

Total Synthesis of Sperabillin A and C

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Received 22 August 2005

Dedicated to Prof. Dr. Dr. h.c. Fritz Eiden on the occasion of his 80th birthday.

Abstract: The first total synthesis of sperabillin A and an improved total synthesis of sperabillin C have been achieved in 11 steps from *N*-Boc-*O*-methyl-L-tyrosine. The stereoselective pathway to the core (3*R*,5*R*)-3,6-diamino-5-hydroxyhexanoic acid involves an Arndt–Eistert homologation, an asymmetric Henry reaction and a ruthenium tetroxide-catalyzed oxidative degradation of a benzene ring as key steps.

Key words: sperabillins, antibiotics, pseudo-peptides, Henry reaction, ruthenium tetroxide

(3*R*,5*R*)-3,6-Diamino-5-hydroxyhexanoic acid (**1**, Figure 1) is the core fragment of the pseudo-peptide antibiotics negamycin (**2**) and sperabillin A and C (**3a** and **3c**, respectively). Negamycin (**2**), isolated in 1970 from culture filtrates of strains closely related to *Streptomyces purpeofuscus* by Umezawa et al., shows potent growth-inhibitory activity against Gram-negative bacteria.¹ By contrast sperabillin A (**3a**), the main member of the sperabillin family of antibiotics, which was isolated in 1986 from the culture broth of *Pseudomonas fluorescens* YK-437 by researchers at Takeda Chemical Industries Ltd., exerts promising in vitro and in vivo antibacterial activity especially against Gram-positive pathogens, including multiresistant strains of *Staphylococcus aureus*.² While there are numerous reports in the literature concerning the synthesis of negamycin (**2**), both in racemic and optically active form,³ little attention has been given to the synthesis of sperabillin A (**3a**) and C (**3c**) incorporating the same core β,ε-diamino acid. Only one total synthesis of sperabillin C (**3c**; 15 steps starting from *N*-Boc-glycine, 2.5% overall yield) has been reported to date.^{4,5}

In this communication, we describe the first total synthesis of sperabillin A (**3a**) as well as an improved total synthesis of sperabillin C (**3c**). Our approach toward these antibiotics and analogues with modified fragments attached to the N^ε- and C-terminal regions is based on the orthogonally protected β,ε-diamino acid **4**. As outlined in Scheme 1 building block **4** was envisioned to be readily accessible from *N*-Boc-L-tyrosine derived aldehyde **6** by two key transformations: (a) stereoselective Henry reaction to give β-nitro alcohol **5** and (b) oxidative degradation of the phenyl ring, which serves as a masked

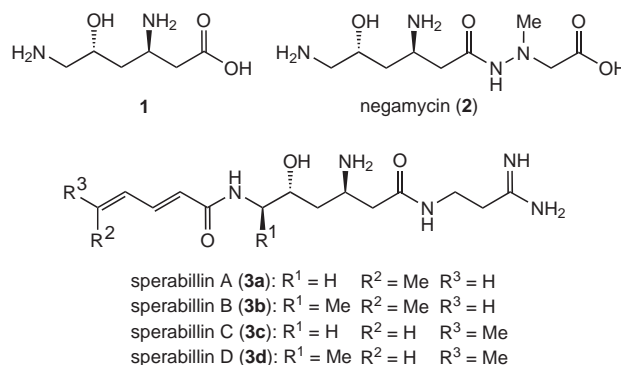
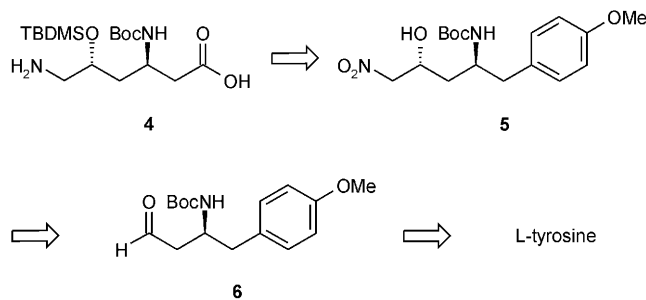


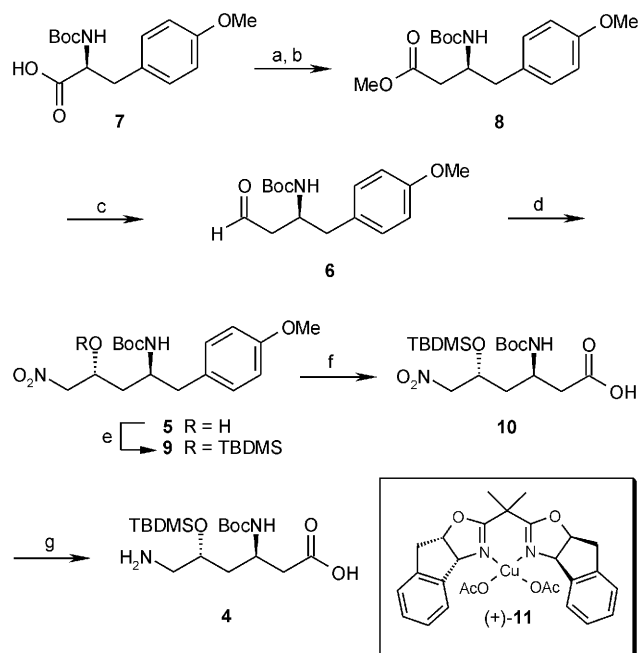
Figure 1

carboxylic acid, and subsequent reduction of the nitro group.

Aldehyde **6** was readily obtained from commercially available *N*-Boc-*O*-methyl-L-tyrosine (**7**) in two steps (Scheme 2). Arndt–Eistert homologation of the α-amino acid **7** following a standard protocol⁶ afforded β-amino ester **8** {[α]_D²³ –9.3 (c 0.8 in CH₂Cl₂); lit.⁷ (R)-**8**: [α]_D²³ +9.2 (c 1.2 in CH₂Cl₂)}, which subsequently was reduced with DIBAL-H (toluene, –85 °C) to give **6** in 73% overall yield. Previous studies conducted in this and other laboratories have indicated that it is difficult to achieve control of stereochemistry for Henry reactions of nitromethane with chiral aldehydes like **6** bearing the inducing stereogenic center in the β-position.⁸ Therefore we envisioned the application of a chiral catalyst, namely Evans' bis(oxazoline) copper(II) acetate-based catalyst (+)-**11**,⁹ to control the configuration of the newly created stereogenic center at C-5.^{8c} Indeed, Henry reaction of nitromethane with aldehyde **6** in the presence of 5 mol% of (+)-**11**



Scheme 1



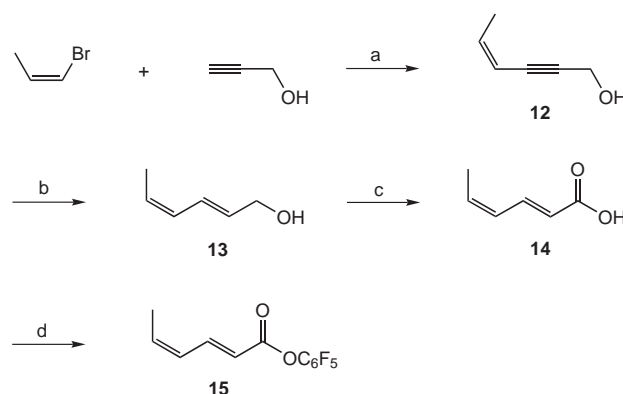
Scheme 2 Reagents and conditions: (a) i) isobutyl chloroformate, NMM, DME, -20°C ; ii) CH_2N_2 , -20°C to r.t.; (b) silver benzoate, Et_3N , MeOH, r.t. (86%, two steps); (c) DIBAL-H, toluene, -85°C (85%); (d) MeNO_2 , (+)-**11** (5 mol%), EtOH, r.t. (83%); (e) TBDMSCl, imidazole, DMF, r.t. (82%); (f) RuO_2 (5 mol%), NaIO_4 , NaHCO_3 , $\text{EtOAc-H}_2\text{O}$, r.t. (67%); (g) HCO_2NH_4 , Pd/C, MeOH, r.t. (90%)

(EtOH, r.t., 3 d) afforded the desired β -nitro alcohol **5** with both high diastereoselectivity (95% de)¹⁰ and yield (83%).

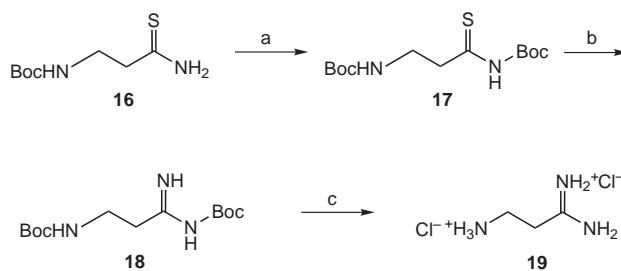
After protecting the secondary alcohol as a *tert*-butyldimethylsilyl (TBDMS) ether we turned our attention to the oxidative degradation of the 4-methoxyphenyl ring. Ruthenium tetroxide-catalyzed degradation of aromatic rings to carboxylic acids has been repeatedly applied as a useful tool in the synthesis of amino acids and peptides.¹¹ However, *N*-acylamines bearing a primary alkyl group at the nitrogen were reported to undergo facile oxidation at the α -position to give the corresponding imides.¹² Accordingly, the aromatic ring has to be cleaved prior to the generation of the C-6 amino group assuming that the nitro group would act as a 'protective group' for amines during ruthenium tetroxide oxidations. Indeed ruthenium tetroxide-catalyzed degradation of **9** afforded the desired ε -nitro carboxylic acid **10** in good yield (67%). Best results were obtained using catalytic amounts of ruthenium dioxide (5 mol%) and an excess of sodium metaperiodate (30 equiv) in a two-phase system of ethyl acetate and water at room temperature according to Yoshifuji et al.^{12,13} To avoid cleavage of the acid labile protecting groups due to pH decrease during the oxidation sodium hydrogen carbonate was added.¹⁴ Finally, catalytic reduction of the nitro group with ammonium formate in the presence of Pd/C afforded the desired amino acid **4**¹⁵ in 90% yield.

(2*E*,4*Z*)-Hexa-2,4-dienoic acid (**14**), representing the N^{ε} -acyl residue of sperabillin A (**3a**) and B (**3b**), was pre-

pared in four steps, starting with a Sonogashira coupling of (*Z*)-1-bromoprop-1-ene and propargylic alcohol $[(\text{Ph}_3\text{P})_2\text{PdCl}_2$ (0.25 mol%), CuI (1.5 mol%), Ph_3P , *n*-PrNH₂, Et₂O, Δ) to give (*Z*)-hex-4-en-2-yn-1-ol (**12**)¹⁶ in 69% yield (*Z*:*E* > 99:1; Scheme 3).¹⁷ This product was stereoselectively reduced with LiAlH_4 (THF, Δ)¹⁸ providing (2*E*,4*Z*)-hexa-2,4-dien-1-ol (**13**) in 94% yield [(2*E*,4*Z*):(2*Z*,4*Z*) > 99:1]. Oxidation of **13** was then accomplished by a one-pot, two-step procedure. First, the allylic alcohol **13** was oxidized with manganese dioxide (pentane, r.t.) to give the corresponding aldehyde, which was further oxidized by the addition of silver nitrate (KOH, EtOH-H₂O, r.t.) to afford diastereoisomerically pure (2*E*,4*Z*)-2,4-hexadienic acid (**14**) in 77% overall yield. Finally, the carboxylic acid **14** was converted into the corresponding pentafluorophenyl ester **15** ($\text{C}_6\text{F}_5\text{OH}$, EDC, CH_2Cl_2 , r.t.) in 94% yield.



Scheme 3 Reagents and conditions: (a) $(\text{Ph}_3\text{P})_2\text{PdCl}_2$, CuI, Ph_3P , *n*-PrNH₂, Et₂O, Δ (69%); (b) LiAlH_4 , THF, Δ (94%); (c) (i) MnO_2 , pentane, r.t. (ii) AgNO_3 , KOH, Et₂O-H₂O, r.t. (77%); (d) $\text{C}_6\text{F}_5\text{OH}$, EDC, CH_2Cl_2 , 0°C (94%)

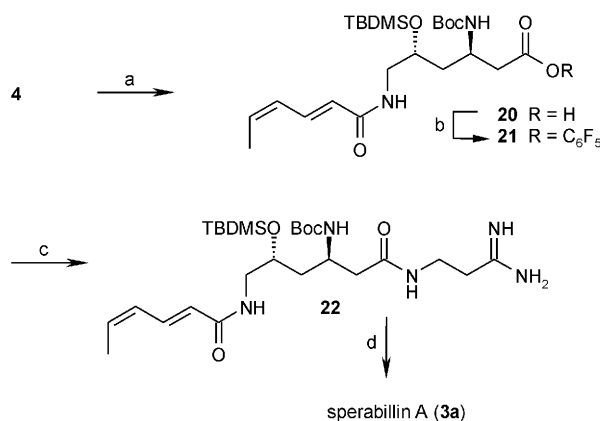


Scheme 4 Reagents and conditions: (a) (i) NaH (2.1 equiv), $(\text{Boc})_2\text{O}$, THF, -78°C to -40°C (88%); (b) NH_3 , EtOH, 0°C (99%); (c) HCl, Et₂O, 0°C to r.t. (quant.)

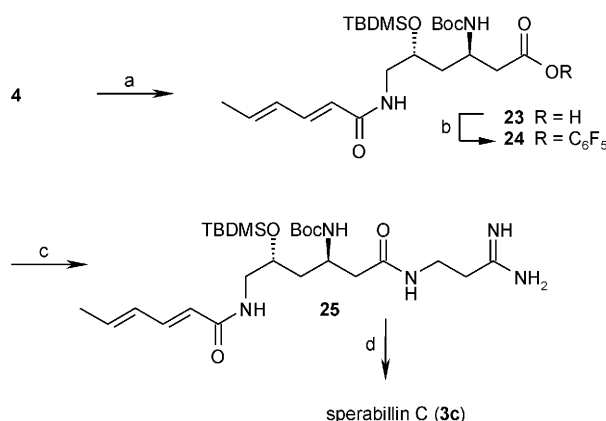
3-Aminopropionamidine, the fragment common to all members of the sperabillin family of antibiotics and also part of a number of other antibiotics (e.g. distamycin A or amidinomycin), was prepared in three steps, starting from readily available *N*-Boc- β -alanine thioamide (**16**,¹⁹ Scheme 4).²⁰ First, the thioamide **16** was acylated with di-*tert*-butyl dicarbonate in the presence of 2.1 equivalents of sodium hydride (THF, -78°C to -40°C)²¹ to afford *N*-Boc thioamide **17** in 88% yield. Secondly, **17** was readily converted into the corresponding *N*-Boc amidine **18** by

treatment with NH_3 in ethanol at 0°C (99% yield). Removal of the *tert*-butoxycarbonyl protecting groups with 2.0 M ethereal HCl then led to 3-aminopropionamidine dihydrochloride (**19**) in quantitative yield.

With the three building blocks **4**, **15** and **19** in hand sperabillin A (**3a**) was readily assembled in four steps (Scheme 5). First, the ϵ -amino group was acylated with pentafluorophenyl ester **15** (Et_3N , DMF, r.t.) to give amide **20** in 71% yield. Subsequently, **20** was activated as a pentafluorophenyl ester ($\text{C}_6\text{F}_5\text{OH}$, EDC, CH_2Cl_2 , r.t.) and then coupled with 3-aminopropionamidine dihydrochloride (**19**) (Et_3N , DMF, r.t.) to afford **22** in 82% yield (two steps). Finally, the acid-labile protecting groups were removed by treatment with TFA (5% H_2O , r.t.) to afford, after anion exchange on Amberlite® IRA 402 Cl resin, sperabillin A dihydrochloride (**3a**)²² in 92% yield. The spectroscopic data for **3a** were in full agreement with those reported for the natural product.^{2c} In the end, sperabillin C (**3c**)²³ was prepared analogously to **3a** in 20% overall yield from **4**, (2*E*,4*E*)-hexa-2,4-dienoic acid pentafluorophenyl ester and **19** (Scheme 6).



Scheme 5 Reagents and conditions: (a) **15**, Et_3N , DMF, r.t. (71%); (b) $\text{C}_6\text{F}_5\text{OH}$, EDC, CH_2Cl_2 , r.t.; (c) **19**, Et_3N , DMF, r.t. (82%, two steps); (d) TFA, H_2O (5%), r.t. (92%)



Scheme 6 Reagents and conditions: (a) (2*E*,4*E*)-hexa-2,4-dienoic acid pentafluorophenyl ester, Et_3N , DMF, r.t. (83%); (b) $\text{C}_6\text{F}_5\text{OH}$, EDC, CH_2Cl_2 , r.t.; (c) **19**, Et_3N , DMF, r.t. (87%, two steps); (d) TFA, H_2O (5%), r.t. (94%)

In summary, the first total synthesis of the pseudopeptide antibiotic sperabillin A (**3a**) as well as a markedly improved total synthesis of sperabillin C (**3c**) have been accomplished [11 steps from *N*-Boc-*O*-methyl-L-tyrosine (**7**), 16% and 20% overall yield, respectively]. Our concise and inherently flexible approach seems to be quite attractive for the generation of libraries of sperabillin A and C analogues for biological evaluation.

Acknowledgment

We are greatly indebted to Prof. Dr. Klaus T. Wanner for his generous support. We would also like to thank Michael Felkel and Alexandra Schüler for technical assistance.

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- (10) The reaction of chiral aldehyde **6** with nitromethane in the presence of catalyst (+)-**11** represents the matched case of double diastereoselection, since application of the enantiomeric catalyst (–)-**11** gave the corresponding epimer of **5** with slightly lower stereoselectivity (91% de).
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- (13) (a) In addition to product **10**, its *N*-formyl derivative 3-*tert*-butoxycarbonylformylamino-5-*tert*-butyldimethylsilyloxy-6-nitrohexanoic acid (5% yield) and trace amounts (<1%) of 4-*tert*-butoxycarbonylamino-2-*tert*-butyldimethylsilyloxyhexanedioic acid, the latter indicating a Nef-type reaction at the nitromethyl group, were obtained. Application of the reaction conditions [2.2 mol% RuCl₄, NaIO₄ (18 equiv), CCl₄-MeCN-H₂O = 2:2:3, r.t.] previously developed by Sharpless and co-workers (see ref. 13b) gave the oxidation product **10** in a 41% yield together with 24% of the *N*-formyl derivative mentioned above. (b) Carlsen, P. H. J.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. *J. Org. Chem.* **1981**, 46, 3936.
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- (15) Spectroscopic data of compound **4**: [α]_D²⁰ +20.8 (*c* 0.95 in CH₂Cl₂). ¹H NMR (500 MHz, MeOH-*d*₄): δ = 0.13 (3 H, s), 0.15 (3 H, s), 0.92 (9 H, s), 1.43 (9 H, s_{br}), 1.78 (2 H, m), 2.31 (1 H, dd, *J* = 14.9, 7.7 Hz), 2.39 (1 H, dd, *J* = 14.9, 4.7 Hz), 3.01 (1 H, dd, *J* = 13.0, 4.1 Hz), 3.06 (1 H, dd, *J* = 13.0, 4.9 Hz), 3.85 (1 H, m), 4.18 (1 H, m), 4.09 (1 H, m). ¹³C NMR (100 MHz, MeOH-*d*₄): δ = -5.0, 18.4, 25.9, 28.3, 40.0, 43.5, 45.2, 45.8, 67.7, 79.6, 157.1, 178.6.
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- (22) Spectroscopic data of sperabillin A (**3a**)^{2c}: [α]_D²³ -11.4 (*c* 0.4 in H₂O), lit.^{2c} [α]_D²³ -11 (*c* 1.1 in H₂O). ¹H NMR (500 MHz, D₂O): δ = 1.77 (1 H, ddd, *J* = 4.5, 10.0, 15.0 Hz), 1.87 (3 H, d, *J* = 7.2 Hz), 1.87 (1 H, m), 2.68 (2 H, t, *J* = 6.6 Hz), 2.74 (2 H, m), 3.33 (1 H, dd, *J* = 6.6, 14.0 Hz), 3.39 (1 H, dd, *J* = 4.5, 14.0 Hz), 3.55 (1 H, dt, *J* = 6.6, 14.0 Hz), 3.59 (1 H, dt, *J* = 6.7, 14.0 Hz), 3.86 (1 H, m), 4.00 (1 H, m), 6.02 (1 H, dq, *J* = 7.2, 10.8 Hz), 6.07 (1 H, d, *J* = 15.1 Hz), 6.23 (1 H, t, *J* = 10.8 Hz), 7.56 (1 H, dd, *J* = 11.8, 15.1 Hz). ¹³C NMR (100 MHz, D₂O): δ = 16.0, 35.3, 38.0, 39.2, 39.8, 47.8, 49.1, 69.1, 125.1, 129.6, 139.4, 139.5, 171.7, 172.4, 174.7.
- (23) Spectroscopic data of sperabillin C (**3c**)^{2c}: [α]_D²³ -10.2 (*c* 0.4 in H₂O), lit.^{2c} [α]_D²⁰ -11 (*c* 0.7 in H₂O). ¹H NMR (500 MHz, D₂O): δ = 1.75 (1 H, ddd, *J* = 4.7, 10.0, 15.0 Hz), 1.83 (3 H, d, *J* = 5.5 Hz), 1.87 (1 H, ddd, *J* = 3.1, 7.5, 15.0 Hz), 2.67 (2 H, t, *J* = 6.7 Hz), 2.73 (2 H, d, *J* = 7.0 Hz), 3.30 (1 H, dd, *J* = 6.5, 14.0 Hz), 3.37 (1 H, dd, *J* = 4.7, 14.0 Hz), 3.57 (2 H, m), 3.85 (1 H, m), 3.98 (1 H, m), 5.97 (1 H, d, *J* = 15.5 Hz), 6.26 (2 H, m), 7.13 (1 H, dd, *J* = 9.7, 15.5 Hz). ¹³C NMR (100 MHz, D₂O): δ = 20.7, 35.3, 38.0, 39.2, 39.8, 47.8, 49.1, 69.1, 122.9, 132.0, 143.2, 145.4, 171.6, 172.5, 174.7.