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SYNTHESES OF1-(5-DEOXY-β-D-arabino-HEXOFURANOSYL)CYTOSINE

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ABSTRACT

1-(5-Deoxy- β -D-arabino-hexofuranosyl)cytosine (4'-homoara-C) (11), a higher homolog of the antileukemic agent ara-C (1- β -D-arabinofuranosylcytosine), was prepared by two independent routes. The first one involved the inversion of configuration at C-2' of the D-*ribo* epimer (1-(5-deoxy- β -D-*ribo*-hexofuranosyl)cytosine, 4'-homocytidine) by the diphenylcarbonate technique; the 5deoxy-D-*ribo*-hexofuranosyl moiety of 4'-homocytidine was obtained by way of an anti-Markovnikov addition of iodine trifluoroacetate to the double bond of 5,6-dideoxy-1,2-O-isopropylidene-3-O-p-tolylsulfonyl- α -D-*ribo*-hex-5-enofuranose and reduction of the resulting iodide(s). In the second approach, 5-deoxy-1,2-O-isopropylidene-3-O-p-tolylsulfonyl- β -D-xylo-hexofuranose was acetolyzed and condensed with 4-acetyl-N-bis(trimethylsilyl)cytosine, and alkaline treatment gave 11 by way of a 2',3'-anhydro intermediate. The structure of 11, in particular the configuration at C-2', was confirmed by its ¹H- and ¹³C-n.m.r. spectra.

INTRODUCTION

1-β-D-Arabinofuranosylcytosine ("cytosine arabinoside", ara-C) is one of the most active antitumor agents in the treatment of acute leukemia¹. After several years of controversy about its mode of action, it seems now generally accepted that ara-C, as its triphosphate (ara-CTP), inhibits the enzymic incorporation of 2'-deoxycytidine triphosphate (dCTP) into DNA by DNA polymerase^{2.3}. An important feature of this analog of 2'-deoxycytidine is its high sensitivity to the pyrimidine nucleoside deaminase, which results into its rapid deamination to 1-β-D-arabinofuranosyluracil (ara-U) devoid of antitumor activity⁴. Furthermore, resistance to "cytosine arabinoside" rapidly develops by deletion of 2'-deoxycytidine kinase, the enzyme responsible for conversion of ara-C into its active phosphorylated form⁵. Numerous modifications of the molecule have been devised to overcome these two limitations. The most important analogs of ara-C are cyclocytidine⁶

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(2.2'-anhydro-1- β -D-arabinofuranosylcytosine) and 2.2'-anhydro-1- β -D-arabinofuranosyl-5-fluorocytosine⁷, both of which exhibit an increased resistance to deamination^{2.8}, and have been studied in clinical trials². Further modifications of ara-C include mostly formation of N or O derivatives, as carboxylic esters, amides^{9–12}, or phosphoric esters^{13,14}, but no alteration of the carbon chain at C-5' has been reported. Owing to the noteworthy biological properties of 9-(5-deoxy- β -D-*ribo*-hexofuranosyl)adenine¹⁵ and other "homonucleosides"^{10–18}, the preparation of 1-(5-deoxy- β -D-*arabino*-hexofuranosyl)cytosine (4'-homoara-C) (11) is of interest, and we describe herein two different approaches to this new analog of ara-C. The first one involves inversion of configuration at C-2' of 4'-homocytidine¹⁹, the 5-deoxy-D-*ribo*-hexose component of which has been prepared by a method developed in this laboratory²⁰; in the second approach, a 1-(5-deoxy-D-*xylo*-hexofuranosyl)cytosine derivative was prepared and used as precursor of 11.

RESULTS AND DISCUSSION

The addition of pseudohalogens to unsaturated carbohydrates has been shown in this laboratory to provide a convenient route to dcoxy sugars²¹; in particular, 5-deoxy-D-xylo- and -L-arabino-hexoses have been prepared by the anti-Markovnikov addition of iodine trifluoroacetate (F_3CCO_2I) to 5.6-unsaturated, 1,4-furanoid precursors and reduction of the resulting iodides, and have been converted into the corresponding adenine nucleosides²⁰. In the present work, the same procedure was used to prepare the desired 5-deoxy-D-ribo-hexose derivatives from the tritosyl derivative 2 of 1,2-O-isopropylidene- α -D-allofuranose²² (1): reductive elimination at C-5–C-6 of 2 with sodium iodide in butanone at reflux to give 3, followed by addition of in situ-generated iodine trifluoroacetate to the double bond of 3, and immediate hydrogenolysis of the resulting iodide(s) in the presence of a Raney nickel catalyst to afford 5-deoxy-1,2-O-isopropylidene-3-O-p-tolylsulfonyl- α -D-*ribo*-hexofuranose (4) in 36.5% overall yield from 1. The structure of 4 was confirmed by the presence, in its ¹H-n.m.r. spectrum, of a 2-proton multiplet centered at δ 1.85 characteristic of the C-5 methylene group. Photochemically induced desulfonylation^{20,23} of 4 proved to be very efficient and gave almost quantitatively 5-deoxy-1,2-O-isopropylidene- α -D-*ribo*-hexofuranose (5). The dibenzoate **6**, a compound that had been prepared by an independent route by David and de Sen- $\pi y e y^{19}$ was obtained by benzovlation of 5. Conversion of 6 into 1-(5-deoxy- β -Dribo-hexopyranosyl)cytosine (4'-homocytidine, 9) was achieved, in 43% overall yield, by acetolysis of 6, condensation of the resulting diacetate 7 with 4-N-acetylbis(trimethylsilyl)cytosine in the presence of tin(IV) chloride, and deacylation of the nucleoside 8 under conditions very similar to those described earlier¹⁹.

Several methods were available for the inversion of configuration at C-2' of cytidine by way of a 2,2'-anhydro intermediate (cyclocytidine), the active form of cytidine being generated with such reagents as thionyl chloride²⁴, a Vilsmeier-Haack salt²⁵, acetic anhydride-boron trifluoride etherate²⁶, or diphenylcarbo-



nate^{27–29}. The diphenylcarbonate procedure is very convenient as three reactions (formation of the 2'.3'-O-carbonyl derivative, cyclization to the 2.2'-anhydro intermediate with loss of carbon dioxide, and hydrolysis to the β -D-arabino nucleoside) take place in one operation. Thus, treatment of 9 with diphenylcarbonate in N, N-dimethylformamide in the presence of sodium hydrogencarbonate afforded, after 25 min at 144° and chromatographic separation, a new compound (49% yield) that exhibited spectroscopic properties in agreement with the expected structure 11 (see later). No 2,2'-anhydro intermediate (10) was observed at the end of the reaction; as expected, this intermediate had been entirely hydrolyzed to 11 by traces of water present in the reaction mixture (see ref. 28).

As an extension of the method developed by Reist *et al.*³⁰, and by Shimizu and Shimizu³¹ for the synthesis of ara-U and of ara-C, respectively, our second approach to 11 involved the preparation of a suitable nucleoside precursor having a 5-deoxy-3-*O*-*p*-tolylsultonyl- β -D-xylo-hexofuranosyl residue. Compound 13, previously reported by this laboratory²⁰, constitutes the ideal starting material for this purpose. Acetolysis of 13 under standard conditions and condensation of the resulting triacetates 14 with 4-*N*-acetyl-bis(trimethylsilyl)cytosine in the presence of tin(IV) chloride afforded the desired cytosine derivative 15 in 36Cé overall yield. In the key step, compound 15 was treated with sodium hydroxide in methanol to give 11 in excellent yield (97%), most probably by way of 4-*N*-acetyl-1-(2.3-anhydro-5deoxy- β -D-*ribo*-hexofuranosyl)cytosine and the 2.2'-anhydro intermediate 10 (or its 4-*N*-acetyl derivative).

The physico-chemical properties of the samples of 11 obtained by the two methods are identical and confirm its structure. Thus, the chemical shift of H-1' (δ 6.15) in its ¹H-n.m.r. spectrum, and the corresponding $J_{1'2'}$ value (5 Hz) indicate clearly that 11 belongs to the β -D-arabino series (corresponding values³² for ara-C: δ (H-1') 6.18, $J_{1'2'}$ 4.8 Hz; for cytidine: δ (H-1') 5.90, $J_{1'2'}$ 3.6 Hz; same values for 4'-homocytidine¹⁹). The ¹³C-n.m.r. spectra of 9, 11, and the parent nucleosides cytidine and ara-C have been recorded for solutions in deuterium oxide and the data are listed in Table I (for ¹³C-n.m.r. spectra of cytidine and ara-C under different conditions, see ref. 33). The signals of the carbon nuclei of the heterocyclic base are readily identified by comparison with the data of the reference nucleosides and analogs³⁴. The signal appearing at $\delta \sim 35$ in the spectra of 9 and 11 corresponds to the C-5' methylene atom and constitutes a further proof of the "homo" (5deoxy-D-hexofuranosyl) constitution of 9 and 11. Inversion of configuration at C-2' in both types of nucleosides is accompanied by characteristic changes of the chemical shifts of C-1' and C-2': the signal of C-1' is shifted upfield (\sim -5 p.p.m.), whereas the signal of C-2' undergoes a slight downfield shift ($\sim +2$ p.p.m.) upon the ribo to arabino configurational change. These effects are very similar to those observed³⁵ for the corresponding methyl β -D-(or -L)-pentofuranosides. Finally, "homologation" is found to alter substantially the chemical shift of C-3' ($\sim +4$ p.p.m.), an effect that is difficult to rationalize on the basis of the available data, whereas the slight upfield shift of C-4' is entirely consistent with the suppression of the electronegative substituent at C-5'.

TABLE I

¹³C-N M R CHEMICAL-SHIFT DATA²

Compound	C-1'	C-2'	C-3'	C-4'	C-5'	C-6'	C-5	C-6
Cytidine	90.52	74.15	69.42	83.92	60.88		96.22	141.71
Ата-С	86.19	75.64	75.64	83.40	60.95		95.38	142.81
9	91.17	73.89 ⁶	73.24 ^b	80.62	35.19	58.55	96.41	141.78
11	86.25	75.70	79.52	80.49	35.06	58.62	95.25	142.94

"Solutions in deuterium oxide containing methanol as internal standard; chemical shifts (δ) converted to the tetramethylsilane scale (δ CH₃OH 49.039). ^bAssignments may be reversed.

EXPERIMENTAL

General methods. — Melting points were determined with a Fisher-Johns melting-point apparatus and are uncorrected. Optical rotations were measured with a Perkin-Elmer model 141 automatic polarimeter. I.r. spectra were recorded with a Perkin–Elmer 180 or a Beckman Acculab 6 spectrophotometer. U.v. spectra were obtained on a Unicam SP 800 spectrophotometer. The ¹H-n.m.r. spectra were recorded at 60 MHz with a Varian EM-360 or a Bruker HX-60 spectrometer for solutions in $({}^{2}H)$ chloroform with tetramethylsilane as the internal standard, or in deuterium oxide with sodium 4,4-dimethyl-4-silapentane-1-sulfonate as the internal standard. The ¹³C-n.m.r. spectra were obtained at 15.09 MHz on a Bruker HX-60 spectrometer equipped with an FT60M Fourier-transform accessory; chemical shifts are given in δ downfield from the signals of the standards. T.l.c. was performed with Silica gel G as the adsorbent in the following solvent systems (v/v): (A) 1:2, (B) 1:1, (C) 4:1, and (D) 9:1 benzene-ethyl acetate; (E) ethyl acetate; (F) 3:1 hexane-ethyl acetate; (G) 20:1 ethyl acetate-ethanol; (II) ethanol; and (I) 2:1 ethanol-chloroform. The developed plates were air dried and compounds located by irradiation with a short-wavelength, u.v. lamp or by spraying with a solution of 1% cerium sulfate and 1.5% molybdic acid in 10% aqueous sulfuric acid and heating the plates at $\sim 150^{\circ}$ or by both methods. Preparative t.l.c. was performed on 20 \times 20 cm plates coated with a 1-mm layer of Silica gel G. Column chromatography was performed on Silica gel 60 (70-230 mesh, Merck). U.v. irradiations were obtained with a 450-W Hanovia high-pressure, mercury-arc lamp (cat. No. 679A-36) contained in a water-cooled, quartz immersion-well; a Vycor 7010 filter-sleeve was employed.

1,2-O-Isopropylidene-3,5,6-tri-O-p-tolylsulfonyl- α -D-allofuranose (2). — A solution of 1,2:5,6-di-O-isopropylidene- α -D-allofuranose²² (9 g) in 70% aqueous acetic acid (90 mL) was kept for 16 h at room temperature. The solvent was then removed under reduced pressure and the remaining crystalline mass was recrystallized (ethanol-hexane) to afford an almost quantitative yield of 1,2-O-isopropylidene- α -D-allofuranose (1), m.p. 128–130° (lit.³⁶ m.p. 129–130°).

To a solution of 1 (6.54 g, 30 mmol) in pyridine (30 mL) was added, at 0°, p-

toluenesulfonyl chloride (23 g, 120 mmol). The mixture was kept at room temperature for 4 days and then poured into ice-water. The precipitated solid mass was removed by filtration and recrystallized from dichloromethane-methanol to give pure 2 (17.4 g, 85.8%) as white needles, m.p. 158–159.5°, $[\alpha]_D^{-3} + 51.7°$ (c 1.3, chloroform); R_F 0.13 (solvent F); $\nu_{\text{MAF}}^{\text{MAF}}$ 1600, 1500, 1380, and 1180 cm⁻¹; ¹Hn.m.r.: δ 1.23 and 1.40 (2 s, 2 × 3 H, CMe₂), 2.45 (s, 9 H, 3 ArMe), 3.63–5.0 (m's, 6 H, H-2, -3, -4, -5, -6a, and -6b), 5.40 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 7.30 and 7.73 (d and m, 12 H, 3 Ar).

Anal. Calc. for $C_{30}H_{34}O_{12}S_3$; C, 52.77; H, 5.02; S, 14.09. Found: C, 52.55; H, 5.02; S, 14.07.

5,6-Dideoxy-1,2-O-tsopropylidene-3-O-p-tolylsulfonyl- α -D-ribo-hex-5-enofuranose (3). — Sodium iodide (37 g, 250 mmol) was added to a solution of **2** (20.4 g, 30 mmol) in butanone (380 mL). The mixture was boiled at reflux for 4 h and then filtered. Fresh sodium iodide (29 g, 190 mmol) was added to the filtrate, and the mixture was boiled at reflux for 14 h. After this time, the salts were removed by filtration, and the filtrate was evaporated. The residue was dissolved in chloroform (350 mL), and the solution washed with aqueous sodium thiosulfate and then dried (MgSO₄). Pure **3** (8.2 g, 83%) was obtained after evaporation of the solvent and recrystallization of the remaining solid from methanol; m.p. 138–139° (dec.), $[\alpha]_{12}^{11}$ +58.6° (c 1.8, chloroform); R_{μ} 0.40 (solvent F); ν_{max}^{HBT} 1640, 1600, 1490, 1375, and 1180 cm⁻¹; ¹H-n.m.r.; δ 1.30 and 1 57 (2 s, 2 × 3 H, CMe₂), 2.47 (s, 3 H, ArMe), 4.07–4.63 (m's, 3 H, H-2, -3, and -4), 4.97–5.63 (m's, 3 H, H-5, -6a, and -6b), 5.73 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 7.33 and 7.83 (2 d, 2 × 2 H, Ar).

Anal. Calc. for $C_{16}H_{20}O_6S$: C, 56.46; H, 5.92; S, 9.42. Found: C, 56.63; H, 5.85; S, 9.48.

5-Deoxy-1.2-O-isopropylidene-3-O-p-tolylsulfonyl- α -D-ribo-hexofuranose (4). -- To a solution of iodine (6.4 g, 25 mmol) in dry 1,2-dimethoxyethane (120 mL) was added a mixture of 3 (6 g, 17.6 mmol) and silver trifluoroacetate (5.5 g. 25 mmol) in dry 1,2-dimethoxyethane (150 mL). After having been vigorously stirred for 15 min in the dark at room temperature, the mixture was filtered through Celite and the filtrate concentrated to dryness. The residue was dissolved in chloroform, the solution washed with aqueous sodium thiosulfate and dried, and the solvent evaporated. The residue was hydrogenated in the presence of W-4 Raney nickel (0.3–0.35 MPa of hydrogen) in methanol (350 mL) containing triethylamine (3 mL) for 3 h. The catalyst was removed by filtration and the solvent evaporated. The residue was dissolved in chloroform (200 mL), the solution washed with 3%aqueous hydrochloric acid, and then with water, and dried. The solution was evaporated and the major component of the remaining syrup was separated by column chromatography (solvent C) which afforded 3.2 g (51%) of crystalline 4, m.p. 110–112° (from ethanol-hexane), $[\alpha]_D^{24}$ +88.7° (c 1.2, chloroform); R_I 0.17 (solvent C); ν_{max}^{KBr} 3480, 1600, 1360, and 1190 cm⁻¹; ¹H-n.m.r. (after D₂O exchange): δ 1.28 and 1.53 (2 s, 2 × 3 H, CMe₂), 1.60–2.10 (m, 2 H, H-5a, -5b), 2.47 (s, 3 H, ArMe), 3.68 (t, 2 H, H₂-6), 4.13–4.63 (m/s, 3 H, H-2, -3, and -4), 5.75 (d, 1 H, J_{1,2} 3.5 Hz, H-1), 7.38 and 7.89 (2 d, 2 × 2 H, Ar).

Anal. Calc. for C₁₆H₂₂O₇S: C, 53.62; H, 6.19; S, 8.95. Found: C, 53.58; H, 6.13; S, 8.91.

5-Deoxy-1,2-O-isopropylidene- α -D-ribo-hexofuranose (5).— Compound 4 (3.23 g, 9 mmol) in anhydrous methanol (250 mL) containing sodium methoxide (from 230 mg of sodium) was submitted to u.v. irradiation for 10 h at 5–20°. The solution was decolorized with charcoal, the mixture filtered, and the filtrate concentrated. The residue was extracted with hot ethyl acetate (2 × 100 mL), the combined extracts evaporated, and the residual syrup purified by column chromatography (solvent *B*) to give 1.68 g (91%) of pure 5, m.p. 76.5–77.5° (from ether-petroleum ether), $[\alpha]_{D}^{24}$ +50.3° (c 1.5, chloroform); $R_{\rm F}$ 0.20 (solvent *A*); $\nu_{\rm max}^{\rm KBr}$ 3440 and 3290 cm⁻¹.

Anal. Calc. for C₉H₁₆O₅: C, 52.93; H, 7.90. Found: C, 52.89; H, 7.90.

I-(5-Deoxy-β-D-ribo-hexofuranosyl)cytosine (homocytidine, 9) from 5. — Compound 5 was treated with benzoyl chloride in pyridine to afford the dibenzoate 6 in 89% yield after usual processing and column chromatography (19:1, v/v, benzene–ethyl acetate); m.p. 89° (from ethanol–hexane) (lit.¹⁹ m.p. 87°). Compound 6 was converted into 9 in three steps under conditions similar to those described by David and de Sennyey¹⁹: acetolysis of 6 to afford 7 (83%), condensation of 7 with 4-*N*-acetylbis(trimethylsilyl)cytosine in the presence of tin(IV) chloride to give 8 [62%; m.p. 188.5–189.5° (from ethanol) (lit.¹⁹ m.p. 183°)], and deacylation of 8 to give 9 (84% yield) m.p. 114–116°(from ethanol–water) (lit.¹⁹ m.p. 116–117°).

1,2,6-Tri-O-acetyl-5-deoxy-3-O-p-tolylsulfonyl-D-xylo-hexofuranoses (14). — To a mixture of 5-deoxy-1,2-O-isopropylidene-3-O-p-tolylsulfonyl- α -D-xylo-hexofuranose²⁰ (13), acetic anhydride (23 mL), and glacial acetic acid (23 mL) was added slowly, at 0°, conc. sulfuric acid (1.4 mL). The mixture was stirred for 3 days at room temperature and then cooled in an ice bath. Sodium acetate (4 g) was added in portions and the mixture stirred for 1 h. The solvent was removed *in vacuo* and the residue dissolved in water (100 mL) and dichloromethane (200 mL). The separated organic layer was washed successively with saturated aqueous sodium hydrogencarbonatc and with water, and dried (MgSO₄). The solvent was evaporated, and the remaining, light-brown syrup purified by column chromatography (solvent *C*) to give 1.02 g (78%) of pure, syrupy 14, $[\alpha]_{10}^{24} + 37.5^{\circ}$ (c 2.2, chloroform); $R_{\rm F}$ 0.30 (solvent *C*); $\nu_{\rm max}^{\rm film}$ 1750, 1600, 1370, 1230, 1180, and 1020 cm⁻¹.

Anal. Calc. for C₁₉H₂₄O₁₀S: C, 51.35; H, 5.44; S, 7.21. Found: C, 51.22; H, 5.63; S, 7.09.

 $4-N-Acetyl-1-(2,6-di-O-acetyl-5-deoxy-3-O-p-tolylsulfonyl-\beta-D-xylo-hexofu$ ranosyl)cytosine (15). — To a solution of 14 (1.51 g, 3.4 mmol) in dry 1,2-dichloroethane (100 mL) was added, with stirring, a solution of tin(IV) chloride (1.2mL, 3 equiv.) in 1,2-dichloroethane and, after 10 min, a solution of 4-N-acetylbis(trimethylsilyl)cytosine (1.3 g, 1.2 equiv.) in dichloroethane (20 mL). After 4.5h at room temperature, additional equivalents of tin(IV) chloride (0.4 mL) and 4-N-acetyl-bis(trimethylsilyl)cytosine (0.87 g) in dichloroethane (5 mL) were added to the mixture. After having been stirred overnight at room temperature, the mixture was diluted with dichloromethane (100 mL) and poured into a vigorously stirred, saturated aqueous sodium hydrogenearbonate solution. The precipitated solids were removed by filtration through Celite; the organic layer of the filtrate was separated, washed with water (100 mL), and dried (MgSO₄). The solvent was evaporated and the major component of the remaining syrup was separated by column chromatography (50:1, v/v. ethyl acetate–ethanol) to afford pure 15 as a foam (0.84 g, 46%). $[\alpha]_D^{24} + 50.9^\circ$ (c 1.1, chloroform); $R_F 0.34$; ν_{max}^{him} 3240, 1740, 1665, 1555, 1490, 1375, 1225, 1180, and 1105 cm⁻¹; λ_{max}^{hiOH} 216 (c 11 300), 230 (c 10 300), 249 (c 9750), 298 nm (c 4500); ¹H-n.m.r.: δ 2.08, 2 12, 2.28, 2.40 (4 s and overlapped m, 14 H, H₂-5', 2 AcO, AcN, MeAr), 4.0–4.67 (m, 3 H, H-3', -6'a, and -6'b), 5.05 (s + d, 2 H, H-2', -3'), 5.92 (s, 1 H, H-1'), 7.40 and 7.77 (AB, 2 H, J 8 Hz, H-5, -6), 7.27 and 7.70 (2 d, 4 H, C₆H₄Me).

Anal. Calc. for C₂₃H₂₇N₃O₁₀S: C, 51.39; H, 5.06; N, 7.82; S, 5.96, Found: C, 51.45; H, 5.26; N, 7.69; S, 5.80.

I-(5-Deoxy- β -D-arabino-hexofuranosyl)cytosine (11). — (a) From 9 A mixture of 9 (144 mg, 0.56 mmol), diphenylcarbonate (155.5 mg, 1.3 equiv.), and sodium hydrogencarbonate (7.1 mg, 0.15 equiv.) in dry N,N-dimethylformamide (0.3 mL) was heated for 25 min at 144–145°. After this time, t.l.c. (solvent *I*) indicated that all of the starting material (R_F 0.35) had been converted into a fastermoving compound (R_F 0.48). The mixture was applied to a preparative t.l.c. plate, which was developed successively with 7:3 and 1:5 (v/v) chloroform–ethanol. Collection of the band corresponding to the new product and elution of the product with ethanol afforded, after evaporation of the solvent, homogeneous 11 (71 mg, 49%) as a slightly colored solid, m.p. 185–192° (from ethanol–water).

(b) From 15. To a solution of 15 (0.252 g, 0.47 mmol) in 20' × aqueous methanol (10 mL) was added, with stirring, aqueous M sodium hydroxide (0.8 mL) over a period of 2 h, and the mixture was stirred for an additional 4 h. The methanol was removed *in vacuo* and the remaining solution loaded onto a column of strongly acidic, ion-exchange resin [CGC-240 (H⁺), 20 mL]. The column was washed with water (100 mL) and the product then eluted with 1' ϵ aqueous amonia (150 mL). Evaporation of the eluate to dryness afforded pure 15 (117 mg. 97%), m.p. 218–220° (from ethanol). [α]]⁸/₂ + 164.6° (c 1.1, water): $R_{\rm F}$ 0.48 (solvent *I*); $\nu_{\rm max}^{\rm HeI}$ 3340, 3190, and 1640 cm⁻¹; $\lambda_{\rm max}^{\rm He2}$ 271 (ϵ 8500), 230 sh (ϵ 5900), $\lambda_{\rm mun}^{\rm He2}$ (10 mL); $I_{\rm H-m.r.r.}$ (D₂O): δ 2.07 (broad q, 2 H, $J \sim 6.5$ Hz, H₂-5'), 3.78 (t, 2 H, $J \sim 6.5$ Hz, H₂-6'), ~4.0 (m, 2 H, H-3', -4'), 4.37 (m, 1 H, H-2'), 6.03 (d, 1 H, J_{5,6} 8 Hz, H-5), 6.15 (d, 1 H, J_{1',2'} 5 Hz, H-1'), and 7.70 (d, 1 H, H-6).

Anal. Calc. for C₁₀H₁₅N₃O₅: C, 46.69; H, 5.88; N, 16.33. Found: C, 46.58; H, 5.93; N, 16.32.

4-N-Acetyl-1-(2,3,6-tri-O-acetyl-5-deoxy- β -D-arabino-hexofuranosyl) cytosine (12). — Compound 15 was treated with acetic anhydride in pyridine under standard conditions to give 12 as white needles, after purification by preparative t.l.c. and recrystallization from ethanol; m.p. 187–188°, $|\alpha|_{D}^{22} + 99.6^{\circ}$ (c 0.97, chloroform); $R_{\rm F}$ 0.16 (solvent *E*); $\nu_{\rm max}^{\rm KBr}$ 3220, 1750, 1730, 1660, 1630, 1610, 1550, 1485, and 1235 cm⁻¹; $\lambda_{\rm max}^{\rm EiOH}$ 249 (ϵ 17 400), 299 nm (ϵ 8160); $\lambda_{\rm min}^{\rm EiOH}$ 227 (ϵ 4430), 275 nm (ϵ 4610); ¹H-n.m.r.: δ 1.98, 2.09, 2.17, 2.31 (4 s and overlapped m, 14 H, H₂-5', 3 AcO, AcN), ~4.2 (m, 3 H, H-4', H₂-6'), 5.0 (m, 1 H, $J_{2',3'}$ ~1, $J_{3',4'}$ ~3 Hz, H-3'), 5.48 (dd, 1 H, $J_{1',2'}$ 4.0 Hz, H-2'), 6.30 (d, 1 H, H-1'), 7.48 (d, 1 H, $J_{5,6}$ 7.5 Hz, H-5), and 7.90 (d, 1 H, H-6).

Anal. Calc. for C₁₈H₂₃N₃O₉: C, 50.82; H, 5.45; N, 9.88. Found: C, 50.76; H, 5.65; N, 9.69.

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