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First approach to the cycloisodityrosine unit of RA-IV

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Abstract—The syntheses of the methyl (2S,3S)-2-amino-3-(3-hydroxy-4-methoxyphenyl)-3-t-butyldimethylsilyloxypropanoate and a precursor of the cycloisodityrosine unit of RA-IV are described. © 2001 Elsevier Science Ltd. All rights reserved.

Bouvardin 1, a bicyclic hexapeptide isolated from *Bouvardia ternifolia*,¹ represents the initial member of a growing class of potent antitumor antibiotics. To date, 16 congeners (RA-I–RA-XVI) have been identified and their relative and absolute configurations have been determined.² These bicyclic compounds are characterized by an 18-membered peptide ring and a bridged 14-membered cycloisodityrosine unit that constitutes the pharmacophore.³ This cycloisodityrosine unit is composed by two tyrosines for the majority of the compounds of this family. Three compounds include a β -hydroxytyrosine unit: Bouvardin 1, RA-IV 2 and RA-VI 4. Whereas the syntheses of RA-V 3 and RA-VII 5 are well documented,⁴ only a total synthesis of

Bouvardin and RA-VI was reported using Ullman macrocyclisation to obtain the cycloisodityrosine unit.⁵ To our knowledge, the synthesis of the cycloisodityrosine unit of RA-IV has never been described.

A novel cycloetherification methodology based on intramolecular S_NAr reaction has been recently developed⁶ and used for the synthesis of RA-VII 5.^{4e,f}

In connection with our continued work on α -amino β -hydroxy acids,⁷ we report the synthesis of the *anti* β -hydroxytyrosine component of the 14-membered ring of RA-IV and the first preparation of a precursor of its cycloisodityrosine unit **6**.



 $\begin{array}{ll} 1 \mbox{ Bouvardin } R^1 = R^2 = H; R^3 = OH \\ 2 \mbox{ RA-IV } R^1 = Me; R^2 = H; R^3 = OH \\ 3 \mbox{ RA-V } R^1 = R^2 = R^3 = H \\ 4 \mbox{ RA-VI } R^1 = Me; R^2 = OH; R^3 = H \\ 5 \mbox{ RA-VII } R^1 = Me; R^2 = R^3 = H \end{array}$

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CH₃CN (85%). (c) H₂, 10 bar, 1% (S)-Binap Ru Br₂, CH₂Cl₂, room temperature, 78h (83%, ee=90%). (d) MeZnBr, LDA, DBAD, -78°C then aq. NH₄Cl (50%, de>95%). (e) TBDMSOTf, 2,6-lutidine ; H₂, Pd/C 10%, MeOH (96%). (f) CF₃CO₂H, CH₂Cl₂ ; H₂, Raney Ni, ultrasounds, MeOH (50%).

6 could be obtained by a peptide coupling reaction between the protected β-hydroxytyrosine **7** and the *N*-Boc protected (*S*)-4-fluoro-3-nitrophenylalanine **8**⁸ followed by an S_NAr macrocyclisation. The key steps to obtain **7** are the asymmetric hydrogenation of a βketoester⁹ and the diastereoselective electrophilic amination of the resulting β-hydroxyester.⁷

OMe

The synthesis started from the commercially available 3-hydroxy-4-methoxybenzoic acid **9** which was homologated to the β -ketoester **10**¹⁰ after protection of the phenolic functionality as a benzyl ether. **10** was hydrogenated enantioselectively at room temperature and low pressure in the presence of 1% of (*S*)-Binap RuBr₂. The (*R*)- β -hydroxyester **11** was obtained in 83% yield and 90% ee.¹¹ The zinc enolate of **11** was then aminated with *t*-butylazodicarboxylate: the electrophilic amination was highly diastereoselective, providing the *anti* α -hydrazino- β -hydroxyester **12** as the only detectable diastereomer.¹² To achieve the synthesis of **7**, the alcohol was protected as a *t*-butyldimethylsilylether and the benzyl ether hydrogenolyzed quantitatively. After deprotection of the *t*-butyl carbamates, the N–N bond of the hydrazine was cleaved with H_2 in the presence of Raney Ni under ultrasounds. (2*S*,3*S*)-7 was purified by silica gel flash chromatography and isolated in 50% yield and 90% enantiomeric excess.¹³

The peptide coupling reaction of 7 with optically pure (S)-8 was performed under classical conditions. The major diastereomer of the dipeptide 13 was obtained in 80% yield after silica gel flash chromatography.

A first attempt at the intramolecular S_NAr reaction was run in the presence of K_2CO_3 and 3 Å molecular sieves in DMSO at room temperature for 1 h. The cycloetherification occured under these conditions with a retro-Aldol reaction giving 14. During the S_NAr reaction, fluoride ions are liberated leading to *t*-butyldimethylsilyl ether deprotection and subsequent retro-Aldol cleavage.

A similar observation was reported by Boger et al. for the synthesis of the 16-membered D–E ring system of vancomycin. A successful macrocyclisation was



(a) EDCI, HOBt, CH_2Cl_2 , room temperature, 1h (80%). (b) K_2CO_3 (4 equiv.), 3Å molecular sieves, DMSO (0.01M), room temperature, 1h (45%).

described using a mixture of K_2CO_3 -CaCO₃: CaCO₃ being as an efficient scavenger of the liberated fluoride anion.¹⁴

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(a) K_2CO_3 -CaCO₃ (4 equiv./5 equiv.), 3Å molecular sieves, DMSO (0.01M), room temperature, 1h (10%).

These conditions were applied to the synthesis of the 14-membered ring of RA-IV. Treatment of a 0.01 M solution of the dipeptide 13 with K_2CO_3 -CaCO₃ (4 equiv./5 equiv.) in the presence of 3 Å molecular sieves in DMSO for 1 h at room temperature afforded the desired macrocyle 6.¹⁵ After preparative thin layer chromatography, (8*S*,9*S*,12*S*)-6 was isolated in 10% yield as an inseparable mixture of atropoisomers.

In conclusion, an efficient synthesis of the dipeptide unit 13 has been developed. In spite of the low macrocyclisation yield, this synthesis represents the first approach via an intramolecular S_NAr to a 14-membered ring system including a β -hydroxytyrosine 6, which is the direct precursor of the cycloisodityrosine unit of RA-IV.

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- 11. The asymmetric hydrogenation of β -ketoesters in the presence of the Ru(II) catalyst bearing atropoisomeric ligands proceeds in a predictable sense, as shown below:



11); 0.50 (m, 51); 5.15 (s, 21); 5.04 (dd, J = 6.3, 4.5 Hz, 1H); 3.88 (s, 3H); 3.71 (s, 3H); 2.73 (dd, J = 16.3; 8.5 Hz, 1H); 2.63 (dd, J = 16.3; 4.5 Hz, 1H). e.e. = 90% measured by ¹H NMR in the presence of 0.1 equiv. of (+)-[Eu(Tcf)₃]. $[\alpha]_{\rm D}^{20}\!=\!+24$ (c 1.1, CHCl₃). Anal. calcd for C₁₈H₂₀O₅: C, 68.34; H, 6.37. Found: C, 68.37; H, 6.44.

12. The *anti* diastereoselectivity of the electrophilic amination was explained as earlier described via the chelated zinc enolate:



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(2*S*,3*S*)–**12**: ¹H NMR (200 MHz, CDCl₃), δ (ppm): 7.32 (m, 5H); 6.95 (m, 3H); 5.20 (m, 1H); 5.13 (s, 2H); 4.80

(m, 1H); 3.87 (s, 3H); 3.70 (sl, 3H); 1.44 (s, 9H); 1.41 (s, 9H). e.e. = 90%. $[\alpha]_{D}^{20} = -44$ (*c* 1, EtOH). Anal. calcd for $C_{28}H_{38}N_2O_9$: C, 61.52; H, 7.00; N, 5.12. Found: C, 61.50; H, 6.99; N, 5.06.

- 13. (2*S*,3*S*)-7: ¹H NMR (200 MHz, CDCl₃), *δ* (ppm): 6.8 (m, 3H); 4.71 (d, *J*=6.6 Hz, 1H); 3.88 (s, 3H); 3.71 (s, 3H); 3.62 (d, *J*=6.6 Hz, 1H); 0.86 (s, 9H); 0.02 (s, 3H); -0.16 (s, 3H). e.e. =90%. [α]_D²⁰ = +59 (*c* 0.6, CHCl₃). Anal. calcd for C₁₇H₂₉NO₅Si: C, 57.44; H, 8.22; N, 3.94. Found: C, 57.42; H, 8.19; N, 4.00.
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- 15. **6**: ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.82 (s, 1H); 7.5 (m, 1H); 7.45 (m, 1H); 6.76 (d, J=8.2 Hz, 1H); 6.62 (d, J=8.2 Hz, 1H); 5.76 (m, 1H); 5.52 (m, 1H); 5.32 (m, 1H); 4.83 (d, J=6.2 Hz, 1H); 4.3 (d, J=6.2 Hz, 1H); 4.11 (m, 1H); 4 (s, 3H); 3.65 (m, 1H); 3.45 (dd, J=13 and 4 Hz, 1H); 3.29 (s, 3H); 1.46 (s, 9H); 0.8 (s, 9H); 0.05 (s, 3H); -0.13 (s, 3H). MS m/z (CI) 646 (M⁺+1).