TOTAL SYNTHESIS OF DIHYDROTELEOCIDIN B-4 (DIHYDROTELEOCIDIN B)

Hideaki Muratake, Kazuaki Okabe, and Mitsutaka Natsume\* Research Foundation Itsuu Laboratory 2-28-10 Tamagawa, Setagaya-ku, Tokyo 158, Japan

Abstract: A potent tumor promoter, dihydroteleocidin B-4 (=dihydroteleocidin B) (1) was synthesized from L-valine derivative 8 using an acid-catalyzed cyclization of 15 to 16a to construct the teleocidin B structure unit.

Dihydroteleocidin B was first described in the literature as the sole crystalline derivative of "teleocidin B," a highly toxic and skin-irritating substance obtained from the mycelia of Streptomyces mediocidicus. 1) The structure was studied extensively by chemical means, 1, 2, 3 and finally the Xray crystallographic analysis of its monobromoacetate revealed it to be 1.4) Later, dihydroteleocidin B was found to have a potent tumor promoting activity,<sup>5)</sup> and so-called "teleocidin" from the above species was reinvestigated thoroughly. As a result, it was concluded that "teleocidin B" is actually composed of four stereoisomers, teleocidins B-1 (2), B-2 (3), B-3 (4), and B-4 (5),<sup>6)</sup> whose structures are now unambiguously established, including the  $\tilde{absolute}$  configuration.<sup>7,8)</sup> Thus dihydroteleocidin B-4 (dihydro-5) is the above dihydroteleocidin B (1), which was obtained by chance from the catalytic hydrogenation mixture of "teleocidin B" due to its nice crystalline nature. Here we report a total synthesis of 1 in the optically active form utilizing our efficient synthetic pathway for teleocidins A-1 (6) and A-2 (7). $^{9)}$  Total synthesis of  $(\pm)$ -teleocidin B-3 and B-4 has already been reported.<sup>10)</sup>

Our synthetic plan relied upon the Friedel-Crafts type of cyclization reaction  $(15 \rightarrow 16a)$  to construct the desired carbon framework, the 6,7,8,9-tetrahydrobenzo[g]indole ring system, carrying the requisite alkyl substituents, the methyl, ethyl, and isopropyl groups at the proper locations. For this purpose,



preparation of the 4-aminoindole derivative 15 with a 1,4,5-trimethyl-1-ethyl-4-hexenyl side chain was necessary and this was achieved in the following way.

L-Valine derivative  $8^{9}$  was treated with 3,6,7-trimethylocta-2,6-dienyl bromide<sup>11)</sup> and Mg in THF to afford  $9^{12}$  in 70% yield, which was successively dehydrated with a catalytic amount of p-TsOH in boiling benzene to give  $10^{12}$ in 90% yield. 10 was converted to  $11^{12}$  with Lawesson's reagent<sup>13)</sup> in refluxing THF in 74% yield, and the subsequent indole formation from 11 was readily attained with MeI in DMF at room temperature to provide 4-amino-7-alkylindole derivatives  $12^{12}$  and 13,<sup>12)</sup> mp 68-70°C, in 38% and 25% yields, respectively, accompanied by the formation of a by-product 14 in 30% yield. Stereochemistry of the side chain at the C-7 position of 12 and 13 was tentatively assigned by analogy with the corresponding intermediates for teleocidins A synthesis. Catalytic hydrogenation of the major isomer 12 on PtO<sub>2</sub> proceeded without difficulty in MeOH at room temperature to furnish  $15^{12}$  in 92% yield.

The acid-mediated cyclization reaction of 15 was examined by using a variety of Lewis and protonic acids to find a reaction condition to afford the highest yield of the isomer 16a, which has the correct stereochemical arrangement of the alkyl groups for the synthesis of 1. Treatment of 15 in CH<sub>2</sub>Cl<sub>2</sub> with *ca*. 30 equiv. of 95% H<sub>2</sub>SO<sub>4</sub> at 0°C for 4 h was found to be the best reaction condition, and 16a,<sup>12)</sup> mp 134-136°C,  $[\alpha]_D^{24}$  -102° (*c*=0.495, CH<sub>2</sub>Cl<sub>2</sub>) was obtained in 40% yield along with 16b,<sup>12)</sup> mp 195-196°C,  $[\alpha]_D^{24}$  -179° (*c*=0.500, CH<sub>2</sub>Cl<sub>2</sub>) in 10% yield. Formation of undesired by-products, 17a,<sup>12)</sup> mp 100-101°C, and 17b<sup>12</sup> was inevitable and these were produced in 15% and 8% yields, respectively. BF<sub>3</sub>.OEt<sub>2</sub> (*ca*. 40 equiv., 0°C+r.t., 7 h) and SnCl<sub>4</sub> (*ca*. 50 equiv., -20--10°C, 1.3 h) in CH<sub>2</sub>Cl<sub>2</sub> were less satisfactory in terms of the ratio of 16a and 16b.



a: 3,6,7-trimethylocta-2,6-dienyl bromide, Mg, THF, Ar, 0°C, 3 h. b: p-TsOH-H<sub>2</sub>O, PhH, Ar, reflux, 2 min. c: Lawesson's reagent, THF, Ar, reflux, 1 h. d: MeI, DMF, r.t., 4 h. e: H<sub>2</sub> (1 atm.), PtO<sub>2</sub>, MeOH, r.t., 3 h.

f: 95%  $H_2SO_4$ ,  $CH_2Cl_2$ , 0°C, 4 h.

Other acids such as  $EtAlCl_2$ ,  $CF_3SO_3H$  and p-TsOH gave only disappointing results.

With the important intermediate 16a in hand, the indolactam V part of 1 was constructed according to our standard procedure developed in the synthesis of teleocidins A.9) The functionalized three-carbon unit was introduced into the C-3 position of 16a using one equiv. of ethyl 3-bromo-2-hydroxyiminopropanoate<sup>14)</sup> in the presence of Na<sub>2</sub>CO<sub>3</sub> (2 equiv.) with the recovery of 16a in 11% yield. The desired compound 18 was obtained in 57% yield together with two 1:2 adducts 19 and 20 in 9% and 2.5% yields, respectively. With the increased ratio of the reagent to the substrate, the by-products were formed in much higher yields. Al amalgam reduction<sup>15)</sup> of the hydroxyimino function of 18 readily afforded a mixture of amino ester compounds. Fortunately these were separated by applying preparative silica gel TLC [hexane-EtOAc (1:4)] to give 21 (40% yield) and 22 (39.5% yield). This made it easy to purify the final compound from a supposed by-product originating from the partial racemization of the valinate residue. The ethyl ester group of 21 was selectively reduced with NaBH, in the presence of LiCl<sup>16)</sup> to yield 23, which was submitted to the next step without further purification, since the attempted purification by silica gel or alumina chromatography caused a great loss of the material. The crude 23 was hydrolyzed with 10% KOH in MeOH-H2O (4:1) under reflux and the nine-membered lactam ring was constructed with diphenylphosphoryl azide (DPPA), <sup>17)</sup> bearing in mind the series of pre-treatments described in the previous report,<sup>9)</sup> to afford dihydroteleocidin B-4 (1),<sup>12)</sup> mp 154-156°C and 232-235°C (decomp.),<sup>18)</sup> in 18% yield from 21 after recrystallization from



- a: Ethyl 3-bromo-2-hydroxyiminopropanoate, Na<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 16 h. b: Al-Hg, THF-H<sub>2</sub>O (9:1), r.t., 4.5 h. c: NaBH<sub>4</sub>, LiCl, EtOH-THF (4:3), r.t., 23 h.
- d: i) 10% KOH in MeOH-H<sub>2</sub>O (4:1), Ar, reflux, 48 h; ii) Et<sub>3</sub>N·HCl, 0°C, 2 min and r.t., 10 min; iii) evaporation in vacuo and dryness over P<sub>2</sub>O<sub>5</sub>; iv) DPPA, Et<sub>3</sub>N, DMF, r.t., 62 h.

6270

acetone-H<sub>2</sub>O. In the same manner, 9-epidihydroteleocidin B-4 (25),<sup>12)</sup> mp 259-261°C (decomp.) was obtained from 22 by way of 24 in 13% yield. The synthetic 1 was completely identified with the authentic specimen derived from natural teleocidin B-4 (5) in all respect (mixed mp determination, MS, IR, <sup>1</sup>H NMR, CD spectra, mobility of HPLC, and biological assay<sup>19</sup>).

Acknowledgment — The authors' heartiest thanks are due to Professor Shinichiro Sakai of Chiba University for his generous gift of dihydroteleocidin B-4 and for the measurement of CD spectrum. We are also indebted to Dr. Hirota Fujiki of National Cancer Center Research Institute for the assay of biological activity. REFERENCES AND NOTES

- 1) M. Takashima, H. Sakai, and K. Arima, Agric. Biol. Chem., 26, 660 (1962).
- 2) M. Takashima, H. Sakai, R. Mori, and K. Arima, Agric. Biol. Chem., 26, 669 (1962).
- H. Nakata, H. Harada, and Y. Hirata, Tetrahedron Lett., 1966, 2515; H. Harada, H. Nakata, and Y. Hirata, Nippon Kagaku Zasshi, 87, 86 (1966).
- 4) N. Sakabe, H. Harada, Y. Hirata, Y. Tomiie, and I. Nitta, *Tetrahedron Lett.*, 1966, 2523; H. Harada, N. Sakabe, Y. Hirata, Y. Tomiie, and I. Nitta, *Bull. Chem. Soc. Japan*, 39, 1773 (1966). The structural formula is erroneously printed in these literatures. Regarding this fact, *confer* the comment of footenote 10) in the reference 8).
- 5) H. Fujiki, M. Mori, M. Nakayasu, M. Terada, T. Sugimura, and R. E. Moore, Proc. Natl. Acad. Sci. U. S. A., 78, 3872 (1981).
- 6) H. Fujiki and T. Sugimura, *Cancer Surveys*, 2, 539 (1983); H. Fujiki, M. Suganuma, T. Tahira, A. Yoshioka, M. Nakayasu, Y. Endo, K. Shudo, S. Taka-yama, R. E. Moore, and T. Sugimura, "Cellular Interactions by Environmental Tumor Promoters," p. 37, Jpn. Sci. Soc. Press, Tokyo/VNU Science Press, Utrecht, 1984.
- 7) Y. Hitotsuyanagi, H. Fujiki, M. Suganuma, N. Aimi, S. Sakai, Y. Endo, K. Shudo, and T. Sugimura, Chem. Pharm. Bull., 32, 4233 (1984).
- 8) S. Sakai, N. Aimi, K. Yamaguchi, Y. Hitotsuyanagi, C. Watanabe, K. Yokose,
  Y. Koyama, K. Shudo, and A. Itai, Chem. Pharm. Bull., 32, 354 (1984).
- 9) H. Muratake and M. Natsume, Tetrahedron Lett., 28, 2265 (1987).
- 10) S. Nakatsuka, T. Masuda, and T. Goto, Tetrahedron Lett., 28, 3671 (1987).
- 11) G. E. Muntyan and R. N. Vaskan, Izv. Akad. Nauk Mold. SSR, Ser. Biol. Khim. Nauk, 1978, 65 [Chem. Abst., 89, 75405g (1978)]. We prepared this compound in the following alternative way: i) Wittig reaction of 5,6-dimethyl-5hepten-2-one with ethyl diethylphosphonoacetate; ii) reduction with LiAlH<sub>4</sub>, 56% overall yield; iii) bromination with CBr<sub>4</sub>-Ph<sub>3</sub>P, 51% yield.
- 12) The structure was verified by correct elementary analysis (C, H, N) or HRMS as well as reasonable PMR, IR and mass spectra.
- S. Scheibye, B. S. Pedersen, and S. O. Lawesson, Bull. Soc. Chim. Belg., 87, 229 (1978).
- 14) T. L. Gilchrist, D. A. Lingham, and T. G. Roberts, J. Chem. Soc., Chem. Commun., 1979, 1089.
- 15) D. J. Drinkwater and P. W. G. Smith, J. Chem. Soc. (C), 1971, 1305.
- 16) Y. Hamada and T. Shioiri, Chem. Pharm. Bull., 30, 1921 (1982).
- 17) T. Shioiri, K. Ninomiya, and S.-i. Yamada, J. Am. Chem. Soc., 94, 6203 (1972).
- 18) Solidified once at ca. 170°C. See reference 1) above.
- 19) Irritation on mouse ear and inhibition of specific <sup>3</sup>H-TPA binding.

(Received in Japan 21 April 1988)