

Bioscience, Biotechnology, and Biochemistry

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/tbbb20>

Syntheses and Antibacterial Activities of Gramicidin S Analogs Containing L-Ornithine in Place of L-Valine

Yasuhiro Soejima^a, Atsuko Hashiguchi^{ab} & Nobuo Izumiya^a

^a Laboratory of Biochemistry, Faculty of Engineering, Kyushu Sangyo University, Matsukadai, Higashi-ku, Fukuoka 813, Japan

^b Laboratory of Biochemistry, Faculty of Science, Kyushu University, Hakozaki, Higashi-ku, Fukuoka 812, Japan

Published online: 12 Jun 2014.

To cite this article: Yasuhiro Soejima, Atsuko Hashiguchi & Nobuo Izumiya (1994) Syntheses and Antibacterial Activities of Gramicidin S Analogs Containing L-Ornithine in Place of L-Valine, *Bioscience, Biotechnology, and Biochemistry*, 58:5, 826-829, DOI: [10.1271/bbb.58.826](https://doi.org/10.1271/bbb.58.826)

To link to this article: <http://dx.doi.org/10.1271/bbb.58.826>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

Syntheses and Antibacterial Activities of Gramicidin S Analogs Containing L-Ornithine in Place of L-Valine

Yasuhiro SOEJIMA, Atsuko HASHIGUCHI,* and Nobuo IZUMIYA†

Laboratory of Biochemistry, Faculty of Engineering, Kyushu Sangyo University, Matsukadai, Higashi-ku, Fukuoka 813, Japan

*Laboratory of Biochemistry, Faculty of Science, Kyushu University, Hakozaki, Higashi-ku, Fukuoka 812, Japan

Received August 24, 1993

A gramicidin S analog ([Orn^{1,1'}]GS·4HCl) containing L-ornithine in place of L-valine at the 1,1' positions was synthesized by the conventional solution method in order to examine whether this analog had antibacterial activity toward Gram-negative bacteria. In the synthesis of [Orn^{1,1'}]GS·4HCl, two intermediate analogs ([Orn^{1,1'}, Orn(For)^{2,2'}]GS·2HCl and [Orn(Z)^{1,1'}]GS·2HCl) were obtained. [Orn^{1,1'}]GS·4HCl and [Orn^{1,1'}, Orn(For)^{2,2'}]GS·2HCl showed no activity toward either Gram-negative or Gram-positive bacteria, whereas [Orn(Z)^{1,1'}]GS·2HCl showed appreciable activity toward only Gram-positive bacteria.

Gramicidin S (GS) is a cyclic decapeptide, cyclo-(Val-Orn-Leu-D-Phe-Pro-Val-Orn-Leu-D-Phe-Pro-), which is antibiotic toward Gram-positive bacteria, but inactive to Gram-negative ones. After the early synthesis of a GS analog [L-Lys^{2,2'}]GS·2HCl by Schwyzer and Sieber,¹⁾ a large number of analogs have been synthesized to study the relationships between the structure and activity.²⁾ Nishino *et al.* recently found that [L-Amy^{3,3'}]GS·2HCl was inactive due to the bulky side chains of the L-Amy residues.³⁾ Izumiya *et al.* has reported that [D-Dpr^{4,4'}]GS·4HCl showed appreciable antibacterial activity toward several Gram-negative bacteria, suggesting that the degree of activity would be influenced by the positive charges of the molecule.⁴⁾

In this paper, we report the synthesis and antibacterial activity of [Orn^{1,1'}]GS·4HCl (**14**) and its intermediates, [Orn(Z)^{1,1'}]GS·2HCl (**12**) and [Orn^{1,1'}, Orn(For)^{2,2'}]GS·2HCl (**13**) as shown in Fig. 1.

Firstly, a linear pentapeptide (**5**) was synthesized, in which the δ-amino groups of two L-Orn residues were protected by Z and For, respectively (Fig. 2).

Boc-5-OH (**5**) was transformed to Boc-5-ONSu (**6**) and H-5-OH (**7**), and these pentapeptides were then coupled to produce Boc-10-OH (**8**). Compound **8** was converted to H-10-ONSu·CF₃COOH (**10**) and then treated with pyridine at a high dilution. The reaction product was purified by column chromatography, using ion-exchange resins and Sephadex LH-20, the desired cyclic decapeptide (**11**) being obtained in a 24% yield (Fig. 3).

Hydrolysis of **11** with an HCl solution in MeOH gave crystalline [Orn(Z)^{1,1'}]GS·2HCl (**12**) in an 84% yield. Hydrogenolysis of **11** produced crystalline [Orn^{1,1'}, Orn(For)^{2,2'}]GS·2HCl (**13**) in an 81% yield. The important analog [Orn^{1,1'}]GS·4HCl (**14**) was obtained as colorless crystals in a 75% yield. The purity of each of these analogs was ascertained by TLC, amino acid analysis, and paper

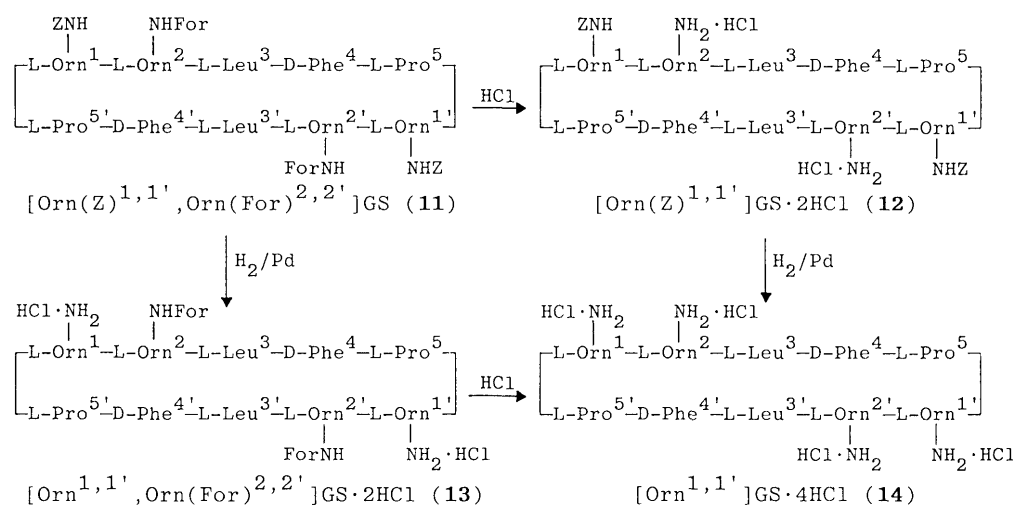


Fig. 1. Structures and Synthetic Scheme for the GS Analogs.

† To whom correspondence should be addressed.

Abbreviations according to IUPAC-IUB Commission, *Eur. J. Biochem.*, **138**, 9–37 (1984), are used throughout.

Additional abbreviations: Amy, α-aminomyristic acid; Boc, *tert*-butoxycarbonyl; CD, circular dichroism; DCHA, dicyclohexylamine; Dpr, 2,3-diaminopropanoic acid; EDC, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide; For, formyl; GS, gramicidin S; HONSu, *N*-hydroxysuccinimide; Orn, L-ornithine; Z, benzyloxycarbonyl.

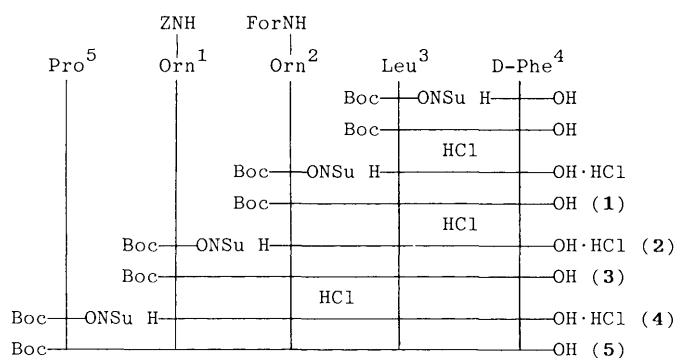


Fig. 2. Synthesis of Boc-pentapeptide-OH 5.

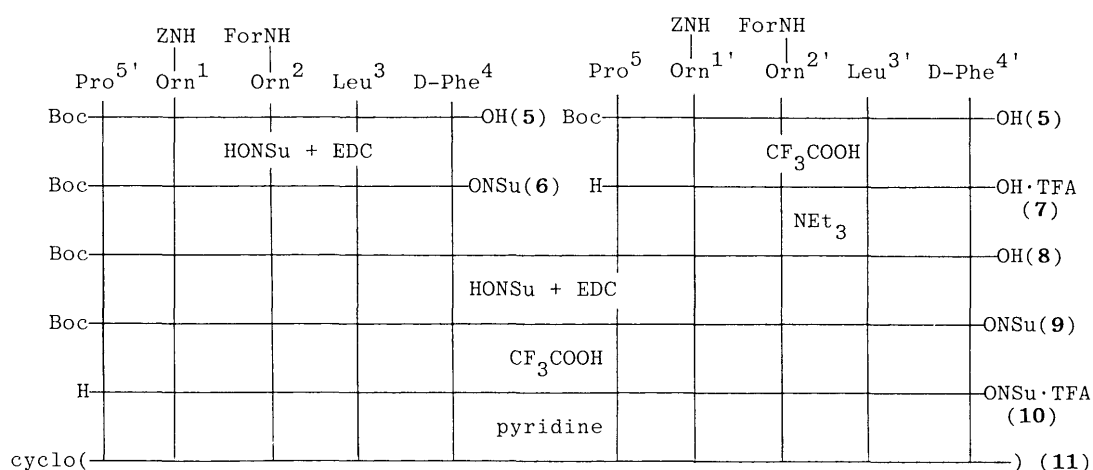


Fig. 3. Synthesis of cyclo-(protected decapeptide-) 11.

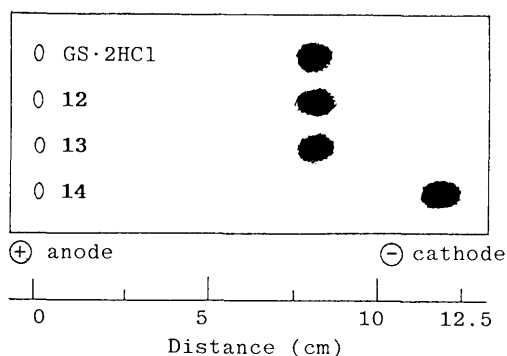


Fig. 4. Paper Electrophoresis of GS and Its Analogs. The conditions are described in the Experimental section.

Table Antibacterial Activity of GS and Its Analogs

Peptide	Minimum Inhibitory concentration (μg/ml)		
	<i>B. subtilis</i> ^a	<i>S. aureus</i> ^a	<i>E. coli</i> ^b
GS·2HCl	6.25	12.5	>100
[Orn(Z) ^{1,1'}]GS·2HCl (12)	12.5	25	>100
[Orn ^{1,1'} , Orn(For) ^{2,2'}]-GS·2HCl (13)	>100	>100	>100
[Orn ^{1,1'}]GS·4HCl (14)	>100	>100	>100
[Gly ^{1,1'}]GS·2HCl ^c	100	>100	>100
[D-Dpr ^{4,4'}]GS·4HCl ^d	50	100	25

^a Gram-positive bacteria.^b Gram-negative bacteria.^c From ref. 5.^d From ref. 4.

electrophoresis. Analogs **12** and **13** showed almost the same mobility as GS·2HCl, whereas the mobility of **14** was greater than that of GS (Fig. 4).

The CD spectrum of **12** was similar to that of GS and showed double-minima at *ca.* 205 and 215 nm, although the ellipticity of **12** was lower than that of GS (Fig. 5). The CD spectra of **13** and **14** showed no distinct ellipticity at 215 nm. The molecular shape of **14** may be different from that of GS due to the presence of two cationic charges at the 1- and 1'-positions.

Compound **12** showed appreciable antibacterial activity toward Gram-positive bacteria as GS did (Table). It should be noted that [Gly^{1,1'}]GS was inactive.⁵⁾ These

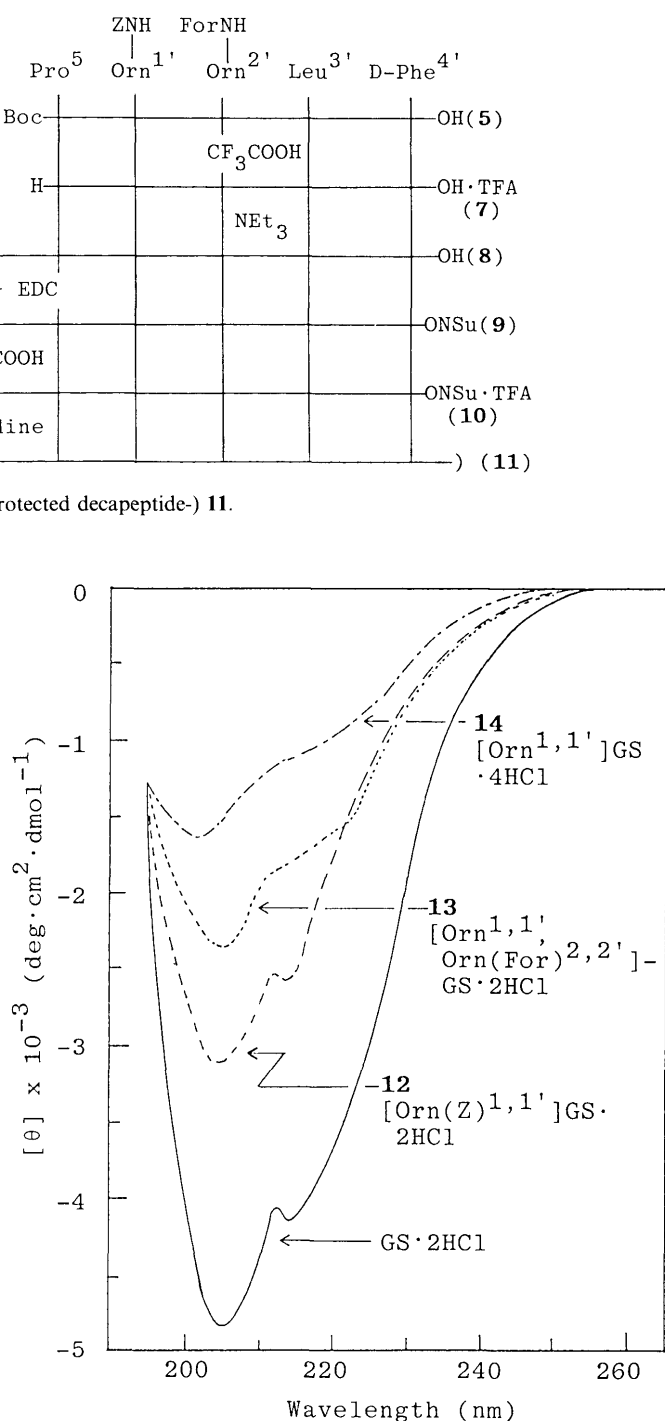


Fig. 5. CD Curves for GS and Its Analogs. Methanol was used as the solvent.

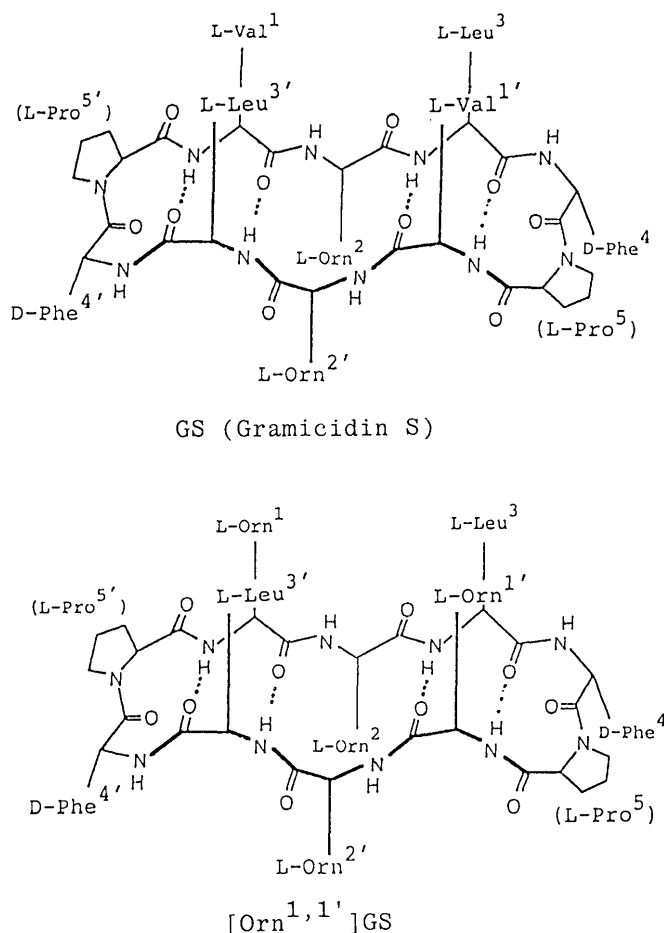


Fig. 6. Conformation of GS (top) and the Assumed Conformation of [Orn^{1,1'}]GS (bottom).

results indicate that the bulkiness of the side chain of 1,1'-L-Orn(δ -Z) residues may be important for displaying antibacterial activity. Compound **14** showed no activity against Gram-negative bacteria nor against positive ones. Assuming that **14** and GS had a similar conformation as shown in Fig. 6, the four alkyl side chains of L-Val and L-Leu may form the hydrophobic face of GS, and this face would be changed to a hydrophilic face by the two 1,1'-L-Orn residues in **14**.

Experimental

TLC was carried out on silica gel G (Merck) with the following solvent systems (vol. %): R_f^1 , CHCl₃-MeOH (5:1); R_f^2 , CHCl₃-MeOH-AcOH (50:10:2); R_f^3 , *n*-BuOH-AcOH-pyridine-H₂O (4:1:1:2). Optical rotation values were measured with a Horiba SEPA-200 polarimeter, while amino acid analyses were performed with a Hitachi KLA-5 analyzer. Paper electrophoreses were carried out on Toyo Roshi No. 51 paper with HCOOH-AcOH-MeOH-H₂O (1:3:6:10, pH 1.9) for 3 h at 600 V. CD measurements were taken with a Jasco J-40A spectropolarimeter, using a cuvette with a 0.01 cm light-path width.

H-Leu-D-Phe-OH·HCl. A solution of Boc-Leu-D-Phe-OH·DCHA (22.4 g, 40 mmol)⁶⁾ in EtOAc was treated with 10% aqueous citric acid, and the organic layer was washed with water and dried with Na₂SO₄. The filtrate was evaporated *in vacuo*, and the remaining oil was dissolved in 3.5 M HCl in dioxane (120 ml). The solution was left to stand at room temperature for 2 h and then evaporated. The residue that had solidified during standing was collected with the help of ether: yield, 9.3 g (74%); mp 185–188°C; R_f^2 0.71. H-Leu-D-Phe-OH·2H₂O has been synthesized by hydrogenating Z-Leu-D-Phe-OH.⁷⁾

Boc-Orn(For)-Leu-D-Phe-OH (1). H-Leu-D-Phe-OH·HCl (7.9 g, 25

mmol) was dissolved in a mixture of H₂O (50 ml) and NEt₃ (7 ml, 50 mmol). A solution of Boc-Orn(For)-ONSu (9.8 g, 27.5 mmol)⁸⁾ in dioxane (50 ml) was added to this mixture at 5–10°C, and the resulting mixture was stirred overnight at room temperature. After the reaction mixture was evaporated, 10% citric acid was added to the residue. This mixture was extracted with EtOAc, and the organic layer was washed with sat. NaCl and then dried (Na₂SO₄). The resulting filtrate was evaporated, and the solid was collected with ether: yield, 9.1 g (70%); mp 141–143°C; $[\alpha]_D^{20}$ –40.6° (c 1.0, dioxane); R_f^2 0.57, R_f^3 0.77. *Anal.* Found: C, 59.66; H, 7.70; N, 10.64%. Calcd. for C₂₆H₄₀O₇N₄: C, 59.98; H, 7.74; N, 10.76%.

H-Orn(For)-Leu-D-Phe-OH·HCl (2). Compound **1** (7.8 g, 15 mmol) in 3.5 M HCl in dioxane (45 ml) was treated as described for the preparation of H-Leu-D-Phe-OH·HCl: yield of solid **2**, 6.5 g (95%); mp 122–124°C; R_f^3 0.19. *Anal.* Found: C, 53.21; H, 7.37; N, 11.41%. Calcd. for C₂₁H₃₂O₅N₄·HCl·H₂O: C, 53.11; H, 7.43; N, 11.79%.

Boc-Orn(Z)-Orn(For)-Leu-D-Phe-OH (3). Compound **2** (4.57 g, 10 mmol) was dissolved in a mixture of H₂O (100 ml) and NEt₃ (2.8 ml, 20 mmol). A solution of Boc-Orn(Z)-ONSu (5.1 g, 11 mmol)⁹⁾ in dioxane (100 ml) was added to this mixture, which was then reacted as described for **1**. After evaporating, the solid was collected with ether, and recrystallized from MeOH and ether: yield, 5.46 g (71%); mp 152–156°C; $[\alpha]_D^{20}$ –34.4° (c 1.0, EtOH); R_f^2 0.71, R_f^3 0.75. *Anal.* Found: C, 61.06; H, 7.62; N, 10.81%. Calcd. for C₃₉H₅₆O₁₁N₆: C, 60.92; H, 7.43; N, 10.93%.

H-Orn(Z)-Orn(For)-Leu-D-Phe-OH·HCl (4). Compound **3** (3.85 g, 5 mmol) was dissolved in 3.5 M HCl/dioxane (14 ml), and the solid was collected with ether as described for H-Leu-D-Phe-OH·HCl: yield, 3.22 g (88%); mp 227–229°C (dec.); R_f^2 0.25, R_f^3 0.63. *Anal.* Found: C, 55.43; H, 6.93; N, 11.42%. Calcd. for C₃₄H₄₈O₈N₆·HCl·2H₂O: C, 55.09; H, 7.22; N, 11.34%.

Boc-Pro-Orn(Z)-Orn(For)-Leu-D-Phe-OH (5). Compound **4** (2.93 g, 4 mmol) was dissolved in 50% aqueous dioxane (200 ml) with NEt₃ (1.12 ml, 8 mmol). A solution of Boc-Pro-ONSu (1.37 g, 4.4 mmol)¹⁰⁾ in dioxane (20 ml) was then added, and the resulting mixture was stirred overnight at room temperature. The mixture was evaporated, the residue was triturated with 10% citric acid, and the solid was collected: yield, 2.85 g (81%); mp 182–185°C; $[\alpha]_D^{20}$ –84.5° (c 0.5, EtOH); R_f^1 0.85. Amino acid hydrolyzate of **5**: Orn 1.92, Leu 1.03, Phe 1.00, Pro 0.88. *Anal.* Found: C, 59.69; H, 7.37; N, 11.04%. Calcd. for C₄₄H₆₃O₁₁N₇·H₂O: C, 59.79; H, 7.43; N, 11.10%.

Boc-Pro-Orn(Z)-Orn(For)-Leu-D-Phe-ONSu (6). A solution of **5** (1.77 g, 2 mmol), EDC·HCl (0.767 g, 4 mmol) and HONSu (0.46 g, 4 mmol) in DMF (15 ml) was stirred at 0°C for 2 d. After evaporating, the residue was triturated with cold water, collected, and dried in a desiccator: yield, 1.69 g (86%); R_f^2 0.25.

H-Pro-Orn(Z)-Orn(For)-Leu-D-Phe-OH·TFA (7). A solution of **5** (1.77 g, 2 mmol) in CF₃COOH (20 ml) was left to stand at 0°C and then at room temperature for 30 min each. The mixture was evaporated, and the solid was dried in a desiccator: yield, 1.76 g (100%); R_f^2 0.31.

Boc-(Pro-Orn(Z)-Orn(For)-Leu-D-Phe)₂-OH (8). To a solution of **7** (1.94 g, 2.2 mmol) and NEt₃ (0.66 ml, 4.4 mmol) in DMF (20 ml) was added a solution of **6** (1.76 g, 2 mmol) in DMF (20 ml) at 0°C. The mixture was left to stand at 0°C for 1 h and then overnight at room temperature, and evaporated. The residue was triturated with 10% citric acid, and the solid was collected (3.13 g). This solid material was purified in a Sephadex LH-20 column (1.8 × 180 cm), MeOH being used as the eluent. The fractions containing **8** were evaporated, and the residue was recrystallized from MeOH-ether: yield, 1.79 g (52%); mp 147–151°C; $[\alpha]_D^{20}$ –48.5° (c 1.0, DMF); R_f^2 0.73. *Anal.* Found: C, 60.16; H, 7.25; N, 11.97%. Calcd. for C₈₃H₁₁₆O₁₉N₁₄·2H₂O: C, 60.42; H, 7.35; N, 11.89%.

Boc-(Pro-Orn(Z)-Orn(For)-Leu-D-Phe)₂-ONSu (9). A solution of **8** (3.43 g, 2 mmol), HONSu (0.46 g, 4 mmol) and EDC·HCl (0.77 g, 4 mmol) in DMF (20 ml) was treated in the same way as that described for **6**: yield, 3.24 g (95%); R_f^2 0.73.

H-(Pro-Orn(Z)-Orn(For)-Leu-D-Phe)₂-ONSu·TFA (10). A solution of **9** (1.71 g, 1 mmol) in TFA (10 ml) was left to stand for 2 h at 0°C, and then evaporated. The solid was collected by ether: yield, 1.66 g (96%); R_f^2 0.42.

*cyclo(-Orn(Z)-Orn(For)-Leu-D-Phe-Pro-)*₂ (**11**). A solution of **10** (1.66 g, *ca.* 0.95 mmol) dissolved in DMF (20 ml) was added to pyridine (300 ml) at room temperature. The mixture was stirred for 2 d and then evaporated. The residue was dissolved in a mixture of MeOH and H₂O (5:1), and this solution was put into a column of Dowex 50 × 8 (1.8 × 10 cm), eluting with the same solvent. The eluate was treated in a column of Dowex 1 × 8 (1.8 × 10 cm) and evaporated, the residue then being dissolved in MeOH and purified by LH-20 as described for the purification of **8**. Recrystallization from MeOH–ether afforded pure **11** in a 0.366 g (24%) yield; mp 132–134°C; $[\alpha]_D^{20}$ –215 (*c* 0.2, MeOH); R_f^1 0.55. Amino acid hydrolyzate of **11**: Orn 2.04, Leu 0.97, Phe 1.00, Pro 0.94. *Anal.* Found: C, 58.73; H, 7.35; N, 12.41%. Calcd. for C₇₈H₁₀₆O₁₆N₁₄·6H₂O: C, 58.41; H, 7.43; N, 12.23%.

[Orn(Z)^{1,1'}]GS·2HCl (**12**). A solution of **11** (80 mg, 0.05 mmol) in 0.5 M HCl in MeOH (10 ml) was left to stand for 3 d at room temperature. The solution was then evaporated, and the residual crystals were collected with the help of a mixture of MeOH and ether (1:4): yield, 68 mg (84%); mp 162–165°C (dec.); R_f^1 0.22, R_f^3 0.78. *Anal.* Found: C, 57.56; H, 7.34; N, 12.31%. Calcd. for C₇₆H₁₀₆O₁₄N₁₄·2HCl·4H₂O: C, 57.60; H, 7.27; N, 12.38%.

[Orn^{1,1'}, Orn(For)^{2,2'}]GS·2HCl (**13**). Compound **11** (80 mg, 0.05 mmol) was dissolved in a mixture of MeOH (10 ml), AcOH (1 ml) and H₂O (1 ml), and then hydrogenated on Pd black. The resulting filtrate was evaporated, and the crystalline residue was dried in a desiccator. The crystals were dissolved in MeOH (5 ml) containing HCl (0.1 mmol), and the solution was evaporated. Crystals of **13** were collected with a mixture of MeOH and ether (1:4): yield, 58 mg (81%); mp 167–170°C (dec.); R_f^1 0.07, R_f^3 0.71. *Anal.* Found: C, 54.09; H, 7.50; N, 14.09%. Calcd. for C₆₂H₉₄O₁₂N₁₄·2HCl·4H₂O: C, 54.26; H, 7.51; N, 14.29%.

[Orn^{1,1'}]GS·4HCl (**14**). Compound **11** (160 mg, 0.1 mmol) was dissolved in a mixture of MeOH–AcOH–H₂O (10:1:1), and then hydrogenated. The resulting filtrate was evaporated, and the residue was treated with 0.5 M HCl in MeOH (20 ml) as described for **12**: yield, 112 mg (75%); mp

240–242°C (dec.); R_f^1 0.04, R_f^3 0.53. *Anal.* Found: C, 49.22; H, 7.53; N, 13.28%. Calcd. for C₆₀H₉₄O₁₀N₁₄·4HCl·8H₂O: C, 49.31; H, 7.60; N, 13.42%.

A small amount of **14** was also obtained by the route *via* **11** and **12**.

Antibacterial activity. The minimum amounts of peptides necessary for completely inhibiting growth were determined by a dilution method, using a bouillon agar medium. The results are shown in the Table.

Acknowledgments. We thank Prof. Eiji Kimoto of Fukuoka University, and Prof. Norikazu Nishino of Kyushu Institute of Technology for their valuable discussions. We also thank the staff at Central Research Institute of Takeda Chem. Industries Ltd. for the bioassays.

References

- 1) R. Schwyzler and P. Sieber, *Helv. Chim. Acta*, **41**, 2186–2189 (1958).
- 2) N. Izumiya, T. Kato, H. Aoyagi, M. Waki, and M. Kondo, in "Synthetic Aspects of Biologically Active Cyclic Peptides—Gramicidin S and Tyrocidines," Kodansha, Tokyo, and J. Wiley & Sons, New York, 1979, Chapter 4.
- 3) H. Mihara, N. Nishino, I. Ogawa, N. Izumiya, and T. Fujimoto, *Bull. Chem. Soc. Jpn.*, **65**, 228–233 (1992).
- 4) S. Ando, T. Kato, and N. Izumiya, *Int. J. Peptide Protein Res.*, **25**, 15–26 (1985).
- 5) M. Kondo and N. Izumiya, *Bull. Chem. Soc. Jpn.*, **40**, 1975–1980 (1965).
- 6) K. Sato, H. Abe, T. Kato, and N. Izumiya, *Bull. Chem. Soc. Jpn.*, **50**, 1999–2004 (1977).
- 7) R. L. M. Synge, *Biochem. J.*, **44**, 99–104 (1948).
- 8) K. Hofmann, J. P. Visser, and F. M. Finn, *J. Am. Chem. Soc.*, **92**, 2900–2909 (1970).
- 9) S. F. Sernosek, M. Wells, and G. D. Fasman, *Israel J. Chem.*, **12**, 47–66 (1974).
- 10) G. W. Anderson, J. E. Zimmerman, and F. M. Callahan, *J. Am. Chem. Soc.*, **86**, 1839–1842 (1964).