal TMS; *J* values are quoted in Hz. DMSO, Et₃N and CH₂Cl₂ were distilled from CaH₂ and stored over molecular sieves. Benzene and Et₂O

were dried over sodium and benzophenone. All reactions were carried out under N_2 using dry glassware.

Methyl (2*R*)-2-(*tert*-Butoxycarbonyl)amino-3-(4-methoxy-phenyl)propanoate (4)

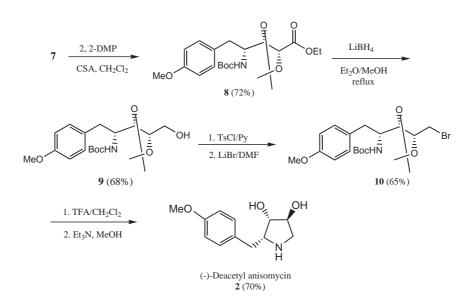
A solution of D-tyrosine (3 g, 16.5 mmol) in a mixture of dioxane (30 mL), H₂O (20 mL) and aq 1 M NaOH (20 mL) was stirred and cooled in an ice-water bath. Di-tert-butyl dicarbonate (3.97 g, 18.2 mmol) was added and stirring was continued at r.t. for 30 min. The solution was concentrated in vacuo to about 10-15 mL, cooled in an ice-water bath, covered with a layer of EtOAc (30 mL) and acidified with dilute aq KHSO₄ solution to pH 2–3. The aqueous phase was extracted with EtOAc (2 \times 50 mL). The EtOAc extracts were pooled, washed with H₂O (2 × 30 mL), dried (Na₂SO₄) and evaporated in vacuo. To a solution of this crude N-Boc protected amino acid (3 g, 10.6 mmol) in acetone (100 mL) were added anhyd K₂CO₃ (7.35 g, 53.3 mmol) and MeI (4.55 g, 32 mmol). The resulting mixture was refluxed for 8 h, cooled to r.t. and filtered. The filtrate was concentrated and purified by column chromatography (hexane-EtOAc, 7:3) to give 4 (2.3 g, 70%) as a colorless viscous oil; $[\alpha]_D^{25}$ –58.4 (*c* = 1, CHCl₃).

¹H NMR (CDCl₃): $\delta = 1.45$ (s, 9 H), 3.05 (d, J = 4.65 Hz, 2 H), 3.7 (s, 3 H), 3.8 (s, 2 H), 4.05 (m, 1 H), 4.9 (br d, 1 H, NH), 6.8 (d, J = 8.3 Hz, 2 H), 7.05 (d, J = 8.3 Hz, 2 H).

EIMS: $m/z = 192 (M^+ - 117), 121, 57.$

(2R)-2-(*tert*-Butoxycarbonyl)amino-3-(4-methoxyphenyl)propan-1-ol (5)

To a stirred solution of ester 4 (2 g, 6.4 mmol) in anhyd Et_2O (100 mL) were added LiBH₄ (0.57 g, 25.5 mmol) and MeOH (10 mL) at



Scheme 3

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0 °C. The reaction mixture was then refluxed for 5 h and quenched by adding an aq sat. NH₄Cl solution. The aqueous phase was extracted with EtOAc (2 × 50 mL) and the combined organic layers were washed with brine (2 × 25 mL), and dried (Na₂SO₄). The volatiles were removed under reduced pressure and the residue purified by column chromatography (hexane–EtOAc, 6.5:3.5) to afford **5** (1.3 g, 72%) as a syrupy liquid; $[a]_D^{25}$ +12.58 (c = 1, CHCl₃).

¹H NMR (CDCl₃): δ = 1.45 (s, 9 H), 2.48 (m, 1 H), 2.78 (d, *J* = 4.55 Hz, 2 H), 3.55 (m, 2 H), 3.8 (s, 3 H), 4.75 (br d, 1 H, NH), 6.8 (d, *J* = 6.8 Hz, 2 H), 7.1 (d, *J* = 6.8 Hz, 2 H).

HRMS: m/z calcd for $C_{15}H_{23}NO_4$: 281.1627 (M⁺), observed: 281.1628 (M⁺).

Ethyl (2*E*,4*R*)-4-(*tert*-Butoxycarbonyl)amino-5-(4-methoxy-phenyl)pent-2-enoate (6)

To a stirred solution of oxalyl chloride (1.2 g, 9.6 mmol) in CH₂Cl₂ (15 mL) at -78 °C under N₂ was added DMSO (0.75 mL, 10 mmol) dropwise. After stirring for 30 min, a solution of the amino alcohol **5** (1.5 g, 5.33 mmol) in CH₂Cl₂ (25 mL) was added over 15 min. The mixture was warmed to -45 °C and stirring was continued for 1 h at this temperature, then Et₃N (2.7 g, 26.6 mmol) was added. The reaction mixture was brought to 0 °C and maintained at this temperature for 15 min, then a solution of (ethoxycarbonylmethylene)triphenylphosphorane (2.2 g, 6.4 mmol) in benzene (15 mL) was added and the resulting solution was stirred for 15 h at r.t. The solvent was removed under vacuum; the residue was washed with H₂O, brine and dried (Na₂SO₄). The residue was purified by column chromatography (hexane–EtOAc, 8:2) to yield the amino α , β -unsaturated ester **6** (1.35 g, 73%) as a pale yellow viscous liquid; [α]_D²⁵ – 5.25 (c = 2, CHCl₃).

¹H NMR (CDCl₃): $\delta = 1.3$ (t, J = 5.1 Hz, 3 H), 1.4 (s, 9 H), 2.85 (d, J = 4.0 Hz, 2 H), 3.8 (s, 3 H), 4.2 (q, J = 5.1 Hz, 2 H), 4.45 (m, 1 H), 4.55 (br d, 1 H, NH), 5.82 (d, J = 13.5 Hz, 1 H), 6.85 (m, 3 H), 7.08 (d, J = 6.2 Hz, 2 H).

FABMS: $m/z = 372 (M^+ + 23)$.

Ethyl (2*R*,3*S*,4*R*)-4-(*tert*-Butoxycarbonyl)amino-2,3-dihydroxy-5-(4-methoxyphenyl)pentanoate (7)

A mixture of **6** (0.4 g, 1.15 mmol) and the Sharpless AD-mix- α (1.6 g) modified by additional (DHQ)₂PHAL (35.65 mg) and potassium osmate (3.45 mg) in *tert*-butyl alcohol (11.5 mL) and H₂O (11.5 mL) was stirred at r.t. for 18 h. The mixture was treated with Na₂SO₃ (2.3 g). After 0.5 h, EtOAc (100 mL) was added and the organic phase separated and washed twice with aq 1 M KHSO₄ (100 mL) and aq 5% NaHCO₃. The organic phase was filtered through silica gel and dried (MgSO₄). The residue was purified by column chromatography (hexane–EtOAc, 6:4) to yield **7** (0.26 g, 60%) as a semi-solid; $[\alpha]_D^{25}$ +9.12 (*c* = 1, CHCl₃).

¹H NMR (CDCl₃): δ = 1.3 (t, *J* = 3.1 Hz, 3 H), 1.4 (s, 9 H), 2.85 (m, 2 H), 3.35 (m, 2 H), 3.68 (m, 1 H), 3.75 (s, 3 H), 3.92 (m, 1 H), 4.2 (m, 3 H), 4.9 (m, 1 H, NH), 6.8 (d, *J* = 6.1 Hz, 2 H), 7.1 (d, *J* = 6.1 Hz, 2 H).

HRMS m/z calcd for C₁₉H₂₉NO₇: 383.1944 (M⁺), observed: 383.1946 (M⁺).

Ethyl (1*R*,4*R*,5*R*)-5-[1-(*tert*-Butoxycarbonyl)amino-2-(4-meth-oxyphenyl)ethyl]-2,2-dimethyl-1,3-dioxolane-4-carboxylate (8)

To a stirred solution of **7** (0.25 g, 2.4 mmol) in CH₂Cl₂ (10 mL) were added 2,2-dimethoxypropane (1.25 g, 12 mmol) and a catalytic amount of camphorsulfonic acid (15 mg) at 0 °C. The mixture was left stirring at r.t. for 2–3 h under N₂. The progress of the reaction was monitored by TLC. The solvent was removed under vacuo and the residue was purified by column chromatography (hexane–EtOAc, 8:2) to give the desired compound **8** (0.2 g, 72%) as a yellow viscous oil; $[\alpha]_D^{25}$ +14.22 (c = 1, CHCl₃).

¹H NMR (CDCl₃): δ = 1.12 (t, J = 4.75 Hz, 3 H), 1.30 (s, 6 H), 1.35 (s, 9 H), 2.8 (m, 3 H), 3.7 (s, 3 H), 4.1 (m, 4 H), 4.7 (br d, 1 H, NH), 6.75 (d, J = 7.1 Hz, 2 H), 7.1 (d, J = 7.1 Hz, 2 H).

FABMS: m/z = 424 (M+1).

(1*R*,4*S*,5*R*)-5-[1-(*tert*-Butoxycarbonyl)amino-2-(4-methoxyphenyl)ethyl]-2,2-dimethyl-1,3-dioxolan-4-ylmethanol (9)

To a stirred solution of the protected ester **8** (0.5 g, 1.18 mmol) in anhyd Et₂O (50 mL) were added LiBH₄ (0.1 g, 4.72 mmol) and MeOH (5 mL) at 0 °C. The reaction mixture was then refluxed for 5 h and quenched by the addition of aq sat. NH₄Cl solution. The aqueous phase was extracted with EtOAc (2 × 20 mL) and the combined organic layers were washed with brine (2 × 10 mL), dried (Na₂SO₄). The volatiles were removed under reduced pressure and the residue was purified by column chromatography (hexane– EtOAc, 7:3) to afford **9** (0.375 g, 68%) as a syrupy liquid; $[\alpha]_D^{25}$ +7.75 (*c* = 1.5, CHCl₃).

¹H NMR (CDCl₃): $\delta = 1.4$ (2 s, 9 H + 6 H), 2.65 (m, 1 H), 2.8 (d, J = 5.1 Hz, 2 H), 3.68 (m, 2 H), 3.78 (s, 3 H), 3.88 (m, 2 H), 4.86 (br d, 1 H, NH), 6.85 (d, J = 5.5 Hz, 2 H), 7.17 (d, J = 5.5 Hz, 2 H). FABMS: m/z = 382 (M + 1).

(1*R*,4*R*,5*R*)-1-[*N*-(*tert*-Butoxycarbonyl)]-1-(5-bromomethyl-2,2-dimethyl-1,3-dioxolan-4-yl)-2-(4-methoxyphenyl)ethan-1-amine (10)

To a solution of compound **9** (0.2 g, 5.3 mmol) in anhyd CH₂Cl₂ (15 mL) containing anhyd pyridine (0.062 g, 7.9 mmol) at 0 °C under N₂ was added a solution of tosyl chloride (0.12 g, 6.3 mmol) in CH₂Cl₂ (5 mL) and the reaction mixture was stirred for 10 h at r.t.. The mixture was washed with aq sat. CuSO₄ solution (20 mL) and extracted with CH₂Cl₂ (2 × 20 mL). The organic phase was dried (Na₂SO₄) and concentrated under reduced pressure. To the above obtained crude tosyl compound (0.2 g, 0.37 mmol) in anhyd DMF (15 mL) was added LiBr (0.1 g, 1.12 mmol) and the reaction mixture was stirred for 10 h at 80 °C. The mixture was extracted with Et₂O (2 × 25 mL) and dried (Na₂SO₄) and concentrated. The crude product was purified by column chromatography (hexane–EtOAc, 8.5:1.5) to afford the bromo compound **10** (0.107 g, 65%) as a yellow liquid; $[\alpha]_D^{25}$ +6.65 (c = 0.5, CHCl₃).

¹H NMR (CDCl₃): δ = 1.35 (2 s, 9 H + 6 H), 2.85 (m, 3 H), 3.38 (d, *J* = 2.9 Hz, 2 H), 3.76 (s, 3 H), 3.95 (m, 2 H), 4.8 (br d, 1 H, NH), 6.8 (d, *J* = 7.2 Hz, 2 H), 7.15 (d, *J* = 7.2 Hz, 2 H).

FABMS: m/z = 467 (M + 23).

(-)-Deacetylanisomycin (2)

To a solution of compound **10** (0.23 g, 0.6 mmol) in CH₂Cl₂ (5 mL) were added trifluoroacetic acid (1 mL) and H₂O (0.5 mL) and the reaction mixture was stirred at r.t. for 10 h. After concentration of the solvent under reduced pressure, benzene (5 mL) was added to the residue and the solvents were removed under vacuo. The crude residue was dissolved in MeOH (5 mL) and cooled to 0 °C. Et₃N (0.15 g, 1.5 mmol) was added slowly to the mixture and it was stirred at r.t. for 5 h. Removal of solvent under reduced pressure, followed by purification of the crude residue by column chromatography (CH₂Cl₂–MeOH, 9:1) afforded compound **2** (0.07 g, 70%) as a solid; $[\alpha]_D^{25}$ –22.5 (*c* = 1, MeOH).¹³

¹H NMR (DMSO-*d*₆): δ = 2.36 (dd, 1 H, *J* = 2.1, 11.8 Hz, H-5), 2.52 (dd, 1 H, *J* = 6.5, 13.2 Hz, *CH*₂Ar), 2.71 (dd, 1 H, *J* = 7.4, 13.2 Hz, *CH*₂Ar), 2.96 (ddd, 1 H, *J* = 3.5, 6.8, 7.6 Hz, H-2), 3.15 (dd, 1 H, *J* = 5.5, 11.8 Hz, H-5), 3.45–3.47 (m, 1 H, H-3), 3.70 (s, 3 H, OCH₃), 3.82–3.85 (m, 1 H, H-4), 4.56 (br s, 1 H, OH), 4.61 (br s, 1 H, OH), 6.81 (d, 2 H_{arom}, *J* = 8.3 Hz), 7.16 (d, 2 H_{arom}, *J* = 8.3 Hz). HRMS: *m*/*z* calcd for C₁₂H₁₇NO₃: 224.1286 (M + 1), observed: 224.1286 (M + 1).

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