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Design, synthesis and biological evaluation of azithromycin glycosyl derivatives as potential antibacterial agents

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ABSTRACT

A series of 11,12-cyclic carbonate azithromycin-4"-O-carbamoyl glycosyl derivatives were designed, synthesized, and evaluated as antibacterial agents to search for target compounds with excellent activity. The results of preliminary antibacterial tests against eight strains in vitro revealed that all of the title compounds exhibited improved activities with broad spectrum compared with the parent compound. The glycosylated side chains may be the pharmacophores responsible for the improved activity.

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Over the past decade, macrolide antibiotics have played an important role in the clinical treatment of upper and lower respiratory tract infections. These antibiotics act by binding to the ribosomal RNA of bacteria. The previous studies provided strong, detailed evidence of this aforementioned interaction at the molecular level via high-resolution X-ray cocrystal structures.^{1–4} These X-ray studies demonstrated that macrolides inhibit bacterial protein synthesis by sterically blocking the passage of nascent polypeptides through the exit tunnel of the ribosome.⁵

Macrolide antibiotics, such as erythromycin A (ERY), clarithromycin (CLA), and azithromycin (AZI) (Fig. 1), are widely prescribed clinically. However, these antibiotics have various disadvantages. ERY, a first-generation macrolide, is readily degraded under acidic conditions, thus losing its antibacterial activity;⁶ these degraded products are responsible for undesirable gastrointestinal side effects.^{7,8} Second-generation macrolides, such as CLA and AZI, have been widely used for respiratory tract infection due to their superior antibacterial activity, pharmacokinetic properties and fewer gastrointestinal side effects compared to ERY.⁹ However, their clinical uses have been limited by the emergence of drug resistance.¹⁰ The increasing incidence of bacterial resistance is becoming a major threat to the successful treatment of infectious diseases. The most serious threat is the increasing resistance of community-acquired respiratory tract infections to various antimicrobials, which is a pandemic phenomenon.¹¹ Third-generation macrolide ketolides, such as telithromycin, can effectively address bacterial resistance and other issues associated with current macrolide regimens.^{12,13} However, the use of telithromycin suffers from limitations such as hepatotoxicity.¹⁴

With the widespread emergence of antibacterial resistance against macrolides, there is still a need to develop and extend safe chemotherapeutic agents with potent antibacterial activity. Recently, considerable efforts have been focused on discovering novel macrolides to combat the resistance. Research has indicated that compounds containing particular groups in the 4"-position of the cladinose sugar were effective against macrolide resistant strains.¹⁵ Additionally, the C-11,12 carbamate side chain of macrolides may aid in interactions with the target enzyme.¹⁶ So, we have chosen the 4"-position of the cladinose sugar as the modification position.

Glycosyl moieties exist extensively in organisms and exhibit a variety of biological functions, such as diagnostic and therapeutic potential. D-desosamine and L-cladinose, the two monosaccharides in macrolide structures, are important for the binding of macrolides to ribosomes. X-ray analyses of macrolides have revealed that the hydroxy group of a glycosyl moiety was important for hydrogen bond formation.² However, there have been few reports about glycosylation modification of macrolide antibiotics.^{17,18} As we believe that the glycosylation may help for the hydrogen bond formation, we therefore introduced glycosyl moieties to AZI through a carbamoyl group and examined the antibacterial activities of the resultant products.





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By substitution of the 4"-position with various carbamoylgroups, a series of novel 4"-substituted azithromycin derivatives were obtained. Further modification at 11,12-cyclic carbonate azithromycin was performed. We hypothesized that the 11,12-cyclic carbonate azithromycin-4"-O-carbamoyl glycosyl derivatives might enhance the antibacterial activity against resistant strains.

Eight glycosylated intermediates **1f–8f** were prepared from the corresponding commercially available saccharides by following a series of established transformations. We have chosen **1f** as an example (Scheme 1). First, all the hydroxyl groups of glucose (**1a**) were protected with acetyl groups to give the acetylated saccharide (**1b**) in an excellent yield. The treatment of **1b** with 2-azido-ethanol in the presence of BF₃·Et₂O afforded a series of protected azidoethyl-glycosides (**1c**) in the β -configuration with a very good yield. After deprotection of **1c**, we obtained the azidoethyl-glycosides (**1d**). Compound **1d** was protected with benzyl groups to give the benzylated saccharide (**1e**) in an excellent yield. Lastly, **1f** was reduced by Pd/CaCO₃ under a hydrogen atmosphere. The other seven glycosylated intermediates (**2f–8f**) were synthesized in the same manner.

The general synthetic methodology for preparing the compounds of interest (**F1–F8**) is outlined in Scheme 2. Protection of the 2'-hydroxyl group of azithromycin with acetic anhydride provided 2'-acetyl azithromycin (**B**) in 82% yield. 11,12-Cyclic carbonate azithromycin 4"-O-acylimidazolide (**C**) was obtained in 62% yield by treatment of compound **B** with NaH and *N*,*N*'-carbonyldiimidazole (CDI) in DMF. Then, the intermediates (**D1–D8**) were prepared by coupling compound **C**, followed by benzyl protection of the glycosylated group in the presence of 1,8-diazabicyclo [5.4.0] undec-7-ene (DBU). Lastly, intermediates **D1–D8** were reduced by 10% Pd-C under a hydrogen atmosphere. After methanolysis, we obtained compounds **F1–F8**. The yields were within the range of 70–80%.

The in vitro antibacterial activities were reported as minimum inhibitory concentrations (MICs), which were determined using astandard dilution assay as recommended by the NCCLS (National Committee of Clinical Laboratory Standard).¹⁹ The MIC_{80} was defined as the first well with an approximate 80% reduction in growth compared to the growth of the drug-free well. For the assays, the title compounds to be tested were dissolved in dimethyl sulfoxide (DMSO), serially diluted in growth medium, inoculated and incubated at 35 °C.

The selected strains evaluated were methicillin-susceptible *Staphylococcus aureus* MSSA-1 (*S. aureus* MSSA-1), methicillin-resistant *Staphylococcus aureus* MRSA-1 (*S. aureus* MRSA-1), *Staphylococcus aureus* ATCC25923 (*S. aureus* ATCC25923), *Streptococcus pneumoniae* 943 (*S. pneumoniae*943), *Staphylococcus pneumonia* 746 (*S. pneumoniae*746), *Streptococcus pyogenes* 447 (*S. pyogenes* 447), *Escherichia coli* 236 (*E. coli* 236), and *Escherichia coli* ATCC25922 (*E. coli* ATCC25922). ERY and AZI served as the positive control and were obtained from their respective manufacturers. The results of the assays are summarized in Table 1. The data points express the mean of replicate experiments. All of our susceptibility tests were performed three times using each antibacterial agent.

The results indicated that nearly all of the 11,12-cyclic carazithromycin-4"-O-carbamoyl glycosyl bonate derivatives showed moderate activity against all the strains, and someof the derivatives exhibited improved activity compared with AZI and ERY. Notably, the MIC₈₀ values indicate that compounds F1 and F2 showed improved activity against all of the bacterial strains relative to the other compounds, expressing the same or higher antibacterial activities as AZI. Compounds F7 and F8 with the disaccharide side chain showed the least activity; this observation could be due to increasing side chain length. Among the compounds tested, Compounds F1 and F6 showed the same activity against the S. aureus MSSA-1 as AZI, and they exhibited eightfold higher activity than ERY. Particularly, compounds F1 and F2 showed fourfold and eightfold higher activity against the S. pneumoniae 943 than AZI and ERY, respectively. In addition, the activity of compoundsF1 and F2 (MIC₈₀ 2 µg/mL) increased significantly against S. pneumoniae 746, showing eightfold higher activity than ERY. The activity of compounds

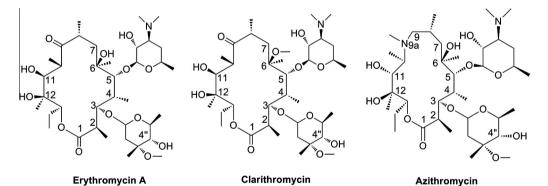
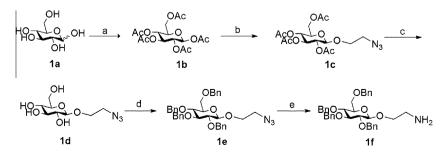
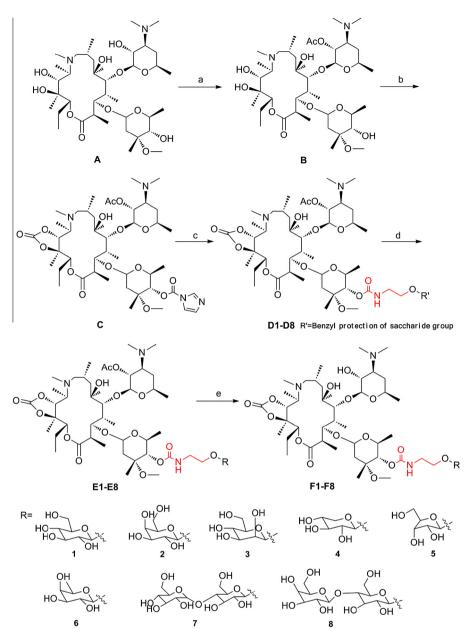


Figure 1. The structures of macrolide antibiotics.



Scheme 1. The synthesis of the saccharide intermediates. Reactions and conditions: (a) Ac₂O, CH₃COONa, reflux, 5 h, in 90% yield; (b) HOCH₂CH₂N₃, BF₃·Et₂O, DCM, Ar, 0 °C, 18 h, in 80% yield; (c) CH₃ONa, CH₃OH, 4 h, in 93% yield; (d) BnBr, NaH, TBAI, DMF, in 52% yield; and (e) Pd/CaCO₃, H₂, CH₃OH, in 98% yield.



Scheme 2. The synthesis of the compounds of interest. Reactions and conditions: (a) Ac₂O, DCM, Et₃N, rt, 24 h, in 82% yield; (b) CDI, NaH, DMF, rt, 24 h, in 62% yield; (c) R'OCH₂CH₂NH₂, DMF, DBU, rt, 12 h, in 21–38% yield; (d) Pd/C, H₂, CH₃OH, in 75–88% yield; and (e) CH₃OH, 55 °C, 20 h, 70–80% yield.

Table 1
Antibacterial activities of the title compounds in vitro (MIC_{80}, $\mu g/mL$)

Compd	S. aureus MSSA-1	S. aureus MRSA-1	S. aureus ATCC25923	S. pneumoniae 943	S. pneumoniae 746	S. pyogenes 447	E. coli 236	E. coli ATCC25922
F1	2	16	8	1	2	16	64	128
F2	4	32	32	1	2	64	128	128
F3	8	8	8	4	4	32	64	128
F4	16	16	8	2	4	16	64	256
F5	8	8	4	4	8	64	64	128
F6	2	8	16	4	8	64	128	128
F7	16	16	32	8	16	32	128	256
F8	32	32	16	8	16	64	128	256
AZI	2	16	16	4	8	32	256	256
ERY	16	16	16	8	16	32	256	256

F1 and **F2** against *E. coli* 236 and *E. coli* ATCC25922 was either maintained or slightly greater.

In conclusion, an efficient method has been developed for the synthesis of 11,12-cyclic carbonate azithromycin-4"-O-carbamoyl glycosyl derivatives. The results of preliminary antibacterial tests against eight strains in vitro demonstrated that these derivatives exhibited improved activities with a broad spectrum compared with the parent compound, which could be attributed to the glycosylated side chains. The monosaccharide side chain may help the target compounds fit into the binding pocket and interact with thebacterial ribosome. However, compounds **F7** and **F8** with disaccharide side chains expressed the lowest activity, indicating that the bulky disaccharide side chain might hinder the interactions of the derivative with the bacterial ribosome. This study has led to the discovery of a series of compounds for further optimization.

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Supplementary data

Supplementary dataassociated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl.2013. 07.042.

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